

Supplementary Materials

Table S1. Describes the different types and specific characteristics of DyPs isolated from various bacterial strains.

Clas	Name	Source	Substrate	λ_{ma} x	Conditio n	k_{cat}/K_m ($s^{-1}M^{-1}$)	Ref
A	TcDyP	<i>Thermomonospor</i> <i>ra curvata</i>	Reactive Blue5	595	pH 3.0	6.6×10^5	(Chen et al., 2015)
				nm	22 °C		
	TfuDyP	<i>Thermobifida</i> <i>fusca</i>	Reactive Blue19	595	pH 3.0	7.8×10^6	(Rahmanpo ur, Rea, Jamshidi, Fülöp, & Bugg, 2016)
				nm	22 °C		
A	CboDy P	<i>Cellulomonas</i> <i>bogoriensis</i>	Reactive Blue 19	595 nm	pH 4.0	1.3×10^4	Rozeboom, & Fraaije, (Habib, 2019)
A	SviDyP	<i>Saccharomonosp</i> <i>ora viridis</i>	Reactive Blue 19		pH 7.0		(Yu et al., 2014)
B	DyPB	<i>Rhodococcus</i> <i>jostiiRHA1</i>	Reactive Blue4			1.4×10^2	(Roberts et al., 2011)
			pyrogallol			6.0×10^2	
			ABTS			2.4×10^3	
B	PpDyP	<i>Pseudomonas</i> <i>putida</i> MET94	Reactive Blue 5		pH 4.2	2.0×10^5	(Mendes et al., 2015)
B	VcDyP	<i>Vibrio cholerae</i>	Reactive Blue 5			2.6×10^4	(Uchida, Sasaki, Ishimori, 2015)

			Reactive Blue4	pH 3.0 22 °C	6.5×10 ⁵	
			Reactive black 5	pH 3.0	3.6×10 ²	
B	BsDyP	<i>Bacillus subtilis</i>	Malachite green	595 nm	pH 4.0	6.3×10 ² (Dhankhar et al., 2020)
			Methyl violet		pH 4.0	3.6×10 ²
			Reactive Blue19	pH 3.5	3.5×10 ⁵	
B	DyPA	<i>Rhodococcus jostiiRHA1</i>	Reactive Blue4 ABTS	pH 4.0	1.3×10 ⁴ 2.0×10 ³	(Roberts et al., 2011)
B	BIDyP	<i>Brevi bacterium linens</i>				(Sutter et al., 2008)
C	SaDyP 2	<i>Streptomyces avermitilis</i>	Acid Blue 324 ABTS	608 nm	pH 4.0 pH 4.5	1.2×10 ⁴ 1.7×10 ³ (Sugawara et al., 2017)
C	AnaPX	<i>Anabaena sp</i>	Reactive Blue5			1.2×10 ⁷ (Yoshida et al., 2016)
C	DyP2	<i>Amycolatopsis sp. 75iv2</i>	Reactive Blue5 ortho-nitrophenol		7.1×10 ⁵	Barros, & Chang, 2012)
D	AauDy P	<i>Auricularia auricula-judae</i>	meta-nitrophenol	pH 4.5	1.2×10 ⁴	
			para-nitrophenol	pH 4.5	2.1×10 ³	
			Reactive Blue 19	pH 4.0	1.1×10 ⁶	(Salvachúa, Prieto,
D	IlDyP	<i>Irpex lacteus</i>	Reactive Black 5 ABTS	pH 3.0	5.9×10 ⁶	Martínez Á, & Martínez, 2013)
				pH 3.0	8.0×10 ⁶	

D	Dyp	<i>Geotrichum candidum</i> Dec 1	Reactive Blue 5	556 nm	pH 5.5 30 °C	4.8×10^6	(Kim & Shoda, 1999)
D	AauDy P2	<i>Auricularia auricula-judae</i>	Reactive Blue 5		pH 4.5	1.7×10^7	(Liers et al., 2013)
D	Pleos-DyP1	<i>Pleurotus ostreatus</i>	Reactive Blue 19		pH 4.5	1.1×10^5	(Fernández-Fueyo et al., 2015)
D	Pleos-DyP4	<i>Pleurotus ostreatus</i>	Reactive Blue 19		pH 4.5	1.9×10^6	
D	rPsaDy P	<i>Pleurotus sapidus</i>	Guaiacol Reactive Blue 5 ABTS			1.0×10^5 7.5×10^5 3.8×10^6	(Lauber et al., 2017)

- Brown, M. E., Barros, T., & Chang, M. C. (2012). Identification and characterization of a multifunctional dye peroxidase from a lignin-reactive bacterium. *ACS Chem Biol*, 7(12), 2074-2081. doi:10.1021/cb300383y
- Büttner, E., Ullrich, R., Strittmatter, E., Piontek, K., Plattner, D. A., Hofrichter, M., & Liers, C. (2015). Oxidation and nitration of mononitrophenols by a DyP-type peroxidase. *Arch Biochem Biophys*, 574, 86-92. doi:10.1016/j.abb.2015.03.003
- Chen, C., Shrestha, R., Jia, K., Gao, P. F., Geisbrecht, B. V., Bossmann, S. H., . . . Li, P. (2015). Characterization of Dye-decolorizing Peroxidase (DyP) from Thermomonospora curvata Reveals Unique Catalytic Properties of A-type DyPs. *J Biol Chem*, 290(38), 23447-23463. doi:10.1074/jbc.M115.658807
- Dhankhar, P., Dalal, V., Mahto, J. K., Gurjar, B. R., Tomar, S., Sharma, A. K., & Kumar, P. (2020). Characterization of dye-decolorizing peroxidase from *Bacillus subtilis*. *Arch Biochem Biophys*, 693, 108590. doi:10.1016/j.abb.2020.108590
- Fernández-Fueyo, E., Linde, D., Almendral, D., López-Lucendo, M. F., Ruiz-Dueñas, F. J., & Martínez, A. T. (2015). Description of the first fungal dye-decolorizing peroxidase oxidizing manganese(II). *Appl Microbiol Biotechnol*, 99(21), 8927-8942. doi:10.1007/s00253-015-6665-3
- Habib, M. H., Rozeboom, H. J., & Fraaije, M. W. (2019). Characterization of a New DyP-Peroxidase from the Alkaliphilic Cellulomonad, *Cellulomonas bogoriensis*. *Molecules*, 24(7). doi:10.3390/molecules24071208
- Kim, S. J., & Shoda, M. (1999). Purification and characterization of a novel peroxidase from *Geotrichum candidum* dec 1 involved in decolorization of dyes. *Appl Environ Microbiol*, 65(3), 1029-1035. doi:10.1128/aem.65.3.1029-1035.1999
- Lauber, C., Schwarz, T., Nguyen, Q. K., Lorenz, P., Lochnit, G., & Zorn, H. (2017). Identification, heterologous expression and characterization of a dye-decolorizing peroxidase of *Pleurotus sapidus*. *AMB Express*, 7(1), 164. doi:10.1186/s13568-017-0463-5
- Liers, C., Pecyna, M. J., Kellner, H., Worrich, A., Zorn, H., Steffen, K. T., . . . Ullrich, R. (2013). Substrate oxidation by dye-decolorizing peroxidases (DyPs) from wood- and litter-degrading agaricomycetes compared to other fungal and plant heme-peroxidases. *Appl Microbiol Biotechnol*, 97(13), 5839-5849. doi:10.1007/s00253-012-4521-2
- Mendes, S., Brissos, V., Gabriel, A., Catarino, T., Turner, D. L., Todorovic, S., & Martins, L. O. (2015). An integrated view of redox and catalytic properties of B-type PpDyP from *Pseudomonas putida* MET94 and its distal variants. *Arch Biochem Biophys*, 574, 99-107. doi:10.1016/j.abb.2015.03.009

11. Rahmanpour, R., Rea, D., Jamshidi, S., Fülöp, V., & Bugg, T. D. (2016). Structure of *Thermobifida fusca* DyP-type peroxidase and activity towards Kraft lignin and lignin model compounds. *Arch Biochem Biophys*, 594, 54-60. doi:10.1016/j.abb.2016.02.019
12. Roberts, J. N., Singh, R., Grigg, J. C., Murphy, M. E., Bugg, T. D., & Eltis, L. D. (2011). Characterization of dye-decolorizing peroxidases from *Rhodococcus jostii* RHA1. *Biochemistry*, 50(23), 5108-5119. doi:10.1021/bi200427h
13. Salvachúa, D., Prieto, A., Martínez Á, T., & Martínez, M. J. (2013). Characterization of a novel dye-decolorizing peroxidase (DyP)-type enzyme from *Irpef lacteus* and its application in enzymatic hydrolysis of wheat straw. *Appl Environ Microbiol*, 79(14), 4316-4324. doi:10.1128/aem.00699-13
14. Sugawara, K., Nishihashi, Y., Narioka, T., Yoshida, T., Morita, M., & Sugano, Y. (2017). Characterization of a novel DyP-type peroxidase from *Streptomyces avermitilis*. *J Biosci Bioeng*, 123(4), 425-430. doi:10.1016/j.jbiosc.2016.12.001
15. Sutter, M., Boehringer, D., Gutmann, S., Günther, S., Prangishvili, D., Loessner, M. J., . . . Ban, N. (2008). Structural basis of enzyme encapsulation into a bacterial nanocompartment. *Nat Struct Mol Biol*, 15(9), 939-947. doi:10.1038/nsmb.1473
16. Uchida, T., Sasaki, M., Tanaka, Y., & Ishimori, K. (2015). A Dye-Decolorizing Peroxidase from *Vibrio cholerae*. *Biochemistry*, 54(43), 6610-6621. doi:10.1021/acs.biochem.5b00952
17. Yoshida, T., Ogola, H. J., Amano, Y., Hisabori, T., Ashida, H., Sawa, Y., . . . Sugano, Y. (2016). *Anabaena* sp. DyP-type peroxidase is a tetramer consisting of two asymmetric dimers. *Proteins*, 84(1), 31-42. doi:10.1002/prot.24952
18. Yu, W., Liu, W., Huang, H., Zheng, F., Wang, X., Wu, Y., . . . Jin, Y. (2014). Application of a novel alkali-tolerant thermostable DyP-type peroxidase from *Saccharomonospora viridis* DSM 43017 in biobleaching of eucalyptus kraft pulp. *PLoS One*, 9(10), e110319. doi:10.1371/journal.pone.0110319