

Supplementary Materials: Toward Tumor Fight and Tumor Microenvironment Remodeling: PBA Induces Cell Cycle Arrest and Reduces Tumor Hybrid Cells' Pluripotency in Bladder Cancer

Carolina Rubio, José Avendaño-Ortiz, Raquel Ruiz-Palomares, Viktoriya Karaivanova, Omaira Alberquilla, Rebeca Sánchez-Domínguez, José Carlos Casavilla-Dueñas, Karla Montalbán-Hernández, Iris Lodewijk, Marta Rodríguez-Izquierdo, Ester Munera-Maravilla, Sandra P. Nunes, Cristian Suárez-Cabrera, Miriam Pérez-Crespo, Víctor G. Martínez, Lucía Morales, Mercedes Pérez-Escavy, Miguel Alonso-Sánchez, Roberto Lozano-Rodríguez, Francisco J. Cueto, Luis A. Aguirre, Félix Guerrero-Ramos, Jesús M. Paramio, Eduardo López-Collazo and Marta Dueñas

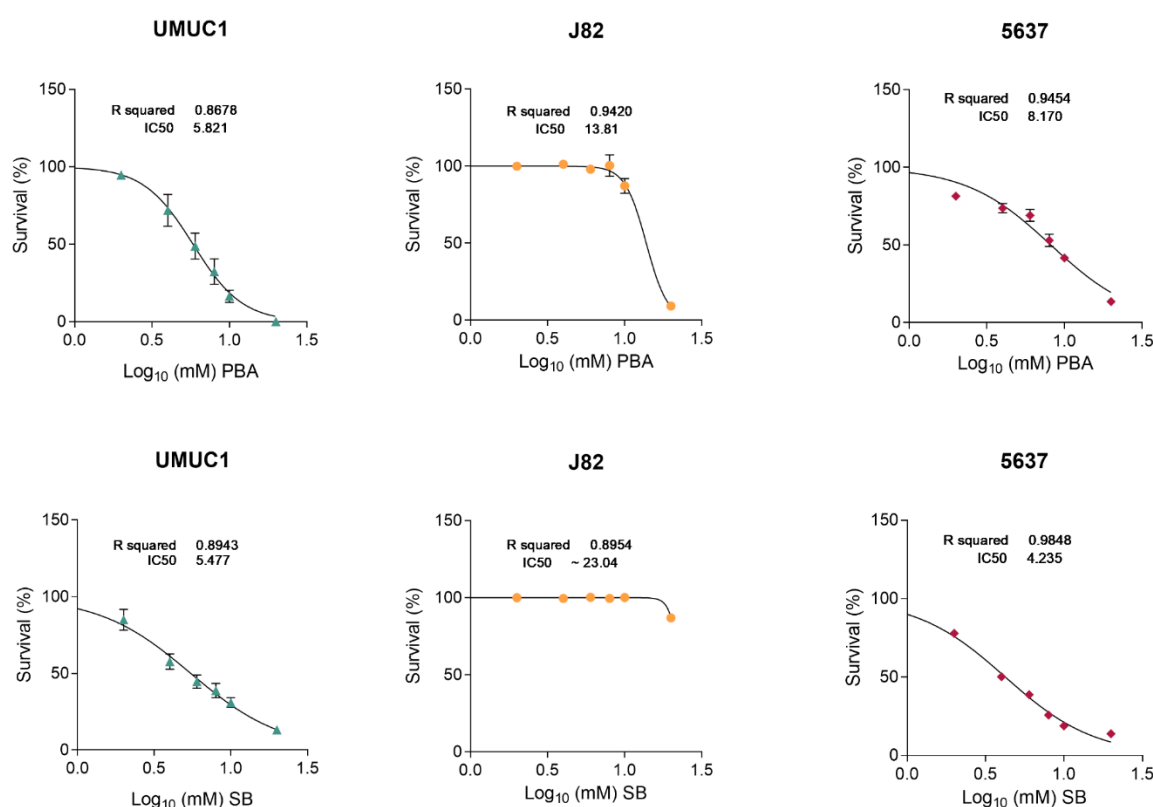


Figure S1. Summary of sensitivity assays of UMUC1, 5637 and J82 BC cells to PBA and SB. Data come from 5 independent experiments for each cell line and are shown as mean \pm SEM for different concentrations of the compounds.

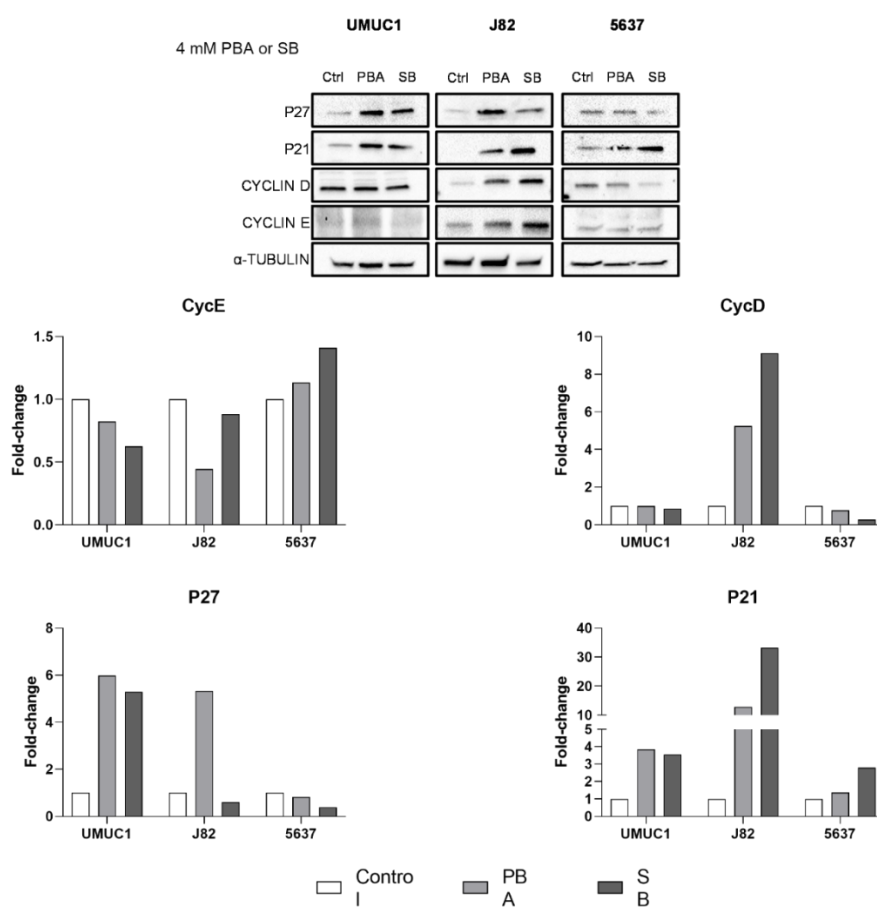


Figure S2. Expression of cell cycle related proteins in UMUC1, J82 and 5637 BC cells without treatment (Ctrl) and after treatment with PBA and SB. Quantification levels of all marks in the same cells is shown, α -TUBULIN was used for loading normalization.



Figure S3. Original Western blots images.

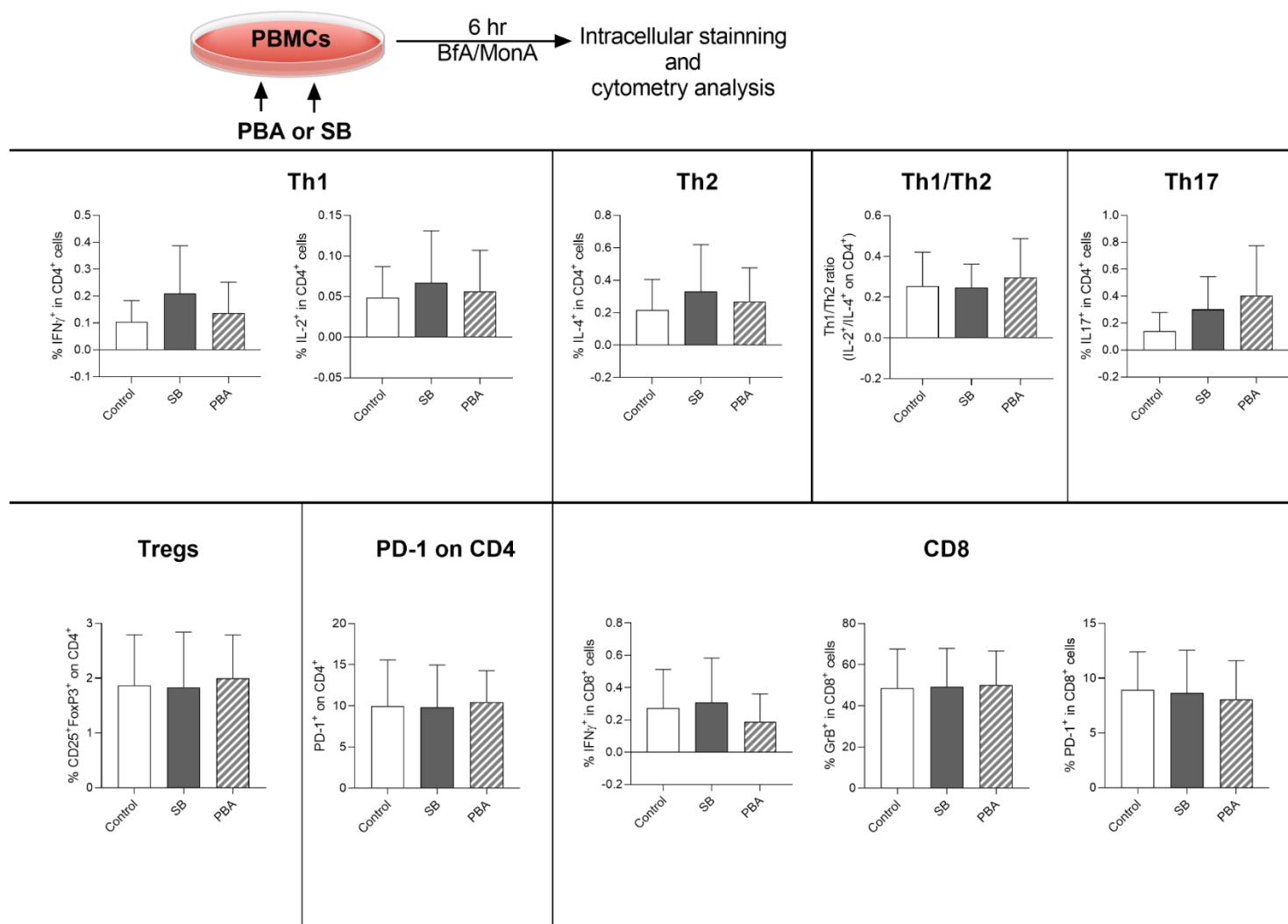


Figure S4. Cytokine expression profiles of PBMC treated with PBA and SB. Brefeldin A and Monensin were used to enhance cytokine detection.

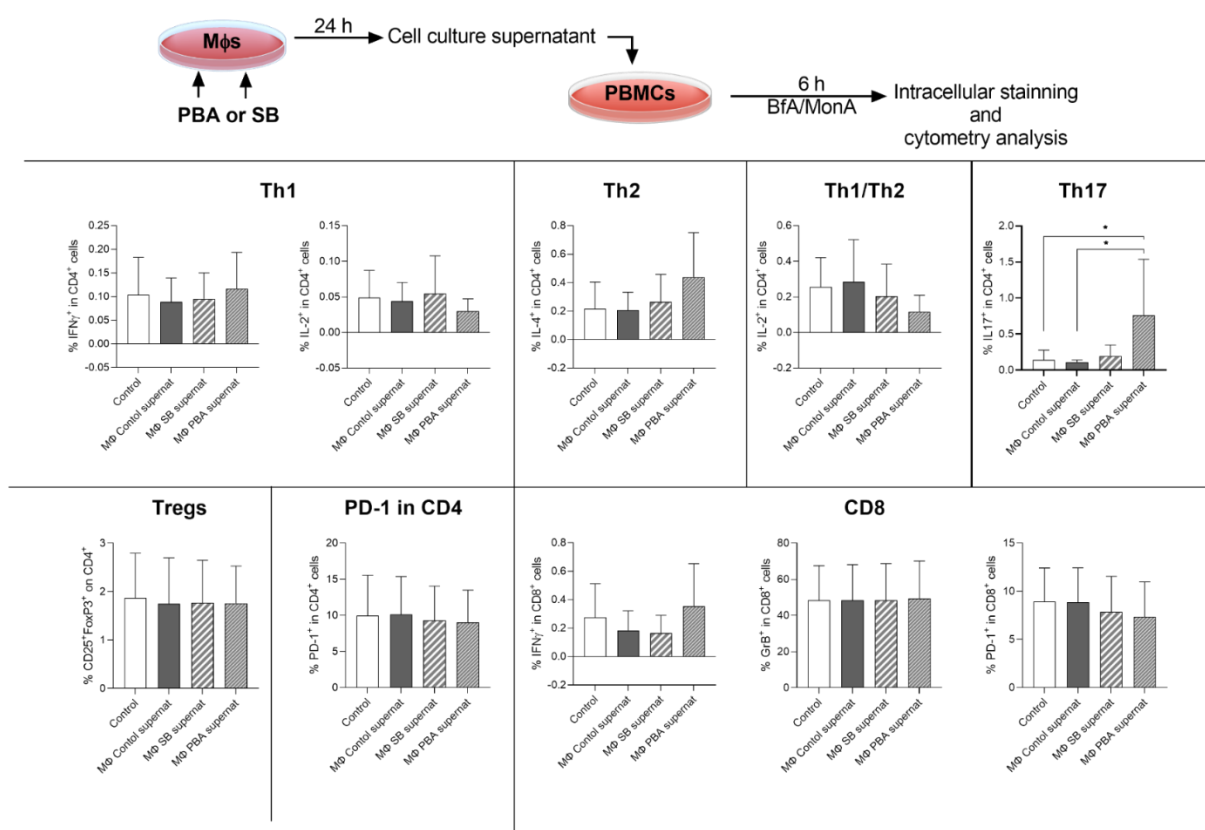


Figure S5. Cytokine expression profiles of PBMC stimulation with conditioned medium from macrophage treated with PBA and SB. Brefeldin A and Monensin were used to enhance cytokine detection.

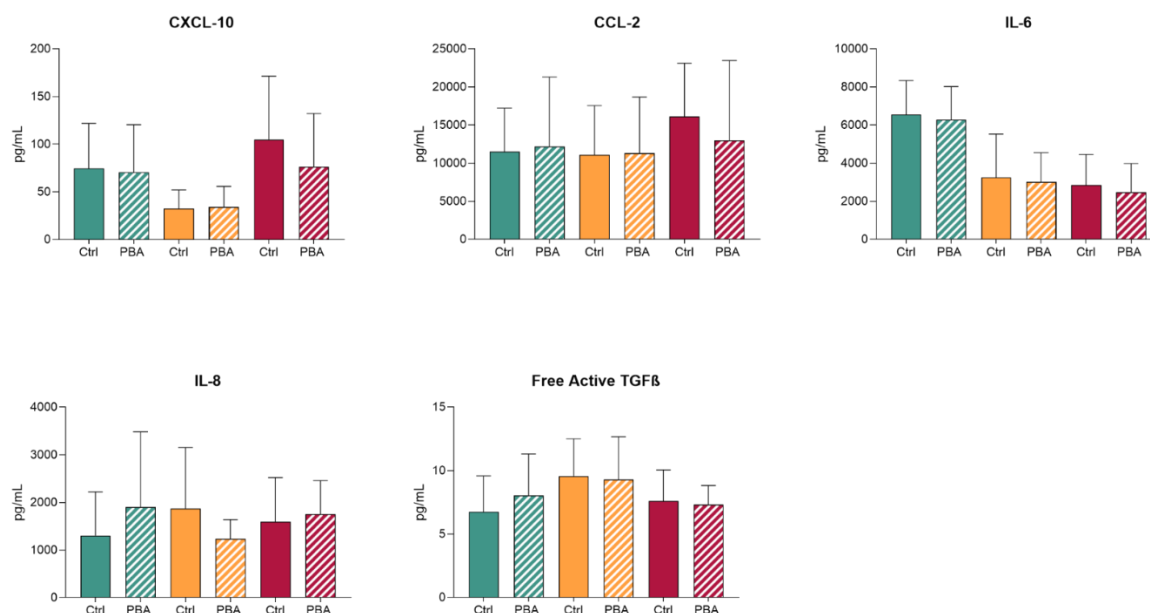


Figure S6. Evaluation of cytokine expression profiles characteristic for M1 or M2 macrophage polarization in co-culture supernatants. CXCL10, CCL2, IL-6, IL-8 and free active TGF β cytokine expression profiles are shown for single-culture and co-culture supernatants, Supernatants of macrophages and BC cell single cultures were included as a control.

Table S1. Antibodies employed in the present study.

Antibodies for Western Blot				
Target	Clone	Source	Dilution	
Acetyl-Histone H3 (Lys9)		Cell Signaling #9649	1:1000	
H4K20me		Abcam9051	1:1000	
H3K4me3		Abcam 8580	1:1000	
Acetyl-H3	-	Millipore 06-599	1:1000	
H3	96C10	Cell Signaling #3638	1:10000	
Acetyl-H4	-	Millipore 06-598	1:100	
H4	62-141-13	Millipore 05-858	1:1000	
p27	-	Abcam ab7961	1:500	
p21	F-5	Santa Cruz sc-6246	1:500	
Cyclin D1	H-295	Santa cruz sc-753	1:200	
Cyclin E	-	Upstate biotechnology 06-134	1:500	
α -Tubulin	FL-335	Sigma T5168	1:1000	
Secondary antibody for Western Blot				
Target	Clone	Conjugated	Source	Dilution
Anti-Rabbit IgG	Donkey	HRP	GE Healthcare NA934	1:5000
Anti-Mouse IgG	Donkey	HRP	JACKSON N° 715-035-151	1:5000
Antibodies for Flow Cytometry				
Target	Clone	Source	Dilution	
CD11b	-	Beckman Coulter IM0530	1 μ l per million cells in 100 μ l	
EpCAM	9C4	BioLegend 324208	5 μ l per million cells in 100 μ l	
List of antibodies for extracellular and intracellular staining for T cell stimulation by Flow Cytometry				
Target	Clone	Source	Fluorochrome	
Extracellular markers				
CD3	UCHT1	Biolegend 300436	BV570	
CD4	SK3	Biolegend 344608	PerCP/Cy5.5	
CD8	SK1	BD biosciences 612889	BUV805	
CD25	2A3	BD biosciences 347643	FITC	
PD-1	EH12.2H7	Biolegend 329929	BV785	
Intracellular markers				
Granzyme B	QA16A02	Biolegend 372214	PE/Cy7	
IFN γ	4S.B3	Biolegend 502540	BV711	
FoxP3	236A/E7	BD biosciences 563955	PE-Dazzle594	
IL-2	MQ1-17H12	BD biosciences 565136	APC-R700	
IL-17	SCPL1362	BD biosciences 560439	Alexa Fluor 647	
IL-4	MP4-25D2	BD biosciences 566274	BV421	
TNF α	MAb11	BD biosciences 563418	BV650	

Table S2. Primer sequences for RT-qPCR.

Primer name	Sequence 5' -> 3'
<i>TBP-F</i>	AGTGAAGAACAGTCCAGACTG
<i>TBP-R</i>	CCAGGAAATAACTCTGGCTCAT
<i>TBP-RT</i>	TGCCTTTGTTGCTCTT
<i>CDK1NA(p21)-F</i>	CCTCATCCCGTGTCTCCTTT
<i>CDK1NA(p21)-R</i>	ACTTGTCGCTGGGTGGTAC
<i>CDK1NA(p21)-RT</i>	GGTGAATTCATAACCG
<i>CDKN1B (p27)-F</i>	CGCTTTGTTTTGTTGCGTTT
<i>CDKN1B (p27)-R</i>	CACTCGCACGTTTGACATCT
<i>CDKN1B (p27)-RT</i>	GTCCCGGGTTAACTCTTCGT
<i>MYC-F</i>	AATGAAAAGGCCCCCAAGGTAGTTATCC
<i>MYC-R</i>	GTCGTTTCCGCAACAAGTCCTCTTC
<i>MYC-RT</i>	GTTAGAAGGAATCG
<i>E2F1-F</i>	TCCAAGAACCACATCCAGTG
<i>E2F1-R</i>	CTGGGTCAACCCCTCAAG
<i>E2F1-RT</i>	GTATAAATTAAATGTTTCCA
<i>BMP4-F</i>	TCCACAGCACTG GTCTTG
<i>BMP4-R</i>	TGGGATGTTCTCCAGATG
<i>BMP4-RT</i>	GGG TG TGCTGAGGTTA
<i>PD-L1-F</i>	CCATACAGCTGAATTGGTCATC
<i>PD-L1-R</i>	CAGAATTACCAAGTGAGTCCTTTCA
<i>PD-L1-RT</i>	GTCTCCTCCAAA
<i>TLR4-F</i>	CCTGCGTGAGACCAGAAAG
<i>TLR4-R</i>	TTCAGCTCCATGCATTGATAA
<i>TLR4-RT</i>	TGTCAATATTAAGGTAGAGA
<i>BMPR-2-F</i>	TCTGGATCTTTCAGCCACAA
<i>BMPR-2-R</i>	TGCCATCTTGTTGACTCAC
<i>BMPR-2-RT</i>	AGTGGAGATGACCC
<i>SOX2-F</i>	CGGAGGAGAAGGAAAGTCG
<i>SOX2-R</i>	CCAGCAAACCTCCAGCAGAC
<i>SOX2-RT</i>	GCT TGGAGACTAGC
<i>NANOG-F</i>	ACCTTCCAATGTGGAGCAAC
<i>NANOG-R</i>	ACTGGATGTTCTGGGTCTGG
<i>NANOG-RT</i>	GTTGGTCAGCACAGGAGAAAATGCC
<i>IL-10-F</i>	GATGCCTTCAGCAGAGTGAA
<i>IL-10-R</i>	GCAACCCAGGTAACCCTTAAA
<i>IL-10-RT</i>	GTCTGGGTCTT
	GTCTGGGTCTT