

# **Exo2D™ - for EV isolation**

(Urine, Culture media, Saliva, Ascites fluid)

## **User manual**

**Store kit at +25 C on receipt**

**A limited-use label license covers this product. By use of this product, you accept the terms and conditions outlined in the Licensing and Warranty Statement contained in this user manual.**

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## List of components

| Item                  | Volume |
|-----------------------|--------|
| EXO2D™ - L solution A | 15 ml  |

Exo2D™ are shipped at room temperature and should be stored at +25°C upon receipt. Properly stored kits are stable for 6 month from the date received.

Exo2D™ can be used for urine, saliva, spinal cord, ascites, culture media, and etc.

Range of bio fluid volume is 50 ul to 100 ml.

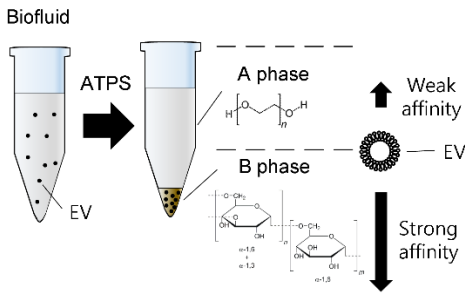
# Exo2D™ – for RNA analysis

## I. Overview

Exosomes, small membrane vesicles secreted by cell, are found in blood, urine, amniotic fluid, malignant ascite fluids and contain distinct subsets of microRNAs and proteins. Although exosome have increasingly received attention by the field of diagnosis, regeneration, delivery, and etc., existing isolation methods including commercialized kit did not meet the users' needs because of low isolation efficiency and poor purity. To solve the unmet needs, we developed a new exosome isolation kit which mechanism is different from the previous ones. We guarantees almost perfect exosome isolation efficiency and highest purity. This may make impossible assays to possible assays.

- No time-consuming ultracentrifugation
- No incubation time
- Total process time is less than 1 h
- Almost perfect exosome isolation efficiency
- Purity of exosome can be improved by additional process
- High ability of RNA and miRNA PCR
- High ability of NGS

## II. Principle



We developed a new exosome isolation methods, which is different from the conventional methods.

Following is a main mechanism of the conventional methods.

**Ultracentrifugation:** Use high g-force

**Polymeric precipitation:** Aggregate the exosome

**Magnetic bead:** Antibody-antigen reaction

**Chromatography:** Size exclusion or antibody-antigen reaction

Unlike the methods, we use surface property of exosomes.

We make an aqueous two phase, which is similar to oil and water phase.

Using the interactions between surface of exosome and each phase, we almost perfectly isolate exosome to one of phases.

# III. PROTOCOL

Material delivered

## A11 Incoming Inspection

- . Check the labels
- . Visual check : Damage, impurity, leakage during delivery

Urine, Culture media,  
Saliva, Ascites fluid

Site stored at 25°C

Exo2D™-L Reagent A

## A21 Pre-cleaning

- . Centrifuge at 3000×g for 15 min to remove cells and cell debris.
- . Transfer supernatant to a new tube

## A22 Inspection

- . Visual check : **pellet**

## A23 Sample collection

- . Transfer supernatant to a new tube

## A41 Injection & Mixing

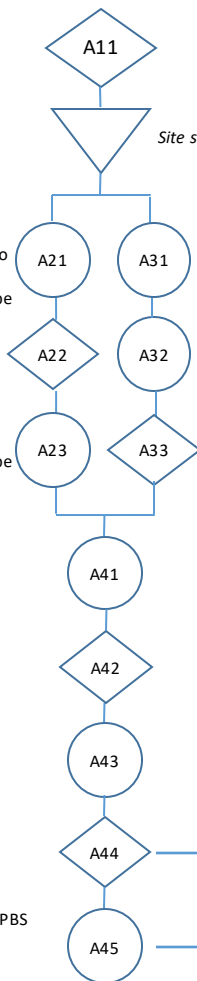
- . Immediately, add to 10 ml of sample per 2 ml of Exo2D™-L reagent A
- . Mixing well by inverting or flicking

## A44 Removal of supernatant

- . Exosomes are dissolved in very small volume of aqueous phase of Exo2D™
- . The phase appears as white pellets. The remainder should be eliminated.

## A45 Resuspending

- . Resuspend bottom solution using PBS



## A31 Incubation

- . Incubate at 37°C for 15 min.

## A32 Homogenization

- . Vortexing, Inverting or flicking the tube

## A33 Inspection

- . Visual check : Evenly **OPAQUE**
- . Shake every 5 min. during the process to keep optically opaque

## A42 Inspection

- . The solution was incubated at 4°C for 30 min.
- . Mixing well by inverting or flicking
- . Visual check : Evenly **OPAQUE**

## A43 Precipitation

- . Centrifuge at 3,000 × g for 30 min. at 4°C.
- . Aqueous phase of Exo2D™ grabs exosomes and precipitated

*Optional*  
Go to step B in case you  
need highly purified  
exosome

*Optional*  
Go to step C in case you  
use trizol based RNA  
isolation



#### B11 Purification

- . Add 5 ml purification solution
- . Mix well by inverting or flicking the tube

#### B12 Inspection

- . Visual check : pellet was dissolved in purification solution

#### B21 Centrifugation

- . Centrifuge the sample at  $3,000 \times g$  for 30 minutes at  $4^{\circ}\text{C}$ .
- . Remove the supernatant.

#### B12 Inspection

- . Visual check : pellet

#### A23 Resuspending

- . Resuspend bottom solution using PBS

### III. PROTOCOL



C11

C21

C22

C31

C41

C51

#### C11 Lysis and Phase separation

*You can use your protocol in this step*

- . add 1000  $\mu\text{L}$  of **Lysis Buffer** per 200  $\mu\text{L}$  of the isolated sample.
- . Shake tube vigorously by vortex for 15 seconds.
- . Incubate for 2 min at room temperature
- . Add 300  $\mu\text{L}$  of chloroform per 1 mL of **Lysis Buffer**.
- . Incubate for 2 min at 4°C

Shake tube vigorously by vortex for 15 seconds . . Centrifuge the sample at  $12,000 \times g$  for 15 min at 4°C.

#### C21 Purification

- . Place the aqueous phase into a new tube, and add 1ml of chloroform.
- . Shake the tube vigorously by vortex for 10 seconds.
- . Place the sample on ice, and shake the sample for 30 min

#### C22 Sample extraction

- . Centrifuge at  $1000 \times g$  for 5 minutes.
- . Separate upper phase by pipette.

#### C31 RNA precipitation

*You can use your protocol in this step*

- . Add same volume of isopropanol to the upper phase
- . Incubate overnight at -80°C or incubate at -20°C for 30 min
- . Centrifuge at  $16,000 \times g$  for 10 minutes at 4°C
- . Remove the supernatant from the tube, leaving only the RNA pellet

#### C41 RNA purification

- . Wash the pellet, with 1 mL of 70% ethanol (invert the tube) If there is white pellet in the bottom of tube, shake the tube by vortexing for 10 sec. The white material protects RNA and assists precipitation of RNA.

- . Centrifuge at  $16,000 \times g$  for 10 minutes at 4°C
- . Repeat step C41, after first purification, you don't need to vortexing the tube.

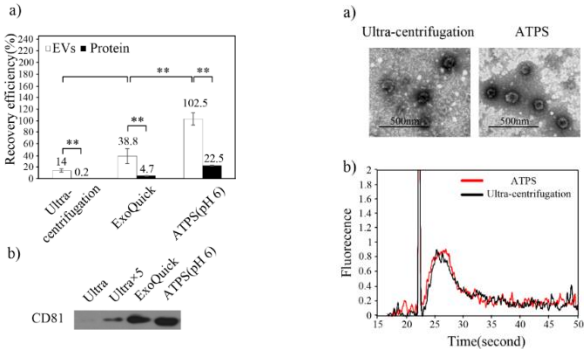
#### C51 Resuspending

*You can use your protocol in this step*

- . Air dry the RNA pellet
- . Resuspend the RNA pellet in 10  $\mu\text{L}$  RNase-free water

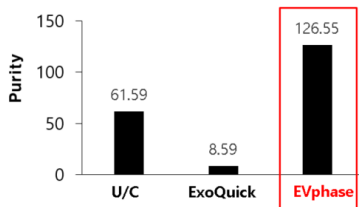
## IV. Sample data and applications

### A. Isolation efficiency of exosome



- Exosome isolation efficiency of ATPS is almost perfect.

### B. Purity of exosome



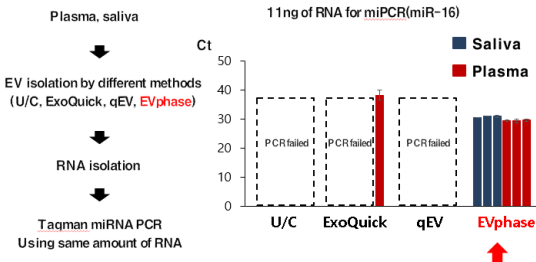
- Purity of isolated exosome by EVphase is highest among the methods.



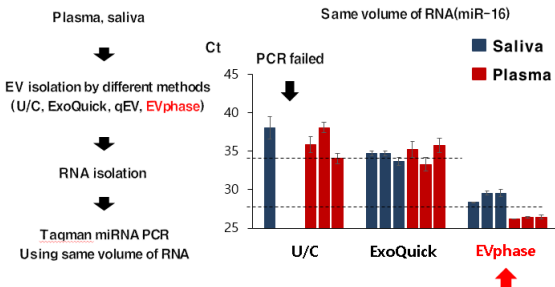
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### C. mi-RNA PCR

#### 1. High miRNA performance – High purity



#### 2. High miRNA performance – High yield



- Highest miRNA performance
- High reliability