

Supporting Information for Oxidized Forms of Ergothioneine Are Substrates for Mammalian Thioredoxin Reductase[†]

Kaelyn A. Jenny^{1,3}, Gracyn Mose^{1,3}, Daniel J. Haupt², and Robert J. Hondal^{*1, 2}

¹Department of Biochemistry, 89 Beaumont Ave, Given Laboratory, Room B413, Burlington, VT 05405

²Department of Chemistry, 82 University Place, Innovation Hall, Room E340, Burlington, VT 05405

³These authors contributed equally.

[†]These studies were supported by National Heart, Lung, and Blood Institute grant HL141146 to Robert J. Hondal.

*To whom correspondence should be addressed. Department of Biochemistry, University of Vermont, College of Medicine. 89 Beaumont Ave, Given Laboratory, Room B413, Burlington, VT 05405. Tel: 802-656-8282. FAX: 802-862-8220. E-mail: Robert.Hondal@uvm.edu

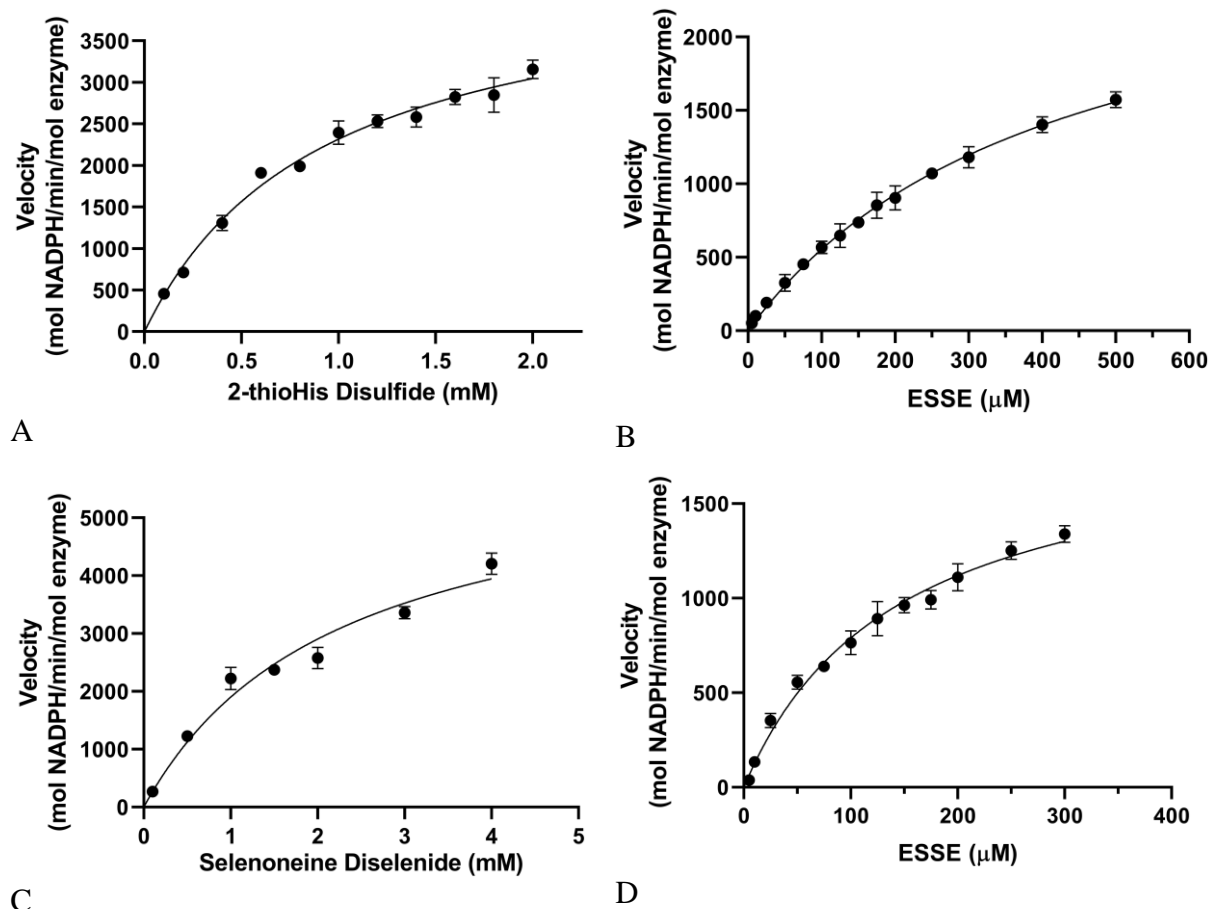


Figure S1: Michaelis-Menten activity plots for the disulfides/diselenides of 2-thioHis, EGT, and selenoneine with Sec-TrxR. A) Assay of 2-thioHis oxidized with H_2O_2 . B) Assay of EGT oxidized with H_2O_2 . C) Assay of 2-thioHis oxidized with H_2O_2 . D) Assay of EGT oxidized by H_2O_2 in the presence of $10\ \mu\text{M}$ *E. coli* Trx. The data points in D) represent the corrected data with the activity of $10\ \mu\text{M}$ *E. coli* Trx alone subtracted from the activity in which both ESSE and Trx are present.

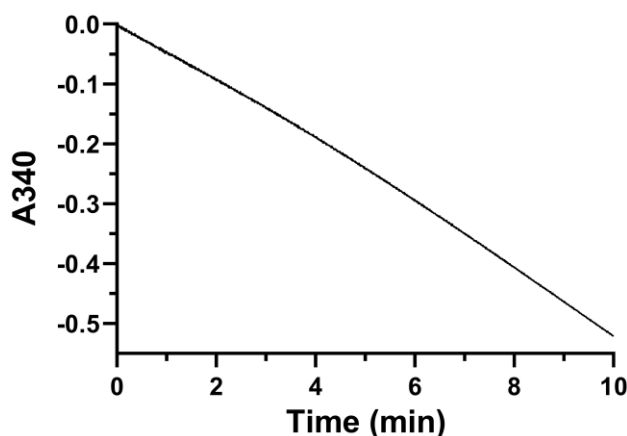


Figure S2: Absorbance versus time for GR/GSH, NADPH with 1 mM ESSE. To a cuvette, 200 μ M GSH, 200 μ M NADPH, 1 mM ESSE, and 0.60 nM GR were added and the reaction was followed for 10 min. The activity increases with time as substrate is reduced.

NMR experiments with EGT and H₂O₂: To quantify the extent of desulfurization of EGT following treatment with H₂O₂ to produce ESSE we performed quantitative ¹H-NMR experiments. The hercynine C2 proton in samples of EGT in D₂O was found to have a long T1 relaxation time of 14.42 seconds. Using this T1 value, quantitative ¹H-NMR with 2 dummy scans and 8 scans over ~12 min was performed on two samples: 10 mM EGT dissolved in D₂O and 10 mM EGT with 5 mM H₂O₂ in D₂O (**Figure S3**). A ¹H-NMR spectrum was obtained 10 min after adding H₂O₂ to the sample. The hercynine C2 proton at 8.34 ppm is well resolved in ¹H-NMR so we used this as the main indicator of desulfurization. With the addition of H₂O₂ to EGT the C2 proton signal increased from 0.02 to 0.03, indicating an increase from 2% to 3% desulfurization upon a 10 min incubation with 5 mM H₂O₂ (**Figure S3**). Several new peaks appeared in the oxidized sample at 3.34, 3.89, 7.15, 7.21, and 7.26 ppm indicating the presence of oxidized species of EGT such as ESSE, hercynine, or other oxidized forms. There was also notable peak broadening and slight peak shifting in the oxidized sample, most evident in the peak at 6.82 ppm (compare to 6.72 ppm peak in unoxidized sample on the left in **Figure S3**) indicating the presence of multiple EGT species of different oxidation states and possibly the

formation and break-down of these species on the NMR time scale. It has been shown that when EGT forms ESSE under aqueous conditions, it can form *S*-monoxide disulfide and *S*-dioxide disulfide, which break down into the sulfenic and sulfinic acids, followed by desulfurization to hercynine, EGT, and sulfurous acid.^{9,13} This NMR experiment leads us to conclude that the desulfurization of ESSE producing EGT and hercynine is not extensive in the time-frame of the enzyme kinetics assays.

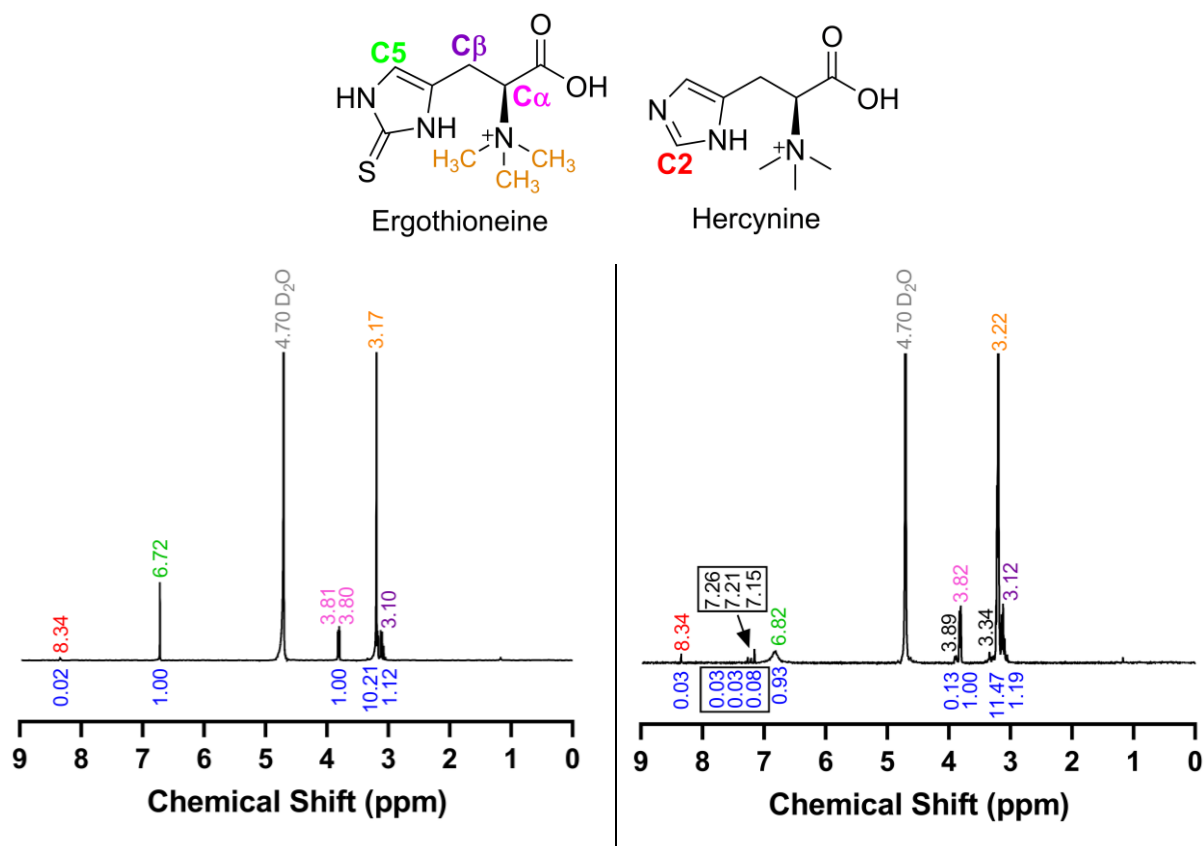


Figure S3: ¹H-NMR spectra of EGT before and after addition of H₂O₂ with structures of EGT and hercynine for reference. The carbons of EGT and hercynine are labeled for reference. (left) ¹H-NMR spectrum of 10 mM EGT with no H₂O₂. (right) ¹H NMR spectrum of 10 mM EGT with 5 mM H₂O₂.

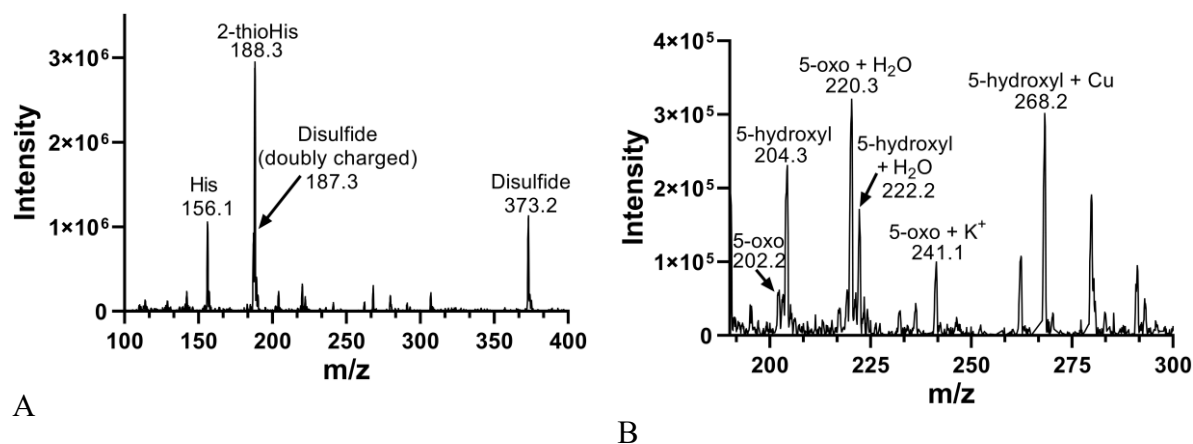


Figure S4: MS analysis of 2-thioHis oxidized with ¹O₂. A) Full MS spectrum of 2-thioHis following oxidation with ¹O₂. B) MS spectrum of 2-thioHis oxidized with ¹O₂ zoomed in to show the 5-oxo product peaks. The main oxidized species of 2-thioHis present was the disulfide form with only small amounts of the 5-oxo and 5-hydroxyl products.

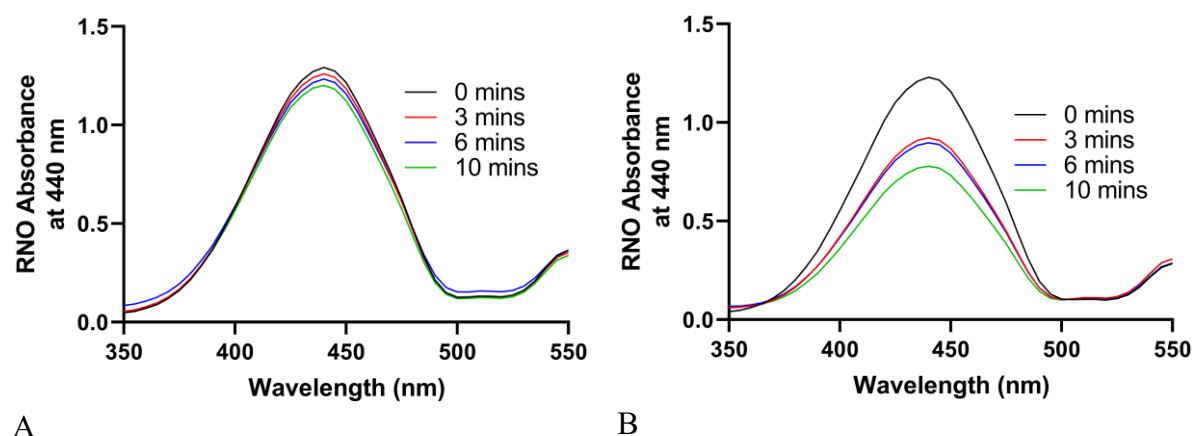


Figure S5: RNO bleaching experiments. A) 10 μM rose bengal with 50 μM RNO in 90% D₂O incubated under light for 20 min. B) 10 μM rose bengal with 50 μM RNO and 8 mM imidazole in 90% D₂O incubated under light for 20 min. The absorbance of RNO at 440 nm decreases dramatically when imidazole, a singlet oxygen scavenger, is added to the reaction.

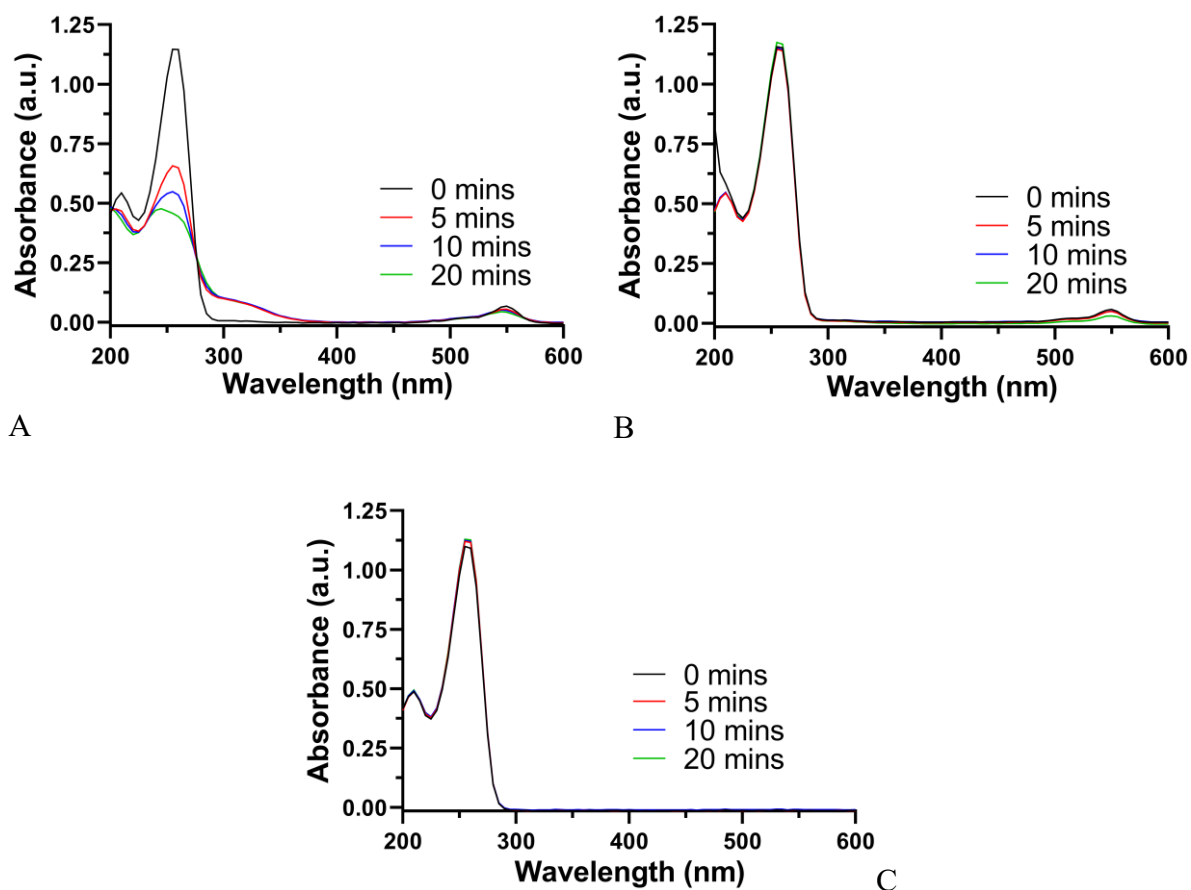


Figure S6: Absorbance spectra of 2-thioHis with and without rose bengal and light. A) 1 mM 2-thioHis and 10 μ M rose bengal, irradiated with light for 20 min. B) 1 mM 2-thioHis and 10 μ M rose bengal, incubated in the dark at room temperature for 20 min. C) 1 mM 2-thioHis irradiated with light for 20 min. Absorbance of a diluted solution (75 μ M 2-thioHis) was measured at 0, 5, 10, and 20 mins.

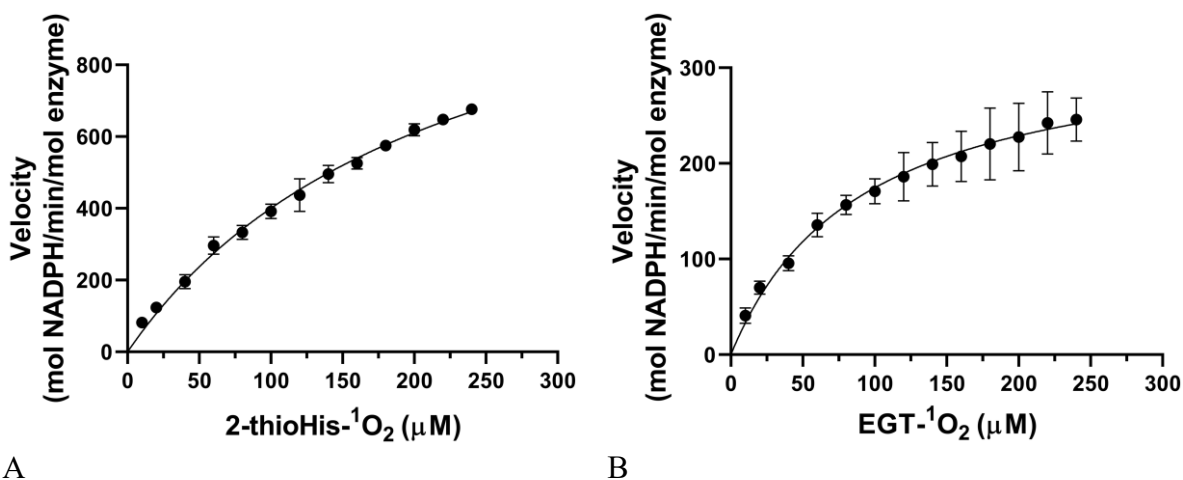


Figure S7: Michaelis-Menten plots of the reduction of aliquots of 2-thioHis/EGT oxidized by ¹O₂ catalyzed by Sec-TrxR. A) Plot for ¹O₂ oxidized 2-thioHis and Sec-TrxR. B) Plot for ¹O₂ oxidized EGT and Sec-TrxR.

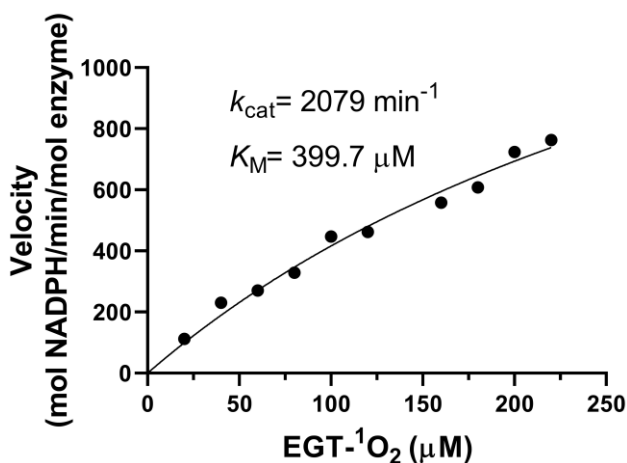


Figure S8: Michaelis-Menten curve for EGT-¹O₂ in pH 8.0 ammonium bicarbonate with 200 μM NADPH and 20 nM Sec-TrxR. These are the conditions used for MS analysis of the enzymatic recycling of 5-oxo-EGT. This experiment was performed to ensure that Sec-TrxR had activity in pH 8.0 ammonium bicarbonate buffer prior to MS analysis.

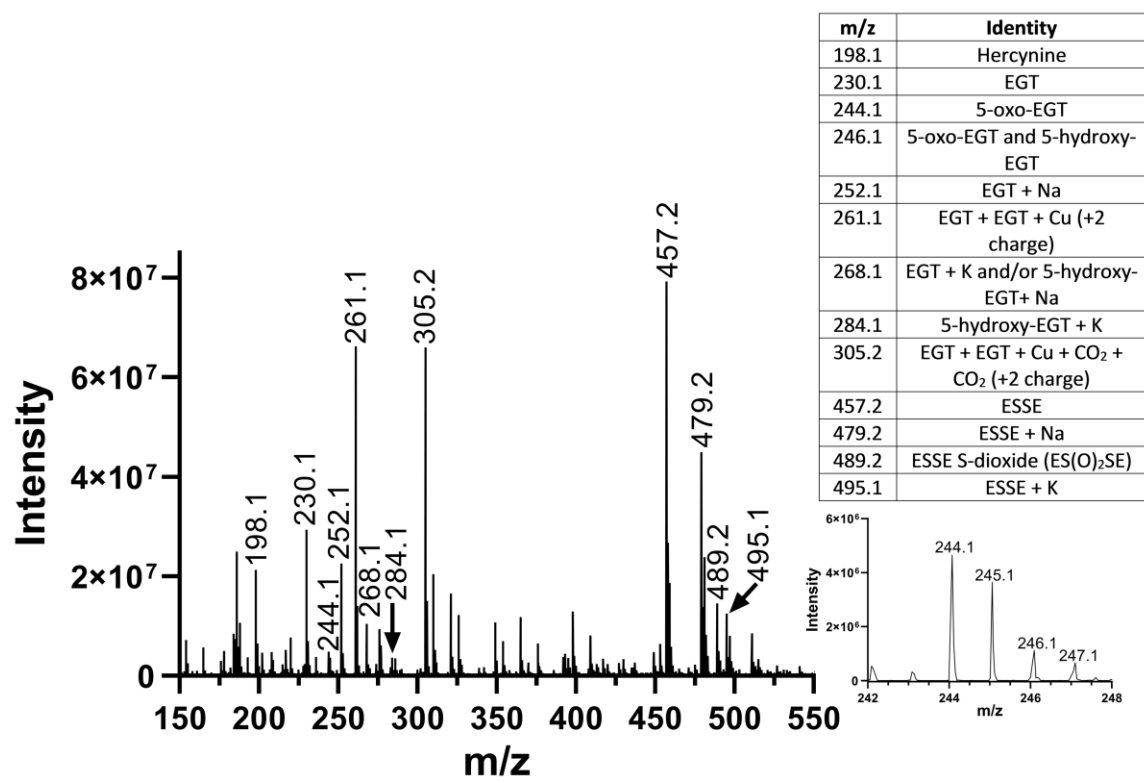


Figure S9: Control MS experiment showing that 5-oxo-EGT, 5-hydroxy-EGT, and ESSE are stable to ultrafiltration. The inset shows a close-up of 5-oxo-EGT and 5-hydroxy-EGT.