

Table S1. Antibodies used for flow cytometric analysis of PBMCs.

Fluorochrome	CD No. Name	Clone	Company	Order No.
FITC	pan-KIR: CD158a/h, b/j, e1/2, i	11PB6	Miltenyi	130-099-209
FITC	Granzyme K	GM6C3	SantaCruz	sc56125-FTIC
PE	CD56	N901	Beckman Coulter	A07788
PE	CD159c/NKG2C	REA205	Miltenyi	130-103-635
BV785	CD16	3G8	Biolegend	302046
ECD	CD20	B9E9	Beckman Coulter	IM3607U
PerCP/Cy5.5	CD226 DNAM-1	11A8	Biolegend	338314
PC7	CD159a/NKG2A	Z199	Beckman Coulter	B10246
PECy7	Granzyme A	GzA-3G8.5	eBiosciences	25-9177-42
APC	CD3	Hit3a	Biolegend	300312
BV510	CD4	OKT4	Biolegend	317444
BV605	CD45RO	UCHL1	Biolegend	304238
APC	CD336/NKp44	P44-8	Biolegend	325110
eFluor660	Granzyme M	4B2G4	eBiosciences	50-9774-42
APC	CD336/NKp44	P44-8	Biolegend	325110
APC-A700	CD8	B9.11	Beckman Coulter	PN A66332
PE	Granzyme B	REA226	Miltenyi	130-101-351
APC-A700	NKG2D		R&D	FAB139N-100
APC/Cy7	CD244/2B4	C1.7	Biolegend	329518
PacB	CD57	HCD57	Biolegend	322316
VioBlue	Perforin	REA1061	Miltenyi	130-096-671
Biotin	CD337/NKp30	AF29-4D12	Miltenyi	130-092-553
BV650	CD335/NKp46	9,00E+02	Biolegend	331927

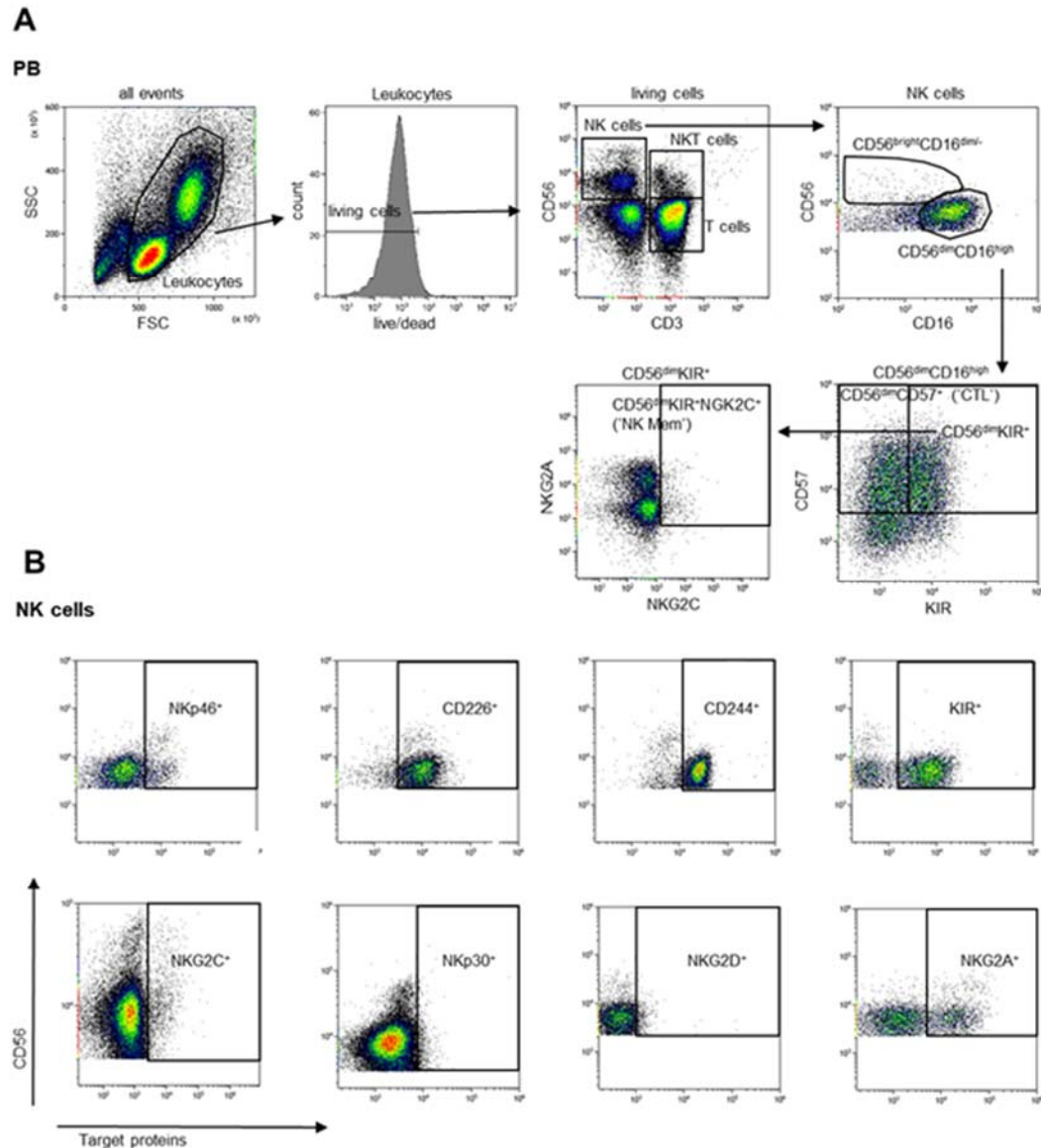


Figure S1. Gating strategy. (A) Single cells from the peripheral blood (PB) were simultaneously analyzed by flow cytometry. Total leukocytes were identified by forward scatter (FSC) and sideward scatter (SSC). Live/dead discrimination was determined using the amine reactive dye Aqua. From these cells, we identified lymphocyte subsets based on CD3 signal and CD56 expression. Next, we selected natural killer (NK) cell subsets based on CD56 and CD16 expression. $CD56^{dim}CD16^{high}$ were further differentiated based CD57 und KIR expression into cytotoxic T cell subsets (CTL) and memory NK cells (memory NK). (B) NK cells, $CD56^{bright}CD16^{dim/-}$, $CD56^{dim}CD16^{high}$ CTL and memory NK cells were differentiated based on their surface marker expression in NKp46, CD226, CD244, KIR, NKG2C, Nkp30, NKG2D and NKG2A positive cells (here exemplarily shown at total NK cells).