

Supplementary Materials:

Deregulation of Exo70 Facilitates Innate and Acquired Cisplatin Resistance in Epithelial Ovarian Cancer by Promoting Cisplatin Efflux

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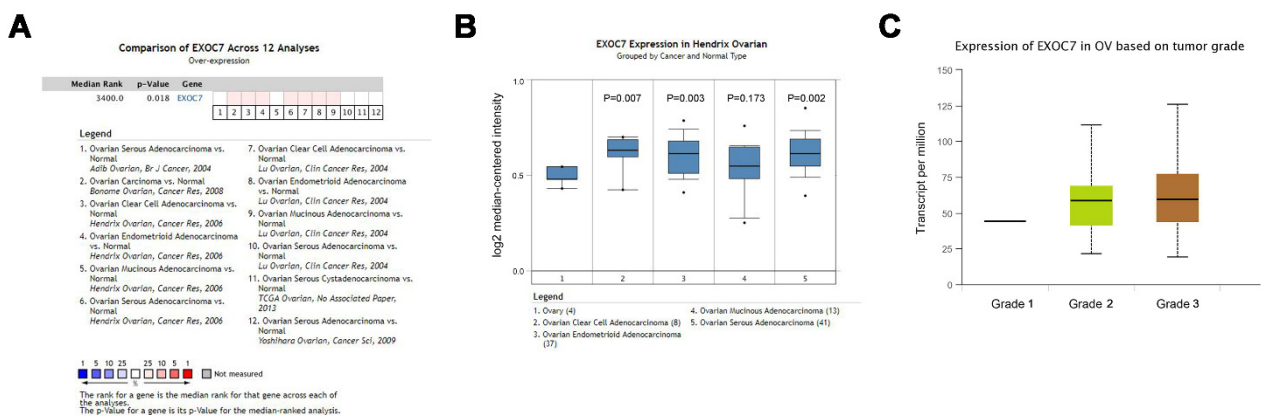


Figure S1. EXOC7 was highly expressed in EOC samples. (A) Comparison of EXOC7 between ovarian cancer and normal tissue across 12 analyses by Oncomine database. (B) EXOC7 was highly expressed in EOC samples compare to normal ovary tissue. Data were generated from the Hendrix Ovarian dataset (Serous vs normal $P = 0.007$; Mucinous vs normal $P = 0.003$; Endometrioid vs. normal $P = 0.173$; Clear cell vs. normal $P = 0.002$). (C) The expression level of EXOC7 increased with the EOC progressions. Data were generated from the UALCAN database.

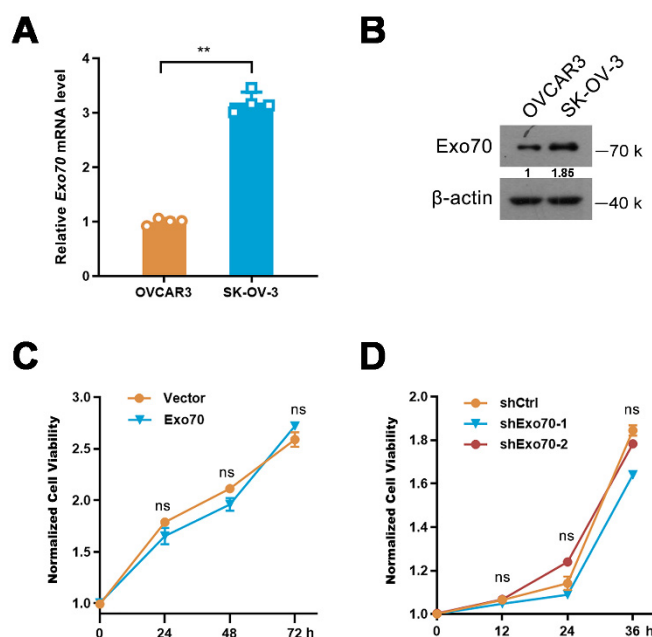


Figure S2. Exo70 expression did not affect the growth ratio of EOC cells. (A) RT-PCR showed the Exo70 mRNA levels in OVCAR3 and SK-OV-3 cells. Data were expressed as mean \pm SEM, $n = 3$. **, $P < 0.01$. (B) Western blotting showed the Exo70 protein expressions in OVCAR3 and SK-OV-3 cells. (C) CCK8 assay measured the effect of overexpressing Exo70 on the proliferation of OVCAR3 cells; Data were expressed as mean \pm SEM, $n = 3$. ns: not significant. (D) CCK8 assay was carried out to measure the effect of knockdown Exo70 on the proliferation of SK-OV-3 cells. Data were expressed as mean \pm SEM, $n = 3$. ns: not significant.

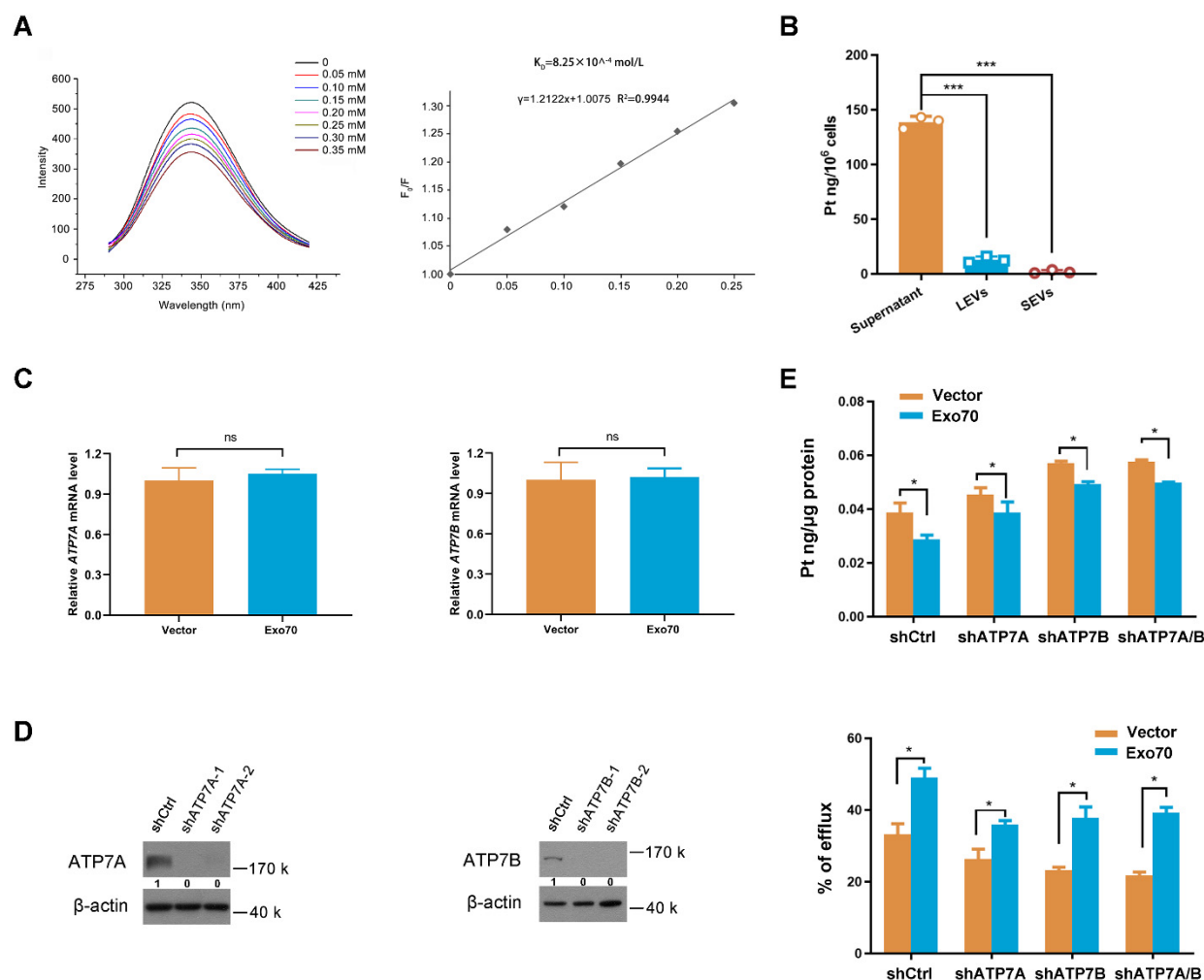


Figure S3. Exo70 efflux cisplatin to supernatant independent of ATP7A and ATP7B. (A) Fluorescence quenching assay was carried out to exam the binding ability of Exo70 protein and cisplatin. $K_D = 8.25 \times 10^{-4} \text{ mol/L}$. (B) ICP-MS assay was carried out to test the cisplatin distributions in different components of the culture medium. Data were expressed as mean \pm SEM, $n = 3$. ***, $P < 0.001$. LEVs: large extracellular vesicles, SEVs: small extracellular vesicles. (C) RT-PCR showed the ATP7A and ATP7B mRNA levels in cells. Data were expressed as mean \pm SEM, $n = 3$. ns: not significant. (D) The knock-down of ATP7A or ATP7B by shRNA in OVCAR3 cells were shown by western blotting. (E) ICP-MS assays were carried out to test the effects of knockdown ATP7A or ATP7B on the intracellular and extracellular cisplatin distributions in OVCAR3 cells. Data were expressed as mean \pm SEM, $n = 3$. *, $P < 0.05$.

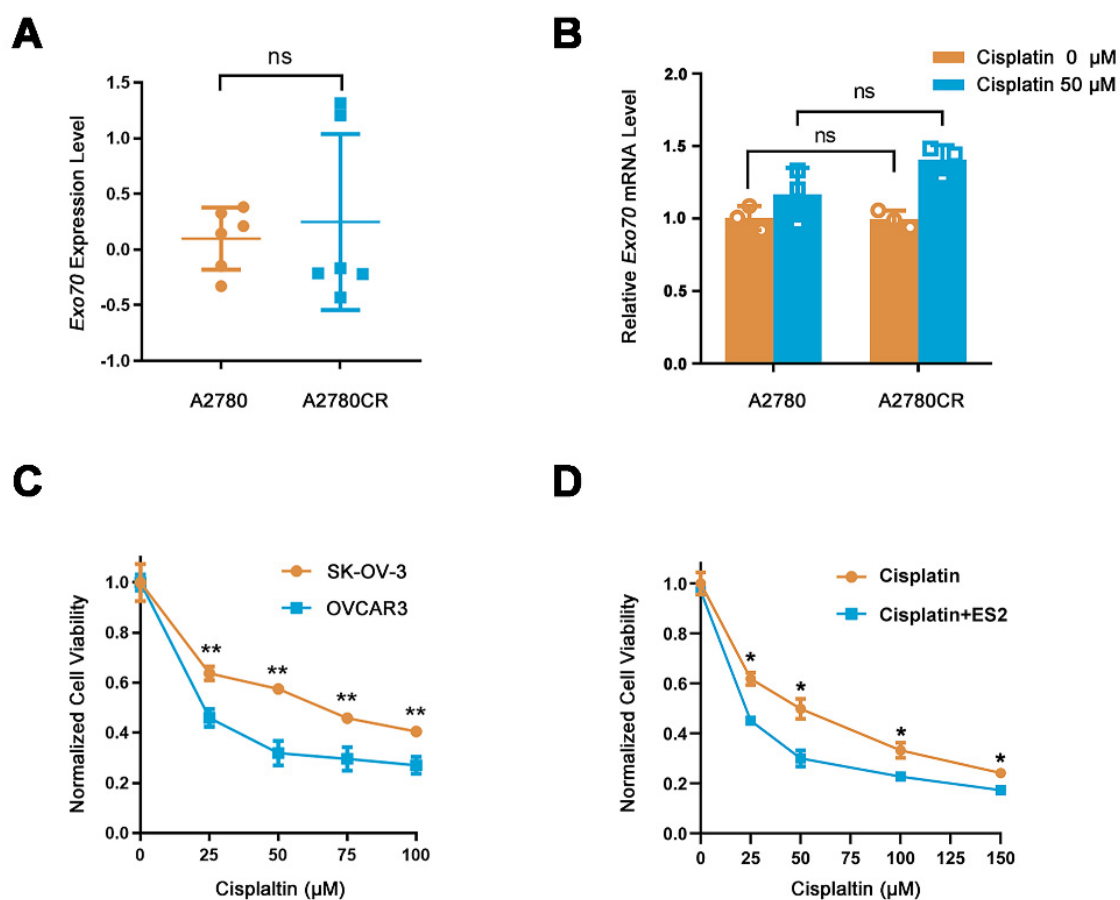


Figure S4. Exo70 affected the sensitivity of cells to cisplatin. **(A)** Exo70 mRNA expression levels were evaluated in datasets from GEO (GSE 33482) by comparing cisplatin-sensitive A2780 cells vs cisplatin-resistant A2780 cells. **(B)** RT-PCR showed the Exo70 mRNA expression levels in A2780 and A2780CR cells. Data were expressed as mean \pm SEM, $n = 3$. ns: not significant. **(C)** CCK8 assay showed the cytotoxicity of cisplatin in OVCAR3 and SK-OV-3 cells. Cells were treated with different doses of cisplatin for 24 hours. **, $P < 0.01$. **(D)** CCK8 assay was carried out to examine the cytotoxicity of cisplatin in SK-OV-3 cells with or without ES2 (100 μ M) treatment. Data were presented as mean \pm SEM, $n = 4$. **, $P < 0.01$.

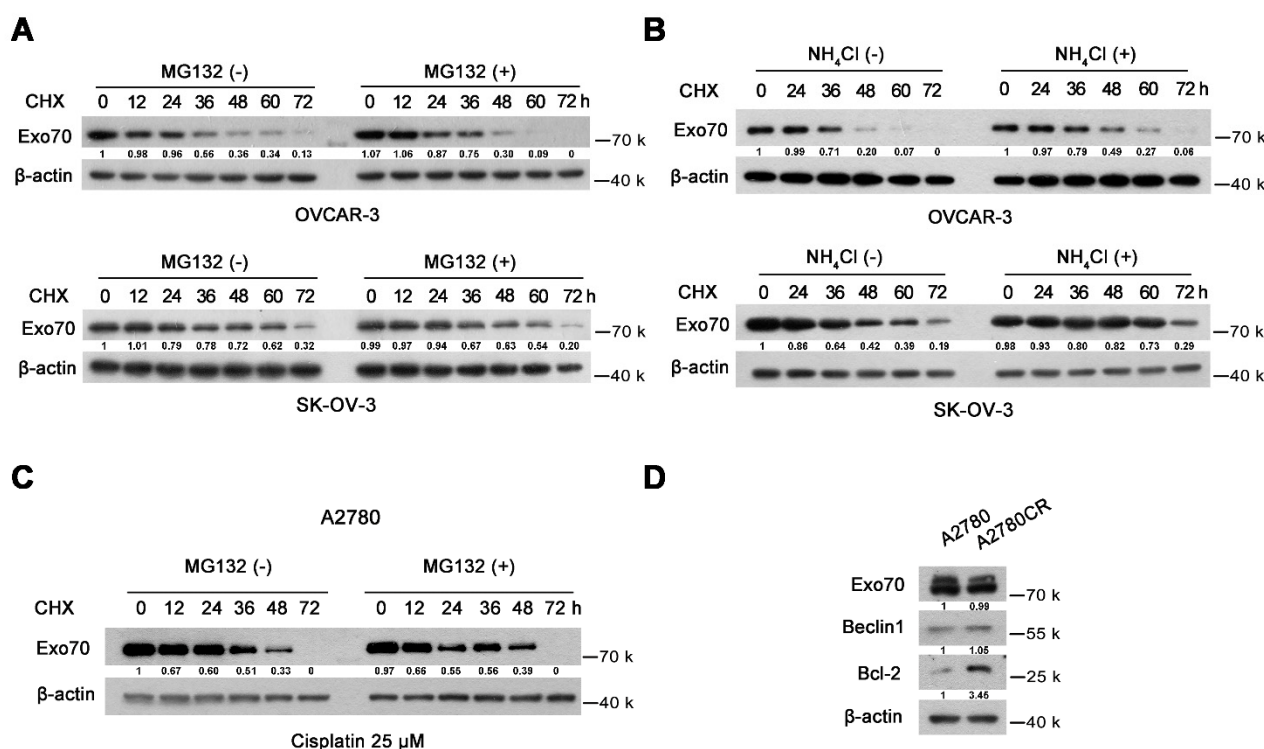


Figure S5. Autophagy affected the protein stability of Exo70. **(A)** Proteasome inhibitor MG132 did not alter the protein stability of Exo70 in OVCAR3 and SK-OV-3 cells. Cells were treated with 10 μM CHX together with or without 20 μM MG132. One of three experiments was shown. **(B)** Lysosome inhibitor NH₄Cl altered the protein stability of Exo70 in OVCAR3 and SK-OV-3 cells. Cells were treated with 10 μM CHX together with or without 10 mM NH₄Cl. One of three experiments was shown. **(C)** Western blotting was used to detect the half-lives of Exo70 in A2780 cells with 25 μM cisplatin treatment together with or without MG132. **(D)** Western blotting showed the expression level of Beclin1 and Bcl-2 in A2780 and A2780CR cells. One of three experiments was shown.

Table S1. Information of Antibodies.

Antibody	Company	Product Number	RRID
Anti-Exo70 (Rabbit)	Abcam	118792	RRID:AB_10901372
Anti-Exo70 (Goat)	Abcam	77352	RRID:AB_1523581
IgG-Isotype (Goat)	Abcam	172730	RRID:AB_2687931
Anti-LC3B (Rabbit)	Novus	NB100-2220	RRID:AB_10003146
Anti-phospho-Histone H2A.X (Rabbit)	CST	9720	RRID:AB_10692910
Anti-mTOR (Rabbit)	CST	2972	RRID:AB_330978
anti-phospho-mTOR (Ser2448) (Rabbit)	CST	5536	RRID:AB_10691552
anti-AMPKα (Rabbit)	CST	2532	RRID:AB_330331
anti-phospho-AMPKα (Thr172) (Rabbit)	CST	2535	RRID:AB_331250
Anti-Ubiquitin (Rabbit)	CST	3933	RRID:AB_2180538
Anti-ATG5 (Rabbit)	CST	12994	RRID:AB_2630393
Anti-ATG7 (Rabbit)	CST	8558	RRID:AB_10831194
Anti-ATG12 (Rabbit)	CST	4180	RRID:AB_1903898
Anti-ATG16L1 (Rabbit)	CST	8089	RRID:AB_10950320
Anti-Beclin1 (Rabbit)	CST	3495	RRID:AB_1903911
Anti-BCL2 (Mouse)	CST	15071	RRID:AB_2744528
Anti-PIK3R4 (Rabbit)	CST	14580	RRID:AB_2798523
Anti-ATP7A (Mouse)	Santa Cruz Biotechnology	sc-376467	RRID:AB_11150485

Anti-ATP7B (Mouse)	Santa Cruz	sc-373964	RRID:AB_10920224
Anti-Sec3 (Rabbit)	Proteintech	11690-1-AP	RRID:AB_2231500
Anti-Sec5 (Rabbit)	Proteintech	12723-1-AP	RRID:AB_10734442
Anti-Sec15 (Rabbit)	Proteintech	18420-1-AP	RRID:AB_10694431
Anti-Flag (Mouse)	Sigma-Aldrich	F3165	RRID:AB_259529
Anti-HA (Mouse)	Sigma-Aldrich	H9658	RRID:AB_260092
Anti-c-Myc (Mouse)	Sigma-Aldrich	M4439	RRID:AB_439694
Anti-Cisplatin DNA adducts (Rabbit)	Sigma-Aldrich	MABE416	

Table S2. Sequences of oligonucleotide primers for shRNA construction.

Target	Sequence
shExo70#1	5'-GCACAGAGACCGTGGGCAA-3'
shExo70#2	5'-GCTGGTGTGTACATGAA-3'
shATG5#1	5'-ATATCAGGATGAGATAAC-3'
shATG5#2	5'-GCAGTGGCTGAGTGAACATCT-3'
shATG7#1	5'-GCTGCCTGTCCTGAAATAGAT-3'
shATG7#2	5'-GCAGAACAATTCCTTGCATTC-3'
shSec3#1	5'-TTTGGAGTTCCTCAGCTT-3'
shSec3#2	5'-GAGTGGCTAAAGAGTACAGAT-3'
shSec5#1	5'-GCCTGGTATCTTATAGAGAAT-3'
shSec5#2	5'-CAATGTGCTTCAGCGATTTAA-3'
shSec15#1	5'-GCAGTTAACTTAGATGTCAT-3'
shSec15#2	5'-CCCTTTCAAGATCCAGACCTT-3'
shATP7A#1	5'-GGAAGGGATGACCTGCCATTC-3'
shATP7A#2	5'-GCAGCCCAAGTACCTCAAATT-3'
shATP7B#1	5'-GCAGTGCCACTGTGAAATATG-3'
shATP7B#2	5'-GGGACCATGTAAATGACATGG-3'

Table S3. Sequences of primers for real-time PCR.

Gene	Primers
<i>GAPDH</i>	Forward: 5'-TGCACCACCAACTGCTTAGC-3'
	Reverse: 5'-GGCATGGACTGTGGTCATGAG-3'
<i>EXOC7</i>	Forward: 5'-GGAGTATTTCCAGGACAACAGC-3'
	Reverse: 5'-AAGATGAGCACGGGCGAGA-3'
<i>ATG5</i>	Forward: 5'-AACTGAAAGGGAAGCAGAACCA-3'
	Reverse: 5'-ATGCCATTTTCAGTGGTGTGCC-3'
<i>ATG7</i>	Forward: 5'-GCTTGTGCCTCACCAGGTTC-3'
	Reverse: 5'-GATGGTCTCATCATCGCTCATGTC-3'
<i>EXOC1</i>	Forward: 5'-ACCAAATGATGAACGCCTGC-3'
	Reverse: 5'-GGCATCTACCACAGCAAGAT-3'
<i>EXOC2</i>	Forward: 5'-AAGAACTGGTCCCTCGGGTA-3'
	Reverse: 5'-TCGGGTGTCAGGTAAACAGC-3'
<i>EXOC6</i>	Forward: 5'-CGGGTGAATCCAAACACAGC-3'
	Reverse: 5'-TCAGCTGTTTCACGACTGTCT-3'
<i>ATP7A</i>	Forward: 5'-TGGACCATTGAGCAGCAGAT-3'
	Reverse: 5'-GTGACGCCGTAACAGTCAGA-3'
<i>ATP7B</i>	Forward: 5'-TGGTCACCCTCCAACTGAGA-3'
	Reverse: 5'-GGGCTGGTACAAGAAGGGTC-3'

Table S4. Characteristics of the ten ovarian cancer patients.

Platinum-sensitive		
Serial Number	Stage	Pathological Type
1	II c	Serous ovarian cancer
2	III b	Serous ovarian cancer
3	III c	Serous ovarian cancer
4	III c	Serous ovarian cancer
5	III c	Serous ovarian cancer
Platinum-resistant		
Serial Number	Stage	Pathological Type
1	III a	Serous ovarian cancer
2	III c	Serous ovarian cancer
3	III c	Serous ovarian cancer
4	III c	Serous ovarian cancer
5	IV	Serous ovarian cancer

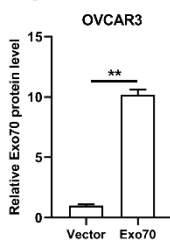
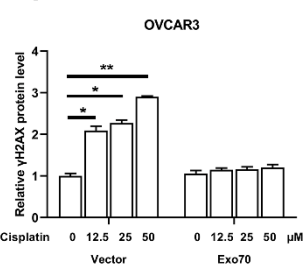
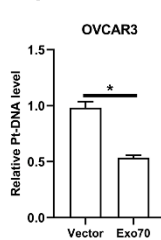
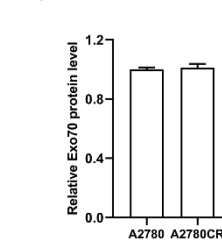
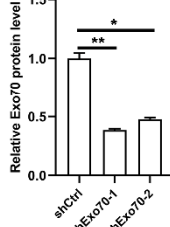
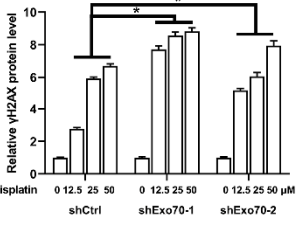
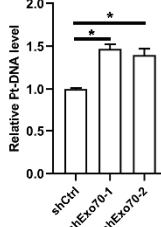
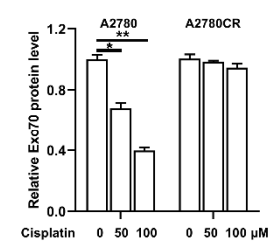
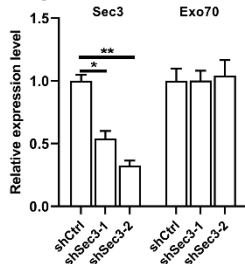
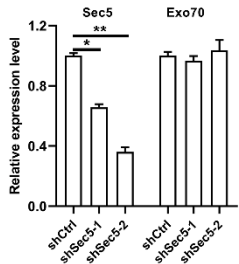
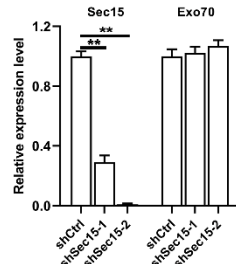
Figure 2A**Figure 2C****Figure 3E****Figure 4B****Figure 3F****Figure 3G****Figure 3H****Figure 4D****Figure 3I****Figure 3J****Figure 3K**

Figure 5A

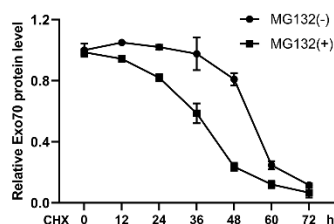


Figure 5B

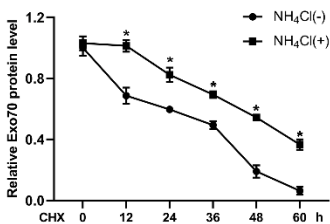


Figure 5D

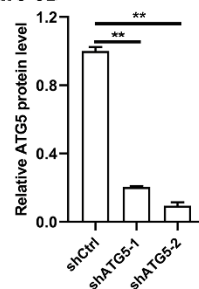


Figure 5C

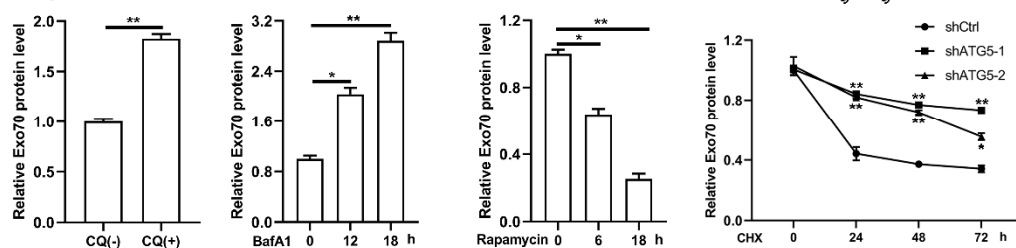


Figure 5F

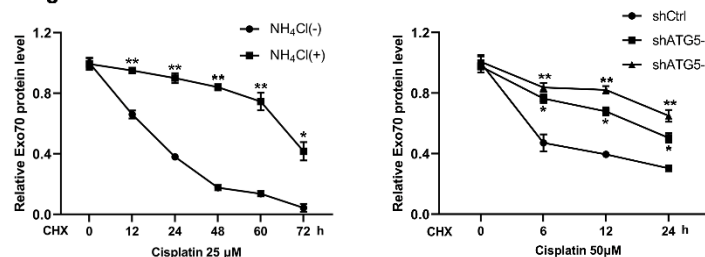


Figure 5H

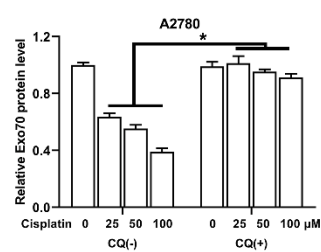


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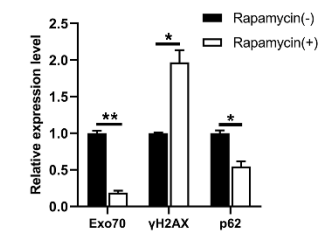


Figure 5I

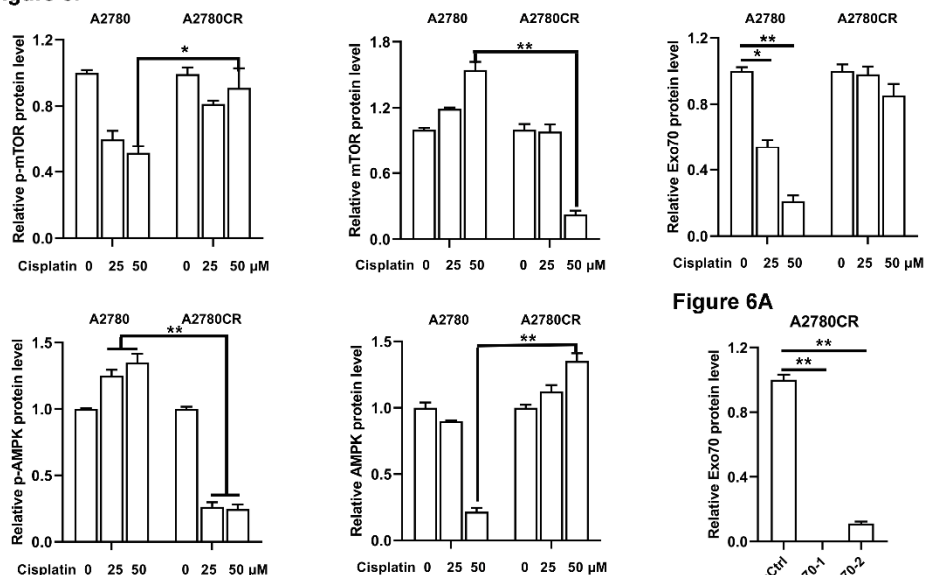
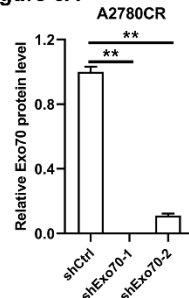


Figure 6A



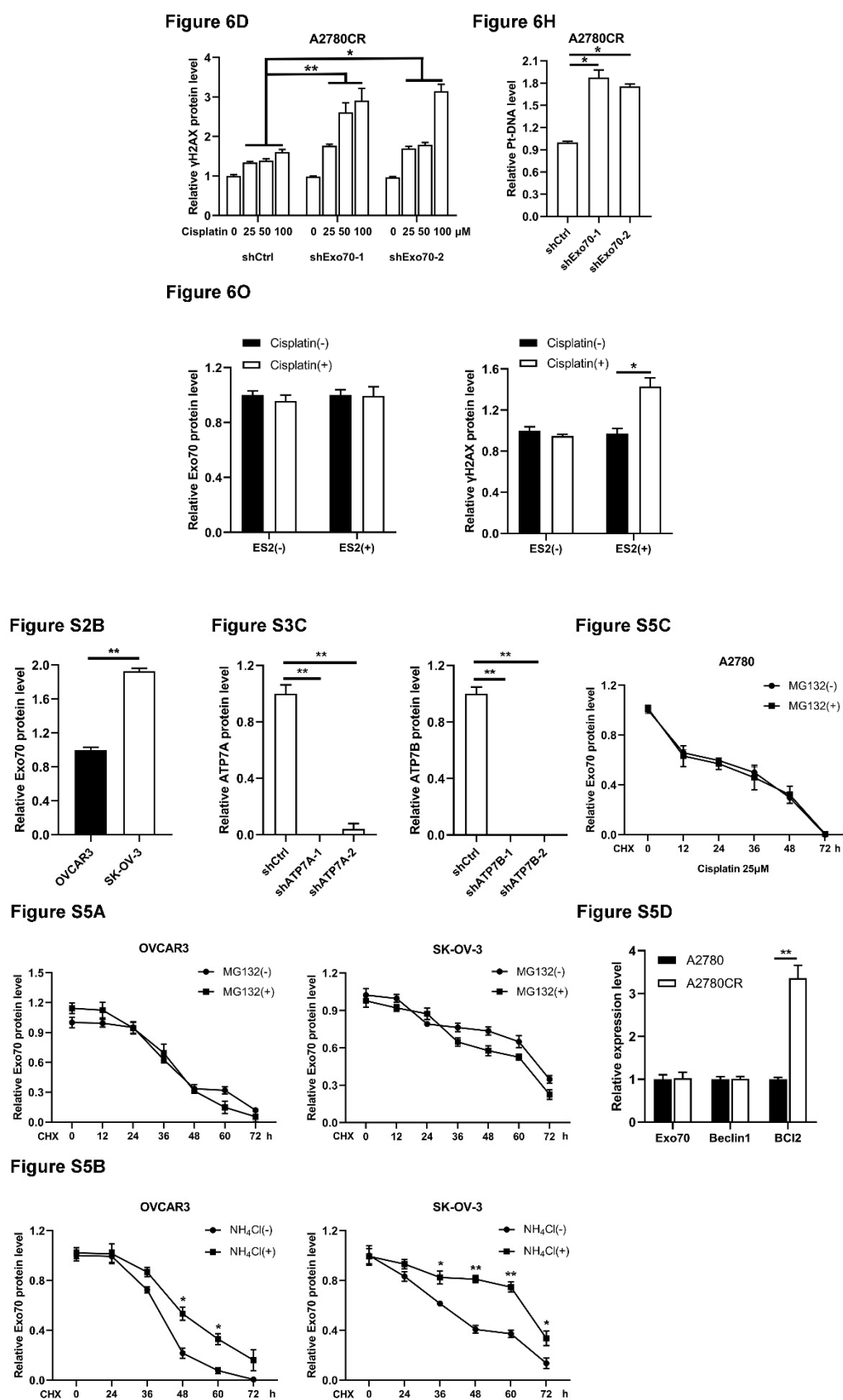


Figure S6. Quantification of Western Blot. Data were expressed as mean \pm SEM, $n = 3$. * $P < 0.05$, ** $P < 0.01$.