



## Supplementary Material

**Table S1.** Comparison of *Toxoplasma gondii* results for heart fluid samples using serological methods (ELISA and IFAT) relative to magnetic capture real-time PCR to determine the best dilution for cut-off values..

Dilutions	ID	ELISA S/P%				IFAT			
		ND	1:2	1:4	1:8	1:2	1:8	1:32	1:128
N	19	33.65	7.76	7.59	2.62	N	N	N	N
	31	112.34	97.30	69.49	46.34	P	P	N	N
	45	24.89	12.16	11.51	2.62	N	N	N	N
LP	2	124.15	109.98	85.92	67.26	P	P	P	N
	16	114.56	121.93	106.50	87.53	P	P	P	P
	17	46.86	59.07	56.97	50.78	P	P	P	P
HP	7	133.22	131.56	109.29	81.69	P	P	P	N
	18	150.35	174.59	163.30	142.89	P	P	P	P
	24	211.51	216.04	198.30	182.87	P	P	P	P

Positive results: ELISA  $\geq 40\%$  and IFAT  $\geq 1+$ ; N: negative; LP: low positive on magnetic capture ( $C_p$  value  $> 30$ ); HP: high positive on magnetic capture ( $C_p$  value  $\leq 30$ ); ND: no dilution; ELISA: enzyme-linked immunosorbent assay; IFAT: indirect fluorescent antibody test.

**Table S2.** Comparison of optimized serological methods (ELISA and IFAT) for *Toxoplasma gondii* relative to magnetic capture qPCR.

	MC-qPCR		
	Positive	Negative	Total
ELISA			
Positive	16	6	22
Negative	0	38	38
Total	16	44	60
McNemar test ( $p$ -value)	0.041 *		
Relative sensitivity	100		
Relative specificity	86.36		
IFAT			
Positive	16	11	27
Negative	0	33	33
Total	16	44	60
McNemar test ( $p$ -value)	0.003*		
Relative sensitivity	100		
Relative specificity	75		

MC-qPCR: magnetic capture qPCR; ELISA: enzyme-linked immunosorbent assay; IFAT: indirect fluorescent antibody test; \*: significant  $p$ -value  $< 0.05$ ; Note: A subset of randomly chosen lynx ( $n = 60$ ) were tested.