

## **Supplementary material**

# **Adsorption of Macrolide Antibiotics and a Metabolite onto Polyethylene Terephthalate and Polyethylene Microplastics in Aquatic Environments**

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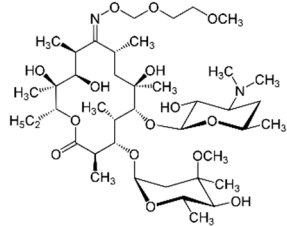
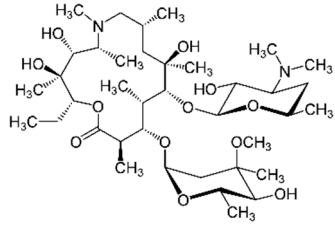
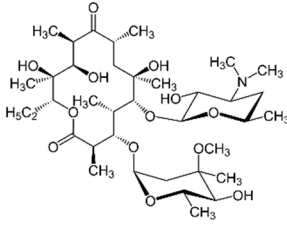
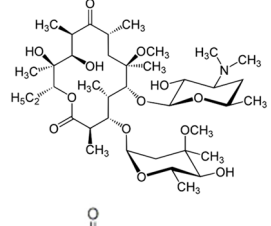
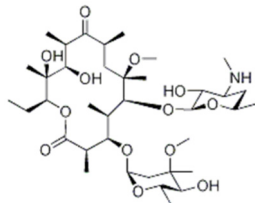
**Table S1.** Surface area and particle size of PE and PET particles before and after adsorption process.

	PE particles 300 $\mu\text{m}$ before adsorption	PE particles 300 $\mu\text{m}$ after adsorption	PET particles 300 $\mu\text{m}$ before adsorption	PET particles 300 $\mu\text{m}$ after adsorption
BET surface area ( $\text{m}^2/\text{g}$ )	0.0686	0.0726	0.1345	0.1444
Single point surface area ( $\text{m}^2/\text{g}$ )	0.0555	0.0601	0.1137	0.1213
Particle size ( $\mu\text{m}$ )	252.630	242.985	118.847	125.620

**Table S2.** Adsorption kinetic parameters.

Compound	PFO model			PSO model		
	R <sup>2</sup>	q <sub>e</sub> (mg/g)	k <sub>1</sub>	R <sup>2</sup>	k <sub>2</sub>	q <sub>e</sub> (mg/g)
<b>PE</b>						
CLM	0.661	0.001	0.001	0.968	0.004	0.001
DM-CLM	0.760	0.002	0.002	0.979	0.005	0.002
ERY	0.868	0.001	0.001	0.983	0.004	0.001
RXM	0.756	0.001	0.001	0.987	0.004	0.001
AZM	0.662	0.001	0.001	0.976	0.004	0.001
<b>PET</b>						
CLM	0.663	0.004	0.002	0.969	0.006	0.004
DM-CLM	0.667	0.005	0.003	0.977	0.008	0.005
ERY	0.576	0.004	0.002	0.973	0.006	0.004
RXM	0.764	0.004	0.002	0.977	0.006	0.004
AZM	0.865	0.004	0.002	0.984	0.007	0.004

**Table S3.** Physical-chemical properties of the target compounds.

Compound	Molecular weight (g mol <sup>-1</sup> )	p <i>K</i> <sub>a</sub>	Log <i>K</i> <sub>ow</sub>	Structure
<b>Roxithromycin (RXM)</b>	837.0	9.08 <sup>a</sup> , 12.45 <sup>a</sup>	2.75 <sup>c</sup>	
<b>Azithromycin (AZM)</b>	749.0	9.57 <sup>a</sup> , 12.43 <sup>a</sup>	4.02 <sup>c</sup>	
<b>Erythromycin (ERY)</b>	733.9	8.38 <sup>a</sup> , 12.44 <sup>a</sup>	3.06 <sup>c</sup>	
<b>Clarithromycin (CLM)</b>	747.9	8.38 <sup>a</sup> , 12.46 <sup>a</sup>	3.16 <sup>c</sup>	
<i>N</i> - desmethylclarithromycin (DM-CLM)	733.9	13.08 <sup>b</sup>	-	

Parent compounds are marked in bold; Abbreviations are written in brackets; -: Not data found. <sup>a</sup>: <https://go.drugbank.com/>; <sup>b</sup>: <https://www.chemicalbook.com/>; <sup>c</sup>: Li et al., 2018.

**Table S4.** Water sources characterisation.

<b>Water sources</b>	<b>pH</b>	<b>Cl<sup>-</sup> (M)</b>	<b>P<sub>T</sub> (mg/L)</b>	<b>COD (mg/L)</b>	<b>N<sub>T</sub> (mg/L)</b>	<b>σ (mS/cm)</b>
Tap water	6.59	8.2 E-4	<0.1	22.3	<20	0.44
Surface water	8.15	2.8 E-4	<0.17	31.0	<20	2.07
Effluent wastewater	8.99	2.5 E-3	0.42	32.0	21.1	0.94
Influent wastewater	9.20	2.5 E-3	2.98	171	35.9	1.11

COD: Chemical oxygen demand.

**Table S5.** Matrix correlation of macrolides and metabolite adsorption onto PE MPs with water sources physical-chemical characteristics.

Variable	pH	Cl <sup>-</sup> (M)	PO <sub>4</sub> <sup>3-</sup> (mg/L)	COD (mg/L)	N <sub>T</sub> (mg/L)	σ (mS/cm)	AZM Ads (%)	CLM Ads (%)	DM-CLM Ads (%)	ERY Ads (%)	RXM Ads (%)
pH	<b>1.00</b>	<b>0.99</b>	<b>0.62</b>	0.59	0.59	0.37	<b>-0.95</b>	<b>-0.97</b>	<b>-0.97</b>	<b>-0.89</b>	0.17
Cl <sup>-</sup> (M)		<b>1.00</b>	<b>0.60</b>	0.57	0.57	0.24	<b>-0.98</b>	<b>-0.98</b>	<b>-0.98</b>	<b>-0.90</b>	0.03
PO <sub>4</sub> <sup>3-</sup> (mg/L)			<b>1.00</b>	<b>1.00</b>	<b>1.00</b>	-0.03	<b>-0.69</b>	<b>-0.76</b>	<b>-0.75</b>	<b>-0.89</b>	-0.16
COD (mg/L)				<b>1.00</b>	<b>1.00</b>	0.02	<b>-0.65</b>	<b>-0.73</b>	<b>-0.73</b>	<b>-0.86</b>	-0.10
N <sub>T</sub> (mg/L)					<b>1.00</b>	-0.04	<b>-0.66</b>	<b>-0.73</b>	<b>-0.73</b>	<b>-0.87</b>	-0.17
σ (mS/cm)						<b>1.00</b>	-0.06	-0.20	-0.20	-0.07	<b>0.98</b>
AZM Ads (%)							<b>1.00</b>	<b>0.98</b>	<b>0.98</b>	<b>0.94</b>	0.15
CLM Ads (%)								<b>1.00</b>	<b>1.00</b>	<b>0.97</b>	0.01
DM-CLM Ads (%)									<b>1.00</b>	<b>0.97</b>	0.00
ERY Ads (%)										<b>1.00</b>	0.12
RXM Ads (%)											<b>1.00</b>

Ads: adsorption; COD: Chemical oxygen demand.

**Table S6.** Matrix correlation of macrolides and metabolite adsorption onto PET MPs with water sources physical-chemical characteristics.

Variable	pH	Cl <sup>-</sup> (M)	PO <sub>4</sub> <sup>3-</sup> (mg/L)	COD (mg/L)	N <sub>T</sub> (mg/L)	σ (mS/cm)	AZM Ads (%)	CLM Ads (%)	DM-CLM Ads (%)	ERY Ads (%)	RXM Ads (%)
pH	<b>1.00</b>	<b>0.99</b>	<b>0.62</b>	0.59	0.59	0.37	<b>-0.92</b>	<b>-0.98</b>	<b>-0.98</b>	<b>-0.97</b>	<b>-0.97</b>
Cl <sup>-</sup> (M)		<b>1.00</b>	<b>0.60</b>	0.57	0.57	0.24	<b>-0.96</b>	<b>-0.95</b>	<b>-0.94</b>	<b>-0.96</b>	<b>-0.94</b>
PO <sub>4</sub> <sup>3-</sup> (mg/L)			<b>1.00</b>	<b>1.00</b>	<b>1.00</b>	-0.03	<b>-0.75</b>	<b>-0.65</b>	-0.52	<b>-0.78</b>	<b>-0.75</b>
COD (mg/L)				<b>1.00</b>	<b>1.00</b>	0.02	<b>-0.71</b>	<b>-0.64</b>	-0.51	<b>-0.76</b>	<b>-0.74</b>
N <sub>T</sub> (mg/L)					<b>1.00</b>	-0.04	<b>-0.72</b>	<b>-0.62</b>	-0.49	<b>-0.76</b>	<b>-0.72</b>
σ (mS/cm)						<b>1.00</b>	0.00	-0.50	-0.55	-0.29	-0.41
AZM Ads (%)							<b>1.00</b>	<b>0.87</b>	<b>0.82</b>	<b>0.95</b>	<b>0.90</b>
CLM Ads (%)								<b>1.00</b>	<b>0.99</b>	<b>0.97</b>	<b>0.99</b>
DM-CLM Ads (%)									<b>1.00</b>	<b>0.93</b>	<b>0.95</b>
ERY Ads (%)										<b>1.00</b>	<b>0.99</b>
RXM Ads (%)											<b>1.00</b>

Ads: adsorption; COD: Chemical oxygen demand.

### LC-MS/MS determination

Chromatographic determination was performed using an Agilent 1290 Infinity II liquid chromatographic system (Agilent, USA) coupled to a 6495-triple quadrupole (QqQ) mass spectrometer (MS) equipped with an electrospray ionization source (ESI).

Chromatographic separation was carried out in a Zorbax RRHD Eclipse Plus C18 (150 mm × 3.0 mm i.d., 1.8 µm particle size) column (Agilent, Santa Clara, CA, USA), protected with a Zorbax RRHD Eclipse Plus C18 (3.0 mm i.d., 1.8 µm particle size) guard column (Agilent, Santa Clara, CA, USA) and thermostated at 35 °C. The mobile phase was composed by 10 mM ammonium formate buffer containing 0.05% of formic acid (solvent A) and methanol (solvent B). Elution was carried out at a flow rate of 0.4 mL min<sup>-1</sup>. Elution started with 5% of solvent B, held 1 min. Solvent B was linearly increased to 30% in 3 min, then to 60% in 8 min and, finally, to 100% in 2 min, held for 2 min. Back to initial conditions was carried out in 2 min and held for 2 min for equilibration. Total run time was 20 min. LC-MS/MS parameters of each compound is presented in Table S7.

The LC system was coupled to a 6410 triple quadrupole mass spectrometer (MS/MS) equipped with an electrospray ionisation source operated in positive mode. The following settings were used: fragmentor, 166 V; capillary voltage, 3000 V; nebuliser pressure, 40 psi; drying gas flow rate, 9 L min<sup>-1</sup> and gas temperature, 350 °C.

**Table S7.** LC-MS/MS parameters.

MS/MS parameters								
Compound	Ionization mode	Precursor ion ( <i>m/z</i> )	Product ions (MRM1/MRM2) ( <i>m/z</i> )	CE (V)	RT (min)	Ion ratio	LOD (ng/L)	LOQ (ng/L)
CLM	Positive	749.0	158.1/590.4	28/16	18.128	49.5	0.01	0.03
DM-CLM	Positive	734.9	144.1/576.4	144.1/576.4	18.161	23.5	0.02	0.06
ERY	Positive	734.5	83.0/576.4	83.0/576.4	18.140	81.4	0.02	0.06
RXM	Positive	838.1	158.1/679.4	32/20	18.214	72.9	0.01	0.03
AZM	Positive	750.0	591.5/116.1	28/44	18.138	56.1	0.01	0.03

CE: collision energy; RT: retention time; LOD: limit of detection; LOQ: limit of quantification



### Data analysis

The amount of adsorbed macrolide ( $q$ ) was calculated as the difference between its concentration before and after the adsorption experiment according to Eq. (S1):

$$(S1) \quad q = (C_i - C_e) \times V/m$$

where  $m$  is the MPs weight (g),  $V$  (L) is the solution volume,  $C_i$  (mg/L) and  $C_e$  (mg/L) are the concentrations of the macrolides at the beginning and at the end of the adsorption, respectively. Eq. (S2) was used to calculate adsorption percentage (%):

$$(S2) \quad \text{Adsorption (\%)} = (C_i - C_e)/C_i \times 100$$

Different mathematical models including Langmuir, Freundlich and Linear were evaluated to determine the adsorbent performance to experimental isotherms. Pseudo-first and pseudo-second order kinetic models were evaluated. Supplementary material, Table S8, shows a summary, including equations and parameter definition.

**Table S8.** Adsorption isotherms and kinetic models studied.

Isotherm	Model Equation
Langmuir	$q = \frac{q_{max} K_L C_e}{1 + (C_e K_L)}$ <p><math>C_e</math>: Equilibrium concentration of pollutants (mg/L);  <math>q</math>: Equilibrium adsorption capacity (mg/g);  <math>q_{max}</math>: maximum amount adsorbed within a monolayer (mg/g);  <math>K_L</math>: Langmuir dissociation constant (L/mg), which is related to the adsorption energy.</p>
Freundlich	$q = K_F C_e^{1/n}$ <p><math>C_e</math>: Equilibrium concentration of pollutants (mg/L);  <math>q</math>: Equilibrium adsorption capacity (mg/g);  <math>K_F</math>: Freundlich constant (L/mg), which is related to the affinity of the adsorbent to the adsorbate;  <math>1/n</math>: dimensionless parameter, which indicates how adsorption varies as a function of the concentration.</p>
Linear	$q = K_d \times C_e$ <p><math>C_e</math>: Equilibrium concentration of pollutants (mg/L);  <math>q</math>: Equilibrium adsorption capacity (mg/g);  <math>K_d</math>: solution-soil distribution coefficient (L/g).</p>
Kinetic	Model equation
Pseudo-first order (PFO)	$\ln (q_e - q_t) = \ln q_e - k_1 \times t$

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$q_e$  and  $q_t$ : amounts of compounds (mg/g) adsorbed at equilibrium and at a  $t$  time;

$t$ : time (min);

$k_1$ : PFO kinetic constants.

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Pseudo-second order (PSO)

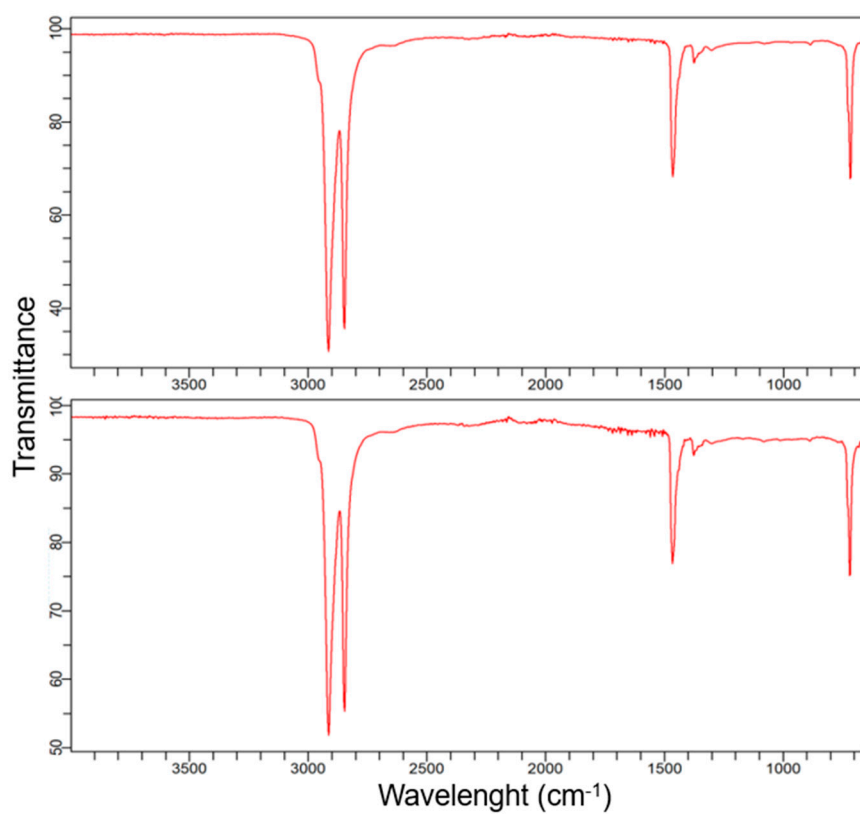
$$t/q_t = 1/(k_2 \times q_e^2) + t/q_e$$

$q_e$  and  $q_t$ : amounts of compounds (mg/g) adsorbed at equilibrium and at a  $t$  time;

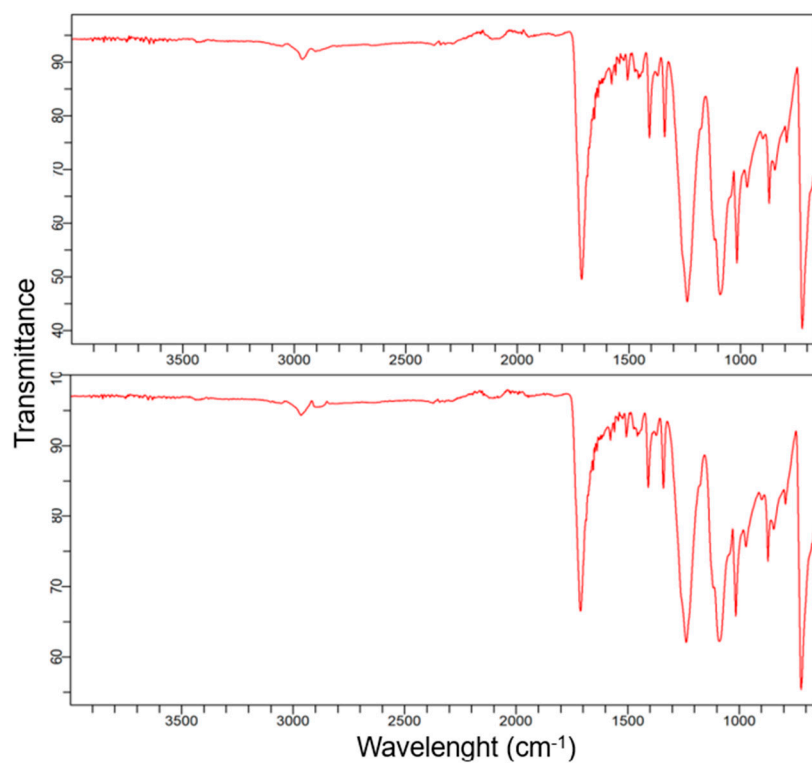
$t$ : time (min);

$k_2$ : PSO kinetic constants.

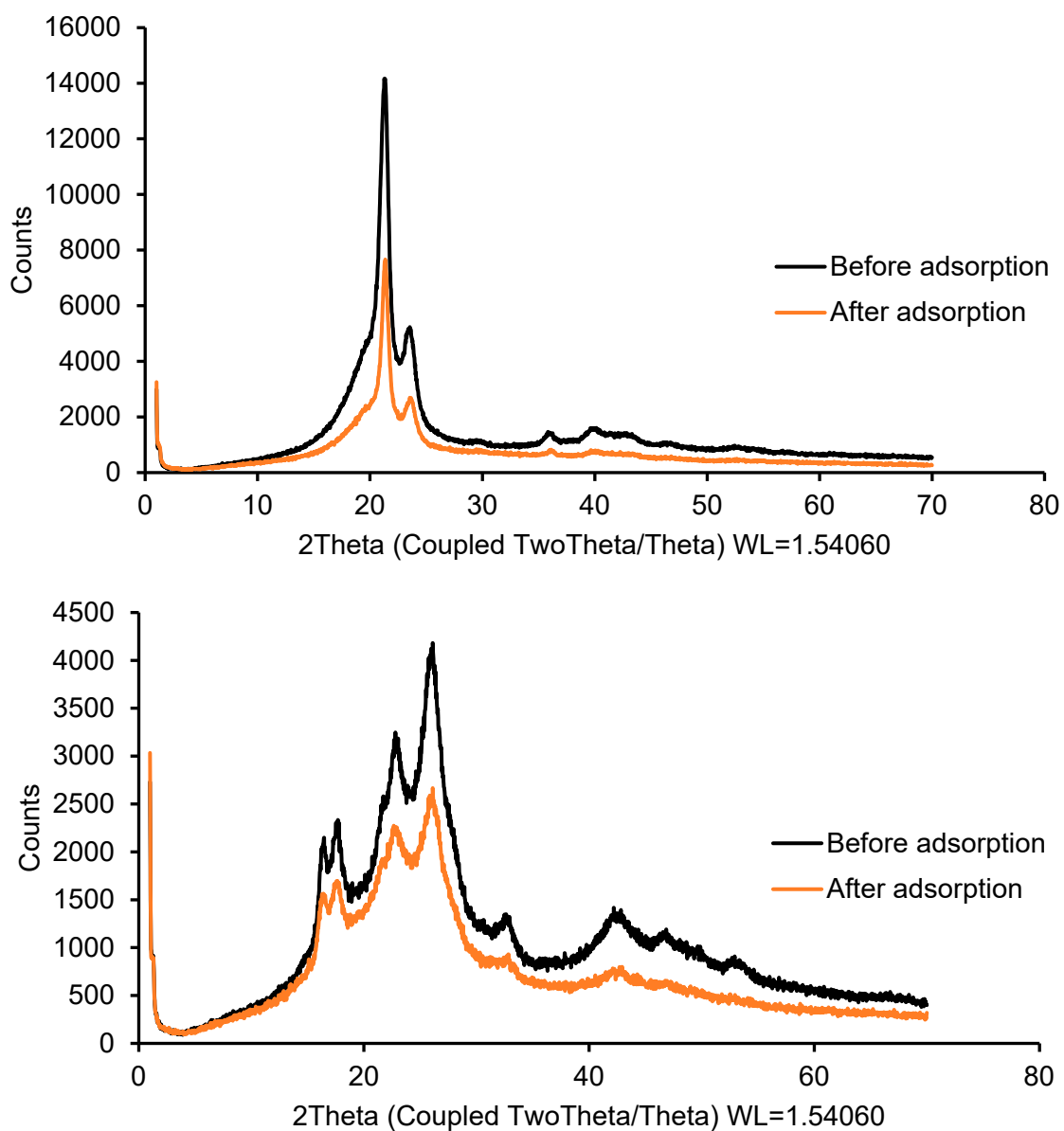
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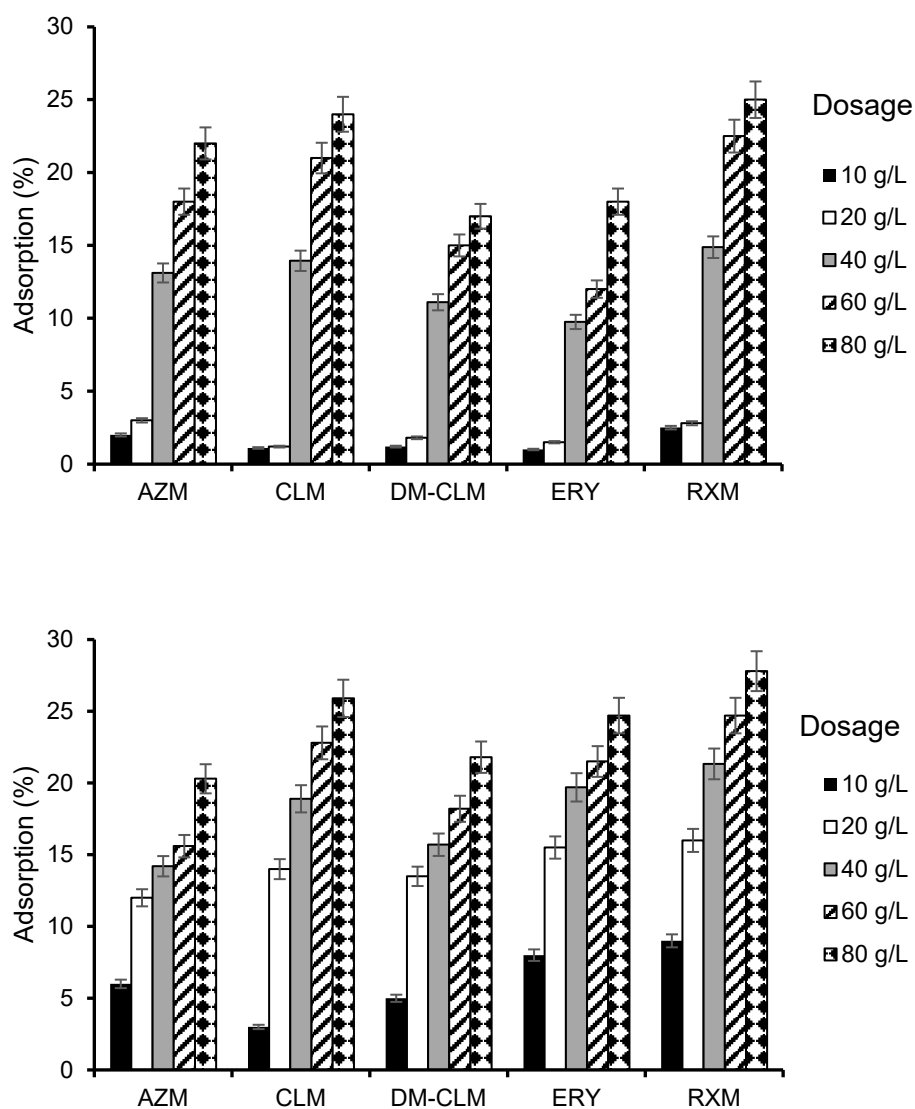
**Figure S1.** FTIR analysis of PE before (upper) and after (down) the adsorption process.



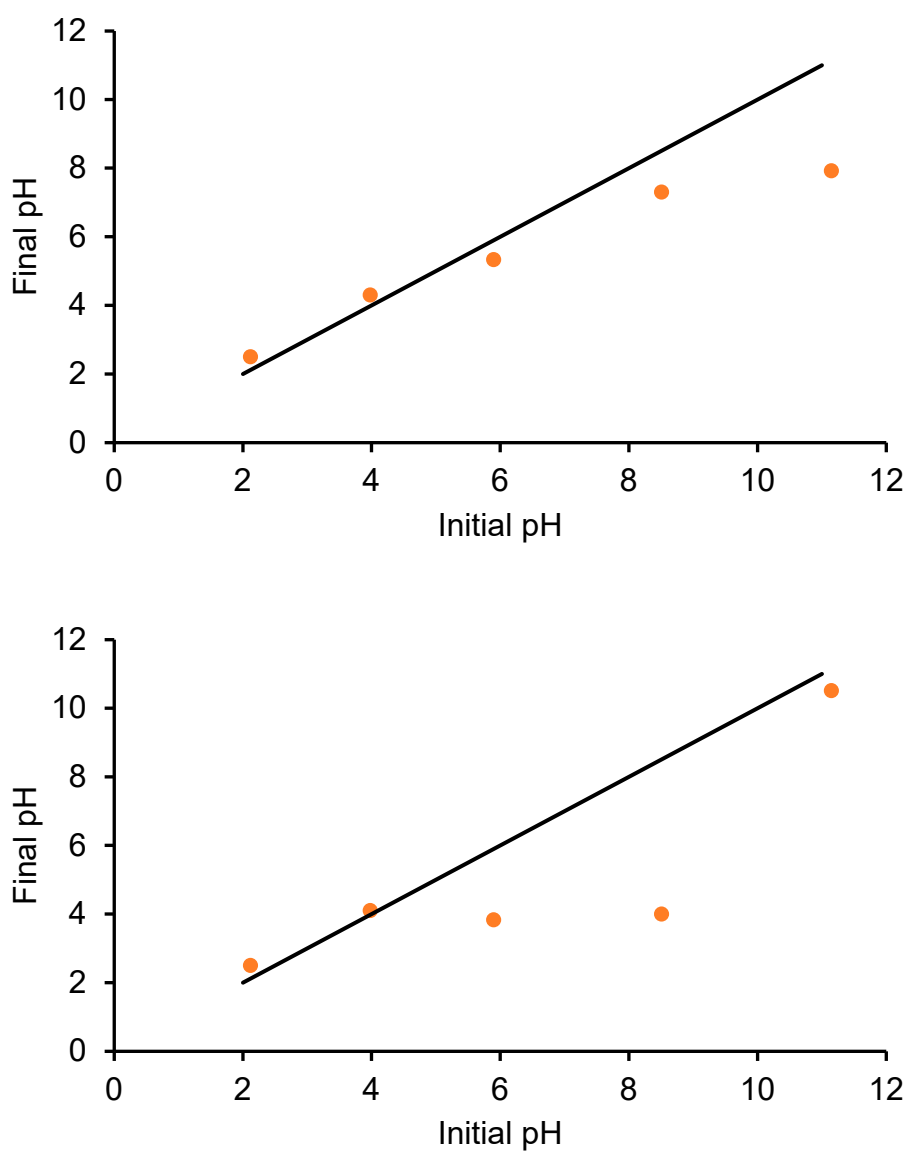
**Figure S2.** FTIR analysis of PET before (upper) and after (down) the adsorption process.



**Figure S3.** XRD analysis for PE before and after the adsorption process (upper) and for PET before and after the adsorption process (down).



**Figure S4.** Adsorption percentage (%) of macrolides and metabolite on PE (upper) and PET (down) MPs in the evaluation of microplastic dosage.



**Figure S5.** Final pH with PE after 48h of agitation versus initial pH without PE (upper) and final pH with PET after 48h of agitation versus initial pH without PET (down).

#### Supplementary material references

Chemical Book. Sourcing and Integrating Center of Chemicals Materials in China. Available on: <https://www.chemicalbook.com/>

DrugBank Online. Database for Drug and Drug Target Info. Available on: <https://go.drugbank.com/>