


Proceedings

Synthesis and Evaluation of New Nitron-Based Benzoxazin Antioxidants [†]

Stefania Marano ¹, Cristina Minnelli ², Giovanna Mobbili ², Emiliano Laudadio ¹ and Pierluigi Stipa ^{1,*}

¹ Dipartimento S.I.M.A.U., Università Politecnica delle Marche, via Brecce Bianche, 60131 Ancona, Italy; s.marano@staff.univpm.it (S.M.); e.laudadio@staff.univpm.it (E.L.)

² Dipartimento di Scienze della Vita e dell'Ambiente (DISVA), Università Politecnica delle Marche, via Brecce Bianche, 60131 Ancona, Italy; c.minnelli@staff.univpm.it (C.M.); g.mobbili@staff.univpm.it (G.M.)

* Correspondence: p.stipa@staff.univpm.it; Tel.: +39-071-2204409

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Abstract: Oxidative stress is often the cause of a wide range of human chronic pathologies including, but not limited to, stroke and cardiovascular and neurodegenerative diseases. In this setting, increasing efforts are currently being devoted to the design and synthesis of new derivatives with enhanced antioxidant efficacy. Among all the potential molecules of interest, synthetic nitron spin-traps have attracted a great deal of research attention, particularly due to their dual function as effective inhibitors of oxidative stress and damage and as analytical tools for the detection and characterization of free radicals by means of the electron paramagnetic resonance (EPR) spectroscopy spin trapping technique. In this study, two derivatives of benzoxazin nitrones (3-aryl-2H-benzo[1,4]oxazin-N-oxides) bearing an electron-withdrawing methyl-acetate group on the benzo moiety (in *para* and *meta* positions with respect to the nitronyl function) were synthesized. Their in vitro antioxidant activity was evaluated by means of the α,α -diphenyl- β -picrylhydrazyl radical (DPPH) scavenging assay, and their inhibitory effects on the erythrocyte hemolysis induced by the water-soluble free radical initiator 2,2'-azobis(2-amidinopropane) dihydrochloride (AAPH) compared. In addition, EPR was employed to monitor the decay profiles of DPPH to evaluate the kinetic behavior of the different antioxidants tested. Results showed that the presence and the position of the electron-withdrawing methyl-acetate group strongly affects the radical scavenging activity of nitrones. In particular, the newly synthesized *para*-substituted derivative, when compared to both the *meta*-substituted isomer and the unsubstituted parent compound, acts as a more effective antioxidant both in cell and cell-free systems. Overall, these results clearly show that the introduction of an electron-withdrawing group on the phenyl moiety significantly increased the antioxidant capacity of benzoxazin nitrones, thus showing exciting opportunities in the search for new therapeutic agents in the treatment of diseases associated with oxidative stress.

Keywords: nitron; DPPH; antioxidant; electron paramagnetic resonance



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1. Introduction

The ability of nitrones to act as good radical traps has been widely researched over the last few decades [1–5]. For years, nitrones have been used as efficient analytical tools for the detection and characterization of free radicals using the EPR spin trapping technique, based on their ability to quickly trap short-lived free radicals and afford persistent and identifiable aminoxyl spin adducts [6–9]. Due to this unique ability, nitrones have attracted considerable attention as potential therapeutic agents for the treatment of many pathologies where reactive oxygen species (ROS) are implicated, such as cardiovascular diseases, cancer, aging, cerebral ischemia and various neurodegenerative diseases [10–12]. Although it would be reasonable to attribute their biological action to their spin trapping activity, a clear picture of the actual mechanism of action of nitrones has not yet been reported. On the

contrary, a large number of investigations have pointed out that more than one mechanism may be involved and the free radical trapping activity is not so likely to be their primary action [10,13–15]. As part of our ongoing research on the reactivity of benzoxazinic nitrones, a series of 3-aryl-2H-benzo[1,4]oxazin-N-oxides have been previously synthesized [16], and their reactivity towards several carbon- and oxygen-centered radicals widely explored [3,10]. Results revealed that they act as efficient spin traps for a wide range of free radicals and their activity has been demonstrated to be higher than the commercially available N-tert-Butyl- α -phenylnitron (PBN) [10]. Since, over the past few years, several *para*-substituted nitrones [15,17,18] bearing electron-withdrawing groups have shown improved radical detection and antioxidant properties, in this study, we synthesized two constitutional isomers of benzoxazinic nitrones bearing a methyl-acetate group on the benzo moiety (in *para*- and *meta*-position) with the aim of enhancing the reactivity of the already-studied unsubstituted parent compound. Moreover, this study aims to obtain additional insights into their antioxidant mechanism of action.

2. Materials and Methods

2.1. Materials and Characterization

2,2-Diphenyl-1-picrylhydrazyl (DPPH) and 2,2'-Azobis(2-methylpropionamidine) dihydrochloride (AAPH) were obtained from Merck Co. (Steinheim, Germany). All the other reagents and chemicals were of analytical grade for biochemical purposes or HPLC grade. All the solutions were prepared in ultrapure MilliQ water to prevent metal contamination. Electrospray ionization mass spectra were recorded on a Finnigan Navigator LC/MS single-quadrupole mass spectrometer in EI⁺ mode, cone voltage 25 V and capillary voltage 3.5 kV, by injecting samples previously dissolved in methanol. The ¹H and ¹³C NMR spectra were recorded on a Varian Gemini 400 spectrometer in CDCl₃ at 400 and 100 MHz, respectively. Chemical shifts are reported in ppm relative to residual solvent signals (δ = 7.24 and 77.30 ppm for ¹H and ¹³C NMR, respectively) and coupling constants (J) in Hz. FT-IR spectra were collected on a PerkinElmer Spectrum GX FT-IR spectrophotometer equipped with ATR single reflection diamond. Measurements were performed with a resolution of 2 cm⁻¹, 16 scans over 4000–500 cm⁻¹ range. Samples were directly placed on the measuring surface without requiring any preparation. Background adsorption spectrum was recorded each time for correction. Spectra were analyzed on Spectrum 5.3.1 (Perkin-Elmer) operating software. Isotropic X-band EPR spectra were recorded on a Bruker EMX/Xenon spectrometer system equipped with a microwave frequency counter and an NMR Gauss meter for field calibration; for g-factor determination, the whole system was standardized with a sample of perylene radical cation in concentrated sulfuric acid (g-factor = 2.00258).

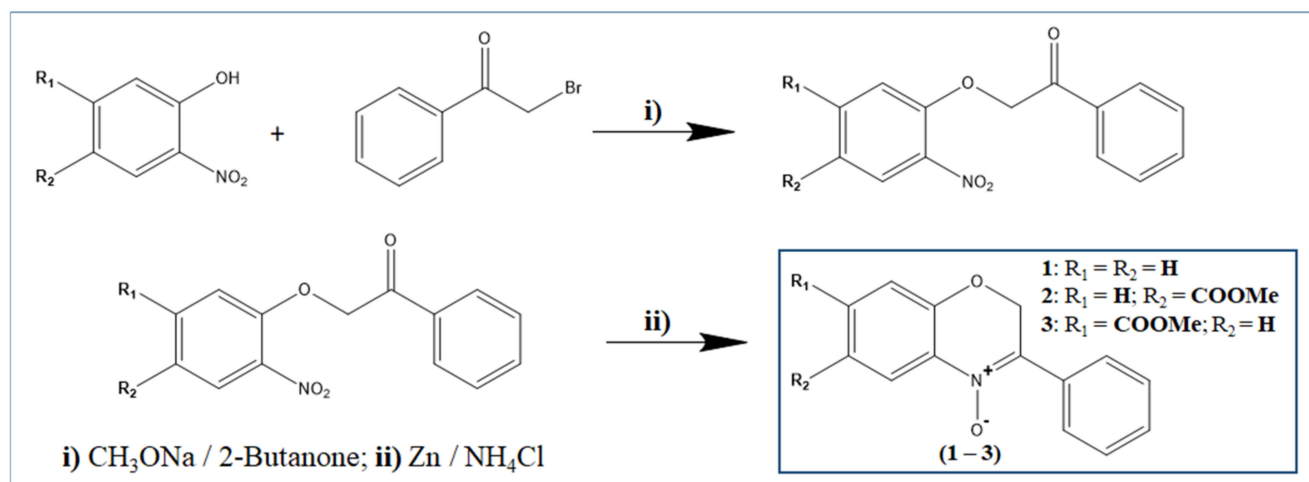
2.2. Synthetic Procedure

The two constitutional isomers of benzoxazinic nitrones (3-aryl-2H-benzo[1,4]oxazin-N-oxides) (2–3) bearing a methyl-acetate group on the benzo moiety and the unsubstituted derivative (1) were prepared with a slight modification of the previously reported two-step method [16,19], as illustrated in Scheme 1: alkaline condensation between the appropriate 2-nitrophenol and the 2-bromoacetophenone, followed by reductive cyclization (Zn/NH₄Cl).

6-(methoxycarbonyl)-3-phenyl-2H-benzo[b][1,4]oxazine 4-oxide (2): ¹H NMR (400 MHz, CDCl₃) δ (ppm): 3.91 (s, 3H, 6-COOCH₃), 5.38 (s, 2H, 2-CH₂), 7.03 (d, J = 8.49 Hz, 1H, C8), 7.24 (CDCl₃), 7.48 (d, J = 7.15 Hz, 1H, 8-CH), 8.03 (d, J = 8.47 Hz, 5H, 3-CH), 8.22 (d, J = 8.06 Hz, 2H, 5-7-CH). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 66.56, 76.77, 76.99, 77.31 (CDCl₃), 115.99, 121.04, 122.64, 128.06, 128.51, 130.68, 131.17, 149.55. Mp: 118–120. IR (KBr, cm⁻¹): 1714 (ν (C=O) ester), 1435 (δ (CH₃) methoxy), 1618 (ν (C=N)), 1266 (ν (N-O) N-oxide). ESI-MS (m/z) of C₁₆H₁₃NO₄: Mr = 283.278, found 283.322.

7-(methoxycarbonyl)-3-phenyl-2H-benzo[b][1,4]oxazine 4-oxide (3): ¹H NMR (400 MHz, CDCl₃) δ (ppm): 3.92 (s, 3H, 7-COOCH₃), 5.35 (s, 2H, 2-CH₂), 7.24 (CDCl₃), 7.48 (m, J = 7.35 Hz,

¹H, 8-CH), 7.65–7.81 (m, J = 8.47 Hz, 5H, 3-CH), 8.23 (d, J = 8.40 Hz, 2H, 6-5-CH). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 66.67, 76.67, 76.98, 77.30 (CDCl₃), 117.44, 121.17, 123.79, 128.13, 131.19, 149.27. Mp: 124–125. IR (KBr, cm^{−1}): 1713 (ν(C=O) ester), 1437 (δ(CH₃) methoxy), 1608 ν(C=N), 1265 (ν(N-O) N-oxide). ESI-MS (*m/z*) of C₁₆H₁₃NO₄: [Mr + H⁺] = 284.29, found 284.57.



Scheme 1. Synthesis of the target benzoxazinic nitrones (1–3).

2.3. DPPH Spectrophotometric Radical Scavenging Assay

The free radical scavenging activity of nitrones was determined by their ability to reduce the free DPPH radical, as previously described [20], with minor modifications. For the spectrophotometric evaluations, antioxidant and DPPH solutions in methanol were mixed (antioxidant 50 μM, DPPH 100 μM, FC), shaken vigorously and incubated for 30 min and 60 min in the dark. The absorbance at 517 nm was determined with a spectrophotometer (BioTek Synergy HT MicroPlate Reader Spectrophotometer) against a blank that lacked DPPH.

The percent inhibition of the DPPH radical by antioxidants was calculated according to Equation (1) as follows:

$$\text{Inhibition ratio (\%)} = [(A_{\text{control}} - A_{\text{sample}}) / A_{\text{control}}] \times 100 \quad (1)$$

where A_{control} is the absorbance of the control obtained by adding into the DPPH methanol solution a methanol aliquot equal to the antioxidant solution volume. A_{sample} is the absorbance of the reaction solution at 30 min. Each sample was measured in triplicate. Mean and standard deviation ($n = 3$) were calculated.

2.4. DPPH Signal Quenching by EPR

The antioxidant ability of nitrones was evaluated using the DPPH quenching approach by EPR spectroscopy. All the experiments were carried out in the dark to avoid photochemical effects on the otherwise stable DPPH radical. EPR spectra were recorded on a Bruker EMX spectrometer system operating at 100 KHz, equipped with an NMR microwave frequency counter. The incubation mixtures contained DPPH at a constant concentration of 800 μM and nitrones in two different nitron:DPPH molar ratios (1:2 and 1:4) in a total volume of 200 μL benzenic solution. The kinetic profiles were obtained following the decay in the EPR signal intensity as a function of time (7200 s) using the 1D_TimeSweep spectra collection mode. As the decay in the DPPH spectra intensity occurs, the EPR-silent diamagnetic counterpart is formed; the test is therefore representative of the capacity of nitrones to react with stable organic radicals. The amplitude of the first line of

DPPH spectra was taken for registration of kinetics. Measurements were run at least in triplicate.

2.5. Determination of Erythrocyte Hemolysis Induced by AAPH

Erythrocyte-oxidative hemolysis induced by AAPH and its inhibition in the presence of nitrones was performed as previously described [21]. First, 5 mL of human blood in EDTA was washed three times in PBS (pH 7.4) by centrifugation at 1000 g for 10 min. The resulting pellet was resuspended with 6-fold PBS (pH 7.4). Then, 0.5 mL of erythrocyte suspension was added to 0.25 mL of PBS nitron solution, obtaining a final antioxidant concentration of 0.012, 0.5 and 0.2 mM. The mixtures were incubated for 1 h at 37 °C and then 0.25 mL of PBS was added with or without AAPH (final concentration AAPH, 50 mM) followed by incubation for 12 h at 37 °C in a gently shaking incubator.

Next, 100% lysis was determined by adding 0.15 mL of a 1% Triton X-100 solution, while only PBS was added in the negative control. After sedimentation of the unlyzed erythrocytes by centrifugation (1000 g, 15 min, 4 °C), the supernatant was transferred to a microtiter plate and diluted 1:4. The hemoglobin absorption was determined at 540 nm using the Synergy HT microplate reader spectrophotometer (BioTek, Winooski, VT, USA). The percent hemolysis was determined using Equation (2) as follows:

$$\% \text{Hemolysis} = [(A - A_0) / (A_{\text{total}} - A_0)] \times 100 \quad (2)$$

where A is the absorbance of the test well, A_0 is the absorbance of the negative control, and A_{total} is the absorbance of the positive control. The mean values of three replicates were reported.

2.6. DFT Calculations

Density functional theory (DFT) calculations were carried out using the GAUSSIAN 09 suite of programs [22], taking advantage of the resources available at the Cineca Supercomputing Center [23]. All calculations on paramagnetic species were carried out with the unrestricted formalism, giving $S^2 = 0.7501 \pm 0.0003$ for spin contamination (after annihilation). Thermodynamic quantities were computed at 298 K by means of frequency calculations performed employing the M06-2X functional in conjunction with the 6-31 + G(d,p) basis set, starting from molecular geometries computed at the B3-LYP/6-31G(d) level of theory. In frequency calculations, no negative values (imaginary frequencies) were found, demonstrating that all quantities refer to geometry minima. All calculations refer to the gas phase.

2.7. Statistical Analysis

Data are presented as mean \pm S.D. Statistical comparison of differences between antioxidants tested was carried out using Student's *t*-test. Values of $p < 0.05$ were considered statistically significant.

3. Results and Discussion

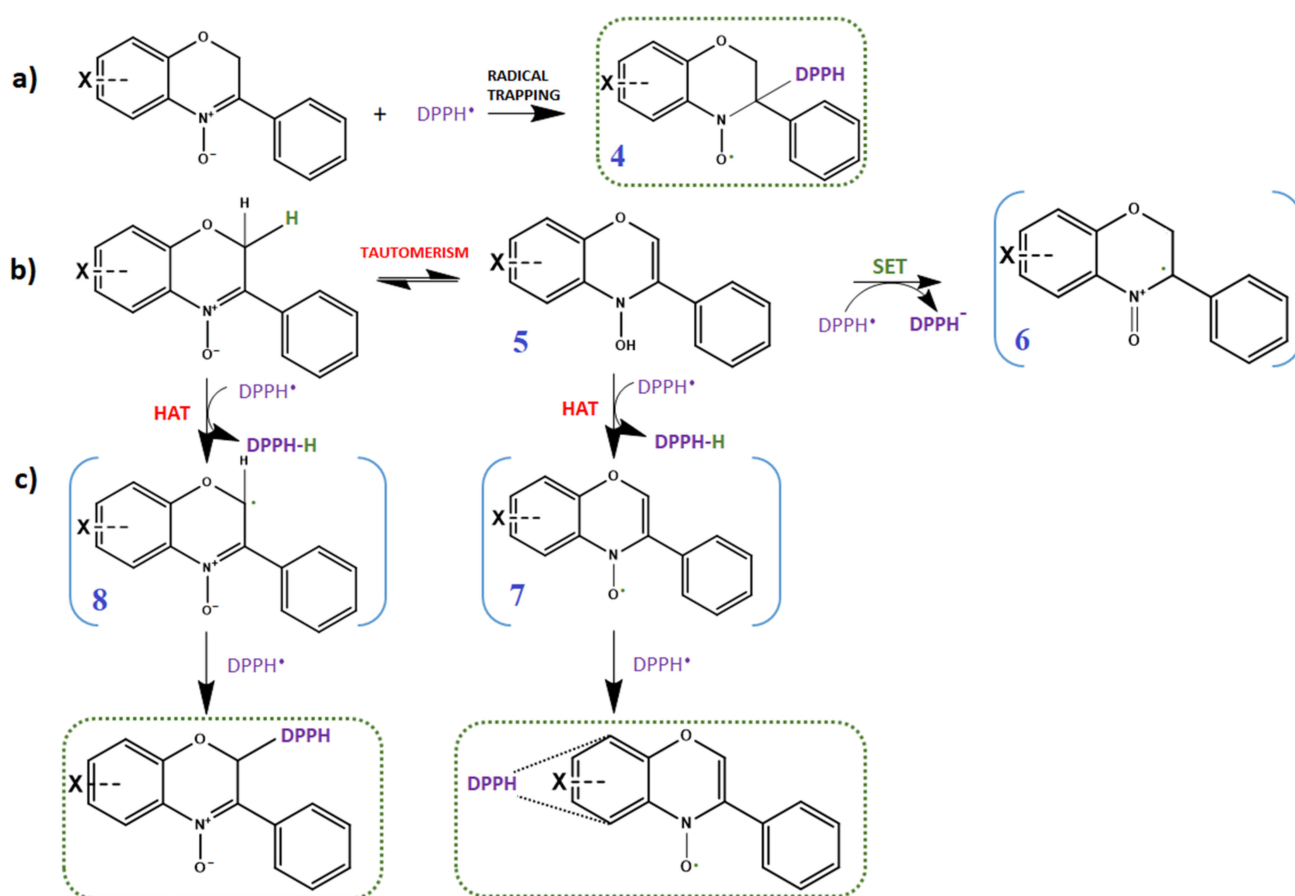
3.1. Synthesis and Structural Characterization

The unsubstituted nitron, 3-phenyl-2H-benzo[1,4]oxazine 4-oxide, is a known compound; thus, both the details of its synthesis and a full structural characterization can be found in the literature [16,19]. In this study, two derivatives of this compound, bearing an electron-withdrawing methyl-acetate group on the benzo moiety, were synthesized and their structure confirmed by mass spectrometry (MS), infrared spectroscopy (IR) and nuclear magnetic resonance spectroscopy (^1H -NMR and ^{13}C -NMR) (see Section 2.2).

3.2. DPPH Scavenging Activity of Nitrones

Nitrones are known spin traps that are capable of trapping free radicals (carbon-, oxygen- and nitrogen-centered radicals) via addition reactions at the C=N double bond of the nitronyl function, thus, encouraging their use as pharmacological agents against

oxidative stress-mediated pathologies. However, the mechanism by which nitrones exert their antioxidant activity is still not completely clear. Besides the well-documented radical trapping ability of nitrones, a large body of investigations suggest that more than one mechanism may be involved [10,13–15]. In the specific case of DPPH, chosen as model radical, Scheme 2 summarizes all the possible reaction mechanisms that could take place between DPPH and the nitrones under investigation. In a typical DPPH assay, nitrones are therefore expected to quench DPPH free radicals via a spin trapping mechanism following pathway (a), giving rise to a nitroxide spin adduct (compound 4). However, quenching of DPPH radical by either sequential electron transfer (SET) (pathway (b)) and/or hydrogen atom transfer (HAT) (pathway (c)) are also possible. The H-donating ability could be associated to either the potentially removable hydrogens in position 2 (i.e., oxygen α position) or the hydrogen from the N-OH group of the corresponding hydroxylamine tautomer (compound 5); such hydrogen atoms could be abstracted by DPPH radicals affording either radical compound 8 or the nitroxide compound 7, likely followed by addition of another DPPH molecule. The relative reactivity of nitrones towards DPPH radicals was evaluated by both EPR and spectrophotometry.



Scheme 2. An overview of the potential mechanisms involving nitrones and DPPH; (a) formation of DPPH spin adduct; (b) formation of benzoxazine radical cation by SET (Single Electron Transfer); (c) HAT (Hydrogen Atom Transfer) pathways, eventually undergoing radical-radical recombination involving C-2 or the benzo-ring.

3.2.1. DFT Calculations

The computed values collected in Table 1 indicate that all the benzoxazinic nitrones under investigation showed comparable reactivity (i.e., similar spin trapping ability) toward all the radicals tested, demonstrating an easier reaction with C-radicals compared to peroxy or alkoxy radicals. This analysis has been already reported for the unsubstituted

parent compound [10]. However, differences could be found in the IP values (in the gas phase) between the two isomers (~13 Kcal smaller for the *meta*-substituted), indicating that a SET pathway (b) may potentially be easier for the *meta*-substituted compound compared to the *para*-substituted one.

Table 1. Computed Gibbs free energy changes (ΔG) in radical addition reactions to nitrones and ionization potential (IP); all quantities are reported in Kcal/mol and refer to the gas phase.

Nitrone	Radical (ΔG)					IP
	Me	OMe	OOMe	OH	OOH	
1	−42.51	−38.69	−16.30	−54.34	−25.06	174.19
2	−42.59	−36.91	−19.29	−52.79	−24.66	163.28
3	−43.10	−39.72	−19.27	−54.60	−24.88	176.41

3.2.2. DPPH Quenching by EPR and UV Assay

Figure 1A–C show the EPR kinetic profiles for the reaction between nitrones and DPPH in the two different molar ratios (1:2 and 1:4). As can be clearly seen from the corresponding traces, *para*- and *meta*-substituted nitrones (Figure 1A,B) are able to quench DPPH and the extent of the signal reduction is dose-dependent, achieving up to approximately 37% and 15% decreases in the DPPH activity at the highest concentrations (1:2 nitrone:DPPH molar ratio), respectively. On the other hand, the unsubstituted nitrone 1 shows no activity toward DPPH and its addition leads to an initial clear growth in the DPPH signal's amplitude, up to ~5% for the 1:2 molar ratio, suggesting a pro-oxidant behavior taking place at the beginning (C). Regarding derivatives 2 and 3, the presence and the position of the electron-withdrawing methyl-acetate group appears to strongly affect their activity. In particular, derivative 3, bearing the substituent in *para* position with respect to the nitronyl function, exhibits higher anti-DPPH radical activity than the *meta*-substituted isomer.

In order to see whether the DPPH scavenging activity behaviors of nitrones were reproducible and independent by potential solvent–solute interactions, the decay profiles in the absorbance of the reaction mixtures at 1:2 nitrone:DPPH molar ratio were also measured at 517 nm in a polar solvent (methanol). Figure 1D shows the percentage inhibition of DPPH based on the measured absorbance values taken at two different incubation times (at 30 and 60 min). It is clear that both the decay trend of the DPPH radical for nitrones 2 and 3 and the signal growth for 1 observed by UV/VIS spectroscopy are in good agreement with those observed by EPR in benzene. This indicates the good reproducibility of the result from the two methods employed, despite the very different polarities of the solvents used (methanol and benzene).

In order to obtain additional insights into the potential mechanisms underlying the DPPH scavenging activity of nitrones, the reaction mixtures were analyzed by liquid chromatography–mass spectrometry (LC–MS). Results indicated that, only for nitrone 3, there was clear evidence of spin adduct formation with DPPH (ions at m/z 701 [$M_{7-COOMe} + M_{DPPH} + Na$] and 985 [$2M_{7-COOMe} + M_{DPPH} + Na$]), while no additional peaks were found in the mass spectra of the other two nitrone–DPPH reaction mixtures. As mentioned before, potential spin adducts could arise from either direct trapping of the free radical at its α -carbon atom of the N-oxide ($C=N^+-O^-$) group, giving rise to a paramagnetic nitroxide adduct (pathway (a)), or by hydrogen atom transfer (HAT) and the subsequent formation of spin adducts in position 2 and/or at the phenyl ring (pathway (c)). Since no additional paramagnetic species were detected during the EPR experiment, pathway (a) may be excluded as nitroxide adducts are typically very stable and persistent, implying that further insights should be sought within pathway (c) for the *para*-substituted nitrone. The lower activity of nitrone 2 compared to nitrone 3 could be linked to its higher tendency to be oxidized (see Section 3.2.1): in this case, since the nitrone could be partially oxidized, its amount available for a further possible reaction with DPPH could be less than expected.

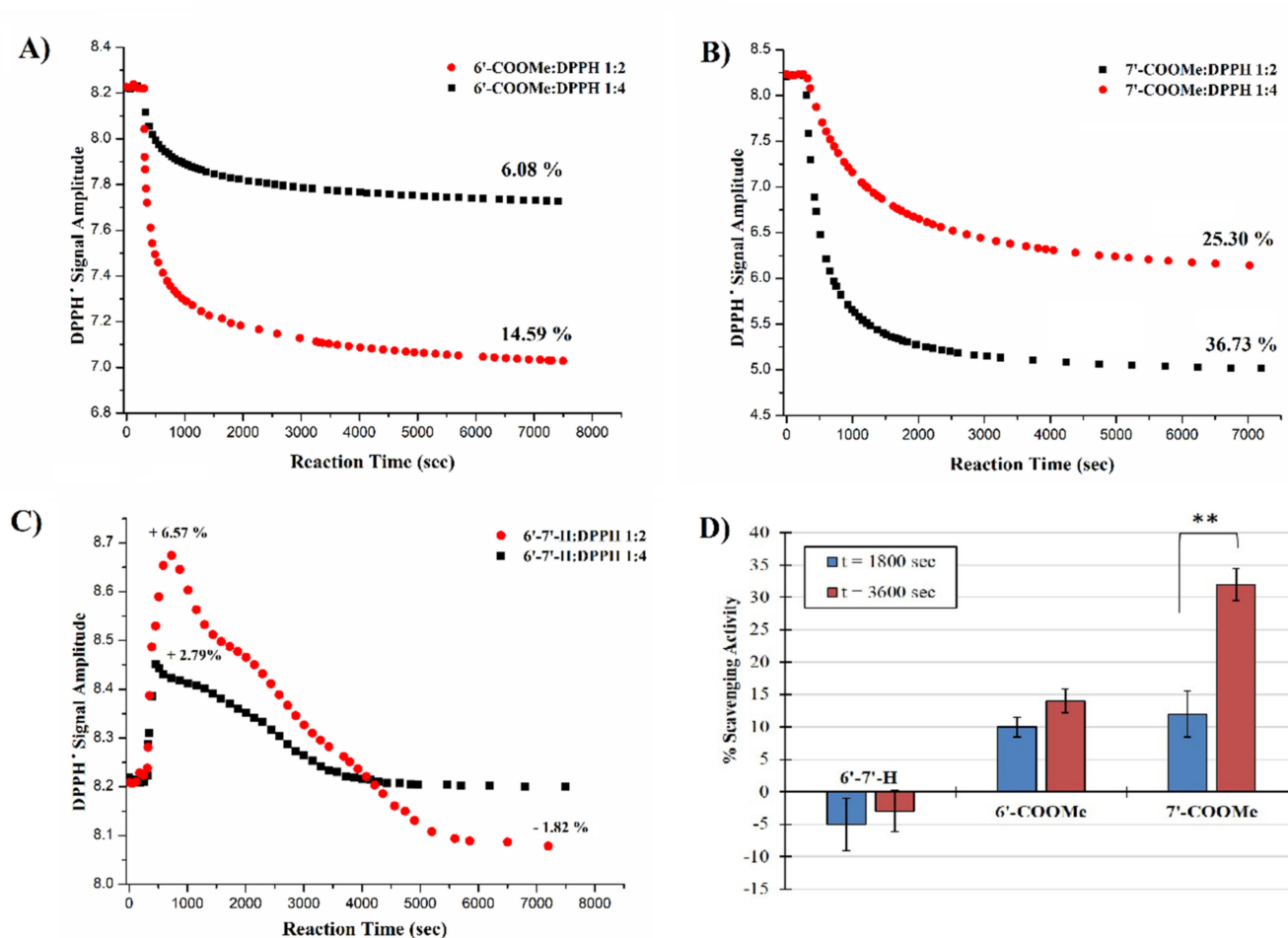


Figure 1. (A–C) Kinetic profiles for the reaction of 200 and 400 μM nitrone solutions with 800 μM DPPH in benzene at 23 $^{\circ}\text{C}$; (D) spectrophotometric evaluation of free radical scavenging activity of nitrones by DPPH assay. Results are expressed as the mean \pm standard deviation ($n = 5$). ** $p < 0.001$.

3.3. Protection against AAPH-Induced Hemolysis

The hemolysis of erythrocytes has been extensively used as an *ex vivo* model in the study of oxidative stress-induced alteration of cell membranes. AAPH is a water-soluble radical initiator, which generates, after thermal decomposition, a constant flux of peroxy radicals, responsible for inducing lipid peroxidation and oxidation of membrane proteins. This leads to the formation of hemolytic holes in the membrane. Erythrocytes were therefore pre-treated with antioxidant compounds followed by the addition of AAPH. As shown in Figure 2, the percentage of survival of the erythrocyte population was related to the concentration of the antioxidant tested, with the *para*-substituted nitrone having the highest ability to protect the erythrocyte cells from AAPH-induced hemolysis, confirming what was observed in the previous paragraphs.

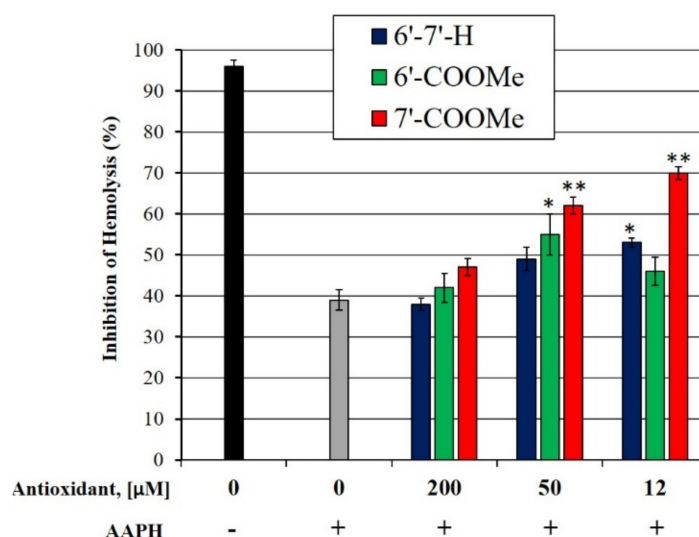


Figure 2. Hemolysis of human erythrocytes induced by AAPH. RBC suspensions were treated with nitrones followed by incubation with 50 mM AAPH at 37 °C. * $p < 0.05$ and ** $p < 0.001$ with respect to untreated RBCs.

4. Conclusions

In this study, two new derivatives of the benzoxazinic nitron family were synthesized and the effects of electron-withdrawing substituents on the benzo-ring on their antioxidant activity were explored in vitro. More in details, the reactivity of the two derivatives bearing a methyl-acetate group in benzoxazine positions 6 and 7 was evaluated using both a typical DPPH scavenging assay and an oxidative hemolysis inhibition test with human erythrocytes. Results demonstrated that, in both cases, the presence and the position of such a substituent appears to affect the nitrones' antioxidant activity. In particular, the derivative substituted in the *para* position with respect to the nitronyl function exhibited higher anti-DPPH radical potency and a higher percentage of hemolysis inhibition than both the *meta*-substituted isomer and the unsubstituted parent compound. Relative to the DPPH activity, MS data of the reaction mixtures showed evidence of spin adduct formation only for the *para*-substituted compound, which may at least account for its higher activity against DPPH radicals. On the other hand, even if less active, the *meta*-substituted isomer showed a lower (computed) ionization potential, indicating a greater tendency to be oxidized and reduce DPPH radicals via a SET mechanism. Although the antioxidant activity trends for all compounds are in fair accordance in both assays, further investigations and analyses are now in progress and will be reported in due course. In this regard, it is worth pointing out that care should be taken while estimating the antioxidant activity of newly synthesized compounds by means of the DPPH assay, whose radical nature could interfere with the antioxidants under investigation.

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Conflicts of Interest: The authors declare no conflict of interest.

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