

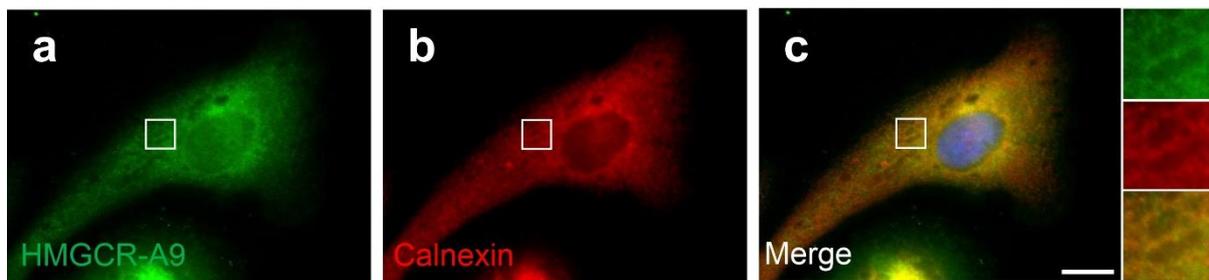
Supplementary Material

Peroxisomal Localization of a Truncated HMG-CoA Reductase under Low Cholesterol Conditions

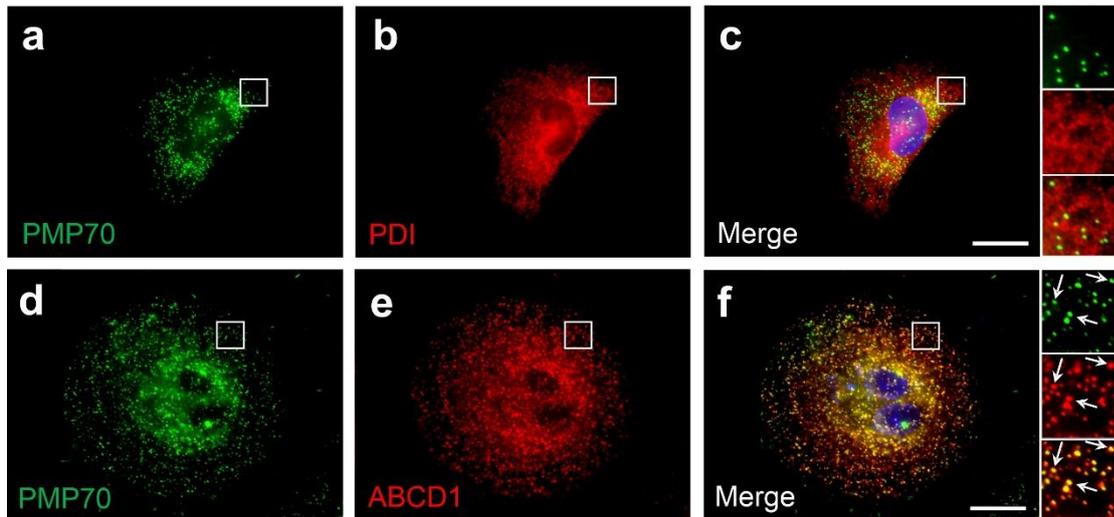
Jianqiu Wang, Markus Kunze, Andrea Villoria-González, Isabelle Weinhofer and Johannes Berger

List of supplementary materials:

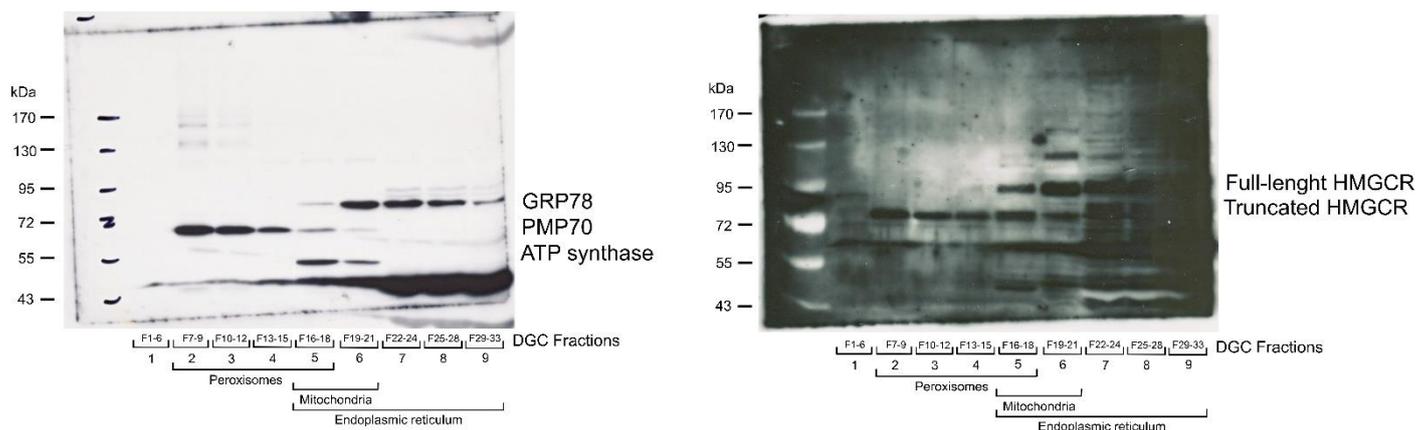
- Supplementary Figure S1. The monoclonal anti-HMGCR antibody (anti-HMGCR-A9) stains predominantly the ER.
- Supplementary Figure S2. Specificity controls for the antibodies in THP-1 cells.
- Supplementary Figure S3. Unprocessed full-size images of Figure 3.
- Supplementary Figure S4. Independent density gradient and western blot analysis of peroxisomal HMGCR in differentiated THP-1 cells grown under low cholesterol conditions and lovastatin treatment.



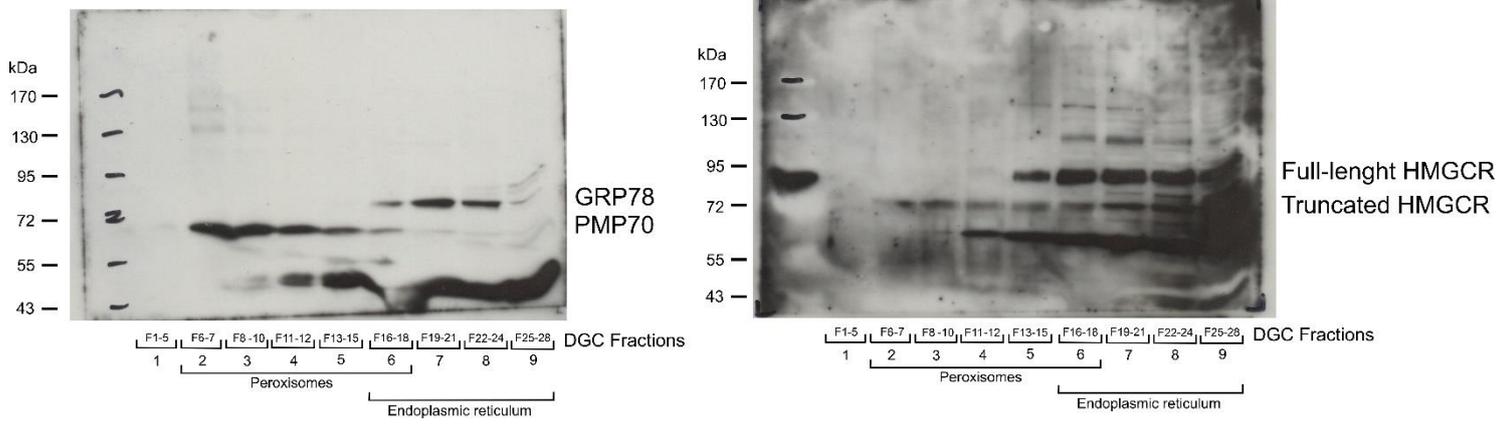
Supplementary Figure S1. The monoclonal anti-HMGCR antibody (anti-HMGCR-A9) stains predominantly the ER. (a-c) Cells of the human monocytic cell-line THP-1 were differentiated using 40 ng/ml PMA and cultivated in lipid-depleted medium for 7 days, with 5 μ M lovastatin added for the last 16 hours to increase endogenous HMG-CoA reductase (HMGCR) protein levels. The subcellular distribution of HMGCR was investigated by immunofluorescence microscopy: (a) Monoclonal α -HMGCGR-A9 (green), (b) polyclonal α -Calnexin (red, labeling the ER) and (c) overlay. The co-localization demonstrates that the monoclonal HMGCGR-A9 antibody detects HMGCR in the ER. Scale bar, 20 μ m.



Supplementary Figure S2. Specificity controls for the antibodies in THP-1 cells. (a-f) Immunofluorescence microscopic analysis of PMA-differentiated THP-1 cells (7 days) under low cholesterol condition. The signals obtained for the peroxisomal marker protein, PMP70 (green) and the ER marker protein, PDI (red), showed no co-localization (**a-c**); while PMP70 (green) and another peroxisomal marker protein, ABCD1 (red), showed very strong co-localization mutually confirming their peroxisomal localization (**d-f**). White arrows: punctate, peroxisomal staining pattern. Scale bars, 20 μm .



Supplementary Figure S3. Unprocessed full-size images of Figure 3. In differentiated THP-1 cells grown under low cholesterol condition and lovastatin treatment a truncated form of HMGCR was found in peroxisome enriched fractions obtained by density gradient centrifugation (DGC). 40 flasks of THP-1 cells were differentiated and incubated in lipid depleted medium for 7 days. 5 μ M lovastatin was provided for the last 16 h before harvesting (low cholesterol condition). DGC was used to enrich organelles and separate fractions with different composition according to their density. Fractions were then pooled and western blots were performed using α -GRP78, α -PMP70 and α -ATP synthase antibodies to identify fractions enriched in ER (fraction 16-33, lane 5-9), peroxisome (F7-18, lane 2-5) and mitochondria (F16-21, lane 5-6), respectively. Whereas full length HMGCR with a size of about 97 kDa was found in fractions enriched in the ER (Lanes 5-9), a truncated form HMGCR with a size of about 76 kDa was found in peroxisome enriched fractions (Lane 2-5). The size of the molecular weight marker is indicated on the left side.



Supplementary Figure S4. Independent density gradient and western blot analysis of peroxisomal HMGCR in differentiated THP-1 cells grown under low cholesterol conditions and lovastatin treatment. As in Figure 3 and Supplementary Figure 3, a shorter (~76 kDa) HMGCR fragment was detected also in the PMP70-positive gradient fractions containing the peroxisomes, whereas the full-length HMGCR was confined to the GRP78-positive, ER-containing microsomal fractions.