

Table S1. Snake venom components as novel drug candidates to eliminate drug resistant bacteria.

No.	Bacteria strains	Toxins or enzymes	Snake	References
1	MRSA, MDR <i>Escherichia coli</i> , MDR <i>Pseudomonas aeruginosa</i>	OH-CATH30 and OH-CM6 (cathelicidins)	<i>Ophiophagus hannah</i>	[37]
2	MRSA and VRSA	CATHPb1 (cathelicidin)	<i>Python bivittatus</i>	[38]
3	MDR <i>Acinetobacter baumannii</i> and MRSA	OH-CATH30 (cathelicidins)	<i>Ophiophagus hannah</i>	[39]
4	MRSA	Cath-A (cathelicidin)	<i>Bungarus fasciatus</i>	[40]
5	MDR <i>Burkholderia pseudomallei</i>	Crotoxin B and daboia toxin (phospholipase A2)	<i>Crotalus durissus terrificus, Daboia russelli siamensis</i>	[41]
6	MDR <i>B. pseudomallei</i>	metalloproteinase	<i>Agkistrodon halys</i>	[42]
7	MDR <i>B. pseudomallei</i>	VipTx-II (phospholipase A2)	<i>Daboia russelli russelli</i>	[43]
8	MDR <i>Mycobacterium tuberculosis</i>	vgf-1 peptide	<i>Naja atra</i>	[44]
9	MRSA	pC-CoaTxII phospholipase A2	<i>Crotalus oreganus abyssus</i>	[45]

MRSA: methicillin-resistant *Staphylococcus aureus*; VRSA: vancomycin-resistant *Staphylococcus aureus*; MDR: multi-drug-resistant.

Table S2. Anticancer properties of snake venom and its components.

No.	Cell lines	Toxins or enzymes	Snake	References
Metastasis inhibition				
1	human melanoma M24met	contortrostatin (disintegrin)	<i>Agkistrodon contortrix contortrix</i>	[46]
2	human breast cancer MDA-MB-435	contortrostatin (disintegrin)	<i>Agkistrodon contortrix contortrix</i>	[47]
3	human glioblastoma T98G, U87, A172, and U138	contortrostatin (disintegrin)	<i>Agkistrodon contortrix contortrix</i>	[48]
4	human hepato-carcinoma HepG2 and SMMC-7721	agkihpin (arginine esterase)	<i>Gloydius halys Pallas</i>	[49]
5	human gingival carcinoma Ca9-22	cardiotoxin III	<i>Naja naja atra</i>	[50]
6	human breast cancer MDA-MB-231	Vaa-Dis (disintegrin)	<i>Vipera ammodytes ammodytes</i>	[51]
Induction of apoptosis				
7	human gastric cancer MKN-45, colorectal	l-amino acid oxidases (LAAO)	<i>Bothrops leucurus</i>	[52]

No.	Cell lines	Toxins or enzymes	Snake	References
	cancer HUTU, colon carcinoma RKO			
8	human ovarian cancer CAOV3 and OVCAR3	LAAO	<i>Crotalus adamanteus</i>	[1]
9	human colorectal cancer LoVo, HTC-116, and HT-29	SVT	<i>Echis coloratus</i>	[54]
10	human leukemia HL60, JURKAT, murine melanoma B16F10	LAAO	<i>Bothrops atrox</i>	[55]
11	human prostate cancer PC-3 and DU145	SVT	<i>Vipera lebetina turanica</i>	[56]
12	human breast cancer MCF-7	rusvinoxidase (LAAO)	<i>Daboia russelii russelii</i>	[57]
13	human colon adenocarcinoma HCT116 and HT-29	SVT	<i>Vipera lebetina turanica</i>	[58]
14	human teratocarcinoma PA-1 and human ovarian cancer SK-OV3	SVT	<i>Vipera lebetina turanica</i>	[59]
15	human neuroblastoma SK-N-MC and SK-N-SH	SVT	<i>Vipera lebetina turanica</i>	[60]
16	human lung cancer A549	ACTX-6 (cytotoxin)	<i>Agkistrodon acutus</i>	[61]
17	colorectal adenocarcinoma LS174, HCT116, and HT-29	lebein (disintegrin)	<i>Macrovipera lebetina</i>	[62]
18	human leukemia HL-60 and hepatocellular carcinoma HepG2	CR-LAAO	<i>Calloselasma rhodostoma</i>	[63]
19	HL-60 and HepG2	BthTX-I (PLA2s)	<i>Bothrops jararacussu</i>	[64]
20	HepG2, MCF-7, and DU145	SVT	<i>Naja naja oxiana</i>	[65]
21	HCT-8, MDA-MB-231	SVT	<i>Bitis arietans, C. gasperettii, E. coloratus, and Echis pyramidum</i>	[66]
22	human endo-cervical carcinoma Ca Ski, C33A, A549, MCF-7	SVT	<i>Vipera lebetina turanica</i>	[67]
23	human cervical cancer SiHa and HeLa	SVT	<i>Bothrops jararaca and Bothrops erythromelas</i>	[68]
24	MCF7	Phospholipase A2 (PLA2)	<i>Bungarus fasciatus</i>	[69]
25	HeLa	Pllans-II (PLA2)	<i>Porthidium lansbergii lansbergii</i>	[70]
26	human melanoma SK-MEL-28 and LU-1205	lebein (disintegrin)	<i>Macrovipera lebetina</i>	[71]
27	MCF-7, SK-BR-3, and MDA-MB-231	BthTX-I (PLA2)	<i>Bothrops jararacussu</i>	[72]
28	MCF-7	SVT	<i>Cerastes cerastes and Vipera lebetina</i>	[73]

No.	Cell lines	Toxins or enzymes	Snake	References
29	HCT-116, SW-480, and HT-29	LAAO	<i>Crotalus adamanteus</i>	[74]
Cell adhesion inhibition				
30	human fibrosarcoma HT1080, IGR39, HT29-D4, K562	MVL-PLA2	<i>Macrovipera lebetina transmediterranea</i>	[75]
31	human melanoma SK-Mel-28	colombistatin (disintegrin)	<i>Bothrops colombiensis</i>	[76]
32	human melanoma A-375 and A-549	tzabcanin (disintegrin)	<i>Crotalus simus tzabcan</i>	[77]
<i>Anti-proliferative activities</i>				
33	MCF-7 and A549	LAAO	<i>Ophiophagus hannah</i>	[78]
34	glioblastoma HCB151	L-amino acid oxidase (LAAOcdt)	<i>Crotalus durissus terrificus</i>	[79]
35	ovarian carcinoma MDAH 2774	saxatilin (disintegrin)	<i>Gloydius saxatilis</i>	[80]
36	Hep2 and HeLa	SVT	<i>Lapemis curtus</i>	[81]
37	HeLa	disintegrin	<i>Echis multiquamatus</i>	[82]
38	HeLa and Hep2	SVT	<i>Hydrophis spiralis</i>	[83]
39	MCF-7 and MDA-MB-231	SVT	<i>Vipera raddei kurdistanica</i>	[84]
Inhibition of angiogenesis				
40	lung cancer NCI-H460	saxatilin (disintegrin)	<i>Gloydius saxatilis</i>	[85]
41	HT-1080 and MDA-MB-231	vixapatin (C-type lectin-protein)	<i>Vipera xantina palestinae</i>	[86]
42	MDA-MB-231	bothropoidin (metalloproteinase)	<i>Bothrops pauloensis</i>	[87]

Table S3. Elapidae and Viperidae families as representatives of the well-characterized proteomics of snake venoms.

No.	Snake	References
Elapidae		
1	<i>Aipysurus laevis</i>	[135]
2	<i>Austrelaps labialis</i>	[136]
3	<i>Bungarus caeruleus</i>	[137]
4	<i>Bungarus candidus</i>	[138]
5	<i>Bungarus fasciatus</i>	[138,139]
6	<i>Bungarus multicinctus</i>	[139,140]
7	<i>Dendroaspis angusticeps</i>	[141]
8	<i>Dendroaspis polylepis</i>	[142]
9	<i>Drysdalia coronoides</i>	[143]
10	<i>Hydrophis cyanocinctus</i>	[144]
11	<i>Hydrophis schistosus</i>	[145]
12	<i>Micrurus aleni</i>	[146]
13	<i>Micrurus altirostris</i>	[147]
14	<i>Micrurus fulvius</i>	[148]
15	<i>Micrurus mipartitus</i>	[149]
16	<i>Micrurus mosquitensis</i>	[146]
17	<i>Micrurus nigrocinctus</i>	[150]
18	<i>Micrurus tschudii tschudii</i>	[151]
19	<i>Naja atra</i>	[140,152]
20	<i>Naja haje</i>	[153]

No.	Snake	References
21	<i>Naja kaouthia</i>	[154,155]
22	<i>Naja katiensis</i>	[156]
23	<i>Naja melanoleuca</i>	[157]
24	<i>Naja mossambica</i>	[156]
25	<i>Naja naja</i>	[158,159]
26	<i>Naja nigricollis</i>	[156]
27	<i>Naja nubiae</i>	[156]
28	<i>Naja pallida</i>	[156]
29	<i>Oxyuranus scutellatus</i>	[160]
30	<i>Pelamis platura</i>	[161]
31	<i>Pseudechis papuanus</i>	[162]
32	<i>Toxicocalamus longissimus</i>	[144]
Viperidae		
33	<i>Agkistrodon contortrix</i>	[163,164]
34	<i>Atropoides nummifer</i>	[165]
35	<i>Atropoides picadoi</i>	[165]
35	<i>Bitis caudalis</i>	[166]
37	<i>Bitis gabonica</i>	[166]
38	<i>Bitis nasicornis</i>	[166]
39	<i>Bothriechis aurifer</i>	[168]
40	<i>Bothriechis bicolor</i>	[168]
41	<i>Bothriechis lateralis</i>	[168]
42	<i>Bothriechis marchi</i>	[168]
43	<i>Bothriechis nigroviridis</i>	[167]
44	<i>Bothriechis schlegelii</i>	[168]
45	<i>Bothriechis thalassinus</i>	[168]
46	<i>Bothrocophias campbelli</i>	[169]
47	<i>Bothropoides pauloensis</i>	[170]
48	<i>Bothrops atrox</i>	[171–175]
49	<i>Bothrops ayerbeai</i>	[176]
50	<i>Bothrops barnetti</i>	[172]
51	<i>Bothrops caribbaeus</i>	[177]
52	<i>Bothrops colombiensis</i>	[178]
53	<i>Bothrops cotiara</i>	[179]
54	<i>Bothrops diporus</i>	[180]
55	<i>Bothrops erythromelas</i>	[181]
56	<i>Bothrops fonsecai</i>	[179]
57	<i>Bothrops insularis</i>	[182]
58	<i>Bothrops jararaca</i>	[174,183]
59	<i>Bothrops lanceolatus</i>	[177]
60	<i>Bothrops neuwiedi</i>	[174]
61	<i>Bothrops pictus</i>	[172]
62	<i>Bothrops pirajai</i>	[184]
63	<i>Bothrops punctatus</i>	[185]
64	<i>Calloselasma rhodostoma</i>	[186]
65	<i>Cerastes cerastes</i>	[187]
66	<i>Cerrophidion godmani</i>	[188]
67	<i>Crotalus adamanteus</i>	[189]
68	<i>Crotalus atrox</i>	[209]
69	<i>Crotalus basiliscus</i>	[190]
70	<i>Crotalus durissus</i>	[191,192]
71	<i>Crotalus horridus</i>	[193]

No.	Snake	References
72	<i>Crotalus simus</i>	[194]
73	<i>Crotalus tigris</i>	[210]
74	<i>Crotalus viridis</i>	[195]
75	<i>Daboia russelii</i>	[196–198]
76	<i>Gloydius brevicauda</i>	[199]
77	<i>Gloydius intermedius</i>	[200]
78	<i>Lachesis acrochorda</i>	[201]
79	<i>Lachesis melanocephala</i>	[201]
80	<i>Lachesis muta</i>	[211]
81	<i>Lachesis stenophrys</i>	[201]
82	<i>Macrovipera lebetinus</i>	[212]
83	<i>Macrovipera mauritanica</i>	[202]
84	<i>Ovophis okinavensis</i>	[203]
85	<i>Porthidium lansbergii</i>	[204]
86	<i>Porthidium nasutum</i>	[188]
87	<i>Porthidium ophryomegas</i>	[188]
88	<i>Protobothrops flavoviridis</i>	[203]
89	<i>Protobothrops mucrosquamatus</i>	[205]
90	<i>Sistrurus catenatus</i>	[213]
91	<i>Tropidolaemus wagleri</i>	[206]
92	<i>Vipera anatolica</i>	[208]
93	<i>Vipera nikolskii</i>	[207]
94	<i>Vipera orlovi</i>	[207]
95	<i>Vipera renardi</i>	[207]

Table S4. List of snake species for which transcriptomics libraries are available.

No.	Snake	References
Colubridae		
1	<i>Dispholidus typus</i>	[5,224]
2	<i>Liophis miliaris</i>	[224]
3	<i>Liophis poecilogyrus</i>	[5,224]
4	<i>Philodryas olfersii</i>	[5,224,225]
5	<i>Rhabdophis tigrinus</i>	[5,224]
6	<i>Telescopus dhara</i>	[5,224]
7	<i>Thamnodynastes strigatus</i>	[226]
8	<i>Thrasops jacksonii</i>	[5]
9	<i>Trimorphodon biscutatus</i>	[5,224]
Elapidae		
10	<i>Austrelaps labialis</i>	[136]
11	<i>Bungarus flaviceps</i>	[227]
12	<i>Bungarus multicinctus</i>	[228]
13	<i>Demansia vestigiata</i>	[229]
14	<i>Drysdalia coronoides</i>	[143]
15	<i>Hydrophis curtus</i>	[230]
16	<i>Micropoecilis ikaheca</i>	[231]
17	<i>Micrurus altirostris</i>	[147]
18	<i>Micrurus corallinus</i>	[147,232]
19	<i>Micrurus fulvius</i>	[148]
20	<i>Micrurus mipartitus</i>	[233]

No.	Snake	References
21	<i>Naja atra</i>	[228]
22	<i>Oxyuranus microlepidotus</i>	[5]
Homalopsidae		
23	<i>Cerberus rynchops</i>	[234]
24	<i>Enhydris polylepis</i>	[224]
Viperidae		
25	<i>Agkistrodon piscivorus</i>	[235]
26	<i>Atropoides mexicanus</i>	[236]
27	<i>Atropoides picadoi</i>	[236]
28	<i>Azemiops feae</i>	[5,224]
29	<i>Bitis gabonica</i>	[237]
30	<i>Bothriechis lateralis</i>	[236]
31	<i>Bothriechis schlegelii</i>	[236]
32	<i>Bothropoides pauloensis</i>	[170]
33	<i>Bothrops alternatus</i>	[238]
34	<i>Bothrops asper</i>	[236]
35	<i>Bothrops atrox</i>	[239]
36	<i>Bothrops insularis</i>	[182,222]
37	<i>Bothrops jararaca</i>	[240,241]
38	<i>Causus rhombeatus</i>	[5]
39	<i>Cerrophidion godmani</i>	[236]
40	<i>Crotalus adamanteus</i>	[189,242]
41	<i>Crotalus durissus</i>	[243,244]
42	<i>Crotalus horridus</i>	[193]
43	<i>Crotalus simus</i>	[236,245]
44	<i>Deinagkistrodon acutus</i>	[246]
45	<i>Echis carinatus</i>	[247]
46	<i>Echis coloratus</i>	[247]
47	<i>Echis ocellatus</i>	[248,249]
48	<i>Echis pyramidum</i>	[247]
49	<i>Lachesis muta</i>	[250]
50	<i>Sistrurus catenatus</i>	[251]
51	<i>Vipera ammodytes</i>	[252]

Table S5. List of promising antivenom molecules.

No.	Antivenom molecules	Toxins or enzymes	Snake	References
Monoclonal antibody (MA)				
1	MA	toxin alpha	<i>Naja nigricollis</i>	[259]
2	MA	metalloproteinase HR1a	<i>Probothrops flavoviridis</i>	[260]
3	MA	dendrotoxins	<i>Dendroaspis polylepis</i>	[261]
4	MA	crototoxin	<i>Crotalus durissus terrificus</i>	[262]
5	MA	Phospholipases A2	<i>Bothrops jararacussu</i>	[263]
6	MA	alpha-neurotoxin	<i>Naja kaouthia</i>	[264]
7	MA	unspecified	<i>Bothrops jararacussu, Crotalus durissus</i>	[265]
Small molecule inhibitors				
8	varespladib (LY315920)	phospholipase A2	<i>Micruurus fulvius, Vipera berus</i>	[266]
9	LY333013	phospholipase A2	<i>Oxyuranus scutellatus</i>	[267]
10	batimastat and mari-mastat	metalloproteinase	<i>Echis ocellatus</i>	[268]

No.	Antivenom molecules	Toxins or enzymes	Snake	References
11	varespladib	phospholipase A2	<i>Deinagkistrodon acutus</i> , <i>Agkistrodon halys</i> , <i>Bungarus multicinctus</i> , and <i>Naja atra</i>	[269]
12	batimastat	metalloproteinases	<i>Bothrops asper</i>	[270]
13	thiosemicarbazones	metalloproteinases	<i>Bothrops pauloensis</i>	[271]
Nanoparticles				
14	C60 fullerene	unspecified	<i>Crotalus oreganusshelleri</i>	[272]
Antibody fragments				
15	llama single-domain antibodies VHHs (C2 and C20) and VHH-Fc	α -cobratoxin	<i>Naja kaouthia</i>	[273]
16	humanized-single domain antibodies VH/VHH	phospholipase A2	<i>Naja kaouthia</i>	[274]
17	camelid Single-Domain Antibodies (VHHs)	crotoxin	<i>Crotalus durissus terrificus</i>	[275]
18	recombinant single chain antibody, scFv	metalloproteinase	<i>Bothrops asper</i>	[276]
19	single-chain variable fragment (scFv)	unspecified	<i>Agkistrodon acutus</i>	[277]