

Microcarrier-Based Culture of Human Pluripotent Stem-Cell-Derived Retinal Pigmented Epithelium

Mohamed A. Faynus ^{1,2,*}, Jeffrey K. Bailey ², Britney O. Pennington ², Mika Katsura ², Duncan A. Proctor ^{2,3}
Ashley K. Yeh ^{2,3}, Sneha Menon ^{2,3}, Dylan G. Choi ^{2,4}, Jane S. Lebkowski ⁶, Lincoln V. Johnson ⁶
and Dennis O. Clegg ^{2,3,5,6}

¹ Program for Biomolecular Science and Engineering, University of California, Santa Barbara, CA 93106, USA

² Center for Stem Cell Biology and Engineering, Neuroscience Research Institute, University of California, Santa Barbara, CA 93106, USA; jkbailey@ucsb.edu (J.K.B.); bop@ucsb.edu (B.O.P.);

mikakatsura@ucsb.edu (M.K.); duncan@ucsb.edu (D.A.P.); ashleyyeh@umail.ucsb.edu (A.K.Y.);

snehamenon@ucsb.edu (S.M.); dylanchoi@ucsb.edu (D.G.C.); clegg@ucsb.edu (D.O.C.)

³ Department of Molecular, Cellular and Developmental Biology, University of California, Santa Barbara, CA 93106, USA

⁴ College of Creative Studies, Chemistry and Biochemistry, University of California, Santa Barbara, CA 93106, USA

⁵ Program in Biological Engineering, University of California, Santa Barbara, CA 93106, USA

⁶ Regenerative Patch Technologies LLC, Portola Valley, CA 94028, USA; jane@regenerativepatch.com (J.S.L.); linc@regenerativepatch.com (L.V.J.)

* Correspondence: faynus@ucsb.edu

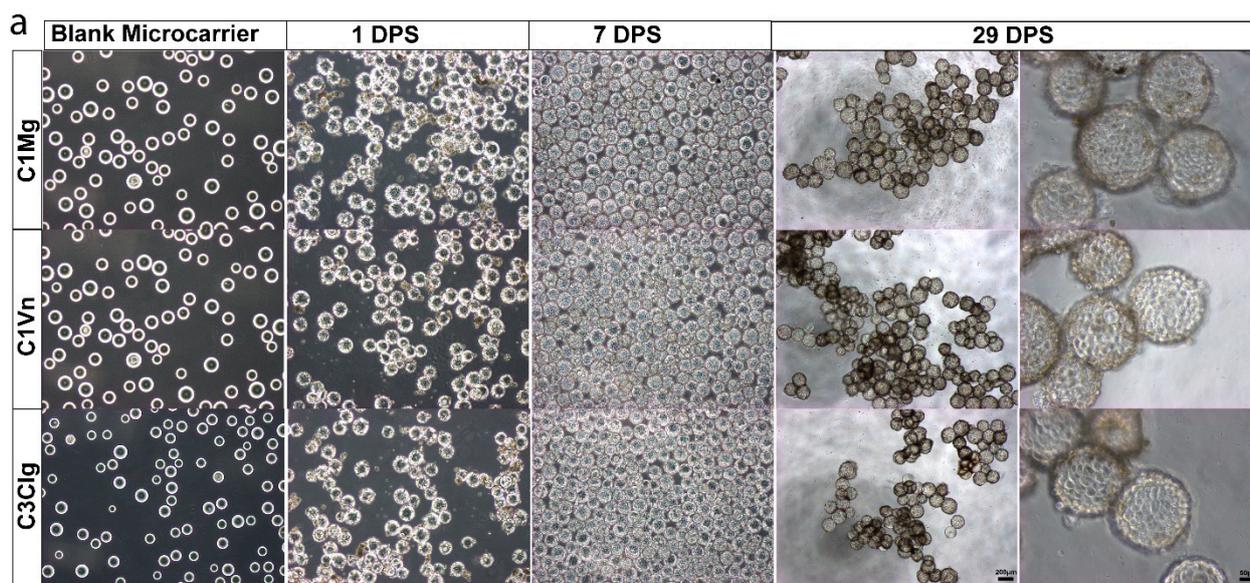


Figure S1. mcRPE mature and pigment over 30-day culture period. Phase contrast images demonstrate attachment of hESC-RPE cells to Matrigel and recombinant human vitronectin coated Cytodex 1 as well as pre-coated collagen Cytodex 3 microcarriers. mcRPE display cobblestone morphology, phase bright borders and pigmentation 29DPS.

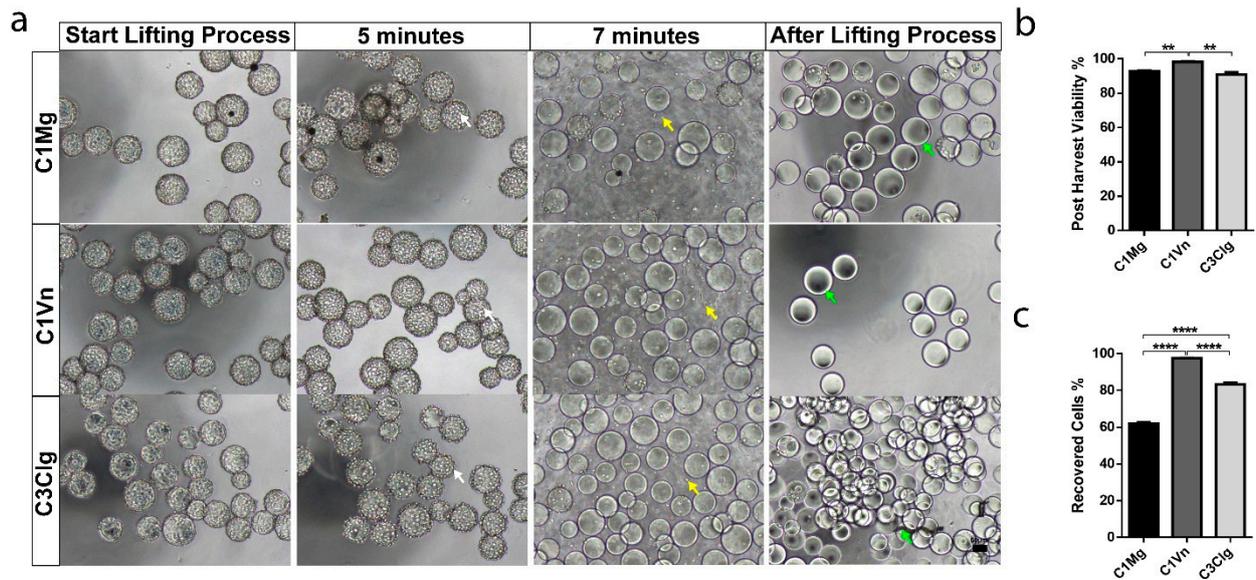


Figure S2. Harvesting hESC-RPE cells from microcarriers using a xeno-free enzymatic disassociation reagent. **a)** mcRPE were collected at a density of 2×10^5 cells per well and exposed to TrypLE enzyme for a period of 7 min. Cells were still attached at 5 min (white arrows) and were completely dissociated after mechanical perturbation at 7 minutes. Dissociated cells were clearly visualized post perturbation (yellow arrow) and harvested by filtration to separate microcarriers from cells. Empty microcarriers (green arrow) after filtration process demonstrates appropriate harvesting of hESC-RPE cells. **b)** Viability of harvested hESC-RPE cells were assessed using Acridine Orange/DAPI exclusion on an NC200. All three conditions demonstrate high viability with C1Vn exhibiting significantly greater viability than C1Mg and C3Clg (** $p < 0.05$). **c)** Cell recovery was quantified using Acridine Orange/DAPI and compared to initial starting density equating to an approximate 2×10^5 cells. All three conditions were amenable to cell recovery but C1Vn exhibited a significantly greater percentage of recovered cells compared to C1Mg and C3Clg. C1Mg was the least favorable coating for cell recovery (**** $p < 0.001$). Statistical analysis, one way ANOVA, Tukey's correction.

Table S1. Calculations to determine the required surface area to meet the estimated patient demand and approaches to achieve.

<i>Surface Area Required to Supply 8×10^9 RPE Cells*</i>	
$\frac{8 \times 10^9 \text{ RPE cells}}{\text{year}}$	$\times \frac{\text{cm}^2}{2.1 \times 10^5 \text{ RPE cells}} \times \frac{1 \text{ year}}{12 \text{ months}} = \frac{3,174 \text{ cm}^2}{\text{month}}$
<i>Approaches to Achieve $\frac{3,174 \text{ cm}^2}{\text{month}}$</i>	
T-75 Flasks	$\frac{3,174 \text{ cm}^2}{\text{month}} \times \frac{1 \text{ flask}}{75 \text{ cm}^2} \cong \frac{42 \text{ flasks}}{\text{month}}$
T-225 Flasks	$\frac{3,174 \text{ cm}^2}{\text{month}} \times \frac{1 \text{ flask}}{225 \text{ cm}^2} \cong \frac{14 \text{ flasks}}{\text{month}}$
Cytodex 1	$\frac{3,174 \text{ cm}^2}{\text{month}} \times \frac{\text{g of Cytodex1}}{4400 \text{ cm}^2} \times \frac{L}{5 \text{ g of Cytodex**}}$ = $\frac{144 \text{ mL Bioreactor Volume}}{\text{month}}$
Cytodex 3	$\frac{3,174 \text{ cm}^2}{\text{month}} \times \frac{\text{g of Cytodex3}}{2700 \text{ cm}^2} \times \frac{L}{5 \text{ g of Cytodex**}}$ = $\frac{235 \text{ mL Bioreactor Volume}}{\text{month}}$

* Estimate of the required cells assumes that only half of the projected number of AMD patients in the United States receives an RPE cell-based therapy [36].

** Microcarrier cultures may contain 1-5g of Cytodex/L [21].