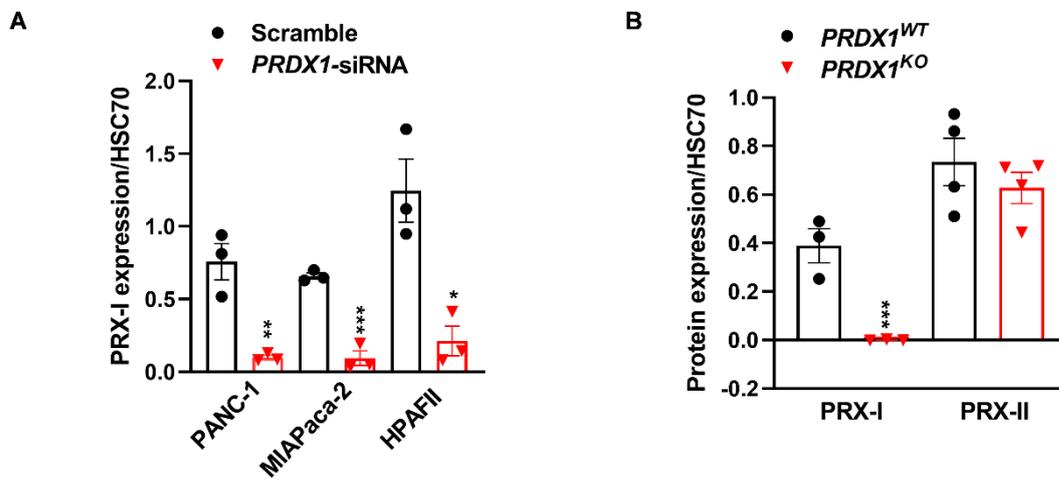
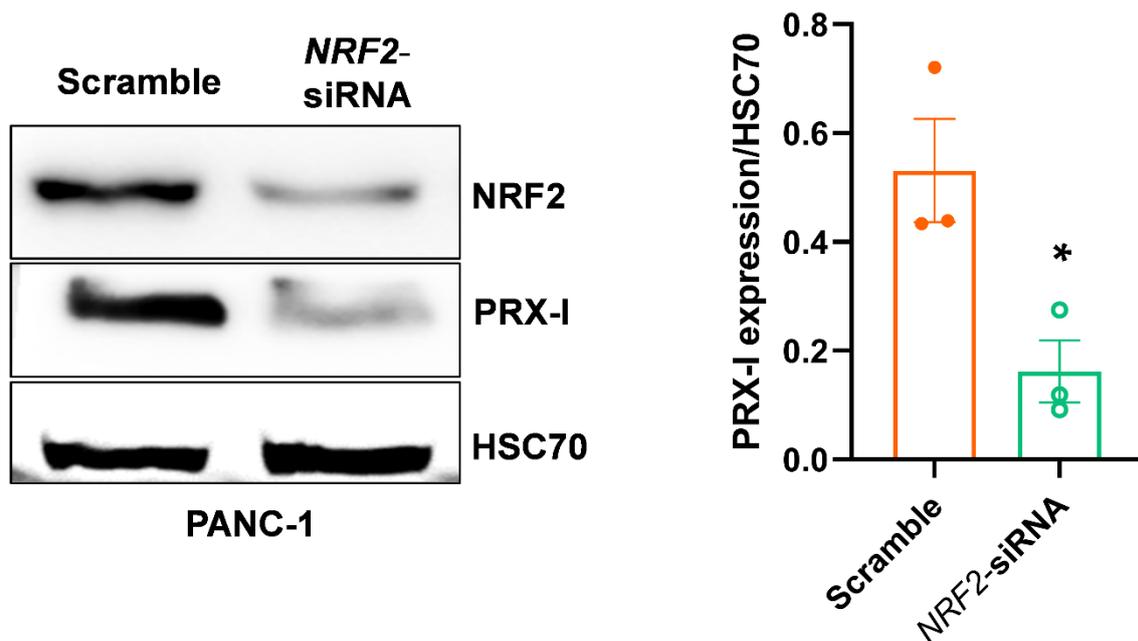


**Figure S1. Illustration of the used tamoxifen and cerulein regimens in *ElastaseCreER-LSLKras<sup>G12D</sup>-p53<sup>R172H</sup>* (KPE) mice.** Adult 6-week-old KPE mice were first treated once a day with tamoxifen (gavage) and 4-hydroxytamoxifen (subcutaneous injection), at days (D) 1, 3 and 5 (red arrows), to recombine the LSL stop cassettes and allow the expression of *Kras<sup>G12D</sup>* and *p53<sup>R172H</sup>* from their respective endogenous locus. After tamoxifen injections, mice were let recover for 1 week. Next, mice started an acute pancreatitis regimen of 1 week consisting of 7 dailies intraperitoneal cerulein injections (1 injection/hour), at days (D) 1, 3 and 5 (red arrows); the dose of each cerulein injection was 125  $\mu\text{g}/\text{Kg}$  in a volume of 100-150  $\mu\text{l}$  (diluted in PBS). At the end of acute treatment, mice started a chronic pancreatitis regimen of 5 weeks. Each week, mice received one daily intraperitoneal cerulein injection at days (D) 1, 2, 3, 4 and 5; the dose of each cerulein injection was 125  $\mu\text{g}/\text{Kg}$  in a volume of 100-150  $\mu\text{l}$  (diluted in PBS). At the end of chronic treatment, mice were kept for 14 additional weeks to allow tumor formation and were then sacrificed by cervical dislocation.

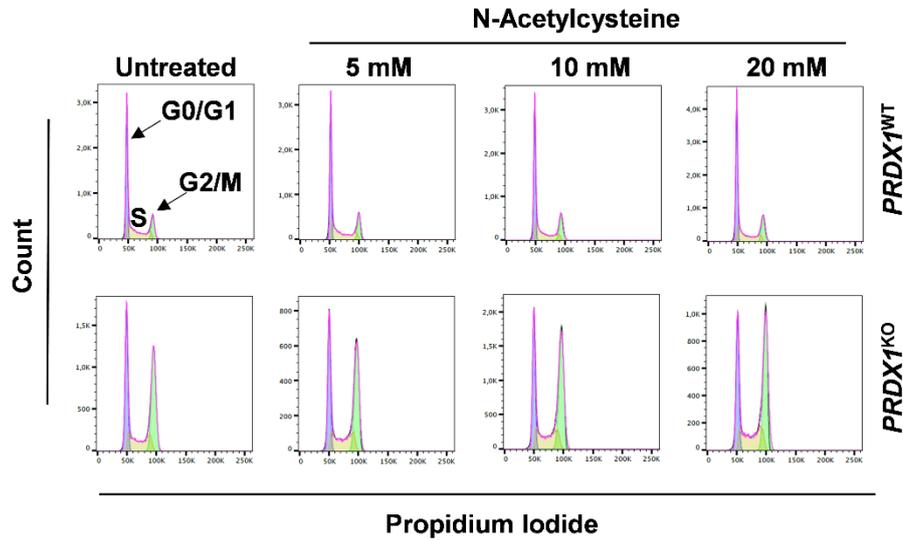


**Figure S2.** Peroxiredoxin-I (PRX-I) expression in human pancreatic ductal adenocarcinoma cells following genetic inactivation of *Peroxiredoxin-1* (*PRDX1*) gene by small interfering RNA (siRNA) or clustered regularly interspaced short palindromic repeats/CRISPR-associated protein 9 (CRISPR/Cas9). (A) Densitometry quantification for western blots shown in **Figure 2A**. Heat-shock cognate 70 (HSC70) was used as a loading control. Experiments were performed on three independent passages (n=3). (B) Densitometry quantification for western blots shown in **Figure 2F**. HSC70 was used as a loading control. Experiments were performed on at least three independent passages (n=3). Data are mean  $\pm$  SEM. Statistical significance was tested by Student t-test (\*  $P < 0.05$ , \*\*  $P < 0.01$ , \*\*\*  $P < 0.001$ ).

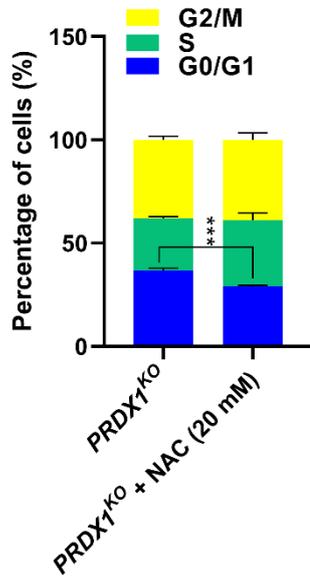


**Figure S3.** Nuclear factor erythroid 2-related factor 2 (NRF2) regulates the expression of Peroxiredoxin-I (PRX-I) in human pancreatic ductal adenocarcinoma cells. Western blot analysis on PANC-1 cells transfected with Scramble or specific *NRF2*-siRNA. The corresponding densitometry analysis is also available; experiments were performed on three independent cultures (n=3). Data are mean  $\pm$  SEM. Statistical significance was tested by Student t-test (\*  $P < 0.05$ ).

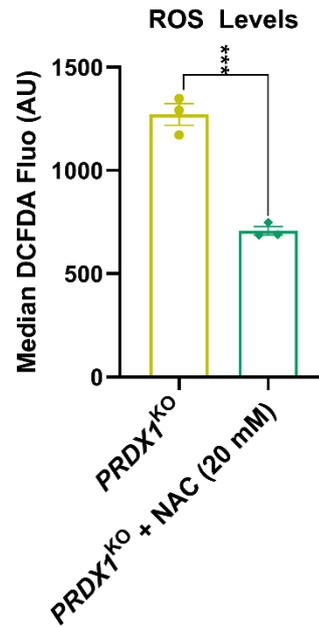
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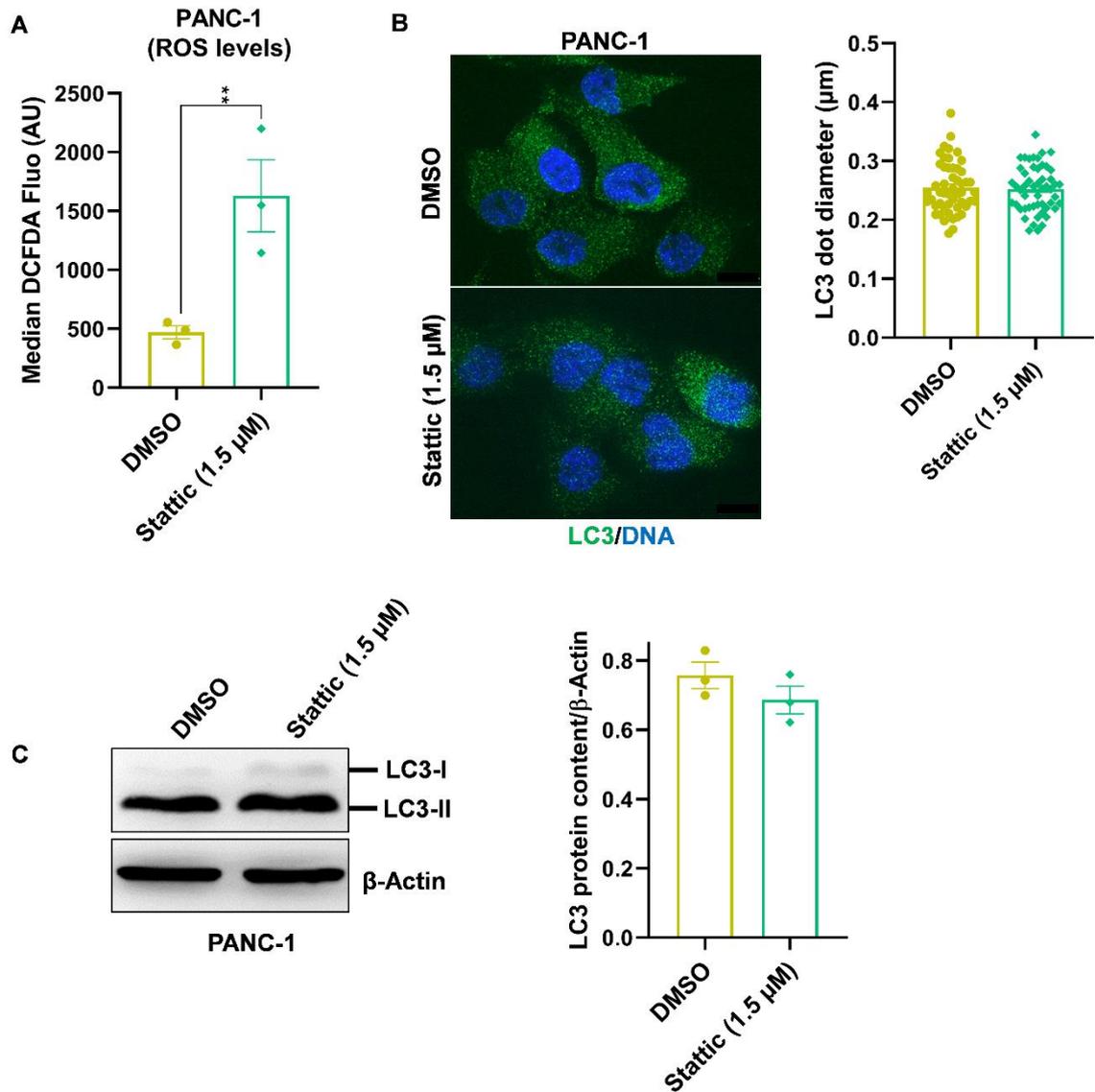
B



C



**Figure S4. N-Acetylcysteine (NAC) treatment does not reverse cell cycle blockade at G2/M in human pancreatic ductal adenocarcinoma cells.** (A) Typical Plots from FACS cell cycle analysis on *PRDX1<sup>WT</sup>* and *PRDX1<sup>KO</sup>* lines untreated or treated with different NAC doses for 16 hours. (B) Percentage of *PRDX1<sup>KO</sup>* cells in the different cell cycle phases following NAC treatment (20 mM) for 16 hours; experiments were performed on three independent cultures (n=3). (C) Reactive oxygen species (ROS) levels, determined by FACS, in *PRDX1<sup>KO</sup>* cells treated or not with NAC (20 mM) for 16 hours; measurements were performed on three independent cultures (n=3). Data are mean  $\pm$  SEM. Statistical significance was tested by Student t-test (\*\*\*)  $P < 0.001$ .



**Figure S5. Inhibition of Signal transducer and activator of transcription 3 (STAT3) induces reactive oxygen species (ROS) production without impacting the autophagic flux in human pancreatic ductal adenocarcinoma cells.** (A) Reactive oxygen species (ROS) levels, determined by FACS, in PANC-1 cells treated with DMSO (0.015%) or Stattic (1.5  $\mu$ M) for 16 hours; measurements were performed on three independent cultures (n=3). AU: Arbitrary Units. (B) Representative confocal images of Microtubule-associated protein 1A/1B-light chain (LC3) immunolabeling on PANC-1 cells treated with DMSO (0.015%) or Stattic (1.5  $\mu$ M) for 16 hours; immunolabeling was performed on two independent cultures (n=2) and 51-to-52 cells were selected randomly from confocal images to measure LC3 dots diameter. (C) Western blots for LC3 and the corresponding densitometry quantification performed on PANC-1 cells treated with DMSO (0.015%) or Stattic (1.5  $\mu$ M) for 16 hours.  $\beta$ -Actin was used a loading control. Western blots were performed on three independent cultures (n=3). Data are mean  $\pm$  SEM. Statistical significance was tested by Student t-test (\*\* $P$ <0.01).