

Supplemental material

Ovicidal Effect on *Haemonchus contortus* of Extract Partitions Shrubby Plants of the Tropical Dry Forest and Potentially Active Compounds Identification by UHPLC-Q/Orbitrap/MS/MS

Néstor Jaime Romero-Jola ¹, Jaime Andrés Cubides-Cárdenas ², Natalia Escobar ^{3*} and Mario J. Simirgiotis ⁴

¹ Departamento de Sanidad Animal, Facultad de Medicina Veterinaria y Zootecnia, Universidad del Tolima, Ibagué 730001, Colombia; njromeroj@ut.edu.co

² Grupo de Investigación e Innovación en Salud y Bienestar Animal, Laboratorio de Salud Animal y Microbiología Pecuaria, Centro de Investigación Tibaitatá, Agrosavia, Mosquera 250047, Colombia

³ Grupo de Investigación Área Verde, Universidad de Cundinamarca, Fusagasugá 252211, Colombia

⁴ Instituto de Farmacia, Facultad de Ciencias, Universidad Austral de Chile, Campus Isla Teja, Valdivia 5090000, Chile

* Correspondence: nataescobar@ucundinamarca.edu.co

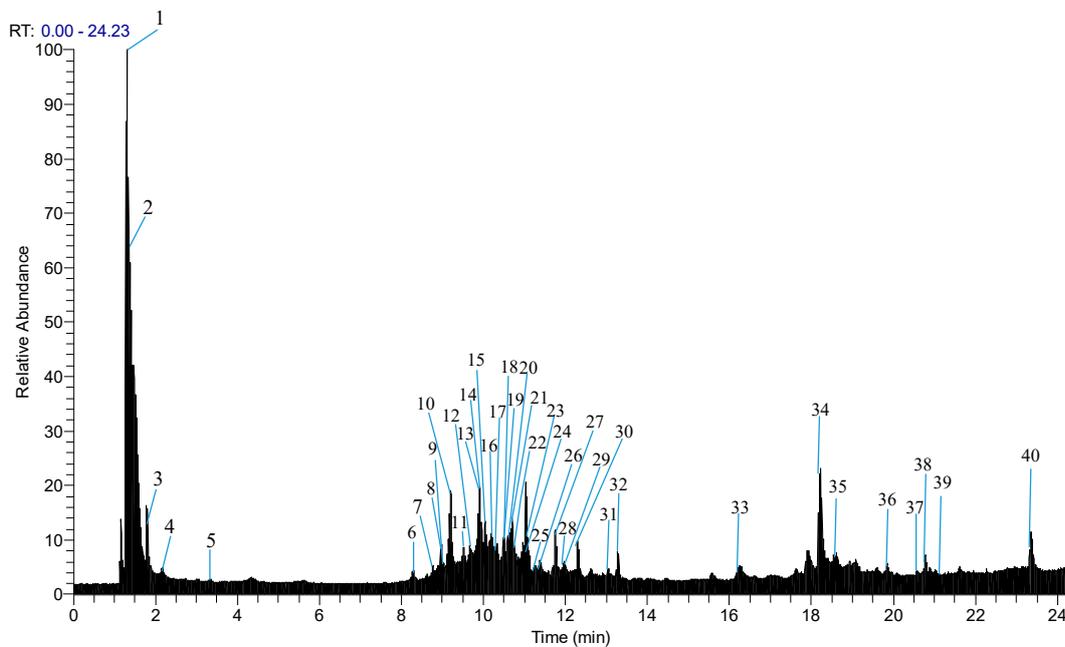


Figure S1. UHPLC TIC (total ion current) chromatogram of *Pithecellobium dulce* hydroethanolic extract partition

Table S1. Tentative identification of compounds in the hydroethanolic extract partition of *Pithecellobium dulce* by UHPLC-Q/Orbitrap/MS/ MS

Peak	Tentative identification	[M-H] ⁻	Retention time (min.)	Theoretical mass (m/z)	Measured mass (m/z)	Accuracy (ppm)	Other Ions (m/z)*	UV
1	Arabinonic acid	C ₅ H ₉ O ₆	1.30	165.0399	165.0400	-0,6	147.0295; 119.0343	251; 271;
2	3-Deoxyhexonic acid	C ₆ H ₁₁ O ₆	1.38	179.0556	179.0558	-1,1	143.0350; 119.0341	268
3	Citric acid	C ₆ H ₇ O ₇	1.80	191.0192	191.0195	-1,6	111.0080	204; 302
4	2,3,5,4'-Tetrahydroxystilbene 2-O-β-D-glucoside	C ₂₀ H ₂₁ O ₉	2.16	405.1186	405.1172	3,5	243.0624; 179.0557; 227.0672; 163.0608	248; 271; 295
5	Protocatechuic acid 4-hexoside	C ₁₃ H ₁₅ O ₉	4.35	315.0716	315.0723	-2,2	153.0221; 152.0165; 137.0237	200; 277; 300
6	Bis(ethoxycarbonyloxymethyl) undecanedioate	C ₁₉ H ₃₁ O ₁₀	8.30	419.1917	419.1926	-2,1	355.0672	216; 276; 315
7	Calaliukiuenoside	C ₂₀ H ₃₅ O ₁₁	8.77	451.2179	451.2188	-2,0	341.1093; 179.0557	233; 278; 315
8	Ebuloside	C ₂₁ H ₃₁ O ₁₀	8.98	443.1917	443.1925	-1,8	179.0555	268; 323
9	Coumaroylquinic acid	C ₁₆ H ₁₇ O ₈	9.09	337.0923	337.0931	-2,4	163.0396; 119.0495; 191.0557; 173.0449	236; 280; 349
10	7-Oxo-tridecanedioic acid	C ₁₃ H ₂₁ O ₅	9.20	257.1394	257.1394	0,0	213.1493	234; 283; 357
11	Unknow	C ₁₅ H ₂₆ O ₉ N	9.51	364.1608	364.1617	-2,5	---	239; 280; 356
12	Ferulic acid-O-hexoside derivative	C ₁₉ H ₂₉ O ₁₂	9.67	449.1659	449.1669	-2,2	355.1038	239; 278; 320
13	Hidroxydioxotridecenoic acid	C ₁₃ H ₁₉ O ₅	9.90	255.1232	255.1238	-2,4	211.1337; 153.0916	240; 278; 320
14	Dihydroxyhexaoxohexacosanoic acid	C ₂₆ H ₃₉ O ₁₀	9.90	511.2543	511.2550	-1,4	465.1440; 323.1114; 255.1238; 153.0914; 211.1337	240; 278; 320
15	Tetrahidroxitetraoxoicosanoic acid	C ₂₀ H ₃₁ O ₁₀	10.05	431.1917	431.1926	-2,1	401.1461; 387.2020	238; 278; 328
16	Heptahydroxyoxohexadecanoic acid	C ₁₆ H ₂₉ O ₁₀	10.20	381.1761	381.1770	-2,4	351.1301	243; 278; 331
17	Tetrahydroxytryoxoicosanoic acid	C ₂₀ H ₃₃ O ₁₀	10.33	433.2074	433.2083	-2,1	387.2028	241; 280; 360
18	Unknow	C ₁₀ H ₁₉ O ₁₄	10.50	363.0775	363.0759	4,4	345.1559; 144.0084; 150.0316	245; 276; 360
19	Verbasoside	C ₂₀ H ₂₉ O ₁₂	10.52	461.1659	461.1666	-1,5	445.1718; 447.1521	245; 277; 360
20	Nepetaside	C ₁₆ H ₂₅ O ₈	10.59	345.1549	345.1559	-2,9	327.1452; 163.0607; 183.1024; 301.1669; 289.1657; 206.0820	242; 280; 320
21	Tetrahydroxytetraoxoicosanoic acid	C ₂₀ H ₃₁ O ₁₀	10.71	431.1917	431.1926	-2,1	389.2185; 417.2127	245; 271; 320
22	Rutin	C ₃₀ H ₂₅ O ₁₄	10.74	609.1482	609.1457	4,1	300.0280; 137.0248; 149.0238	245; 276; 323
23	Ethyl-morrinoside	C ₁₉ H ₂₉ O ₁₁	11.04	433.1710	433.1719	-2,1	375.1665; 179.0557; 255.1236; 417.1762	242; 280; 331
24	Phloretin-di-C-hexoside	C ₂₇ H ₃₃ O ₁₅	11.11	597.1819	597.1823	-0,7	477.1406; 435.1284; 507.1497	280; 331; 368

25	Quercetin-3-glucoside	C ₂₁ H ₁₉ O ₁₂	11.26	463.0877	463.0886	-1,9	179.0556; 163.0610; 151.0395	280; 331; 349
26	Vicenin-2	C ₂₇ H ₂₉ O ₁₅	11.39	593.1506	593.1510	-0,7	117.0380; 533.1301; 159.0295; 163.0401	282; 331; 368
27	Luteolin 7-O-glucoside	C ₂₁ H ₁₉ O ₁₁	11.76	447.0927	447.0936	-2,0	151.0031; 255.0301; 107.0131	283; 368; 379
28	Tridecahydroxypentadecanoic acid	C ₁₅ H ₂₉ O ₁₅	11.90	449.1506	449.1490	3,6	---	283; 368
29	Picrocrocin	C ₁₆ H ₂₅ O ₇	11.97	329.1600	329.1609	-2,7	299.1502; 179.0555; 167.1078; 163.0607	283; 368
30	Kaempferol-3-O- rhamnoside	C ₂₁ H ₁₉ O ₁₀	12.30	431.0978	431.0987	-2,1	255.0299; 285.0420 (kaempferol), 227.0344	283; 368; 379
31	Unknow	C ₁₅ H ₂₁ O ₈ N ₆	13.05	413.1501	413.1490	2,7	---	275
32	Trihydroxyhexaoxohexacosanoic acid	C ₂₆ H ₃₉ O ₁₁	13.29	527.2492	527.2499	-1,3	509.2393	225; 284
33	Unknow	C ₂₅ H ₃₇ O ₁₀	16.26	497.2387	497.2393	-1,2	479.2284; 327.2177	283
34	Dihydroxyhexaoxohexacosanoic acid	C ₂₆ H ₃₉ O ₁₀	18.21	511.2543	511.2548	-1,0	493.2444; 417.2129; 311.1686; 183.1025	283
35	2,5,8,11,14,17-Hexaoxonadecane- 3,19-diol	C ₁₃ H ₂₇ O ₈	18.60	311.1706	311.1688	5,8	251.1481	283
36	Andropanoside	C ₂₆ H ₃₉ O ₉	19.85	495.2594	495.2602	-1,6	333.2075; 317.2134; 399.2376	282; 328
37	Gingerol	C ₁₇ H ₂₅ O ₄	20.57	293.1753	293.1760	-2,4	279.1636; 193.0863; 151.0756	280; 326
38	Phenethyl butyrate	C ₁₂ H ₁₅ O ₂	20.78	191.1072	191.1075	-1,6	---	278
39	Methoxy heptaetilenglicol	C ₁₅ H ₃₁ O ₈	21.28	339.2019	339.2002	5,0	309,2070, 325,1844	276
40	Pentadecatetraenoic acid	C ₁₅ H ₂₁ O ₂	23.38	233.1542	233.1546	-1,7	---	278

* Daughter ions

Compounds identified in *Pithecellobium dulce*

Glycosylated flavonoids.

Five glycosylated flavonoids corresponding to peaks 24, 25, 26, 27, and 30 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 24 with a [M-H]-ion m/z 597.1823 and daughter ions at m/z 477.1406, 435.1284 and 507.1497 were identified as phloretin-di-C-hexoside [1]. Peak 25 with a [M-H]-ion m/z 463.0886 and daughter ions at m/z 179.0556, 163.0610 and 151.0395 were identified as quercetin-3-glucoside [2]. Peak 26 with a [M-H]-ion m/z 463.0886 and daughter ions at m/z 593.1510 and daughter ions at m/z 117.0380, 533.1301, 159.0295 and 163.0401 were identified as vicenin-2 [3]. Peak 27 with a [M-H]-ion m/z 447.0936 and daughter ions at m/z 151.0031, 255.0301 and 107.0131 were identified as luteolin 7-O-glucoside [4]. Peak 30 with a [M-H]-ion m/z 431.0987 and daughter ions at m/z 255.0299, 285.0420 (aglycone kaempferol), 227.0344, and 227.0344 were identified as kaempferol-3-O-rhamnoside [5].

Flavonoids.

One flavonoid we identified using UHPLC-Q/Orbitrap/MS/MS. Peak 22 with a [M-H]-ion m/z 609.1457 and daughter ions at m/z 300.0280, 137.0248, and 149.0238 were identified as rutin [6].

Fatty acids

Eleven fatty acids corresponding to peaks 10, 13, 14, 15, 16, 17, 21, 28, 32, 34 and 40 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 10 with a [M-H]-ion m/z 257.1394 and daughter ion at m/z 213.1493 were identified as 7-oxo-tridecanedioic acid. Peak 13 with a [M-H]-ion m/z 255.1238 and daughter ions at m/z 211.1337 and 153.0916 were identified as hidroxydioxotridecenoic acid. Peak 14 with a [M-H]-ion m/z 511.2550 and daughter ions at m/z 465.1440, 323.1114, 255.1238, 153.0914 and 211.1337 were identified as dihydroxyhexaoxohexacosanoic acid. Peak 15 with a [M-H]-ion m/z 431.1926 and daughter ions at m/z 401.1461 and 387.2020 were identified as tetrahydroxitetraoxoicosanoic acid. Peak 16 with a [M-H]-ion m/z 381.1770 and daughter ion at m/z 351.1301 were identified as heptahydroxyoxohexadecanoic acid. Peak 17 with a [M-H]-ion m/z 433.2083 and daughter ion at m/z 387.2028 were identified as tetrahydroxytryoxoicosanoic acid. Peak 21 with a [M-H]-ion m/z 431.1926 and daughter ions at m/z 389.2185 and 417.2127 were identified as tetrahydroxytetraoxoicosanoic acid. Peak 28 with a [M-H]-ion m/z 449.1490 we identified as tridecahydroxypentadecanoic acid. Peak 32 with a [M-H]-ion m/z 527.2499 and daughter ion at m/z 509.2393 were identified as trihydroxyhexaoxohexacosanoic acid. Peak 34 with a [M-H]-ion m/z 511.2548 and daughter ions at m/z 493.2444, 417.2129, 311.1686, and 183.1025 were identified as dihydroxyhexaoxohexacosanoic acid. Peak 40 with a [M-H]-ion m/z 233.1546 we identified as pentadecatetraenoic acid.

Organooxygen compounds (Carbohydrates, carbohydrate conjugates, alcohols and polyols)

Six organooxygen compounds corresponding to peaks 1, 9, 12, 19, 23, 29 and 35 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 1 with a [M-H]-ion m/z 165.0400 and daughter ions at m/z 147.0295 and 119.0343 were identified as arabinonic acid. Peak 9 with a [M-H]-ion m/z 337.0931 and daughter ions at m/z 163.0396, 119.0495, 191.0557 and 173.0449 were identified as coumaroylquinic acid [7]. Peak 12 with a [M-H]-ion m/z 449.1669 and daughter ions at m/z 355.1038 were identified as ferulic acid-O-hexoside derivative [8]. Peak 19 with a [M-H]-ion m/z 461.1666 and daughter ions at m/z 445.1718 and 447.1521 were identified as verbasoside [9]. Peak 23 with a [M-H]-ion m/z 433.1719 and daughter ions at m/z 375.1665, 179.0557, 255.1236 and 417.1762 were identified as ethyl-morroneiside [10]. Peak 29 with a [M-H]-ion m/z 329.1609 and daughter ions at m/z 299.1502, 179.0555, 167.1078 and 163.0607 were identified as picrocrocin [11]. Peak 35 with a [M-H]-ion m/z 311.1688 and daughter ion at m/z 251.1481 were identified as 2,5,8,11,14,17-Hexaoxonadecane-3,19-diol.

Terpene glycosides

Two terpene glycosides compounds corresponding to peaks 20 and 36 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 20 with a [M-H]-ion m/z 345.1559 and daughter ions at m/z 327.1452, 163.0607, 183.1024, 301.1669, 289.1657 and 206.0820 were identified as nepetaside [12]. Peak 36 with a [M-H]-ion m/z 495.2602 and daughter ions at m/z 333.2075, 317.2134 and 399.2376 were identified as andropanoside [13].

Hydroxy acids and derivatives

One hydroxy acid compound corresponding to peak 2 we identified using UHPLC-Q/Orbitrap/MS/MS. Peak 2 with a [M-H]-ion m/z 179.0558 and daughter ions at m/z 143.0350 and 119.0341 were identified as 3-Deoxyhexonic acid.

Carboxylic acids and derivatives

One carboxylic acid compound corresponding to peak 3 we identified using UHPLC-Q/Orbitrap/MS/MS. Peak 3 with a [M-H]-ion m/z 191.0195 and daughter ion at m/z 111.0080 were identified as citric acid [14].

Stilbenes

One stilbene compound corresponding to peak 4 we identified using UHPLC-Q/Orbitrap/MS/MS. Peak 4 with a [M-H]-ion m/z 405.1172 and daughter ion at m/z 243.0624, 179.0557, 227.0672 and 163.0608 were identified as 2,3,5,4'-Tetrahydroxystilbene 2-O- β -D-glucoside [15].

Benzene and substituted derivatives

One benzene compound corresponding to peak 5 we identified using UHPLC-Q/Orbitrap/MS/MS. Peak 5 with a [M-H]-ion m/z 315.0723 and daughter ion at m/z 153.0221, 152.0165 and 137.0237 were identified as protocatechuic acid 4-hexoside (). Peak 38 with a [M-H]-ion m/z 191.1075 we identified as phenethyl butyrate [16].

Double esters

One double ester compound corresponding to peak we identified using UHPLC-Q/Orbitrap/MS/MS. Peak 7 with a [M-H]-ion m/z 419.1926 and daughter ion at m/z 355.0672 were identified as bis(ethoxycarbonyloxymethyl) undecanedioate.

Carbohydrates and carbohydrate-derivatives

One carbohydrate compound corresponding to peak 7 we identified using UHPLC-Q/Orbitrap/MS/MS. Peak 7 with a [M-H]-ion m/z 451.2188 and daughter ion at m/z 341.1093 and 179.0557 were identified as calaliukiuenoside [17].

Terpenoids

One terpenoid compound corresponding to peak 8 we identified using UHPLC-Q/Orbitrap/MS/MS. Peak 7 with a [M-H]-ion m/z 443.1925 and daughter ion at m/z 179.0555 were identified as ebuloside [18].

Phenols

One phenol compound corresponding to peak 37 we identified using UHPLC-Q/Orbitrap/MS/MS. Peak 37 with a [M-H]-ion m/z 293.1760 and daughter ions at m/z 279.1636, 193.0863 and 151.0756 were identified as gingerol [19].

Glycol alcohols

One glycol compound corresponding to peak 39 we identified using UHPLC-Q/Orbitrap/MS/MS. Peak 39 with a [M-H]⁻ ion m/z 339.2002 and daughter ions at m/z 309.2070 and 325.1844 were identified as methoxy heptaetilenglicol.

Unknown compounds

Peaks 11, 18, 31 and 33 with a [M-H]⁻ ions m/z 364.1617, 363.0759, 413.1490, and 497.2393 respectively correspond to unknown compounds.

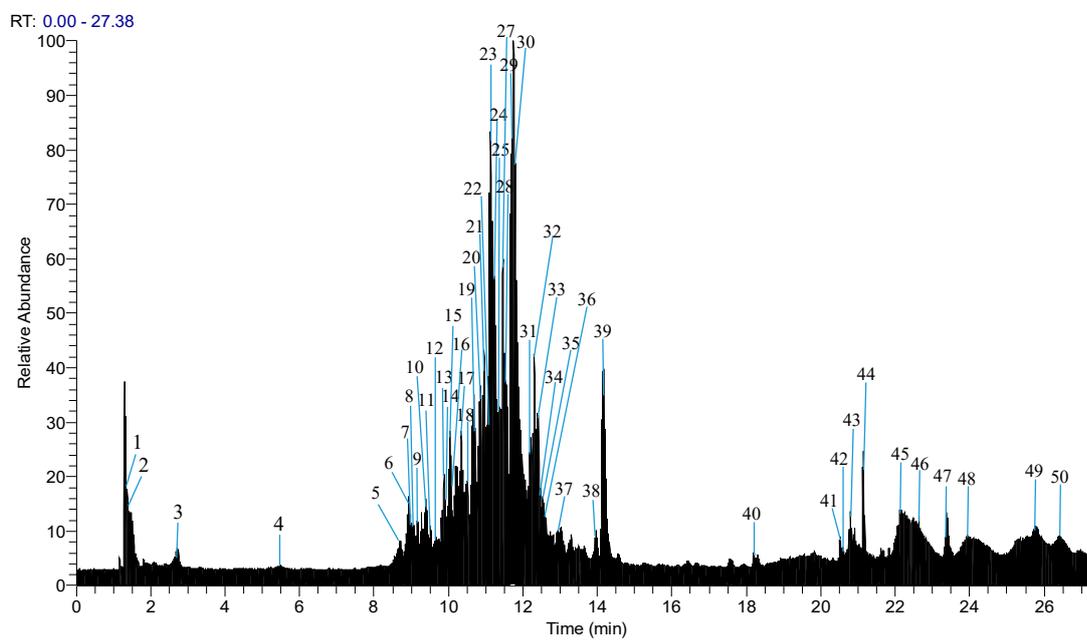


Figure S2. UHPLC TIC (total ion current) chromatogram of *Leucaena leucocephala* ethyl acetate extract partition.

Table S2. Tentative identification of compounds in the ethyl acetate extract partition of *Leucaena leucocephala* by UHPLC-Q/Orbitrap/MS/ MS

Peak	Tentative identification	[M-H] ⁻	Retention time (min.)	Theoretical mass (m/z)	Measured mass (m/z)	Accuracy (ppm)	MS ions m/z)*	UV
1	Quinic acid	C ₇ H ₁₁ O ₆	1.34	191.0556	191.0560	-2,1	173.0452; 117.0189	269
2	Syringaldehyde Syringate O derivate quinic acid.	C ₁₈ H ₁₇ O ₉	1.36	377.0873	377.0861	3,2	347.0755; 349.0911	203; 269
3	Gallic acid	C ₇ H ₅ O ₅	2.70	169.0137	169.0139	-1,2	125.0238	212; 271
4	Gallocatechin isomer	C ₁₅ H ₁₃ O ₇	5.49	305.0661	305.0669	-2,6	153.0188; 109.0287	202; 263; 283
5	Hydroxytrioxopentadecenoic acid	C ₁₅ H ₂₁ O ₆	8.72	297.1338	297.1347	-3,0	233.1446; 125.0236; 177.0201	234; 268
6	Pentahydroxitetraoxoicosanoic acid	C ₂₀ H ₃₁ O ₁₁	8.92	447.1866	447.1875	-2,0	437.1589; 401.1821	235
7	Hydroxytrioxotetradecanoic acid	C ₁₄ H ₂₁ O ₆	9.01	285.1338	285.1346	-2,8	257.1037	236
8	Gallocatechin	C ₁₅ H ₁₃ O ₇	9.18	305.0661	305.0670	-3,0	125.0238; 137.0238; 109.0287; 139.0394; 179.0344; 167.0345	207; 227; 274
9	Digalloyl glucose	C ₂₀ H ₁₉ O ₁₄	9.29	483.0775	483.0783	-1,7	125.0237; 169.0139; 211.0242; 271.0459; 313.0553	222; 278
10	Caffeoylquinic acid	C ₁₆ H ₁₇ O ₉	9.41	353.0873	353.0882	-2,5	191.0559; 161.0239; 179.0344; 135.0444	234; 286; 325
11	Gallic acid derivative I	C ₁₇ H ₂₃ O ₁₁	9.52	403.1240	403.1251	-2,7	125.0238; 177.0177; 109.0288	232; 279
12	Myrsinoside A	C ₂₀ H ₂₁ O ₁₂	9.67	453.1033	453.1043	-2,2	123.0443; 125.0237; 167.0345; 135.0446; 313.0570	232; 279
13	Trihydroxypentaoxooctadecanoic acid	C ₁₈ H ₂₅ O ₁₀	9.89	401.1448	401.1458	-2,5	343.1398	232; 278
14	Unknow	C ₂₄ H ₂₁ O ₈	9.92	437.1236	437.1224	2,7	401.1457; 179.0346	230; 279; 268
15	Tetrahydroxytetraoxoicosanoic acid	C ₂₀ H ₃₁ O ₁₀	10.05	431.1917	431.1925	-1,9	385.1886;	232; 275
16	Catechin	C ₁₅ H ₁₃ O ₆	10.23	289.0712	289.0720	-2,8	125.0237; 137.0238; 151.0395; 165.0553; 271.0610	203; 227; 279
17	Gallocatechin gallate	C ₂₂ H ₁₇ O ₁₁	10.34	457.0771	457.0780	-2,0	125.0237; 169.0135; 335.0775; 289.0718;; 137.0238; 109.0287	207; 276; 368
18	Isorhamnetin -O-glucoside	C ₂₂ H ₂₁ O ₁₂	10.50	477.1033	477.1043	-2,1	151.0025; 163.0396; 300.0284	228; 274; 354
19	Myricetin-3-O-hexoside	C ₂₁ H ₁₉ O ₁₃	10.70	479.0826	479.0833	-1,5	125.0235; 287.0190; 151.0033; 316.0230 (myricetin)	208; 225; 264; 354
20	Myricetin 3-arabinoside	C ₂₀ H ₁₇ O ₁₂	10.85	449.0720	449.0729	-2,0	125.0239; 271.0246; 316.0220 (myricetin)	209; 226; 265; 354
21	Rutin	C ₂₇ H ₂₉ O ₁₆	10.96	609.1499	609.1459	6,6	151.0032; 300.0278; 137.0238	203; 257; 354
22	Myricetin-3-arabinoside isomer	C ₂₀ H ₁₇ O ₁₂	11.02	449.0720	449.0730	-2,2	125.0238; 271.0247; 300.0286; 301.0359	201; 228; 269; 354
23	Myricetin 3-O-rhamnoside (myricitrin)	C ₂₁ H ₁₉ O ₁₂	11.13	463.0877	463.0884	-1,5	271.0249; 316.0226; 178.9982; 151.0031;	208; 262; 351
24	Quercetin-3-O-glucuronide	C ₂₁ H ₁₇ O ₁₃	11.26	477.0669	477.0677	-1,7	301.0355; 151.0031; 285.0404; 177.0192; 125.0237	203; 257; 290; 353
25	Epicatechin gallate	C ₂₂ H ₁₇ O ₁₀	11.36	441.0822	441.0831	-2,0	125.0237; 289.0718; 137.0239;	202; 224; 272; 337

26	Kaempferol-3-O-glucorhamnoside	C ₂₇ H ₂₉ O ₁₅	11.39	593.1506	593.1509	-0,5	285.0403; 125.0237; 255.0302	202; 224; 270; 341
27	Quercetin 3-O-arabioside	C ₂₀ H ₁₇ O ₁₁	11.46	433.0771	433.0780	-2,1	151.0031; 271.0246; 178.9982; 300.0277; 301.0356 (aglycone quercetin)	202; 257; 286; 354
28	Cyanidin-3-arabioside	C ₂₀ H ₁₉ O ₁₀	11.52	419.0978	419.0981	-0,7	285.0414	202; 225; 267; 353
29	Quercetin 3-O-pentoside	C ₂₀ H ₁₇ O ₁₁	11.68	433.0771	433.0779	-1,8	151.0031; 178.9982; 271.0247; 301.0351 (aglycone quercetin); 300.0275	202; 257; 353
30	Quercetin 3-O-Rhamnoside	C ₂₁ H ₁₉ O ₁₁	11.76	447.0927	447.0935	-1,8	151.0031; 178.9982; 255.0296; 300.0276; 301.0351 (aglycone quercetin)	210; 257; 349
31	Kaempferol-3-O-pentoside	C ₂₀ H ₁₇ O ₁₀	12.21	417.0822	417.0832	-2,4	255.0297; 285.0402 (aglycone kaempferol)	228; 270; 337
32	Luteolin-7- rhamnoside	C ₂₁ H ₁₉ O ₁₀	12.31	431.0978	431.0988	-2,3	255.0298; 285.0399 (aglycone luteolin); 109.0286	227; 266
33	Myricetin	C ₁₅ H ₉ O ₈	12.41	317.0297	317.0305	-2,5	151.0031; 137.0237;	226; 270; 368
34	Kaempferol dimethylether Hexoside	C ₂₂ H ₁₉ O ₁₂	12.54	475.0877	475.0886	-1,9	125.0237; 151.0034; 137.0237; 461.0713	272; 367
35	Quercetin galloyl pentoside	C ₂₇ H ₂₁ O ₁₅	12.58	585.0880	585.0886	-1,0	301.0356; 151.0033; 433.0781	271; 268
36	Cinchonain I	C ₂₄ H ₁₉ O ₉	12.65	451.1029	451.1041	-2,7	341.0677; 315.0885; 287.0563; 217.0138	271; 368
37	Apigenin 7-O-glucuronide	C ₂₁ H ₁₇ O ₁₁	12.92	445.0771	445.0782	-2,5	225.0556; 269.0453; 175.0242	273; 368
38	Luteolin	C ₁₅ H ₉ O ₆	13.96	285.0399	285.0408	-3,2	133.0287	195; 221; 284
39	Quercetin	C ₁₅ H ₉ O ₇	14.17	301.0348	301.0356	-2,7	107.0131; 151.0031	195; 220; 284; 368
40	Quercetin pentoside derivate	C ₂₄ H ₂₃ O ₁₁	18.20	487.1240	487.1248	-1,6	255.0293; 271.0255; 300.0276 (aglycone quercetin)	282
41	Gingerol	C ₁₇ H ₂₅ O ₄	20.59	293.1753	293.1761	-2,7	279.1637	278
42	2,5,8,11,14,17-Hexaoxonadecane-3,19- diol	C ₁₃ H ₂₇ O ₈	20.61	311.1706	311.1690	5,1	251.1481	278
43	Phenethyl butyrate	C ₁₂ H ₁₅ O ₂	20.80	191.1072	191.1075	-1,6	121.0653	278
44	Unknow	C ₃₀ H ₅₃ O ₁₄ N ₆	21.13	721.3620	721.3639	-2,6	711.3353;	278; 325
45	Unknow	C ₂₈ H ₄₃ O ₁₁	22.17	555.2805	555.2846	-7,4	---	275
46	Octahydroxyheptacosapentaenoic acid	C ₂₇ H ₄₃ O ₁₀	22.64	527.2856	527.2864	-1,5	---	275
47	Pentadecatetraenoic acid	C ₁₅ H ₂₁ O ₂	23.38	233.1542	233.1547	-2,1	183.0119	278
48	Heptaethylene glycol	C ₁₄ H ₂₉ O ₈	23.94	325.1862	325.1847	4,6	309.1744; 251.1482	275
49	Unknow	C ₃₀ H ₄₁ O ₁₁	25.82	577.2649	577.2690	-7,1	---	275
50	Dihydroxypentadecatetraenoic acid	C ₁₅ H ₂₁ O ₄	26.39	265.1440	265.1482	-15,8	249.1494; 223.0968	275

* Daughter ions

Compounds identified in *Leucaena leucocephala*

Flavonoid glycosides

Sixteen glycosylated flavonoids corresponding to peaks 18, 19, 20, 21, 22, 23, 24, 26, 27, 28, 29, 30, 31, 32, 34 and 40 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 18 with a [M-H]-ion m/z 477.1043 and daughter ions at m/z 151.0025, 163.0396 and 300.0284 were identified as isorhamnetin-O-glucoside [20]. Peak 19 with a [M-H]-ion m/z 479.0833 and daughter ions at m/z 125.0235, 287.0190, 151.0033 and 316.0230 (aglycone myricetin) were identified as myricetin-3-O-hexoside [21]. Peak 20 with a [M-H]-ion m/z 449.0729 and daughter ions at m/z 125.0239, 271.0246, 316.0220 (aglycone myricetin) were identified as myricetin 3-arabinoside [22]. Peak 21 with a [M-H]-ion m/z 609.1459 and daughter ions at m/z 151.0032, 300.0278 and 137.0238 were identified as rutin [23]. Peak 22 with a [M-H]-ion m/z 449.0730 and daughter ions at m/z 125.0238, 271.0247, 300.0286 and 301.0359 were identified as myricetin-3-arabinoside isomer [22]. Peak 23 with a [M-H]-ion m/z 463.0884 and daughter ions at m/z 271.0249, 316.0226, 178.9982 and 151.0031 were identified as myricetin 3-O-rhamnoside (myricitrin) [24]. Peak 24 with a [M-H]-ion m/z 477.0677 and daughter ions at m/z 301.0355, 151.0031, 285.0404, 177.0192 and 125.0237 were identified as quercetin-3-O-glucuronide [25]. Peak 26 with a [M-H]-ion m/z 593.1509 and daughter ions at m/z 285.0403, 125.0237 and 255.0302 were identified as kaempferol-3-O-glucorhamnoside [26]. Peak 27 with a [M-H]-ion m/z 433.0780 and daughter ions at m/z 151.0031, 271.0246, 178.9982, 300.0277 and 301.0356 (aglycone quercetin) were identified as quercetin 3-O-arabinoside [27]. Peak 28 with a [M-H]-ion m/z 419.0981 and daughter ion at m/z 285.0414 were identified as cyanidin-3-arabinoside [28]. Peak 29 with a [M-H]-ion m/z 433.0779 and daughter ions at m/z 151.0031, 178.9982, 271.0247, 301.0351 (aglycone quercetin) and 300.0275 were identified as quercetin 3-O-pentoside [29]. Peak 30 with a [M-H]-ion m/z 447.0935 and daughter ions at m/z 151.0031, 178.9982, 255.0296, 300.0276 and 301.0351 (aglycone quercetin) were identified as quercetin 3-O-rhamnoside [30]. Peak 31 with a [M-H]-ion m/z 417.0832 and daughter ions at m/z 255.0297 and 285.0402 (aglycone kaempferol) were identified as kaempferol-3-O-pentoside [31]. Peak 32 with a [M-H]-ion m/z 431.0988 and daughter ions at m/z 255.0298, 285.0399 (aglycone luteolin) and 109.0286 and were identified as luteolin-7-rhamnoside [32]. Peak 34 with a [M-H]-ion m/z 475.0886 and daughter ions at m/z 125.0237; 151.0034, 137.0237 and 461.0713 were identified as kaempferol dimethylether hexoside [33]. Peak 40 with a [M-H]-ion m/z 487.1248 and daughter ions at m/z 255.0293, 271.0255 and 300.0276 (aglycone quercetin) were identified as quercetin pentoside derivative.

Flavonoids

Nine flavonoids corresponding to peaks 4, 8, 16, 17, 25, 33, 36, 38 and 39 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 4 with a [M-H]-ion m/z 305.0669 and daughter ions at m/z 153.0188 and 109.0287 were identified as gallicocatechin isomer [34]. Peak 8 with a [M-H]-ion m/z 305.0670 and daughter ions at m/z 125.0238, 137.0238, 109.0287, 139.0394, 179.0344 and 167.0345 were identified as gallicocatechin [35]. Peak 16 with a [M-H]-ion m/z 289.0720 and daughter ions at m/z 125.0237, 137.0238, 151.0395, 165.0553, 271.0610 were identified as catechin [36]. Peak 17 with a [M-H]-ion m/z 457.0780 and daughter ions at m/z 125.0237, 169.0135, 335.0775, 289.0718, 137.0238 and 109.0287 were identified as gallicocatechin gallate [34]. Peak 25 with a [M-H]-ion m/z 441.0831 and daughter ions at m/z 125.0237, 289.0718 and 137.0239 were identified as epicatechin gallate [34]. Peak 33 with a [M-H]-ion m/z 317.0305 and daughter ions at m/z 151.0031 and 137.0237 were identified as myricetin [37]. Peak 36 with a [M-H]-ion m/z 451.1041 and daughter ions at m/z 341.0677, 315.0885, 287.0563 and 217.0138 were identified as cinchonain I [38]. Peak 38 with a [M-H]-ion m/z 285.0408 and daughter ion at m/z 133.0287 were identified as luteolin [39]. Peak 39 with a [M-H]-ion m/z 301.0356 and daughter ion at m/z 107.0131 and 151.0031 were quercetin [40].

Fatty acids

Eight fatty acids corresponding to peaks 5, 6, 7, 13, 15, 46, 47 and 50 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 5 with a [M-H]-ion m/z 297.1347 and daughter ions at m/z 233.1446, 125.0236 and 177.0201 were identified as hydroxytrioxopentadecenoic acid. Peak 6 with a [M-H]-ion m/z 447.1875 and

daughter ions at m/z 437.1589 and 401.1821 were identified as pentahydroxitetraoxoicosanoic acid. Peak 7 with a [M-H]-ion m/z 285.1346 and daughter ion at m/z 257.1037 were identified as hydroxytrioxotetradecanoic acid. Peak 13 with a [M-H]-ion m/z 401.1458 and daughter ion at m/z 343.1398 were identified as trihydroxypentaooctadecanoic acid. Peak 15 with a [M-H]-ion m/z 431.1925 and daughter ion at m/z 385.1886 were identified as tetrahydroxytetraoxoicosanoic acid. Peak 46 with a [M-H]-ion m/z 527.2864 we identified as octahydroxyheptacosapentaenoic acid. Peak 47 with a [M-H]-ion m/z 233.1547 and daughter ion at m/z 183.0119 were identified as pentadecatetraenoic acid. Peak 50 with a [M-H]-ion m/z 265.1482 and daughter ions at m/z 249.1494 and 223.0968 were identified as dihydroxypentadecatetraenoic acid.

Organooxygen compounds

Three organooxygen compounds corresponding to peaks 1, 10 and 48 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 1 with a [M-H]-ion m/z 191.0560 and daughter ions at m/z 173.0452 and 117.0189 were identified as quinic acid [41]. Peak 10 with a [M-H]-ion m/z 353.0882 and daughter ions at m/z 191.0559, 161.0239, 179.0344 and 135.0444 were identified as caffeoylquinic acid [42]. Peak 48 with a [M-H]-ion m/z 325.1847 and daughter ions at m/z 309.1744, and 251.1482 were identified as heptaethylene glycol.

Benzene and substituted derivatives

Three benzene and substituted derivatives compounds corresponding to peaks 3, 11 and 43 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 3 with a [M-H]-ion m/z 169.0139 and daughter ion at m/z 125.0238 were identified as gallic acid [43]. Peak 11 with a [M-H]-ion m/z 403.1251 and daughter ions at m/z 125.0238, 177.0177 and 109.0288 were identified as gallic acid derivative I . Peak 43 with a [M-H]-ion m/z 191.1075 and daughter ion at m/z 121.0653 were identified as phenethyl butyrate [16].

Organic acids and related compounds

One organic acid corresponding to peak 2 was identified using UHPLC-Q/Orbitrap/MS/MS. Peak 3 with a [M-H]-ion m/z 377.0861 and daughter ions at m/z 347.0755 and 349.0911 were identified as syringaldehyde syringate or derivate quinic acid [44].

Tannins

One organic acid corresponding to peak 9 was identified using UHPLC-Q/Orbitrap/MS/MS. Peak 9 with a [M-H]-ion m/z 483.0783 and daughter ions at m/z 125.0237, 169.0139, 211.0242, 271.0459 and 313.0553 were identified as digalloyl glucose [45].

Phenolic glycosides

One organic acid corresponding to peak 12 was identified using UHPLC-Q/Orbitrap/MS/MS. Peak 12 with a [M-H]-ion m/z 453.1043 and daughter ions at m/z 123.0443, 125.0237, 167.0345, 135.0446 and 313.0570 were identified as myrsinoside A[46].

Galloylated flavonoid glycoside

One galloylated flavonoid glycoside compound corresponding to peak 35 was identified using UHPLC-Q/Orbitrap/MS/MS. Peak 35 with a [M-H]-ion m/z 585.0886 and daughter ions at m/z 301.0356, 151.0033 and 433.0781 were identified as quercetin galloyl pentoside [47].

Flavonoid-7-o-glucuronides

One flavonoid-7-o-glucuronide compound corresponding to peak 35 was identified using UHPLC-Q/Orbitrap/MS/MS. Peak 37 with a [M-H]-ion m/z 445.0782 and daughter ions at m/z 225.0556, 269.0453 and 175.0242 were identified as apigenin 7-O-glucuronide [48].

Phenols

One phenol compound corresponding to peak 41 was identified using UHPLC-Q/Orbitrap/MS/MS. Peak 41 with a [M-H]-ion m/z 293.1761 and daughter ion at m/z 279.1637 were identified as gingerol [49].

Unknown compounds.

Peaks 14, 44, 45 and 49 with a [M-H] -ions m/z 437.1224, 721.3639, 555.2846 and 577.2690 respectively correspond to unknown compounds.

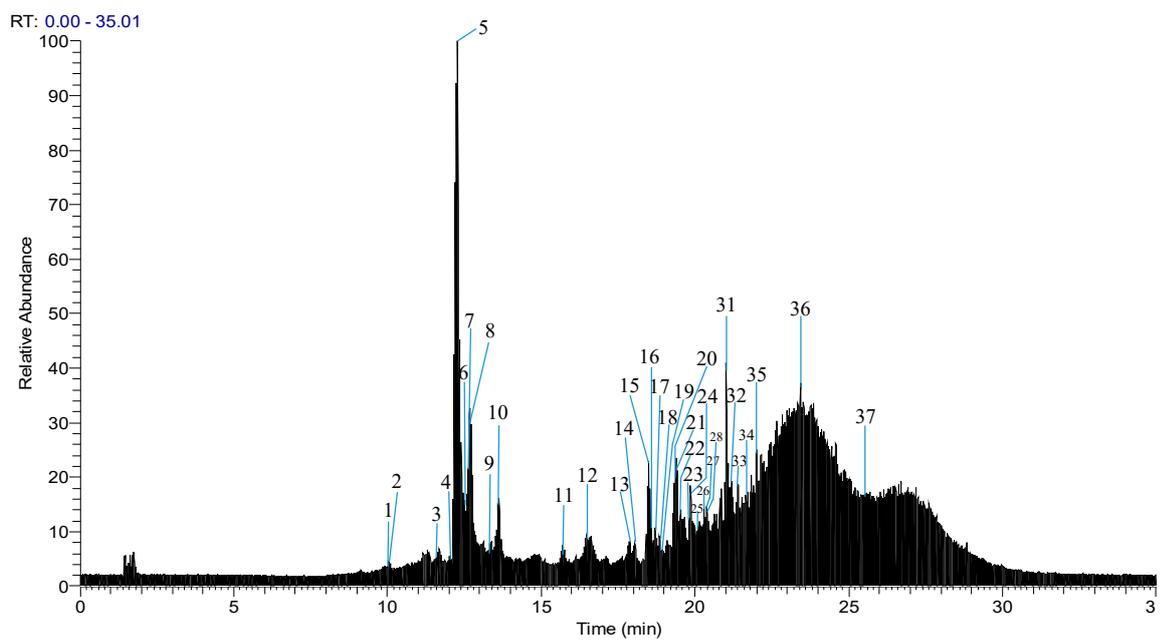


Figure S3. UHPLC TIC (total ion current) chromatogram of *Gliricidia sepium* dichloromethane extract partition.

Table S3. Tentative identification of compounds in the dichloromethane extract partition of *Gliricidia sepium* by UHPLC-Q/Orbitrap/MS/ MS

Peak	Tentative identification	[M-H] ⁻	Retention time (min.)	Theoretical mass (m/z)	Measured mass (m/z)	Accuracy (ppm)	MS ions (m/z)*	UV
1	Esculetin	C ₉ H ₅ O ₄	10.05	177.0188	177.0184	2,3	133.0282; 149.0234	260; 290
2	Caffeic acid	C ₉ H ₇ O ₄	11.15	179.0344	179.0341	1,7	135.0440	246; 291
3	Dihydro-p-coumaric acid isomer	C ₉ H ₉ O ₃	11.52	165.0552	165.0547	3,0	147.0141; 119.0490	228; 273
4	Hydroxycoumarin	C ₉ H ₅ O ₃	12.05	161.0239	161.0234	3,1	133.0283; 117.0335	244; 277
5	Dihydro-p-coumaric acid	C ₉ H ₉ O ₃	12.26	165.0552	165.0546	3,6	121.0648; 106.0412; 147.0440; 119.0490	228; 273
6	Azelaic acid	C ₉ H ₁₅ O ₄	12.49	187.0970	187.0966	2,1	121.0654; 141.0914; 143.1073; 123.0808; 171.1020	246; 267
7	Derivative Tianshic Acid	C ₁₂ H ₁₉ O ₅	12.60	243.1232	243.1228	1,6	225.1122; 199.1328	233; 276; 325
8	p-coumaric acid	C ₉ H ₇ O ₃	12.66	163.0395	163.0391	2,5	147.0440; 119.0492	233; 274; 325
9	Dihydroxydioxooctadecanoic acid	C ₁₈ H ₃₁ O ₆	13.34	343.2121	343.2115	1,7	327.2166	249
10	N-Carbobenzyloxy-L-isoleucine	C ₁₄ H ₁₈ O ₄ N	13.62	264.1236	264.1232	1,5	220.1333; 246.1129	232; 275; 308
11	Leu/dihydro-p-coumaric acid	C ₁₅ H ₂₀ NO ₄	15.70	278.1392	278.1388	1,4	130,0866 (leucine), 119,0495, 121,0651, 165,0552 (dihydro-p-coumaric acid)	247; 266; 329
12	Trihydroxyoctadecadienoic acid	C ₁₈ H ₃₁ O ₅	16.46	327.2171	327.2168	0,9	309.2057; 169.0856; 187.0962	228; 273
13	Phe /Dihydro-p-coumaric acid	C ₁₈ H ₁₈ NO ₄	17.88	312.1236	312.1232	1,3	147.0441; 119.0490; 121.0562; 164.0706 (phenylalanine)	247; 271; 329
14	Dihydroxy-methoxyflavanone	C ₁₆ H ₁₃ O ₅	18.03	285.0763	285.0760	1,1	109.0283; 165.0546; 271.0605; 147.0440; 177.0544	249; 272; 379
15	Dihydroxydodecadienoic acid	C ₁₂ H ₁₉ O ₄	18.50	227.1283	227.1279	1,8	183.1381; 165.1279; 209.1172	249
16	Tianshic acid	C ₁₈ H ₃₃ O ₅	18.61	329.2328	329.2324	1,2	227.1279; 167.1067; 197.1173; 215.1279; 243.1229; 311.2229	248; 273;
17	Dihydro-p-coumaric acid derivative I	C ₂₀ H ₂₂ NO ₅	18.69	356.1498	356.1494	1,1	121.0646; 147.0440; 119.0490; 165.0547; (dihydro-p-coumaric acid); 208.0969	249; 273;
18	Derivative tianshic acid	C ₁₈ H ₃₁ O ₅	18.85	327.2171	327.2167	1,2	301.2010; 227.1279; 197.1173; 215.1279; 243.1229; 167.1067; 311.2229	248; 273
19	Hesperetin	C ₁₆ H ₁₃ O ₆	19.00	301.0712	301.0709	1,0	165.0547; 273.0761; 153.0183; 135.0076	249; 274
20	Dihydro-p-coumaric acid isomer	C ₉ H ₉ O ₃	19.34	165.0552	165.0546	3,6	121.0646; 106.0411; 147.0439; 119.0491	229; 273
21	Tianshic acid	C ₁₈ H ₃₃ O ₅	19.41	329.2328	329.2324	1,2	227.1279; 197.1174; 215.1281; 243.1228; 311.2221	228; 237

22	Dihydroxy-methoxyflavanone derivative I	C ₂₁ H ₂₅ O ₆	19.55	373.1651	373.1646	1,3	121.0647; 147.0439; 165.0543; 285.0768	274
23	Unknow	C ₁₂ H ₁₅ O ₅	19.65	239.0919	239.0916	1,3	---	275
24	Dihydroxy-methoxyflavanone derivative II	C ₂₁ H ₂₇ O ₆	19.86	375.1808	375.1802	1,6	121.0647; 147.0439; 165.0544	283; 311
25	Hydroxy-dimethoxyisoflavan	C ₁₇ H ₁₇ O ₄	20.12	285.1127	285.1122	1,8	147.0441; 107.0490	243; 267
26	Dihydro-p-coumaric acid derivative II	C ₂₉ H ₃₁ O ₈	20.29	507.2019	507.2007	2,4	121.0646; 165.0547; (dihydro- p-coumaric acid) 147.0440; 119.0490;	247; 279
27	Hydroxydioheptadecenoic acid	C ₁₇ H ₂₇ O ₅	20.37	311.1858	311.1855	1,0	193.0863, 195.1021, 197.1169, 237.1122, 239.1278, 267,1236	249; 278
28	Dihydro-p-coumaroyloctanoic acid	C ₁₇ H ₂₃ O ₅	20.40	307.1545	307.1542	1,0	121.0446; 147.0441; 159.1016; 165.0545 (dihydro-p-coumaric acid)	251; 274
29	Dihydro-p-coumaroyl-trioxooctadecanoic acid	C ₂₇ H ₃₉ O ₇	20.84	475.2696	475.2684	2,5	121.0648; 165.0544 (dihydro- p-coumaric acid); 147. 0439; 311.2217	248; 274
30	Phenethyl butyrate	C ₁₂ H ₁₅ O ₂	20.92	191.1072	191.1067	2,6	121.0647;	252; 275
31	Dihydro-p-coumaroylnonanoic acid	C ₁₈ H ₂₅ O ₅	21.02	321.1702	321.1696	1,9	151.0753; 121.0647; 165.0545 (dihydro-p-coumaric acid); 173.1171	274
32	Trihydroxyheptaotriacontadienoic acid	C ₃₀ H ₄₅ O ₁₂	21.18	597.2919	597.2933	-2,3	----	274
33	Dihydro-p-coumaroyl hydroxytrioxooctadecanoic acid	C ₂₇ H ₄₁ O ₇	21.40	477.2852	477.2840	2,5	121.0647; 165.0545 (dihydro- p-coumaric acid); 147.0442; 327.2166; 329.2321	274
34	3,4 -Dihydroxy cinamic acid octacosyl ester derivated	C ₃₀ H ₄₃ O ₄	21.74	467.3161	467.3152	1,9	121.0647; 165.0545; 147.0441	274
35	Dihydro-p-coumaroylhydroxyxohexadecanoic acid	C ₂₅ H ₃₉ O ₆	22.01	435.2747	435.2737	2,3	401.2321; 339.1986; 287.2215	272
36	3,4 -Dihydroxy cinamic acid octacosyl ester derivated isomer	C ₃₀ H ₄₃ O ₄	23.44	467.3161	467.3149	2,6	441.2267; 325.1830	251; 271
37	Octahydroxyoctacosenoic acid	C ₂₈ H ₅₅ O ₁₀	25.48	551.3795	551.3720	13,6	---	271

* Daughter ions

Compounds identified in *Gliricidia seipium*

Phenylpropanoids

Eleven phenylpropanoids compounds corresponding to peaks 3, 5, 11, 13, 17, 20, 26, 28, 29, 31, 33, and 35 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 3 with a [M-H]-ion m/z 165.0547 and daughter ions at m/z 147.0141 and 119.0490 were identified as dihydro-p-coumaric acid isomer [50]. Peak 5 with a [M-H]-ion m/z 165.0546 and daughter ions at m/z 121.0648, 106.0412, 147.0440 and 119.0490 were identified as dihydro-p-coumaric acid [50]. Peak 11 with a [M-H]-ion m/z 278.1388 and daughter ions at m/z 130.0866 (leucine), 119.0495, 121.0651, 165.0552 (dihydro-p-coumaric acid) were identified as Leu/dihydro-p-coumaric acid. Peak 13 with a [M-H]-ion m/z 312.1232 and daughter ions at m/z 147.0441, 119.0490, 121.0562 and 164.0706 (phenylalanine) were identified as Phe /Dihydro-p-coumaric acid. Peak 17 with a [M-H]-ion m/z 356.1494 and daughter ions at m/z 121.0646, 147.0440, 119.0490, 165.0547, (dihydro-p-coumaric acid) and 208.0969 were identified as dihydro-p-coumaric acid derivative I. Peak 20 with a [M-H]-ion m/z 165.0546 and daughter ions at m/z 121.0646, 106.0411, 147.0439 and 119.0491 were identified as dihydro-p-coumaric acid isomer [50]. Peak 26 with a [M-H]-ion m/z 507.2007 and daughter ions at m/z 121.0646, 165.0547, (dihydro-p-coumaric acid), 147.0440 and 119.0490 were identified as dihydro-p-coumaric acid derivative II. Peak 28 with a [M-H]-ion m/z 307.1542 and daughter ions at m/z 121.0446, 147.0441, 159.1016 and 165.0545 (dihydro-p-coumaric acid) were identified as dihydro-p-coumaroyloctanoic acid. Peak 29 with a [M-H]-ion m/z 475.2684 and daughter ions at m/z 121.0648, 165.0544 (dihydro-p-coumaric acid), 147.0439 and 311.2217 were identified as dihydro-p-coumaroyl-trioxooctadecanoic acid. Peak 31 with a [M-H]-ion m/z 321.1696 and daughter ions at m/z 151.0753, 121.0647, 165.0545 (dihydro-p-coumaric acid) and 173.1171 were identified as dihydro-p-coumaroylnonanoic acid. Peak 33 with a [M-H]-ion m/z 477.2840 and daughter ions at m/z 121.0647, 165.0545 (dihydro-p-coumaric acid), 147.0442, 327.2166 and 329.2321 were identified as dihydro-p-coumaroyl hydroxytrioxooctadecanoic acid. Peak 35 with a [M-H]-ion m/z 435.2737 and daughter ions at m/z 401.2321, 339.1986 and 287.2215 were identified as dihydro-p-coumaroyl hydroxytrioxooctadecanoic acid.

Fatty acids

Seven fatty acid compounds corresponding to peaks 6, 7, 9, 12, 15, 16, 18, 21, 27, 32 and 37 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 6 with a [M-H]-ion m/z 187.0966 and daughter ions at m/z 121.0654, 141.0914, 143.1073, 123.0808 and 171.1020 were identified as azelaic acid [51]. Peak 7 with a [M-H]-ion m/z 243.1228 and daughter ions at m/z 225.1122 and 199.1328 were identified as derivative tianshichic acid. Peak 9 with a [M-H]-ion m/z 343.2115 and daughter ion at m/z 327.2166 were identified as dihydroxydioxooctadecanoic acid [52]. Peak 12 with a [M-H]-ion m/z 327.2168 and daughter ion at m/z 309.2057, 169.0856 and 187.0962 were identified as trihydroxyoctadecadienoic acid [52, 53]. Peak 15 with a [M-H]-ion m/z 227.1279 and daughter ion at m/z 183.1381, 165.1279 and 209.1172 were identified as dihydroxydodecadienoic acid. Peak 16 with a [M-H]-ion m/z 329.2324 and daughter ion at m/z 227.1279, 167.1067, 197.1173, 215.1279, 243.1229 and 311.2229 were identified as tianshichic acid [54]. Peak 18 with a [M-H]-ion m/z 327.2167 and daughter ion at m/z 301.2010, 227.1279, 197.1173, 215.1279, 243.1229, 167.1067 and 311.2229 were identified as derivative tianshichic acid. Peak 21 with a [M-H]-ion m/z 329.2324 and daughter ion at m/z 227.1279, 197.1174, 215.1281, 243.1228 and 311.2221 were identified as tianshichic acid [54]. Peak 27 with a [M-H]-ion m/z 311.1855 and daughter ion at m/z 193.0863, 195.1021, 197.1169, 237.1122, 239.1278 and 267.1236 were identified as hydroxydioxoheptadecenoic acid. Peak 32 with a [M-H]-ion m/z 597.2933 was identified as trihydroxyheptacontadienoic acid. Peak 37 with a [M-H]-ion m/z 551.3720 was identified as octahydroxyoctacosenoic acid.

Cinnamic acids and derivatives

Four cinnamic acid and derivatives compounds corresponding to peaks 2, 8, 34, and 36 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 2 with a [M-H]-ion m/z 179.0341 and daughter ion at m/z 135.0440 were identified as caffeic acid [55]. Peak 2 with a [M-H]-ion m/z 163.0391 and daughter ion at m/z 147.0440 and 119.0492 were identified as p-coumaric acid [56]. Peak 34 with a [M-H]-ion m/z 467.3152 and daughter ions at m/z 121.0647, 165.0545 and 147.0441 were identified as 3,4-dihydroxy cinnamic acid octacosyl ester derivated.

Peak 36 with a [M-H]-ion m/z 467.3149 and daughter ions at m/z 441.2267 and 325.1830 were identified as 3,4-dihydroxy cinamic acid octacosyl ester derivated isomer.

Coumarins and derivatives

Two coumarins and derivatives compounds corresponding to peaks 1 and 4 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 1 with a [M-H]-ion m/z 177.0184 and daughter ions at m/z 133.0282 and 149.0234 were identified as esculetin [57]. Peak 4 with a [M-H]-ion m/z 161.0234 and daughter ions at m/z 133.0283 and 117.0335 were identified as hydroxycoumarin [58].

Flavonoids

Five flavonoids compounds corresponding to peaks 14, 19, 22, 24 and 25 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 14 with a [M-H]-ion m/z 285.0760 and daughter ions at m/z 109.0283, 165.0546, 271.0605, 147.0440 and 177.0544 were identified as dihydroxy-methoxyflavanone [59]. Peak 19 with a [M-H]-ion m/z 301.0709 and daughter ions at m/z 165.0547, 273.0761, 153.0183 and 135.0076 were identified as hesperetin [60]. Peak 22 with a [M-H]-ion m/z 373.1646 and daughter ions at m/z 121.0647, 147.0439, 165.0543 and 285.0768 were identified as dihydroxy-methoxyflavanone derivative I. Peak 24 with a [M-H]-ion m/z 375.1802 and daughter ions at m/z 121.0647, 147.0439 and 165.0544 were identified as dihydroxy-methoxyflavanone derivative II. Peak 25 with a [M-H]-ion m/z 285.1122 and daughter ions at m/z 147.0441 and 107.0490 were identified as dihydroxy-methoxyflavanone derivative II.

Amino acids, peptides and analogs

One peptide and analog compound corresponding to peak 10 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 10 with a [M-H]-ion m/z 264.1232 and daughter ions at m/z 220.1333 and 246.1129 were identified as N-Carbobenzyloxy-L-isoleucine.

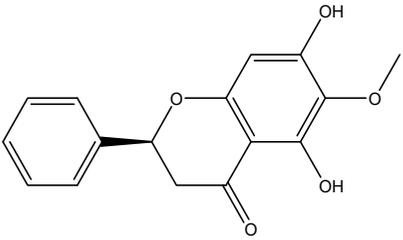
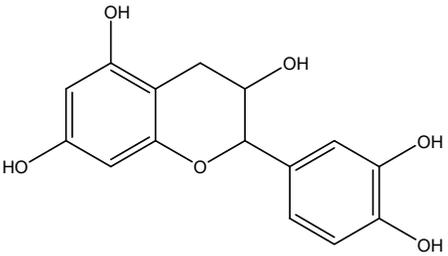
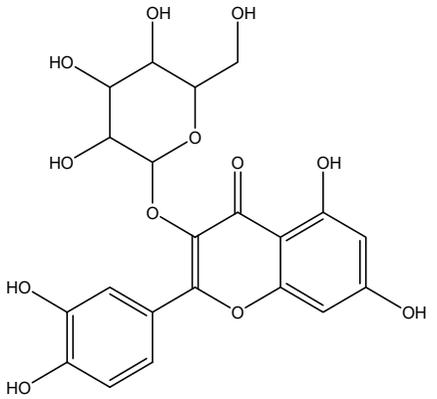
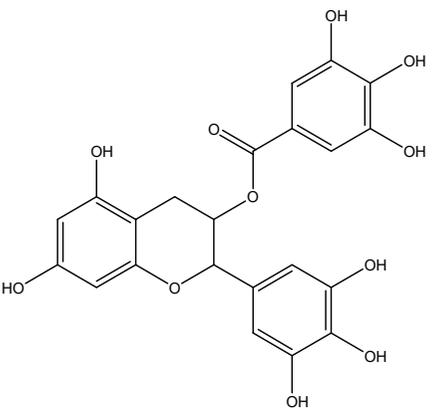
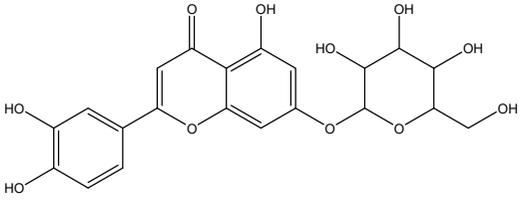
Fatty acid esters

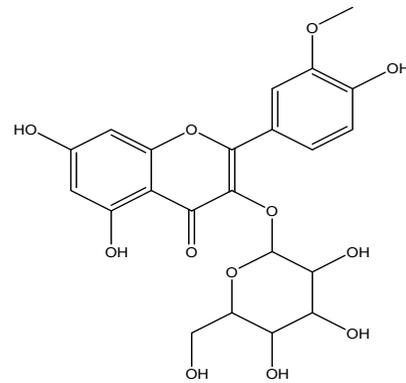
One benzene and substituted derivative compound corresponding to peak 30 were identified using UHPLC-Q/Orbitrap/MS/MS. Peak 30 with a [M-H]-ion m/z 191.1067 and daughter ion at m/z 121.0647 were identified as phenethyl butyrate [16].

Unknown compounds.

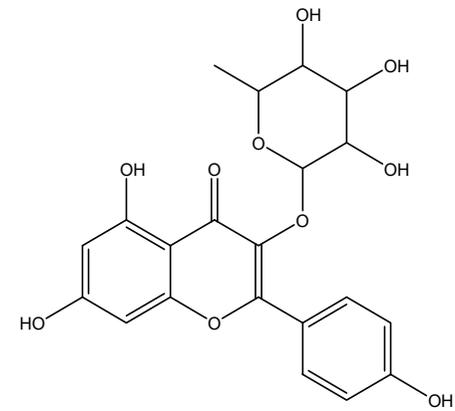
Peak 23 with a [M-H] -ion m/z 239.0916 correspond to unknown compound.

Table S4. Chemical structures of the compounds with potential AA identified

Metabolites	<i>G. sepium</i>	<i>L. leucocephala</i>	<i>P. dulce</i>
	 <p>Dihydroxymethoxy flavanone</p>	 <p>Catechin</p>	 <p>Quercetin-3-glucoside</p>
Flavonoids		 <p>Gallocatechin gallate</p>	 <p>Luteolin 7-O-glucoside</p>

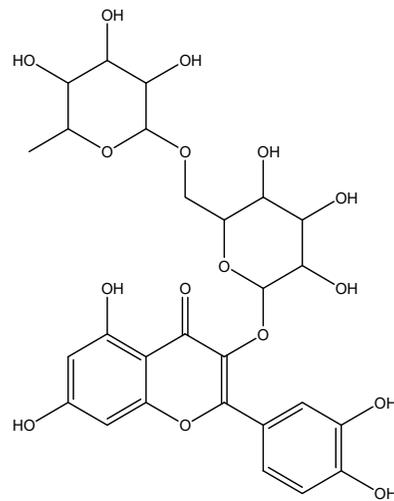


Isorhamnetin-O-glucoside

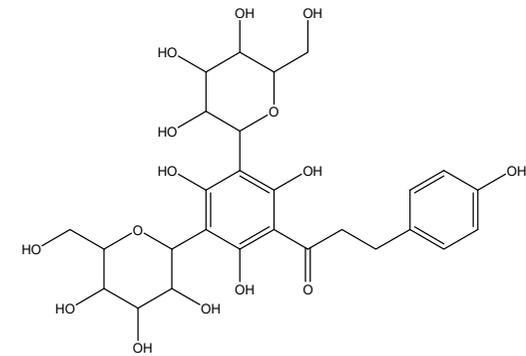


Kaempferol-3-O-rhamnoside

Flavonoids

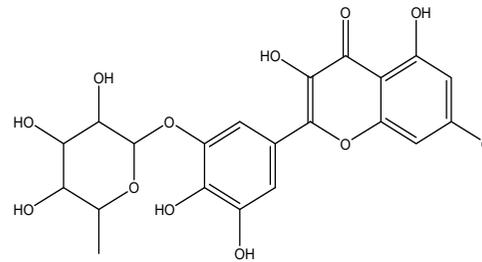


Rutin

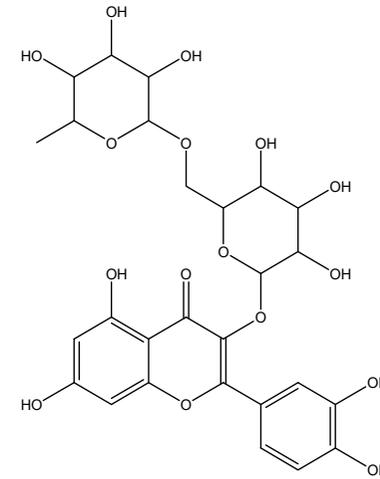


Phloretin-di-C-hexoside

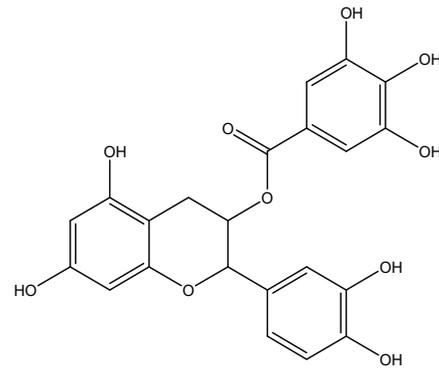
Flavonoids



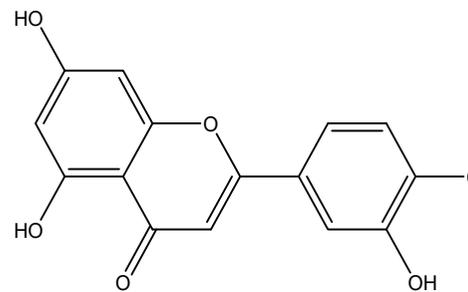
Myricetin 3-O-rhamnoside



Rutin

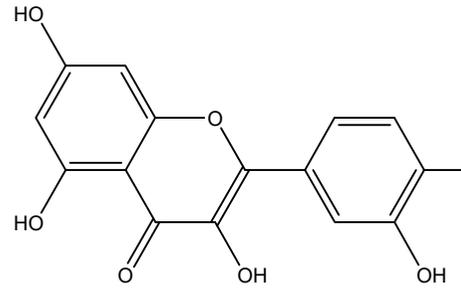


Epicatechin gallate



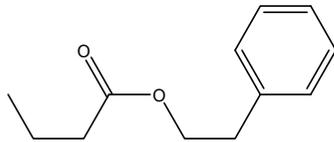
Luteolin

Flavonoids



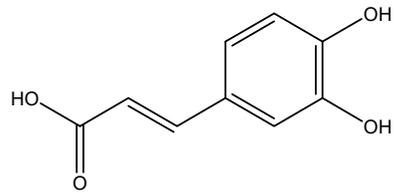
Quercetin

Fatty acid esters

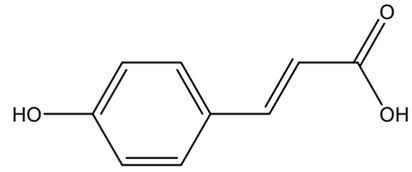


Phenethyl butyrate

Hydroxycinnamic acids

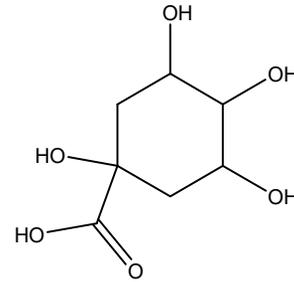


Caffeic acid

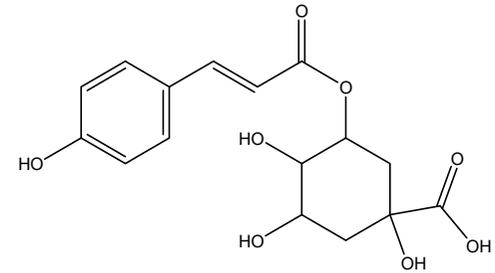


p-coumáric acid

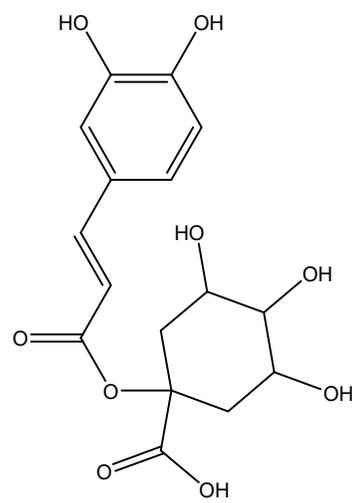
**Organooxygenated
compounds**



Quinic acid

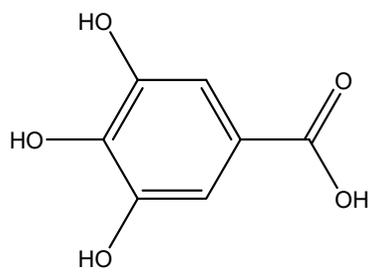


Coumaroylquinic acid



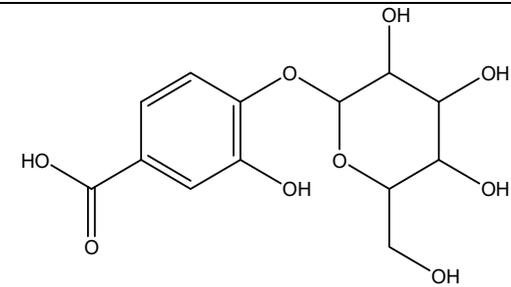
Caffeoylquinic acid

**Benzene and
substituted
derivatives**



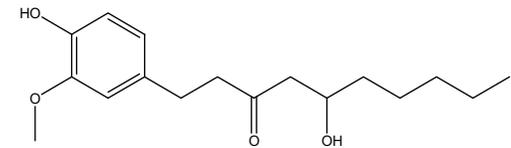
Gallic acid

Phenolic glycosides



Protocatechuic acid 4-hexoside

Phenols



Gingerol

References

1. Vallverdú-Queralt, A.; Jauregui, O.; Medina-Remón, A.; Andrés-Lacueva, C.; Lamuela-Raventós, R. M. Improved characterization of tomato polyphenols using liquid chromatography/electrospray ionization linear ion trap quadrupole Orbitrap mass spectrometry and liquid chromatography/electrospray ionization tandem mass spectrometry. *Rapid Communications in Mass Spectrometry* 2010, 24 (20), 2986-2992. [[CrossRef](#)] [[PubMed](#)]
2. Karar, M. E.; Kuhnert, N. UPLC-ESI-Q-TOF-MS/MS characterization of phenolics from *Crataegus monogyna* and *Crataegus laevigata* (Hawthorn) leaves, fruits and their herbal derived drops (Crataegutt Tropfen). *J. Chem. Biol. Ther* 2015, 1 (102), 2572-0406.1000102. [[CrossRef](#)]
3. Chu, Q.; Yu, L.; Zheng, Z.; Chen, M.; Hua, Z.; Hang, M.; Li, Y.; Li, X.; Liu, Y.; Yang, Y. *Apios americana* Medik flowers extract protects PC12 cells against H₂O₂ induced neurotoxicity via regulating autophagy. *Food and Chemical Toxicology* 2019, 124, 231-238. [[CrossRef](#)]
4. Haq, F. U.; Ali, A.; Akhtar, N.; Aziz, N.; Khan, M. N.; Ahmad, M.; Musharraf, S. G. A high-throughput method for dereplication and assessment of metabolite distribution in *Salvia* species using LC-MS/MS. *Journal of advanced research* 2020, 24, 79-90. [[CrossRef](#)] [[PubMed](#)]
5. Hassan, W. H.; Abdelaziz, S.; Al Yousef, H. M. Chemical composition and biological activities of the aqueous fraction of *Parkinsonia aculeata* L. growing in Saudi Arabia. *Arabian journal of chemistry* 2019, 12 (3), 377-387. [[CrossRef](#)]
6. Karar, M. E.; Quiet, L.; Rezk, A.; Jaiswal, R.; Rehders, M.; Ullrich, M.; Brix, K.; Kuhnert, N. Phenolic profile and in vitro assessment of cytotoxicity and antibacterial activity of *Ziziphus spina-christi* leaf extracts. *Med chem* 2016, 6 (3), 143-156.
7. Fernández-Poyatos, M. d. P.; Ruiz-Medina, A.; Zengin, G.; Llorent-Martínez, E. J. Phenolic characterization, antioxidant activity, and enzyme inhibitory properties of *Berberis thunbergii* DC. leaves: A valuable source of phenolic acids. *Molecules* 2019, 24 (22), 4171. [[CrossRef](#)] [[PubMed](#)]
8. Šuković, D.; Knežević, B.; Gašić, U.; Sredojević, M.; Ćirić, I.; Todić, S.; Mutić, J.; Tešić, Ž. Phenolic profiles of leaves, grapes and wine of grapevine variety vranac (*Vitis vinifera* L.) from Montenegro. *Foods* 2020, 9 (2), 138. [[CrossRef](#)] [[PubMed](#)]
9. Sanz, M.; de Simón, B. F.; Cadahía, E.; Esteruelas, E.; Muñoz, A. M.; Hernández, T.; Estrella, I.; Pinto, E. LC-DAD/ESI-MS/MS study of phenolic compounds in ash (*Fraxinus excelsior* L. and *F. americana* L.) heartwood. Effect of toasting intensity at cooperage. *Journal of Mass Spectrometry* 2012, 47 (7), 905-918. [[CrossRef](#)] [[PubMed](#)]
10. Cao, G.; Zhang, Y.; Feng, J.; Cai, H.; Zhang, C.; Ding, M.; Cong, X.; Cai, B. A rapid and sensitive assay for determining the main components in processed fructus Corni by UPLC-Q-TOF-MS. *Chromatographia* 2011, 73 (1), 135-141. [[CrossRef](#)]
11. Kosar, M.; Demirci, B.; Goger, F.; Kara, I.; Baser, K. Volatile composition, antioxidant activity, and antioxidant components in saffron cultivated in Turkey. *International journal of food properties* 2017, 20 (sup1), S746-S754. [[CrossRef](#)]
12. Ślusarczyk, S.; Cieślak, A.; Yanza, Y. R.; Szumacher-Strabel, M.; Varadyova, Z.; Stafiniak, M.; Wojnicz, D.; Matkowski, A. Phytochemical profile and antioxidant activities of *Coleus amboinicus* Lour. cultivated in Indonesia and Poland. *Molecules* 2021, 26 (10), 2915. [[CrossRef](#)] [[PubMed](#)]
13. Chia, V. V.; Pang, S. F.; Gimbut, J. Mass spectrometry analysis of auxiliary energy-induced terpenes extraction from *Andrographis paniculata*. *Industrial Crops and Products* 2020, 155, 112828. [[CrossRef](#)]
14. Mena, P.; Calani, L.; Dall'Asta, C.; Galaverna, G.; García-Viguera, C.; Bruni, R.; Crozier, A.; Del Rio, D. Rapid and comprehensive evaluation of (poly) phenolic compounds in pomegranate (*Punica granatum* L.) juice by UHPLC-MSn. *Molecules* 2012, 17 (12), 14821-14840. [[CrossRef](#)] [[PubMed](#)]
15. Chen, H. F.; Chen, Y. H.; Liu, C. H.; Wang, L.; Chen, X.; Yu, B. Y.; Qi, J. Integrated chemometric fingerprints of antioxidant activities and HPLC-DAD-CL for assessing the quality of the processed roots of *Polygonum multiflorum* Thunb. (Heshouwu). *Chinese medicine* 2016, 11 (1), 1-12. [[CrossRef](#)] [[PubMed](#)]
16. Nguyen, H. T.; Tran, P. H. An extremely efficient and green method for the acylation of secondary alcohols, phenols and naphthols with a deep eutectic solvent as the catalyst. *RSC advances* 2016, 6 (100), 98365-98368. [[CrossRef](#)]
17. Nwokocha, C.; Palacios, J.; Simirgiotis, M. J.; Thomas, J.; Nwokocha, M.; Young, L.; Thompson, R.; Cifuentes, F.; Paredes, A.; Delgoda, R. Aqueous extract from leaf of *Artocarpus altilis* provides cardio-

- protection from isoproterenol induced myocardial damage in rats: Negative chronotropic and inotropic effects. *Journal of ethnopharmacology* 2017, 203, 163-170. [[CrossRef](#)] [[PubMed](#)]
18. Barrientos, R. E.; Ahmed, S.; Cortés, C.; Fernández-Galleguillos, C.; Romero-Parra, J.; Simirgiotis, M. J.; Echeverría, J. Chemical fingerprinting and biological evaluation of the endemic Chilean fruit *Greigia sphacelata* (Ruiz and Pav.) Regel (Bromeliaceae) by UHPLC-PDA-orbitrap-mass spectrometry. *Molecules* 2020, 25 (16), 3750. [[CrossRef](#)] [[PubMed](#)]
 19. Peng, G.; Guan, H.; Wang, X.; Shi, Y. Simultaneous determination of 14 active constituents of Shengjiang Xiexin decoction using ultrafast liquid chromatography coupled with electrospray ionization tandem mass spectrometry. *Acta pharmaceutica sinica B* 2017, 7 (2), 193-201 [[CrossRef](#)]. [[PubMed](#)]
 20. Olennikov, D. N.; Chirikova, N. K.; Kashchenko, N. I.; Nikolaev, V. M.; Kim, S.-W.; Vennos, C. Bioactive phenolics of the genus *Artemisia* (Asteraceae): HPLC-DAD-ESI-TQ-MS/MS profile of the Siberian species and their inhibitory potential against α -amylase and α -glucosidase. *Frontiers in pharmacology* 2018, 9, 756. [[CrossRef](#)] [[PubMed](#)]
 21. Navarro-González, I.; González-Barrio, R.; García-Valverde, V.; Bautista-Ortín, A. B.; Periago, M. J. Nutritional composition and antioxidant capacity in edible flowers: Characterisation of phenolic compounds by HPLC-DAD-ESI/MSn. *International Journal of Molecular Sciences* 2014, 16 (1), 805-822. [[CrossRef](#)] [[PubMed](#)]
 22. Sojka, M.; Guyot, S.; Kołodziejczyk, K.; Król, B.; Baron, A. Composition and properties of purified phenolics preparations obtained from an extract of industrial blackcurrant (*Ribes nigrum* L.) pomace. *The Journal of Horticultural Science and Biotechnology* 2009, 84 (6), 100-106. [[CrossRef](#)]
 23. Al-Madhagy, S. A.; Mostafa, N. M.; Youssef, F. S.; Awad, G. E.; Eldahshan, O. A.; Singab, A. N. B. Metabolic profiling of a polyphenolic-rich fraction of *Coccinia grandis* leaves using LC-ESI-MS/MS and in vivo validation of its antimicrobial and wound healing activities. *Food & Function* 2019, 10 (10), 6267-6275. [[CrossRef](#)] [[PubMed](#)]
 24. Negri, G.; Tabach, R. Saponins, tannins and flavonols found in hydroethanolic extract from *Periandra dulcis* roots. *Revista Brasileira de Farmacognosia* 2013, 23 (6), 851-860. [[CrossRef](#)]
 25. Falcão, S. I.; Vale, N.; Gomes, P.; Domingues, M. R.; Freire, C.; Cardoso, S. M.; Vilas-Boas, M. Phenolic profiling of Portuguese propolis by LC-MS spectrometry: Uncommon propolis rich in flavonoid glycosides. *Phytochemical Analysis* 2013, 24 (4), 309-318. [[CrossRef](#)] [[PubMed](#)]
 26. Chen, Y.; Yu, H.; Wu, H.; Pan, Y.; Wang, K.; Jin, Y.; Zhang, C. Characterization and quantification by LC-MS/MS of the chemical components of the heating products of the flavonoids extract in pollen typhae for transformation rule exploration. *Molecules* 2015, 20 (10), 18352-18366. [[CrossRef](#)] [[PubMed](#)]
 27. Jang, G. H.; Kim, H. W.; Lee, M. K.; Jeong, S. Y.; Bak, A. R.; Lee, D. J.; Kim, J. B. Characterization and quantification of flavonoid glycosides in the *Prunus* genus by UPLC-DAD-QTOF/MS. *Saudi journal of biological sciences* 2018, 25 (8), 1622-1631. [[CrossRef](#)]
 28. Wang, X.; Cao, X.; Shang, Y.; Bu, H.; Wang, T.; Lyu, D.; Du, G. Preharvest application of prohydrojasmon affects color development, phenolic metabolism, and pigment-related gene expression in red pear (*Pyrus ussuriensis*). *Journal of the Science of Food and Agriculture* 2020, 100 (13), 4766-4775. [[CrossRef](#)] [[PubMed](#)]
 29. Keskes, H.; Belhadj, S.; Jlail, L.; El Feki, A.; Damak, M.; Sayadi, S.; Allouche, N. LC-MS-MS and GC-MS analyses of biologically active extracts and fractions from Tunisian *Juniperus phoenicea* leaves. *Pharmaceutical Biology* 2017, 55 (1), 88-95. [[CrossRef](#)] [[PubMed](#)]
 30. Li, A.; Hou, X.; Wei, Y. Fast screening of flavonoids from switchgrass and *Mikania micrantha* by liquid chromatography hybrid-ion trap time-of-flight mass spectrometry. *Analytical Methods* 2018, 10 (1), 109-122. [[CrossRef](#)]
 31. Shetty, R.; Fretté, X.; Jensen, B.; Shetty, N. P.; Jensen, J. D.; Jørgensen, H. J. L.; Newman, M.-A.; Christensen, L. P. Silicon-induced changes in antifungal phenolic acids, flavonoids, and key phenylpropanoid pathway genes during the interaction between miniature roses and the biotrophic pathogen *Podosphaera pannosa*. *Plant physiology* 2011, 157 (4), 2194-2205. [[CrossRef](#)] [[PubMed](#)]
 32. Sánchez-Rabáneda, F.; Jauregui, O.; Lamuela-Raventos, R. M.; Bastida, J.; Viladomat, F.; Codina, C. Identification of phenolic compounds in artichoke waste by high-performance liquid chromatography-tandem mass spectrometry. *Journal of Chromatography A* 2003, 1008 (1), 57-72. [[CrossRef](#)] [[PubMed](#)]
 33. Barros, L.; Dueñas, M.; Alves, C. T.; Silva, S.; Henriques, M.; Santos-Buelga, C.; Ferreira, I. C. Antifungal activity and detailed chemical characterization of *Cistus ladanifer* phenolic extracts. *Industrial Crops and Products* 2013, 41, 41-45. [[CrossRef](#)]

34. Yuzuak, S.; Ballington, J.; Xie, D.-Y. HPLC-qTOF-MS/MS-based profiling of flavan-3-ols and dimeric proanthocyanidins in berries of two muscadine grape hybrids FLH 13-11 and FLH 17-66. *Metabolites* 2018, 8 (4), 57. [[CrossRef](#)]
35. Sun, J.; Liang, F.; Bin, Y.; Li, P.; Duan, C. Screening non-colored phenolics in red wines using liquid chromatography/ultraviolet and mass spectrometry/mass spectrometry libraries. *Molecules* 2007, 12 (3), 679-693. [[CrossRef](#)] [[PubMed](#)]
36. Ben Said, R.; Hamed, A. I.; Mahalel, U. A.; Al-Ayed, A. S.; Kowalczyk, M.; Moldoch, J.; Oleszek, W.; Stochmal, A. Tentative characterization of polyphenolic compounds in the male flowers of *Phoenix dactylifera* by liquid chromatography coupled with mass spectrometry and DFT. *International journal of molecular sciences* 2017, 18 (3), 512. [[CrossRef](#)] [[PubMed](#)]
37. McNab, H.; Ferreira, E. S.; Hulme, A. N.; Quye, A. Negative ion ESI-MS analysis of natural yellow dye flavonoids—An isotopic labelling study. *International Journal of Mass Spectrometry* 2009, 284 (1-3), 57-65. [[CrossRef](#)]
38. Li, X.; Zhang, Y. F.; Yang, L.; Feng, Y.; Deng, Y. H.; Liu, Y. M.; Zeng, X. Chemical profiling of constituents of *Smilaxis glabrae* using ultra-high pressure liquid chromatography coupled with LTQ Orbitrap mass spectrometry. *Natural Product Communications* 2012, 7 (2), 1934578X1200700213. [[CrossRef](#)] [[PubMed](#)]
39. Hong, Y.; Liao, X.; Chen, Z. Determination of bioactive components in the fruits of *Cercis chinensis* Bunge by HPLC-MS/MS and quality evaluation by principal components and hierarchical cluster analyses. *Journal of pharmaceutical analysis* 2021, 11 (4), 465-471. [[CrossRef](#)] [[PubMed](#)]
40. Enomoto, H. Mass spectrometry imaging of flavonols and ellagic acid glycosides in ripe strawberry fruit. *Molecules* 2020, 25 (20), 4600. [[CrossRef](#)] [[PubMed](#)]
41. Fan, Y.; Li, Y.; Wu, Y.; Li, L.; Wang, Y.; Li, Y. Identification of the chemical constituents in simiao wan and rat plasma after oral administration by GC-MS and LC-MS. *Evidence-Based Complementary and Alternative Medicine* 2017, 2017. [[CrossRef](#)] [[PubMed](#)]
42. Liu, S.; Marsol-Vall, A.; Laaksonen, O.; Korttesniemi, M.; Yang, B. Characterization and quantification of nonanthocyanin phenolic compounds in white and blue bilberry (*Vaccinium myrtillus*) juices and wines using UHPLC-DAD-ESI-QTOF-MS and UHPLC-DAD. *Journal of agricultural and food chemistry* 2020, 68 (29), 7734-7744. [[CrossRef](#)] [[PubMed](#)]
43. Teixeira, N.; Nabais, P.; de Freitas, V.; Lopes, J. A.; Melo, M. J. In-depth phenolic characterization of iron gall inks by deconstructing representative Iberian recipes. *Scientific reports* 2021, 11 (1), 1-11. [[CrossRef](#)] [[PubMed](#)]
44. Beshel, J. A.; Palacios, J.; Beshel, F. N.; Nku, C. O.; Owu, D. U.; Nwokocha, M.; Bórquez, J.; Simirgiotis, M. J.; Nwokocha, C. R. Blood pressure-reducing activity of *Gongronema latifolium* Benth. (Apocynaceae) and the identification of its main phytochemicals by UHPLC Q-Orbitrap mass spectrometry. *Journal of basic and clinical physiology and pharmacology* 2020, 31 (1). [[CrossRef](#)] [[PubMed](#)]
45. Singh, A.; Bajpai, V.; Kumar, S.; Sharma, K. R.; Kumar, B. Profiling of gallic and ellagic acid derivatives in different plant parts of *Terminalia arjuna* by HPLC-ESI-QTOF-MS/MS. *Natural product communications* 2016, 11 (2), 1934578X1601100227. [[CrossRef](#)] [[PubMed](#)]
46. Zou, Y. P.; Tan, C. H.; Wang, B. D.; Zhu, D. Y.; Kim, S. K. Chemical constituents from *Myrsine africana* L. *Helvetica Chimica Acta* 2008, 91 (11), 2168-2173. [[CrossRef](#)]
47. Mekam, P. N.; Martini, S.; Nguefack, J.; Tagliazucchi, D.; Stefani, E. Phenolic compounds profile of water and ethanol extracts of *Euphorbia hirta* L. leaves showing antioxidant and antifungal properties. *South African Journal of Botany* 2019, 127, 319-332. [[CrossRef](#)]
48. Li, F.; Zhang, Y. B.; Wei, X.; Song, C. H.; Qiao, M. Q.; Zhang, H. Y. Metabolic profiling of Shu-Yu capsule in rat serum based on metabolic fingerprinting analysis using HPLC-ESI-MSn. *Molecular Medicine Reports* 2016, 13 (5), 4191-4204. [[CrossRef](#)] [[PubMed](#)]
49. Yan, H.; Zou, D.; Zhou, G.; Yu, H.; Li, P.; Wang, T.; Bao, B.; Guo, S.; Duan, J. Metabolomics of ginger based on ultra-performance liquid chromatography coupled with quadrupole time-of-flight mass spectrometry technology. *Food Quality and Safety* 2021, 5. [[CrossRef](#)]
50. Choi, J.; An, X.; Lee, B. H.; Lee, J. S.; Heo, H. J.; Kim, T.; Ahn, J.-W.; Kim, D.-O. Protective effects of bioactive phenolics from jujube (*Ziziphus jujuba*) seeds against H₂O₂-induced oxidative stress in neuronal PC-12 cells. *Food science and biotechnology* 2015, 24 (6), 2219-2227. [[CrossRef](#)]
51. Zhang, X.; Liang, C.; Li, C.; Bu, M.; Bu, L.; Xiao, Y.; Sun, H.; Zhang, L. Simultaneous qualitative and quantitative study of main compounds in *Commelina communis* linn. by UHPLC-Q-TOF-MS-MS and HPLC-ESI-MS-MS. *Journal of chromatographic science* 2018, 56 (7), 582-594. [[CrossRef](#)] [[PubMed](#)]

52. Taamalli, A.; Arráez-Román, D.; Abaza, L.; Iswaldi, I.; Fernández-Gutiérrez, A.; Zarrouk, M.; Segura-Carretero, A. LC-MS-based metabolite profiling of methanolic extracts from the medicinal and aromatic species *Mentha pulegium* and *Origanum majorana*. *Phytochemical analysis* 2015, 26 (5), 320-330. [[CrossRef](#)] [[PubMed](#)]
53. Simirgiotis, M. J.; Ramirez, J. E.; Hirschmann, G. S.; Kennelly, E. J. Bioactive coumarins and HPLC-PDA-ESI-ToF-MS metabolic profiling of edible queule fruits (*Gomortega keule*), an endangered endemic Chilean species. *Food Research International* 2013, 54 (1), 532-543. [[CrossRef](#)]
54. Lin, H.; Zhu, H.; Tan, J.; Wang, H.; Wang, Z.; Li, P.; Zhao, C.; Liu, J. Comparative analysis of chemical constituents of *Moringa oleifera* leaves from China and India by ultra-performance liquid chromatography coupled with quadrupole-time-of-flight mass spectrometry. *Molecules* 2019, 24 (5), 942. [[CrossRef](#)]
55. Cho, K.; Choi, Y.-J.; Ahn, Y. H. Identification of polyphenol glucuronide conjugates in *Glechoma hederacea* var. *longituba* hot water extracts by high-performance liquid chromatography-tandem mass spectrometry (HPLC-MS/MS). *Molecules* 2020, 25 (20), 4713. [[CrossRef](#)] [[PubMed](#)]
56. Lee, H.; Oh, I.-N.; Kim, J.; Jung, D.; Cuong, N. P.; Kim, Y.; Lee, J.; Kwon, O.; Park, S. U.; Lim, Y. Phenolic compound profiles and their seasonal variations in new red-phenotype head-forming Chinese cabbages. *LWT* 2018, 90, 433-439. [[CrossRef](#)]
57. Zuo, J.; Zhang, W.; Jian, H.; Bou-Chacra, N.; Löbenberg, R. Esculetin as bioactive marker: towards a rational scientific approach for the treatment of hyperuricemia using traditional Chinese medicine. *Brazilian Journal of Pharmaceutical Sciences* 2020, 56. [[CrossRef](#)]
58. Li, S.; Lin, Z.; Jiang, H.; Tong, L.; Wang, H.; Chen, S. Rapid identification and assignment of the active ingredients in fufang banbianlian injection using HPLC-DAD-ESI-IT-TOF-MS. *Journal of Chromatographic Science* 2016, 54 (7), 1225-1237. [[CrossRef](#)] [[PubMed](#)]
59. Simirgiotis, M. J.; Benites, J.; Areche, C.; Sepúlveda, B. Antioxidant capacities and analysis of phenolic compounds in three endemic *Nolana* species by HPLC-PDA-ESI-MS. *Molecules* 2015, 20 (6), 11490-11507. [[CrossRef](#)] [[PubMed](#)]
60. Zhao, H.-Y.; Fan, M.-X.; Wu, X.; Wang, H.-J.; Yang, J.; Si, N.; Bian, B.-L. Chemical profiling of the Chinese herb formula Xiao-Cheng-Qi decoction using liquid chromatography coupled with electrospray ionization mass spectrometry. *Journal of chromatographic science* 2013, 51 (3), 273-285. [[CrossRef](#)] [[PubMed](#)]