

Antimicrobial resistance profiles and genetic typing of *Salmonella* serovars from chicken embryos in China

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Supplementary Figures

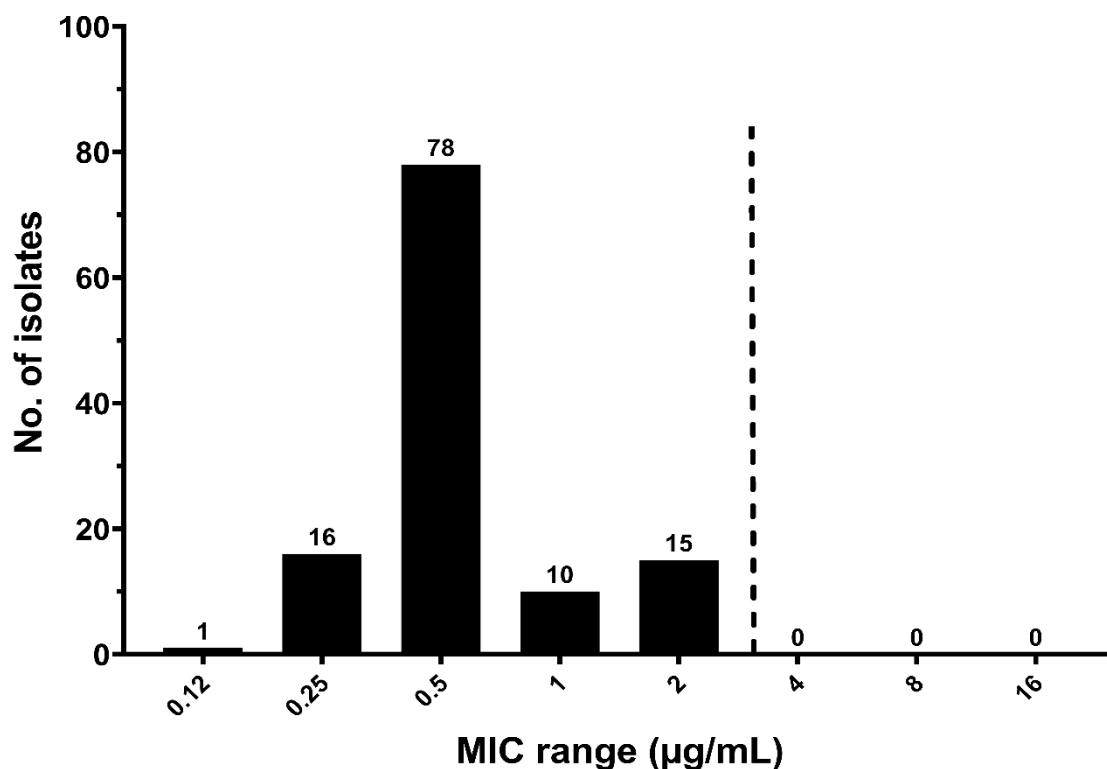


Figure S1. The distribution of minimum inhibitory concentration (MIC) values among the examined *Salmonella* isolates (n=120) against Colistin (CST). The dashed line indicates the cutoff level of the MIC, in which the lowest values correspond to susceptibility and the highest values correspond to the resistance.

Supplementary Tables

Table S1: Primers used for the detection of antimicrobial resistance genes.

Target	Forward primer (5'–3')	Reverse primer (5'–3')	Annealing Temperature (°C)	Product size (bp)	References
<i>β</i>-Lactamases and ESBLs:					
<i>bla</i> _{TEM}	GTATTCAACATTTCCGTGTCG	CCAATGCTTAATCAGTGAGGC	55	854	[1]
<i>bla</i> _{SHV}	CCCTGTTAGCCACCCTGCCG	CGTTGCCAGTGCTCGATCAGC	68	829	[1]
<i>bla</i> _{CTX-M-1}	GCTGTTGTTAGGAAGTGTGCCGC	GCCGCCGACGCTAATACATC	63	798	[1]
<i>bla</i> _{CTX-M-2}	AAATGTGCTGCTCCTTTCGTGAGC	AGGGTTCGTTGCAAGACAAGACTG	55	1122	[2]
<i>bla</i> _{CTX-M-3}	GTTACAATGTGTGAGAAGCAG	CCGTTTCCGCTATTACAAAC	55	1018	[3]
<i>bla</i> _{CTX-M-8}	TAACGCACAGACGCTCTACC	TGGCTGGGTGAAGTAAGTGAC	55	637	[4]
<i>bla</i> _{CTX-M-9}	GATCAGTCAGTGGGATAGTTT	TACTCGGCGTTAACTGATTA	47	671	[5]
<i>bla</i> _{CTX-M-25}	GCGATGTTAATGACGACAGC	AACCGTCGGTGACAATTCTG	55	847	[4]
<i>bla</i> _{OXA-1}	ACACAATACATATCAACTTCGC	AGTGTGTTTAGAATGGTGATC	55	814	[3]
<i>bla</i> _{OXA-2}	TTCAAGCCAAAGGCACGATAG	TCCGAGTTGACTGCCGGGTTG	60	703	[3]
<i>bla</i> _{OXA-10}	CGTGCTTTGTAAAAGTAGCAG	CATGATTTTGGTGGGAATGG	55	389	[3]
<i>bla</i> _{PSE}	TGCTTCGCAACTATGACTAC	AGCCTGTGTTTGAGCTAGAT	55	438	[3]
Cephalosporins:					
<i>bla</i> _{MOX}	GCTGCTCAAGGAGCACAGGAT	CACATTGACATAGGTGTGGTGC	55	520	[6]
<i>bla</i> _{CIT}	TGGCCAGAACTGACAGGCCAAA	TTTCTCCTGAACGTGGCTGGC	55	462	[6]
<i>bla</i> _{DHA}	AACTTTCACAGGTGTGCTGGGT	CCGTACGCATACTGGCTTTGC	55	405	[6]
<i>bla</i> _{ACC}	AACAGCCTCAGCAGCCGGTTA	TTCGCCGCAATCATCCCTAGC	55	346	[6]
<i>bla</i> _{EBC}	TCGGTAAAGCCGATGTTGCGG	CTTCCACTGCGGCTGCCAGTT	55	302	[6]
<i>bla</i> _{FOX}	AACATGGGGTATCAGGGAGATG	CAAAGCGCGTAACCGGATTGG	55	190	[6]
Aminoglycosides:					
<i>armA</i>	CAAATGGATAAGAATGATGTT	TTATTTCTGAAATCCACT	55	777	[7]
<i>rmtA</i>	ATGAGCTTTGACGATGCCCTA	TCACTTATTCCTTTTATCATG	55	756	[7]
<i>rmtB</i>	ATGAACATCAACGATGCCCT	CCTTCTGATTGGCTTATCCA	55	769	[7]
<i>rmtC</i>	CGAAGAAGTAACAGCCAAAG	ATCCCAACATCTCTCCCACT	55	711	[7]
<i>rmtD</i>	CGGCACGCGATTGGGAAGC	CGGAAACGATGCGACGAT	55	401	[7]
<i>rmtE</i>	ATGAATATTGATGAAATGGTTGC	TGATTGATTTCCTCCGTTTTTG	55	819	[7]
<i>npmA</i>	CTCAAAGGAACAAAGACGG	GAAACATGGCCAGAAACTC	55	641	[8]
<i>aac(3)-IV</i>	GTTACACCGGACCTTGGA	AACGGCATTGAGCGTCAG	55	674	[9]

<i>aac6'-Ib</i>	TTGCGATGCTCTATGAGTGGCTA	CTCGAATGCCTGGCGTGTTT	55	482	[10]
<i>aadA1-like</i>	GTGGATGGCGGCCTGAAGCC	ATTGCCCAGTCGGCAGCG	55	527/1916	[9]
Sulphonamides:					
<i>sul1</i>	TGGCGTCGCGACTGCGAAAT	TGGTGACGGTGTTTCGGCATTCT	55	813	[11]
<i>sul2</i>	GTTTCTCCGATGGAGGCCGGT	AGCGAGGTTTCGGGAGCAGC	55	517	[11]
<i>sul3</i>	GATAGTTTTTCCGATGGAGG	GAAGCCCATAACCCGGATCAA	55	495	[11]
Tetracyclines:					
<i>tetA</i>	TGGTCCGGAGGCCAGACGTG	TTCCGAGCATGAGTGCCCGC	55	867	[11]
<i>tetG</i>	TCTTGCAGGAGCCGCAGTCGAT	GGCCGGCATGCCAACACCC	55	721	[11]
<i>tetX</i>	CAATAATTGGTGGTGGACCC	TTCTTACCTTGGACATCCCG	55	468	[11]
<i>tetB</i>	AAATAACAGCAAACAGTAATGG	AAGTAGGGGTTGAGACGCAGCTA	55	493	[12]
<i>tetC</i>	TCTAACAATGCGCTCATCGT	GGTTGAAGGCTCTCAAGGGC	55	589	[12]
<i>tetR</i>	AGGACGACGGTGCTTGCT	ATGAGGACTGGCGGGTGT	55	295	[12]

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