

Review



Microbial Interactions with Particulate and Floating Pollutants in the Oceans: A Review

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Abstract: The Earth's oceans are the final resting place of anthropogenic wastes, mainly plastics, metals, rubber, and fabrics, in order of decreasing abundance. On reaching the sea and the benthos, most of these have assumed fragmented or particulate forms. They become colonized by marine microorganisms and later interact with macroorganisms, leading to potential problems with marine life and the ecosystem. Rapid biodegradation of the polluting materials is a possible, and desirable, result if harmful by-products are not produced or toxic constituents are released. Negative effects are the transport of organisms to other ecosystems, with possible disturbance of the natural biological balance, or transfer of pathogenic organisms. A microbial biofilm can mask unattractive anthropogenic materials, increasing ingestion by marine life, with potentially dangerous results. This article seeks to provide a synthesis of the interactions occurring between oceanic anthropogenic polluting matter in solid and particulate form, and the microbiota present in our seas. It discusses the most important solid and particulate pollutants in the oceans, their sources, adverse effects, interactions with living organisms, mainly microorganisms, and future research for their control. Pollutants included are marine litter (macrodebris), microplastics, engineered nanoparticles, metallic particles, and, finally, sinking particles ("marine snow") as a potential biodegradation "hot spot".

Keywords: marine pollution; plastics; marine litter; nanoparticles; metallic particles; sinking particles; biodegradation; anthropogenic pollutants

1. Introduction

Oceans face global natural and anthropogenic challenges. The United Nations has declared 2021 to 2030 the decade of restoration, promoting, among others, resilience to anthropogenic changes, especially in the oceans [1], which are considered the main sink of anthropogenic contaminants [2].

Detectable only with specific equipment, there is an abundant world of microorganisms inhabiting the oceanic ecosystem with a complexity and diversity that competes with all other forms of life on Earth. This group includes bacteria, viruses, fungi, and other microscopic organisms. Of all the living organisms in the ocean, about 50 percent of the biomass weight consists of microbes [3].

Microorganisms are responsible for the food and nutrient cycling that, in their absence, would not be bioaccessible in the ecosystems [4]. Many are also the guardians of ocean water balance and resulting in healthy ecosystems, cleaning the ocean environments of waste, and preventing the proliferation of opportunistic disease-causing beings. Microorganisms



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Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). inhabit some of the environments considered extreme, such as scalding hydrothermal vents and even underground glacial lakes in Antarctica [5,6]. These were the first organisms to inhabit planet Earth, living in an anoxic environment in a pristine ocean [7].

Several types of surfaces with particular physicochemical and biological characteristics are offered by marine ecosystems; they include living organisms, among them animal and vegetal species. These substrata include many kinds of particles and aggregates, both inert and reactive mineral substrata.

A huge group of aquatic microorganisms shows the capacity to colonize surfaces, resulting in the formation of biofilms and the development of specialized processes within these structures [8,9]. As a survival mechanism, the surface colonization process in aquatic environments has a fundamental role for the microscopic species, since it represents greater access to nutritional resources, higher colony stability, and stronger specimen interactions, in a dynamic environment of low nutrient concentration. Sessile microorganisms (those attached to surfaces in biofilms) have advantages over the planktonic cells in their resistance to adverse conditions and antimicrobial substances, as well as possessing an increased metabolic rate.

Aquatic solid substrates are ideal environments for important biogeochemical activities [10]. Surface colonization and biofilm production protect from predators, viruses, antibiotics, chemical toxins, and other deleterious environmental elements [11–13]. Polluting anthropogenic particles can serve as niches for the survival and replication of marine microorganisms.

Nowadays, however, marine litter is considered a worldwide issue alongside other key environmental threats, such as climate change, ocean acidification, and the loss of biodiversity [14,15]. It is regarded as one of the most significant problems for the marine environment and a major threat to biodiversity [16]. Over the last decades, it has become clear that litter particle pollution presents a global environmental challenge of increasing presence in the oceans. The scientific community has studied the microbial life colonizing particle surfaces of these pollutants, but the general concepts of microbial ecotoxicology have only rarely been involved in the studies [17].

Marine litter results in aesthetically detrimental effects represent a threat to commercial shipping and fishing vessels, can intensify the diffusion of organic and inorganic contaminants, and is harmful to marine higher species and potentially also humans [18]. 30–75% of all marine debris consists of plastic, which pollutes environments from the poles to the equator and from shorelines to the deep-sea [19,20]. Apart from the problems caused when microplastics are ingested by marine animals, the plastics themselves can be degraded under the influence of UV to become toxic and release toxic components such as bisphenols. Marine debris is negatively affecting the global economy, wildlife, and the environment; there is global agreement that it needs to be addressed urgently [21].

Petroleum hydrocarbons resulting from oil spills, shipping disasters, etc. are widely recognized and important marine pollutants. There has been much written about the effects of such contamination on marine ecosystems [22,23] and the potential for its bioremediation using microorganisms [24–26]. The current article will not add to the available information in this area, focussing on less well-studied marine solid particulate pollution of growing importance. The aim is to review the more recent literature that documents the interactions of marine microorganisms with particulate pollutants.

2. Macrodebris

Marine litter, or macrodebris, is one of the sources of microcontaminants. Macrodebris not removed by other means will be broken down by mechanical, chemical, and biological activities in the oceans to add to the already present levels of polluting microparticles. Its vast distribution around the globe and long-life durability make marine litter a critical environmental issue [21,27]. This type of pollutant is present worldwide, from shallow water to the deep sea and from the poles to the equator. The most direct impact of marine litter on marine biota is the mechanical effect resulting from the entanglement of animals,

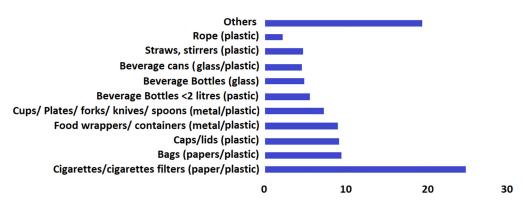
which potentially harms their mobility, feeding, breathing, and reproduction capacities [28], affecting, most of the time, wandering species like fish, marine mammals, sea turtles and seabirds [29]. Sessile species are also potential targets; they are subject to mechanical impacts resulting from the movement of waste, which can affect their body structure [30,31].

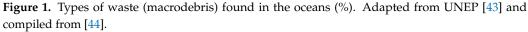
In the deeper layers of the sea, larger-sized anthropogenic litter, which may be composed of plastic, metal, glass, rubber, and fabrics such as rope and clothing, is becoming increasingly common as human beings expand their oceanic activities.

These materials can become colonized by microorganisms within a few days [32], producing biofilms that differ not only according to the physical and chemical environment but also to the man-made substratum. It has been suggested that the main anthropogenic plastic litter in the oceans is associated with fishing activity [30,33–36]. 12.2 million tonnes of plastic are discarded into marine ecosystems per year, 80% being from coastal, 9.4% from fishing, and 4.9% from shipping origin [37,38].

The major type of marine debris affecting the worlds' reefs is derived from fishing activities [39], while in the deep seas most litter is from ships offshore [38]. Plastics are the most frequent materials, followed by metals [40] (Figure 1). *Plastics*, in both macro and micro form, become colonized by cyanobacteria and diatoms within hours of immersion in the sea, these microorganisms later becoming superseded by proteobacteria and, later, by rare fungi (mainly dothideomycetes [41]). The initial adhesion of diatoms and cyanobacteria on polyethylene has been confirmed using the new technique of CLASI-FISH [42]. Diatoms were shown to be associated with surface bacteria of the Bacteroidetes, Rhodobacteriaceae, and Gammaproteobacteria groups. Main colonizers subsequently were found to be members of the Rhodobacteriaceae, which remained predominant over the whole 5 weeks of immersion in the North Atlantic. In the Tropical Atlantic Ocean, the picture was a little different and the microflora more diverse, but Rhodobacteriaceae remained the major colonizers after one week.

Types of Waste found in the oceans (%)





The majority (80%) of litter in coastal areas of the oceans is considered to be landbased in origin [45]. *Metal* litter has been thought to be relatively unimportant in coastal areas of the Mediterranean Sea, since aquatic circulation and meteorological dynamics ensure no permanent impairment of planktonic populations, leading to rapid correction of anthropogenic perturbations in these areas [46]. However, some marine invertebrates have been found rafting on metals, thereby invading other geographical areas [47,48], indicating the importance of this type of litter in ecosystem disturbances. Woodall et al. [49] found that, of all the types of litter collected from the deep waters of the equatorial Atlantic Ocean, metal showed the lowest biofilm diversity compared with the plastic, rubber, glass, and fabric litter investigated and was the only material on which zetaproteobacteria were detected. This is particularly interesting since zetaproteobacteria are iron-oxidizing bacteria that are associated with corroding steel structures [50–52]. Corrosion-associated bacteria had not previously been reported on metallic ocean litter, but in 2019 Muthukrishnan et al. [53] detected higher levels of the anaerobic, heterotrophic sulfate-reducing bacterial genus, Desulforibrio, along with Pseudomonas, in steel biofilms from the Sea of Oman, while the main colonizer in wood biofilms was Corynebacterium. The application of antifouling coatings to metallic surfaces may be another reason for the smaller contribution of waste composed of metals to the colonization and transport of microorganisms. These antifouling coatings can stop or at least reduce the propagation of microorganisms on various surfaces [54]. The use of antimicrobial coatings on metal surfaces is based on the presence of compounds toxic to microorganisms, for example, pigments, solvents, metals, and organic and organometallic biocides [55,56]. Antifouling paints may also, themselves, become pollutants; the release of paint particles during use and cleaning of the vessels thus protected is a threat to aquatic life [57,58], Researchers have pointed to the increasing biocide (antifoulant) concentrations in water and sediment samples collected near boat maintenance areas, posing a threat to aquatic life [59–61]. Hence there is considerable effort being expended to discover and develop antifouling surfaces that are ecologically acceptable [62,63].

Rubber is not commonly considered one of the most important marine pollutants, but it can be significant because of its refractory character. In 2007, a shipload of rubber ducks completed a 15-year-long journey, which began in January 1992 when a cargo ship named Ever Laurel, traveling from Hong Kong to the United States, lost some of its cargo during an ocean storm [64]. The load consisted of approximately 29,000 toys, some of which had reached the Australian and the east coast of the United States by 2007. Others went through the Bering Strait and the Arctic Ocean until arriving in Greenland, the United Kingdom, and Nova Scotia. This represents the considerable potential for the transfer of rafting organisms. Car tire rubber is a major contributor to terrestrial wastes, and its leachates can be found in the oceans. These include cobalt and zinc, both of which inhibit algal growth and mussel reproduction [65]. Most tires are made of polyisoprene rubber and several bacteria and fungi have been shown to degrade isoprene [66], actinomycetes being considered to be the most important [67–69]. Carrión et al. [70] used an isoprene-degrading gene probe (IsoA) to detect degrading microorganisms from various habitats. They found that, in coastal sediments, the gene was associated with the genera *Rhodococcus* and *Variovorax*, while in freshwater sediments Rhodococcus and Sphingopyxis were prevalent. Isoprene degraders had previously been identified in estuarine waters [71].

Textiles also constitute part of marine litter. Microparticles of synthetic and natural fibers are one of the most abundant marine pollutants [72,73]. The increased emphasis on the environmental effects of textiles is linked to an escalation in public focus on plastic pollution [74], of which a significant part is thought to be composed of textile microfibers. Textiles can be produced from various types of plastic polymers, mainly PET, and microfibers can be released during manufacture and normal laundry activities [75,76]. Natural textile polymers, such as silk, cotton, and wool, are less problematic since they are biodegradable and less persistent in the ocean. The pollution associated with textile laundering has recently been reviewed by Gaylarde et al. [75].

The international textile industry is a major booster of global environmental contamination. Global fiber manufacturing exceeded 105 million metric tons in 2018 [77] and it has been estimated that 0.19 million tonnes of microfibers from the production, disposal, and laundering of textiles enter the marine environment annually [76]; indeed, it has been calculated that, for a city of 100,000 inhabitants, approximately 1.02 kg of microfibers will be released into the wastewater treatment system per day from the washing of polyester fleece jackets alone [78].

Fibers can release bioavailable contaminants, like toxic metals and other substances, that have been used during textile production [79]. The Industrievereinigung Chemiefaser [77] has emphasized that the negative environmental influence of the textile industry is centered around the amounts of industrial compounds used during the manufacturing process, as well as the liberation of a variety of pollutants into ecosystems [80–84], including textile

fibers [74,78,85–88]. Many are not degraded by any biochemical and/or natural photochemical process and this may result in bioaccumulation and/or biomagnification.

Even though natural fibers are relatively biodegradable in the marine environment, degradation may release other pollutants, such as dyes, which can carry risks to marine life. Stanton et al. [89] question whether natural fibers are better for the environment than microplastic fibers; in their study, microplastic textile fibers such as polyester and nylon were absent from 82.8% of samples, whereas 'natural' textile fibers were absent from just 9.7% of samples. Nevertheless, it cannot be denied that the resistance of plastic fibers to biodegradation makes them potentially more important pollutants.

Harmful chemical compounds and pollutants can be released from various plastics through UV-linked or biodegradation [31,79,90]. Endocrine disruptors, for example, have negative effects on marine biota even at the extremely low levels produced during the transportation and degradation of the polymer matrix. These pollutants have been identified as major contaminating compounds in wastewater effluents [91] that will be released into the Earth's waters. Endocrine disruptors consist of chemical compounds able to mimic endogenous hormones, as they have a very similar chemical constitution. So a hormone receptor can mistakenly recognize these compounds, activating or blocking the normal functioning of the endocrine system [92,93]. Sex change of organisms exposed to contaminants that mimic estrogen compounds has been recorded in marine ecosystems, with all the specimens becoming female. Imposex (sexual impotence) has also been detected in several marine gastropod groups exposed to TBT (tributyltin).

Girard et al. [94] performed in vitro colonization experiments with mixed textile fibers extracted from intertidal sediments in Indonesia and with HDPE microbeads. They used seawater from a coral reef aquarium as the suspending medium, with its natural microbial populations. The colonizing bacteria (identified by DNA sequencing) differed substantially between the two substrata. HDPE beads became colonized by a community enriched in Alcanivoracaceae, while the bacterial populations on textile fibers (40% cotton, mainly dyed black or blue) became enriched in Kordiimonadaceae and Cellvibrionaceae. According to the authors, oxygen consumption and specific colonization patterns suggested that the fiber and plastic substrata were being biodegraded. They did not consider any possible negative effects of the degradation products on the biota, simply considering that degradation was a positive process removing the fibers from the environment.

3. Microplastics

The increased use of plastic materials over the last years has led to millions of tonnes per year of these recalcitrants being released into our seas. It is acknowledged that they are one of the most important anthropogenic influences on our waters and are even found in the Arctic Sea, arriving there from as far away as the Asian coast [95]. They cause problems through their ingestion by aquatic life [96,97] and references therein. Microplastics also provide a surface on which pollutant molecules and potentially dangerous microorganisms can become concentrated and carried to other locations 18, 19. Once the plastic becomes modified by oceanic organics, a completely new surface, much more attractive as a food source, is presented to living creatures.

There are five plastic accumulation zones in our oceans (Figure 2) [98], perhaps the best known of which is the Great Pacific Garbage Patch, in the north-central Pacific Ocean [41]. However, not all ocean gyres have been equally studied and, indeed, difficulties in standardizing sampling procedures make this a very problematic area [99].

Initially, plastics are broken down in the seas into small fragments by abiotic forces such as u-v and wave action; when these fragments are less than 5 mm in size they are known as microplastics. They rapidly become covered by a thin film of organics present in the water and then by marine microorganisms (Figure 3), forming a surface population known as the plastisphere [100]. This contains a wide variety of prokaryotes and eukaryotes and the exact makeup may depend on the type of plastic, as well as the local environment [101,102], although it has been suggested that the plastisphere is only

specific to plastic-type in conditions of low nutrients and low salinity [103]. A survey of plastisphere biofilms from around the world, and of various degrees of maturity, indicated that the most abundant phyla are Proteobacteria, Cyanobacteria, and Bacteroidetes [42].

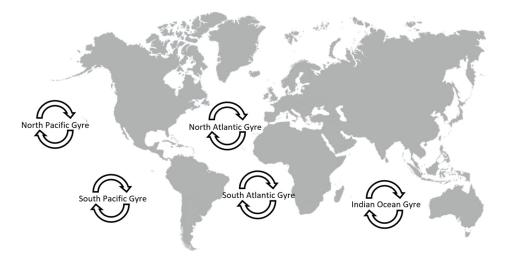


Figure 2. The five swirling ocean garbage patches are called "gyres".

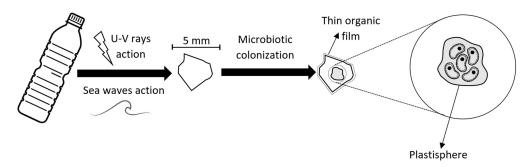


Figure 3. Microplastic colonization.

Colonized microplastics are carried on ocean currents and in spray and so transported to other regions, where the result may be prejudicial to the environment and its biota. It has been suggested that this may be one of the routes whereby metal- and antibiotic-resistant genes are spread around the world [104,105], but pathogenic microorganisms themselves may be transported in this way. For example, opportunistic invasive bacterial populations can enhance coral bleaching [106] and the biofilm in most plastic pools in the Bay of Brest has been shown to contain species related to the potential oyster pathogen, *Vibrio splendidus* [107].

Even if not directly harmful, the introduction of a new microbial population may alter the oceanic ecosystem in unknown ways [108]. Such transfers of local biota are known to occur; plastic debris carrying invasive corals and other living forms can be responsible for the persistence of new species in the ecosystem [109,110]. If such introduced species survive, they may alter the local ecosystems in undesirable ways [111]. In the Antarctic Peninsula Region, 13 species, mainly marine invertebrates, have been identified as presenting a major risk to the native biota [112]. Although these are considered to be transported mainly on ships, the authors point out that the role of floating plastics should not be discounted. Yet another potential problem associated with the biofilm on microplastics is its ability to adsorb contaminants, and potentially toxic, molecules. Biofilms on LDPE have been shown to adsorb Ba, Cs, Fe, Ga, Ni, and Rb and to facilitate the uptake of Cu, Pb, Al, K, U, Co, Mg, and Mn [113].

On the other hand, microorganisms transported on anthropogenic particles may have desirable effects. The surface-associated genus *Phaeobacter*, for example, produces antibi-

otics, including tropodithietic acid, which inhibit a variety of bacteria, including pathogenic *Vibrio* species, and may have protective effects in pisciculture [114] and references therein.

Another positive result of the microbial colonization of plastics is the potential for biodegradation of the material by the adherent organisms. Debroas et al. [115] found that there was an overrepresentation of microorganisms capable of xenobiotics degradation in the plastisphere. Some of the microorganisms that have been detected in the plastisphere are known hydrocarbon-degraders and may be involved in the breakdown of the plastic substrate [115,116]. Enzymes involved in such degradation include lipases, proteinases, cutinases, and dehydrogenases [117]. Table 1 lists some of the plastic-degrading microorganisms that have been identified in the plastisphere and elsewhere, using culture or molecular techniques. Although several fungal species are listed, and, indeed, were among the earliest microorganisms to be demonstrated to have a plastic-degrading ability, few of these plastilytic eukaryotes have been detected in the plastisphere and even less have been assessed for activity under marine conditions. Indeed, fungi are rarely detected in biofilms on benthic marine plastic debris [118,119].

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Table 1. Plastic-degrading microorganisms.

Plastic	Microorganism	Reference
Polyurethane/PE-PU	Comamonas acidovorans	[149]
	Pseudomonas chlororaphis	[150]
	Pseudomonas putida	[151]
	Acinetobacter gerneri	[152]
	Aureobasidium pullulans	[153]
	Fusarium solani	[140]
	Alternaria sp.	[154]
PVC	Pseudomonas putida	[155–157]
Polystyrene	Rhodococcus ruber	[158]
	Azotobacter beijerinckii	[159]

Table 1. Cont.

Abbreviations: PCL—polycaprolactone, PE—polyethylene, PET—polyethylene terephthalate, PHA—polyhydroxyalkanoate, PU—polyurethane, PVC—polyvinylchloride.

Some plastics may become accessible to microbial metabolism only after partial breakdown or modification by non-biotic factors, such as u-v or heat. Such non-hydrolyzable polymers include polyethylene, polypropylene, and polystyrene. Many modern plastics, such as PET, however, are directly susceptible to hydrolysis. Two enzymes jointly capable of degrading PET into environmentally benign monomers have been isolated from the bacterium *Ideonella sakaiensis* in a PET bottle recycling site [160]. [161] has produced a more efficient engineered PET-hydrolase that could form the basis of a bioremediation treatment for environments contaminated with plastics.

Delacuvellerie et al. [116] found that plastic-enriched populations of bacteria from plastispheres contained hydrocarbon-degrading bacteria. These included *Alcanivorax*, *Marinobacter*, and *Arenibacter* on LD (low density) PE and PET. *Alcanivorax borkumensis* from thick biofilms on LDPE was shown, by weight loss, scanning electron microscopy, and ATR-FTIR analysis, to degrade the plastic substratum. Although there was only 3.4% degradation after 80 days, this is worthy of further investigation, since LDPE is a very hydrophobic plastic and thus rather resistant to biodegradation. PET has also been shown, by SEM and FTIR, to degrade and develop surface cracks on exposure to marine biofilms in the Arabian Gulf, while polypropylene and PVC showed cracking under biofilms formed in Chinese coastal waters. Despite the considerable amount of research effort that has been applied to the microbial degradation of plastics, Oberbeckmann and Labrenz [146], in a recent review, conclude that microplastics are recalcitrant substrates for microorganisms and will probably not be biodegraded within a reasonable timescale.

The colonization of microplastics in the oceans leads to an increase in their weight and, because of the production of extracellular polymeric substances (EPS) within the plastisphere, to their agglomeration, resulting in their eventual deposition in the benthos [118], and references therein, where the vast majority of oceanic plastic wastes are deposited [162,163]. Floating microplastic particles have a 50% chance of sinking between 17 and 66 days after they arrive in the water [164] and it is considered that very large amounts of microplastics are removed from surface waters by attachment to organic sinking particles (q.v.) [138]. Once incorporated into the sea-bottom sediments, the final fate and distribution of the plastics are strongly controlled by sea-bed currents [165]. Even here, problems can result from ingestion by benthic organisms [166,167]. Plastic waste has been registered at a depth of almost 11,000 m and plastics have been detected in the hind guts of amphipods from deep ocean trenches of depths 7000 to 10,890 m from the Pacific rim [166]. In order to more accurately assess the degree of plastic contamination in our oceans, more measurements need to be made on this specific ecosystem. Here, any degradation processes must be primarily anaerobic or microaerophilic, given that only low amounts of oxygen will reach the benthos via thermohaline (bottom) currents [165]; the lack of sunlight and low temperature at these depths also drastically reduces non-biological degradation [43]. Very little is known about these biodegradation processes, although several plastic degraders

have been detected in cold marine habitats [168], and the sulfate-reducer *Desulfatitalea tepidiphilia* has been found attached to plastic surfaces in marine sediments in Germany [169]. Since sulfate reduction is the dominant type of bacterial respiration in sediments, this could be important in any biodegradation process. There are no accepted standard tests to assess anaerobic biodegradability in the marine environment. The elucidation of mechanisms for the biodegradation of plastics in the benthos is an important objective for future research.

Upon further breakdown, microplastic particles become nanoplastics, with a size of below 1000 nm according to Gigault et al. [139]. This group of marine pollutants overlaps with the so-called "primary nanoparticles", which are manufactured by industry, and are discussed in the next section (Engineered Nanoparticles). They are subject to the same degradation forces as microplastics but are more readily taken up by marine organisms and have been shown to have adverse effects on a variety of marine species [139,170]. Although it is assumed that nanoplastics will be subject to the same degradation mechanisms as microplastics, there is little direct evidence for this, mainly because of the challenges of isolating and analyzing them [57] and references therein. Gigault et al. [170] discuss these methods in some detail and point out the inherent problems. They consider that innovative tools, such as chemometrics, and newly developed instrumentation will be required for future studies on nanoplastics.

4. Engineered Nanoparticles

Particles of up to 100 nm in diameter are known as nanoparticles. The European Union defines a suspension of nanoparticles as that in which 50% or more particles have one or more external dimensions of 1–100 nm. These suspensions exist in nature (e.g., nanoclays), but in the recent past, they have been manufactured for specific human purposes. Such engineered nanoparticles (ENPs) may be used, for example, in coatings, insulating and magnetic materials, and as antimicrobial additives, principally nanometal oxides, such as TiO₂, which become increasingly antimicrobial under the action of UV In fact, metal and metal oxide nanoparticles are those most produced worldwide, with TiO₂ having the highest produced mass [171], and many, along with silica nanoparticles, are used as antifouling materials for protection of marine and seawater-associated structures [172–174]. The antifouling materials can, of course, also affect the non-target biota in the sea, especially when liberated from the protected structures by sloughing or friction. Toxic effects of the principal metal oxide ENPs in the marine environment decrease in the order Au > Zn > Ag > Cu > Ti > Carbon60 [175].

At some stage in their lifetime, even if not utilized directly in the marine environment, ENPs will be released into their surroundings and end up entering the oceans, where they can exert negative effects on the biota [176,177], including inhibition of movement and metabolic processes, oxidative stress and dysfunctional DNA replication [178]. NanoZnO particles ranging from 30 nm to 2 μ m have been shown to be highly toxic to flounder cells in culture and zebrafish embryos [179]. ENPs can readily pass from terrestrial waters into the ocean and thence into the marine food web [180]. It has been suggested that the main effects of ENPs on coastal marine life forms will be in sediments [181], where they have been shown to have toxic effects on foraminifera [182].

There has been little empirical study of the fate of ENPs in the aquatic environment [183], although it has been suggested that they can cause significant harm to the marine ecosystem [184] and references therein and that they have significant toxic effects on marine phytoplankton [185]. The release of silver NPs has been shown to alter the functioning of the marine food web by hampering important viral and bacterial processes [186]. In a mesocosm experiment, the addition of silver NPs, even at a low dose, affected planktonic communities, especially reducing the growth of the cyanobacterium *Synechococcus*. Viral auxiliary metabolic genes involved in cyanobacterial photosynthesis were also decreased.

It has, however, been suggested recently that ENPs may not be found at sufficiently high concentrations in the natural environment to pose a current problem [187]. More data on the effects and fate of nanoparticles released into the environment are necessary.

Life cycle and ecological risk assessments of ENPs in our oceans are essential to stimulate remediation processes and protect the marine environment.

5. Metallic Particles

Mining of polymetallic nodules, found on the surface of abyssal plains at around 4000 m depth, results in the release into the benthos of sediment plumes and nodule debris. These can be rich in Mn, Ni, Cu, and Co [188]. Fazey and Ryan [164] examined the aerobically grown bacteria present on the surface of nodules and in the overlying sediment, identifying *Halomonas aquamarina*, *H. meridiana*, and *Erythrobacter citreus* in both, but the genera *Arthrobacter*, *Kocuria*, *Loktanella*, *Marinobacter* and *Pseudoalteromonas* only in sediment within 4 cm of the nodule surface. Cho et al. [189] confirmed that the microbiome of nodules differs from the microbial population in the surrounding sediment, but their use of NGS technology led to the detection of a different set of bacteria and Archaea. Thaumarchaeota were found in both sediment and nodule, Mn-oxidizing bacteria (*Hyphomycrobium*, *Aurantimonas*, and *Marinobacter*) were predominant in nodules, and *Idiomarina*, *Erythrobacter*, and *Sulfitobacter* in sediments. Gillard et al. [188], based on their analyses, suggest the use of standard cultivation techniques for monitoring plume propagation; indicator organisms for sediment would be *Diezia maris* and *Pseudoalteromonas shioyasakiensis*, for nodules *Rhodococcus erythropolis* and water *Marinobacter flavimaris*.

Much of the iron found in aerosols over the oceans is anthropogenic in origin, resulting from the burning of fossil and biofuels and fires on land (biomass burning), a situation that is probably mirrored by zinc [190]. Conway et al. [191], using iron-isotope ratios, showed that deposition of anthropogenic Fe could reach almost 100% of the total Fe near highly populated areas. Their model suggested that this effect would be greatest in the Southern and Pacific Oceans, and this was echoed by Hamilton et al. in 2020 [192]. Much of the iron, and, indeed, many metals found in marine particles may be linked to the presence of microorganisms that produce metal-chelating siderophores. Chuang et al. [193] found that hydroxamate siderophores comprised a large part of the sinking particles ("marine snow" q.v.) collected in the Sargasso Sea. One of the important ecological functions of the siderophores produced by microorganisms is the release of iron from sinking particles to supply dissolved iron to the water column [194]. The export of iron from hydrothermal vents in the Southern East Pacific has likewise been linked to particles containing microorganisms [195].

Pollution by mercuric ions is a potential risk to human health, principally through the consumption of fish [196]. The main anthropogenic source of this metal is artisanal and small-scale gold mining, followed by the burning of fossil fuels [197]. The metal is converted to toxic methylmercury and dimethylmercury by microbial activity in the seas [198] and is largely associated with marine particulate matter [199]. The latter authors identified the sulfate-reducing bacterium, *Desulfovibrio desulfuricans*, as important for the uptake or exchange of Hg²⁺ in anaerobic environments. Marine Group II (MGII) archaeal genes associated with assimilatory sulfate reduction have been detected, along with MGII genes involved in surface adhesion, in samples collected from around the world during the Tara Oceans' circumnavigation trip [156]. The authors suggested that archaea MGII could be implicated in the degradation of marine particles, a more positive role for microbial biofilms in our oceans.

Marine microorganisms are also important in the production of metallic compounds. The mineral barite (or baryte), used principally in drilling muds, is produced in the oceans by barium binding initially to phosphate groups in bacterial cells or EPS; the thus concentrated barium is then converted in the marine environment to barite [200].

6. Sinking Particles ("Marine Snow") and Pollution

Marine snow is considered to be composed of heterogeneous agglomerates of living and dead organic matter of >500 um in size, formed by the attachment of organisms to the so-called "transparent exopolymer particles" (TEPs) that consist mainly of acidic polysac-

charides previously produced by phytoplankton and heterotrophic prokaryotes [201] and references therein. The particles contain diverse groups of eukaryotes, which may somewhat resemble, but certainly do not equal, the plankton in the local environment [202]. They have a highly variable composition and there are fundamental differences in particle composition between oligotrophic and eutrophic environments [202]. The microbial taxa associated with the particles are very different from those in the surrounding seawater and may contain increased oil degraders in oil-polluted environments [203,204] or methylmercury genes in saline waters in the North Sea [205]. Hence the particles may be "hot spots" for the degradative activity of surrounding pollutants [206].

Such sinking particles differ from floating particles in the oceans in carrying a changing population of prokaryotic species. Duret et al. [207] identified the prokaryotic populations on both types of particles in the Scotia Sea (Southern Ocean) and suggested that r-strategists, with generalized metabolic activities and rapid substrate consumption, were better adapted to sinking particles, with their changing environment, while K-strategists, specialized for complex organic material degradation, were better adapted to the more stable environment of semi-labile floating particles. So, for instance, pseudomonads and Rhodobacteriales were enriched on sinking particles, Flavobacteriales on floating. Datta et al. [208] had previously shown, using model polysaccharide particles, that the attached bacterial communities underwent rapid metabolic successions, driven by the environment. They suggested that there are 3 phases of colonization: attachment of a highly diverse community, selection of specific metabolic activities by the environment (reducing diversity) and replacement by secondary consumers, metabolizing the products of the second phase cells, and increasing diversity somewhat once more. Liu et al. [209], investigating differences between the two types of particles at low and high pressures in the New Britain Trench, Solomon Sea (Pacific), found that, although there were differences in prokaryotic populations on floating and sinking particles, similar groups participated in the degradation of diatom debris.

Even if similar organisms are involved in degradation, the physical act of sinking, whereby water flows past the particle surface, increases the rate of biodegradation simply by aiding the removal of the degradation products, driving the reaction to the right. This increase in microbiodegradation in sinking, as opposed to static, particles was elegantly demonstrated in a mathematical model developed by Alcolombri et al. [210].

The presence of eukaryotes in marine snow has been less frequently investigated. Bochdansky et al. [211] showed the presence of a fungal biomass equal to that of prokaryotes in bathypelagic particles from the North Atlantic and Arctic seas. Fungi and labyrinthulomycetes (the latter mainly labyrinthulids and thraustochytrids) dominated the biomass. These eukaryotes are tolerant of low temperatures and high pressures and were considered to be potentially important biodegraders in the particles. Schultz et al. [212], using metaproteomics and functional analyses of marine particles, showed that eukaryotes were more abundant in the particles than in the surrounding seawater. Those detected in particles were phytoplankton, Oomycetes, and Fungi. Greater amounts of viral proteins were also found in the particles. They reported rather small differences between bacterial proteins on particles and in the planktonic phase. The relative abundance of eukaryotes and viruses confirmed the results of López-Pérez et al. [213], who investigated the coastal waters off Alicante, Spain. They also found that there was an overrepresentation in the particle-associated microbiome of alpha, delta, and gamma proteobacteria, bacteroidetes (Flavobacteria), Planktomycetes, and Actinobacteria.

Gregson et al. [214] discuss the problems involved in studying oil degradation associated with marine snow. Most experiments have been performed using relatively high concentrations of oil in static or rotating vessels in the laboratory, although decreasing hydrocarbon concentrations as the particles sink from the contaminated site have been well demonstrated in loco. Their excellent review discusses not only the need to employ more relevant conditions in degradation studies but also the influence of dispersants. More research is necessary to determine the structure and function of the sinking, as well as the floating, particle microbiome and its relation to biodegradation activities.

7. Perspectives

A greater understanding of microbial responses to anthropogenic changes in our oceans is required to protect the marine environment. Recently, the number of studies on the biodegradation of synthetic plastics by microorganisms or enzymes has grown exponentially, representing a possibility to develop biological treatment technology for these wastes, of which most are in the public eye at this time.

There is a great need for the development of standard methods for the analysis of the chemistry and environmental effects of particles in the oceans. Much of the research carried out at present use unproven or non-standard methods, and many of the accepted effects of pollutant particles on the environment are based on laboratory experiments, often using simulated particles and conditions, or even using pure cultures of microorganisms. The changes that the particles undergo when exposed to the real environment are insufficiently understood. Life cycle and ecological risk assessments of ENPs and other particulate pollutants in our oceans are essential to stimulate and develop remediation processes. Research into the structure and function of the sinking, as well as floating, particle microbiome and its relation to biodegradation activities, could indicate potential ways of manipulating such activities for a natural "clean-up" of the environment.

Changes in traditional materials and practices to protect materials exposed to the marine environment, such as ships and marine constructions, are necessary to reduce anthropogenic pollution and these are under investigation, in the development of ecologically acceptable antifoulants and non-adhesive surfaces, for example.

Governments can accelerate these activities by introducing relevant legislation aimed at reducing polluting materials and activities, as has been done, for example, in Europe, Thailand, and some of the U.S. states, to combat plastics pollution. As part of the attributions of world managers, there are a series of measures that governments can use to protect the oceans from the harm caused by pollution. Among them can be cited fisheries management, conservation, and restoration of marine ecosystems, investments in research aimed at the environment sector, and effective inspection of the respective coastal areas. But the responsibility should not lie with the government sector alone. The awareness of the population must be used as a tool to maintain the quality of ecosystems. For this, effective educational policies must be applied in the most diverse strata of society and the information collected by the scientific community must be translated to other sectors as a way to mobilize independent and joint initiatives.

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References

- 1. Coleman, M.A.; Wood, G.; Filbee-Dexter, K.; Minne, A.J.P.; Goold, H.D.; Verges, A.; Marzinelli, E.M.; Steinberg, P.D.; Wernberg, T. Restore or redefine: Future trajectories for restoration. *Front. Mar. Sci.* **2020**, *7*, 237. [CrossRef]
- Hochella, M.F., Jr.; Mogk, D.W.; Ranville, J.; Allen, I.C.; Luther, G.W.; Marr, L.C.; McGrail, B.P.; Murayami, M.; Qafoku, N.P.; Yang, Y.; et al. Natural, incidental, and engineered nanomaterials and their impacts on the Earth system. *Science* 2019, 363, eaau8299. [CrossRef] [PubMed]
- 3. Bar-On, Y.M.; Milo, R. The biomass composition of the oceans: A blueprint of our blue planet. *Cell* **2019**, *179*, 1451–1454. [CrossRef] [PubMed]
- 4. Isobe, K.; Ohte, N. Ecological perspectives on microbes involved in N-cycling. *Microbes Environ.* 2014, 29, 4–16. [CrossRef]
- 5. Ando, N.; Barquera, B.; Bartlett, D.H.; Boyd, E.; Burnim, A.A.; Byer, A.S.; Colman, D.; Gillilan, R.E.; Gruebele, M.; Makhatadze, G.; et al. The molecular basis for life in extreme environments. *Annu. Rev. Biophys* **2021**, *50*, 343–372. [CrossRef] [PubMed]
- Poli, A.; Finore, I.; Romano, I.; Gioiello, A.; Lama, L.; Nicolaus, B. Microbial Diversity in Extreme Marine Habitats and Their Biomolecules. *Microorganisms* 2017, 5, 25. [CrossRef]

- Henderson, J.; Salem, H. CHAPTER 1: The atmosphere: Its developmental history and contributions to microbial evolution and habitat. In *Aerobiology: The Toxicology of Airborne Pathogens and Toxins*; RSC Publishing: London, UK, 2016; pp. 1–41. ISBN 978-1-84973-791-3. [CrossRef]
- Costerton, J.W.; Lewandowski, Z.; Caldwell, D.E.; Korber, D.R.; Lappin-Scott, H.M. Microbial biofilms. *Annu. Rev. Microbiol.* 1995, 49, 711–745. [CrossRef]
- 9. Pierce, E.C.; Dutton, R.J. Putting microbial interactions back into community contexts. *Curr. Opin. Microbiol.* **2022**, *65*, 56–63. [CrossRef]
- Roukaerts, A.; Deman, F.; Van der Linden, F.; Carnat, G.; Bratkic, A.; Moreau, S.; Dehairs, F.; Delille, B.; Tison, J.-L.; Fripiat, F. The biogeochemical role of a microbial biofilm in sea ice: Antarctic landfast sea ice as a case study. *Elem. Sci. Anthr.* 2021, 9, 00134. [CrossRef]
- Gadkari, J.; Bhattacharya, S.; Shrivastav, A. Importance and applications of biofilm in microbe-assisted bioremediation. In Development in Wastewater Treatment Research and Processes; Elsevier: Amsterdam, The Nertherland, 2022; pp. 153–173. [CrossRef]
- 12. Serra, D.O.; Hengge, R. Stress responses go three dimensional—the spatial order of physiological differentiation in bacterial macrocolony biofilms. *Environ. Microbiol.* **2014**, *16*, 1455–1471. [CrossRef]
- 13. Tan, D.; Svenningsen, S.L.; Middelboe, M. Quorum sensing determines the choice of antiphage defense strategy in *Vibrio anguillarum. mBio* **2015**, *6*, e00627-15. [CrossRef] [PubMed]
- 14. Bettencourt, S.; Costa, S.; Caeiro, S. Marine litter: A review of educative interventions. *Mar. Pollut. Bull.* **2021**, *168*, 112446. [CrossRef] [PubMed]
- 15. Sutherland, W.J.; Broad, S.; Caine, J.; Clarke, S.J.; Collins, A.M.; Dicks, L.V.; Doran, H.; Esmail, N.; Fleishman, E.; Frost, N.; et al. A horizon scan of global conservation issues for 2016. *Trends Ecol Evol.* **2016**, *31*, 44–53. [CrossRef] [PubMed]
- GEF—Secretariat of the Convention on Biological Diversity and the Scientific and Technical Advisory Panel. *Impacts of Marine Debris on Biodiversity: Current Status and Potential Solutions*; GEF: Montreal, QC, Canada, Technical Series No. 67; 2012. Available online: https://www.cbd.int/doc/publications/cbd-ts-67-en.pdf (accessed on 19 April 2022).
- Jacquin, J.; Cheng, J.; Odobel, C.; Pandin, C.; Conan, P.; Pujo-Pay, M.; Barbe, V.; Meisterzheim, A.-L.; Ghiglione, J.-F. Microbial ecotoxicology of marine plastic debris: A review on colonization and biodegradation by the "plastisphere". *Front. Microbiol.* 2019, 10, 865. [CrossRef]
- GESAMP. Sources, Fate and Effects of Microplastics in the Marine Environment: A Global Assessment; Kershaw, P.J., Ed.; International Maritime Organization: London, UK, 2015; p. 52. Available online: https://ec.europa.eu/environment/marine/good-environmental-status/descriptor-10/pdf/GESAMP_microplastics%20full%20study.pdf (accessed on 19 April 2022).
- 19. Barnes, D.K.A.; Galgani, F.; Thompson, R.C.; Barlaz, M. Accumulation and fragmentation of plastic debris in global environments. *Phil. Trans. R. Soc. B* 2009, *364*, 1985–1998. [CrossRef]
- 20. Ruiz, I.; Abascal, A.J.; Basurko, O.C.; Rubio, A. Modelling the distribution of fishing-related floating marine litter within the Bay of Biscay and its marine protected areas. *Environ. Pollut.* **2022**, 292, 118216. [CrossRef]
- 21. McIlgorm, A.; Raubenheimer, K.; McIlgorm, D.E.; Nichols, R. The cost of marine litter damage to the global marine economy: Insights from the Asia-Pacific into prevention and the cost of inaction. *Mar. Pollut. Bull.* **2022**, *174*, 113167. [CrossRef]
- Camargo, M.; Sandrini-Neto, L.; Carreira, R.; Camargo, M. Effects of hydrocarbon pollution in the structure of macrobenthic assemblages from two large estuaries in Brazil. *Mar. Poll. Bull.* 2017, 125, 66–76. [CrossRef]
- Häder, D.-P.; Banaszak, A.T.; Villafañe, V.E.; Narvarte, N.A.; Gonzalez, R.A.; Helbling, E.W. Anthropogenic pollution of aquatic ecosystems: Emerging problems with global implications. *Sci. Total Environ.* 2020, 713, 136586. [CrossRef]
- 24. Jadhav, S.; Sharma, S.; Sibi, G. Microbial degradation of petroleum hydrocarbons and factors influencing the degradation process. *Bioprocess Eng.* **2019**, *3*, 6–11. [CrossRef]
- Ławniczak, Ł.; Woźniak-Karczewska, M.; Loibner, A.P.; Heipieper, H.J.; Chrzanowski, L. Microbial degradation of hydrocarbons— Basic Principles for bioremediation: A review. *Molecules* 2020, 25, 856. [CrossRef] [PubMed]
- Mahjoubi, M.; Cappello, S.; Souissi, Y.; Jaouani, A.; Cherif, A. Microbial bioremediation of petroleum hydrocarbon-contaminated marine environments. In *Recent Insights in Petroleum Science and Engineering*; Zoveidavianpoor, M., Ed.; InTech: Rijeka, Croatia, 2018. [CrossRef]
- Baptista Neto, J.A.; Gaylarde, C.; da Fonseca, E.M. Microplastics: A pelagic habitat for microorganisms and invertebrates. In Handbook of Microplastics in the Environment; Rocha-Santos, T., Costa, M., Mouneyrac, C., Eds.; Springer: Cham, Switzerland, 2021. [CrossRef]
- Li, W.C.; Tse, H.F.; Fok, L. Plastic waste in the marine environment: A review of sources, occurrence and effects. *Sci. Total Environ.* 2016, 566–567, 333–349. [CrossRef] [PubMed]
- Richardson, K.; Asmutis-Silvia, R.; Drinkwin, J.; Gilardi, K.V.K.; Giskes, I.; Jones, G.; O'Brian, K.; Pragnell-Raasch, H.; Ludwig, L.; Antonelis, K.; et al. Building evidence around ghost gear: Global trends and analysis for sustainable solutions at scale. *Mar. Pollut. Bull.* 2019, 138, 222–229. [CrossRef] [PubMed]
- Angiolillo, M. Debris in Deep Water. In World Seas: An Environmental Evaluation, 2nd ed.; Sheppard, C., Ed.; Academic Press: Cambridge, MA, USA, 2019; Chapter 14; pp. 251–268. [CrossRef]
- 31. Angiolillo, M.; Fortibuoni, T. Impacts of marine litter on Mediterranean reef systems: From shallow to deep waters. *Front. Mar. Sci.* 2020, *7*, 826. [CrossRef]

- 32. Lee, J.-W.; Nam, J.-H.; Kim, Y.-H.; Lee, K.-H.; Lee, D.-H. Bacterial communities in the initial stage of marine biofilm formation on artificial surfaces. *J. Microbiol.* 2008, *46*, 174–182. [CrossRef]
- 33. Caruso, G. Microplastics in marine environments: Possible interactions with the microbial assemblage. *J. Pollut. Eff. Cont.* 2015, 3, e111. [CrossRef]
- Lee, D.-I.; Cho, H.-S.; Jeong, S.-B. Distribution characteristics of marine litter on the sea bed of the East China Sea and the South Sea of Korea. *Estuar Coast. Shelf Sci.* 2006, 70, 187–194. [CrossRef]
- Melli, V.; Angiolillo, M.; Ronchi, F.; Canese, S.; Giovanardi, O.; Querin, S.; Fortibuoni, T. The first assessment of marine debris in a Site of Community Importance in the north-western Adriatic Sea (Mediterranean Sea). *Mar. Pollut. Bull.* 2017, 114, 821–830. [CrossRef]
- Watters, D.L.; Yoklavich, M.M.; Love, M.S.; Schroeder, D.M. Assessing marine debris in deep seafloor habitats off California. *Mar. Pollut. Bull.* 2010, 60, 131–138. [CrossRef]
- 37. Sherrington, C. Plastics in the Marine Environment; Eunomia Research & Consulting Ltd.: Bristol, UK, 2016; p. 16.
- Wei, C.-L.; Rowe, G.T.; Nunnally, C.C.; Wicksten, M.K. Anthropogenic "Litter" and macrophyte detritus in the deep Northern Gulf of Mexico. *Mar. Pollut. Bull.* 2012, 64, 966–973. [CrossRef]
- Carvalho-Souza, G.F.; Llope, M.; Tinôco, M.S.; Medeiros, D.V.; Maia-Nogueira, R.; Sampaio, C.L.S. Marine litter disrupts ecological processes in reef systems. *Mar. Pollut. Bull.* 2018, 133, 464–471. [CrossRef] [PubMed]
- Keller, A.A.; Fruh, E.L.; Johnson, M.M.; Simon, V.; McGourty, C. Distribution and abundance of anthropogenic marine debris along the shelf and slope of the US West Coast. *Mar. Pollut. Bull.* 2010, 60, 692–700. [CrossRef]
- Carson, H.S.; Nerheim, M.S.; Carroll, K.A.; Eriksen, M. The plastic-associated microorganisms of the North Pacific Gyre. *Mar. Pollut. Bull.* 2013, 75, 126–132. [CrossRef] [PubMed]
- Schlundt, C.; Welch, M.J.L.; Knochel, A.M.; Zettler, E.; Amarel-Zettler, L. Spatial structure in the "Plastisphere": Molecular resources for imaging microscopic communities on plastic marine debris. *Mol. Ecol. Resour.* 2019, 20, 620–634. [CrossRef]
- 43. UNEP. Plastic Debris in the Ocean. UNEP Year Book 2014 Emerging Issues Update. 2014. UN Environment Yearbooks/Global Environmental Outlook Yearbooks | UNEP-UN Environment Programme. Available online: https://www.unep.org/resources/ year-books (accessed on 19 April 2022).
- 44. Sherman, D.; Besteiro, S.; Fox, C.; Sherman, K. International Coastal Cleanup Report: 2009; Ocean Conservancy: Washington, DC, USA, 2007. [CrossRef]
- 45. Anfuso, G.; Lynch, K.; Williams, A.T.; Perales, J.A.; Silva, C.P.; Nogueira, M.R.; Maanan, M.; Pretti, C.; Pranzini, E.; Winter, C.; et al. Comments on marine litter in oceans, seas and beaches: Characteristics and impacts. *Ann. Mar. Biol. Res.* **2015**, *2*, 1008.
- 46. Margiotta, F.; Balestra, C.; Buondonno, A.; Casotti, R.; D'Ambra, I.; Di Capua, I.; Gallia, R.; Mazzocchi, M.G.; Merquiol, L.; Pepi, M.; et al. Do plankton reflect the environmental quality status? The case of a post-industrial Mediterranean Bay. *Mar. Environ. Res.* 2020, 160, 104980. [CrossRef]
- Hoeksema, B.W.; Roos, P.J.; Cadée, G.C. Trans-Atlantic rafting by the brooding reef coral Favia fragum on man-made flotsam. Mar. Ecol. Prog. Ser. 2012, 445, 209–218. [CrossRef]
- 48. Kiessling, T.; Gutow, L.; Thiel, M. Marine litter as habitat and dispersal vector. In *Marine Anthropogenic Litter*; Bergmann, M., Gutow, L., Klages, M., Eds.; Springer: Cham, Switzerland, 2015; pp. 141–180. [CrossRef]
- 49. Woodall, L.C.; Jungblut, A.D.; Hopkins, K.; Hall, A.C.; Robinson, L.; Gwinnett, C.; Paterson, G. Deep-sea anthropogenic macrodebris harbours rich and diverse communities of bacteria and archaea. *PLoS ONE* **2018**, *13*, e0206220. [CrossRef]
- Little, B.J.; Lee, J.S. Microbiologically influenced corrosion. In Wiley Series in Corrosion; John Wiley and Sons, Inc.: Hoboken, NJ, USA, 2007.
- 51. McBeth, J.M.; Little, B.J.; Ray, R.I.; Farrar, K.; Emerson, D. Neutrophilic iron-oxidizing "Zetaproteobacteria" and mild steel corrosion in nearshore marine environments. *Appl. Environ. Microbiol.* **2011**, 77, 1405–1412. [CrossRef] [PubMed]
- Tourova, T.P.; Sokolova, D.S.; Nazina, T.N.; Laptez, A.B. Comparative analysis of the taxonomic composition of bacterial fouling developing on various materials exposed to aqueous environments. *Microbiology* 2021, 90, 416–427. [CrossRef]
- 53. Muthukrishnan, T.; Al Khaburi, M.; Abed, R.M.M. Fouling microbial communities on plastics compared with wood and steel: Are they substrate- or location-specific? *Microb. Ecol.* **2019**, *78*, 361–374. [CrossRef] [PubMed]
- 54. Mansour, R.; Elshafei, A. Protection of metal surfaces from microbial colonization. Ann. Res. Rev. Biol. 2017, 14, 1–8. [CrossRef]
- 55. Castro, İ.B.; Westphal, E.; Fillmann, G. Tintas anti-incrustantes de terceira geração: Novos biocidas no ambiente aquático. [Third generation antifouling paints: New biocides in the aquatic environment]. *Quim. Nova* **2011**, *34*, 1021–1031. [CrossRef]
- Thomas, K.V.; Brooks, S. The environmental fate and effects of antifouling paint biocides. *Biofouling* 2010, 26, 73–88. [CrossRef] [PubMed]
- Gaylarde, C.C.; Baptista Neto, J.A.; da Fonseca, E. Paint fragments as polluting microplastics: A brief review. *Mar. Pollut. Bull.* 2021, 162, 111847. [CrossRef]
- Soroldoni, S.; da Silva, S.V.; Castro, Í.B.; Martins, C.M.G.; Pinho, L.; Lopes, G. Antifouling paint particles cause toxicity to benthic organisms: Effects on two species with different feeding modes. *Chemosphere* 2020, 238, 124610. [CrossRef]
- 59. Eklund, B.; Johansson, L.; Ytreberg, E. Contamination of a boatyard for maintenance of pleasure boats. *J. Soils Sed.* **2014**, *14*, 955–967. [CrossRef]
- Soroldoni, S.; Castro, I.B.; Abreu, F.; Duarte, F.A.; Choueri, R.B.; Moller, O.O., Jr.; Fillmann, G.; Pinho, G.L.L. Antifouling paint particles: Sources, occurrence, composition and dynamics. *Water Res.* 2018, 137, 47–56. [CrossRef]

- 61. Takahashi, C.K.; Turner, A.; Millward, G.E.; Glegg, G.A. Persistence and metallic composition of paint particles in sediments from a tidal inlet. *Mar. Pollut. Bull.* **2012**, *64*, 133–137. [CrossRef]
- Jin, H.; Tian, L.; Bing, W.; Zhao, J.; Ren, L. Bioinspired marine antifouling coatings: Status, prospects, and future. *Prog. Mater. Sci.* 2022, 124, 100889. [CrossRef]
- 63. Wang, Y.; Zhao, W.; Han, L.; Tam, K.C. Superhydrophobic surfaces from sustainable colloidal systems. *Curr. Opin. Colloid Interface Sci.* 2022, *57*, 101534. [CrossRef]
- 64. Boxall, S. From rubber ducks to ocean gyres. Nature 2009, 459, 1058–1059. [CrossRef]
- 65. Capolupo, M.; Sørensen, L.; Jayasena, K.D.R.; Booth, A.; Fabbri, E. Chemical composition and ecotoxicity of plastic and car tire rubber leachates to aquatic organisms. *Water Res.* 2020, *169*, 115270. [CrossRef] [PubMed]
- 66. Shah, A.A.; Hasan, F.; Shah, N.; Kanwal, N.; Zeb, S. Biodegradation of natural and synthetic rubbers: A review. *Int. Biodeterior. Biodegrad.* **2013**, *83*, 145–157. [CrossRef]
- 67. Andler, R.; Hiessl, S.; Yücel, O.; Tesch, M.; Steinbuchel, A. Cleavage of poly(*cis*-1,4-isoprene) rubber as solid substrate by culture of *Gordonia polyisoprenivorans*. *New Biotechnol.* **2018**, 44, 6–12. [CrossRef]
- Imai, S.; Ichikawa, K.; Muramatsu, Y.; Kasai, D.; Masai, E.; Fukuda, M. Isolation and characterisation of *Streptomyces, Actinoplanes* and *Methylibium* strains that are involved in degradation of natural rubber and synthetic poly(*cis*-1,4-isoprene). *Enzym. Microb. Technol.* 2011, 49, 526–531. [CrossRef] [PubMed]
- Linos, A.; Berekaa, M.M.; Reichelt, R.; Keller, U.; Schmitt, J.; Flemming, H.; Koppenstedt, R.; Steinbuchel, A. Biodegradation of *cis*-1,4-polyisoprene rubbers by distinct actinomycetes: Microbial strategies and detailed surface analysis. *Appl. Environ. Microbiol.* 2000, *66*, 1639–1645. [CrossRef] [PubMed]
- Carrión, O.; Larke-Mejía, N.L.; Gibson, L.; Ul Haque, M.F.; Ramiro-Garcia, J.; McGenity, T.J.; Murrell, J.C. Gene probing reveals the widespread distribution, diversity and abundance of isoprene-degrading bacteria in the environment. *Microbiome* 2018, *6*, 219. [CrossRef]
- Johnston, A.; Crombie, A.T.; Khawand, M.E.; Sims, L.; Whited, G.M.; McGenity, T.J.; Murrell, J.C. Identification and characterisation of isoprene-degrading bacteria in an estuarine environment. *Environ. Microbiol.* 2017, 19, 3526–3537. [CrossRef]
- 72. Dris, R.; Gasperi, J.; Saad, M.; Mirande, C.; Tassin, B. Synthetic fibers in atmospheric fallout: A source of microplastics in the environment? *Mar. Pollut. Bull.* **2016**, *104*, 290–293. [CrossRef]
- 73. Kwak, J.I.; Liu, H.; Wang, D.; Lee, Y.H.; Lee, J.-S.; An, Y. Critical review of environmental impacts of microfibers in different environmental matrices. *Comp. Biochem. Physiol. Pt C Toxicol. Pharmacol.* 2021, 251, 109196. [CrossRef] [PubMed]
- Pirc, U.; Vidmar, M.; Mozer, A.; Kržan, A. Emissions of microplastic fibers from microfiber fleece during domestic washing. Environ. Sci. Pollut. Res. 2016, 23, 22206–22211. [CrossRef] [PubMed]
- 75. Gaylarde, C.; Baptista Neto, J.A.; da Fonseca, E. Plastic microfibre pollution: How important is clothes' laundering? *Heliyon* **2021**, 7, e07105. [CrossRef] [PubMed]
- Henry, B.; Laitala, K.; Klepp, I.G. Microfibres from apparel and home textiles: Prospects for including microplastics in environmental sustainability assessment. *Sci. Total Environ.* 2019, 652, 483–494. [CrossRef]
- Industrievereinigung Chemiefaser; Statista Inc. Worldwide Production Volume of Chemical and Textile Fibers from 1975 to 2018. 2018. Available online: https://www.statista.com/statistics/263154/worldwide-production-volume-of-textile-fibers-since-1975/ (accessed on 13 November 2021).
- 78. Hartline, N.; Bruce, N.; Karba, S.; Ruff, E.O.; Sonar, S.U.; Holden, P.A. Microfiber masses recovered from conventional machine washing of new or aged garments. *Environ. Sci. Technol.* **2016**, *50*, 11532–11538. [CrossRef]
- 79. Andrady, A. Microplastics in the marine environment. Mar. Pollut. Bull. 2011, 8, 1596–1605. [CrossRef]
- 80. Guo, H.; Qian, Y.; Qiu, P.; Kong, Z.; Zheng, X.; Tang, Z.; Guo, H. Textile wastewater treatment for water reuse: A case study. *Processes* **2019**, *7*, 34. [CrossRef]
- Hassaan, M.A.; El Nemr, A. Health and environmental hazards of synthetic dyes. Am. J. Environ. Sci. Eng. 2017, 1, 64–67. [CrossRef]
- Ho, H.; Watanabe, T. Distribution and removal of nonylphenol ethoxylates and nonylphenol from textile wastewater—A comparison of a cotton and a synthetic fiber factory in Vietnam. *Water* 2017, *9*, 386. [CrossRef]
- 83. Luongo, G. Chemicals in Textiles: A Potential Source for Human Exposure and Environmental Pollution. Ph.D. Thesis, Stockholm University, Stockholm, Sweden, 2015.
- 84. Samchetshabam, G.; Hussan, A. Impact of textile dyes waste on aquatic environments and its treatment. *Environ. Ecol.* **2017**, *35*, 2349–2353. [CrossRef]
- 85. Almroth, B.M.C.; Åström, L.; Roslund, S.; Petersson, H.; Johansson, M.; Persson, M. Quantifying shedding of synthetic fibers from textiles; a source of microplastics released into the environment. *Environ. Sci. Pollut. Res.* **2018**, *25*, 1191. [CrossRef] [PubMed]
- Gago, J.; Carretero, O.; Filgueiras, A.V.; Viñas, L. Synthetic microfibers in the marine environment: A review on their occurrence in seawater and sediments. *Mar. Pollut. Bull.* 2018, 127, 365–376. [CrossRef] [PubMed]
- Ladewig, S.M.; Bao, S.; Chow, A.T. Natural fibers: A missing link to chemical pollution dispersion in aquatic environments. *Environ. Sci. Technol.* 2015, 49, 12609–12610. [CrossRef] [PubMed]
- Stone, C.; Windsor, F.M.; Munday, M.; Durance, I. Natural or synthetic–how global trends in textile usage threaten freshwater environments. *Sci. Total Environ.* 2020, 718, 134689. [CrossRef]

- 89. Stanton, T.; Johnson, M.; Nathanail, P.; Macnaughtan, W.; Gomes, R. Freshwater and airborne textile fibre populations are dominated by 'natural', not microplastic, fibres. *Sci. Total Environ.* **2019**, *666*, 377–389. [CrossRef]
- 90. Avio, C.G.; Gorbi, S.; Regoli, F. Plastics and microplastics in the oceans: From emerging pollutants to emerged threat. *Mar. Environ. Res.* 2017, *128*, 2–11. [CrossRef]
- Sharabatia, M.A.; Abokwiek, R.; Al-Othman, A.; Tawalbeh, M.; Karaman, C.; Orooji, Y.; Karimi, F. Biodegradable polymers and their nano-composites for the removal of endocrine-disrupting chemicals (EDCs) from wastewater: A review. *Environ. Res.* 2021, 202, 111694. [CrossRef]
- 92. Darbre, P.D. What are endocrine disrupters and where are they found? In *Endocrine Disruption and Human Health*; Academic Press: Cambridge, MA, USA, 2022; pp. 3–29. [CrossRef]
- 93. Kabir, E.R.; Rahman, M.S.; Rahman, I. A review on endocrine disruptors and their possible impacts on human health. *Environ. Toxicol. Pharmacol.* **2015**, *40*, 241–258. [CrossRef]
- 94. Girard, E.; Kaliwoda, M.; Schmall, W.W.; Woerdeide, G.; Orsi, W. Textile waste and microplastic induce activity and development of unique hydrocarbon-degrading marine bacterial communities. *bioRxiv* 2020, *2*, 939876. [CrossRef]
- 95. Halsband, C.; Herzke, D. Plastic litter in the European Arctic: What do we know? Emerg. Contam. 2019, 5, 308–318. [CrossRef]
- 96. Setala, O.; Fleming-Lehtinen, V.; Lehtiniemi, M. Ingestion and transfer of microplastics in the planktonic food web. *Environ. Pollut.* **2014**, *185*, 77–83. [CrossRef] [PubMed]
- 97. Rogers, K.L.; Carreres-Calabuig, J.A.; Gorokhova, E.; Posth, N.R. Micro-by-micro interactions: How microorganisms influence the fate of marine microplastics. *Limnol. Oceanogr.* 2020, *5*, 18–36. [CrossRef]
- Cózar, A.; Echevarría, F.; González-Gordillo, J.I.; Irigoien, X.; Ubeda, B.; Hernandez-Leon, S.; Palma, A.T.; Navarro, S.; Garciade-Lomas, J.; Ruiz, A.; et al. Plastic debris in the open ocean. *Proc. Natl. Acad. Sci. USA* 2014, 111, 10239–10244. [CrossRef] [PubMed]
- Montoto-Martínez, T.; Hernández-Brito, J.J.; Gelado-Caballero, M.D. Pump-underway ship intake: An unexploited opportunity for Marine Strategy Framework Directive (MSFD) microplastic monitoring needs on coastal and oceanic waters. *PLoS ONE* 2020, 15, e0232744. [CrossRef] [PubMed]
- 100. Zettler, E.R.; Mincer, T.J.; Amaral-Zettler, L.S. Life in the 'plastisphere': Microbial communities on plastic marine debris. *Environ. Sci. Technol.* **2013**, *47*, 7137–7146. [CrossRef] [PubMed]
- Dudek, K.L.; Cruz, B.N.; Polidoro, B.; Neuer, S. Microbial colonization of microplastics in the Caribbean Sea. *Limnol. Oceanogr.* Lett. 2020, 5, 5–17. [CrossRef]
- Kirstein, I.V.; Wichels, A.; Gullans, E.; Krohne, G.; Gerdts, G. The Plastisphere—Uncovering tightly attached plastic "specific" microorganisms. *PLoS ONE* 2019, 14, e0215859. [CrossRef] [PubMed]
- Oberbeckmann, S.; Kreikemeyer, B.; Labrenz, M. Environmental factors support the formation of specific bacterial assemblages on microplastics. *Front. Microbiol.* 2018, *8*, 2709. [CrossRef] [PubMed]
- 104. Moore, R.E.; Millar, B.C.; Moore, J.E. Antimicrobial resistance (AMR) and marine plastics: Can food packaging litter act as a dispersal mechanism for AMR in oceanic environments? *Mar. Pollut. Bull.* **2020**, *150*, 110702. [CrossRef] [PubMed]
- 105. Yang, Y.; Liu, G.; Song, W.; Ye, C.; Lin, H.; Li, Z.; Liu, W. Plastics in the marine environment are reservoirs for antibiotic and metal resistance genes. *Environ. Int.* 2019, 123, 79–86. [CrossRef]
- 106. Zhou, J.; Lin, Z.-J.; Cai, Z.-H.; Zeng, Y.-H.; Zhu, J.-M.; Dhu, X.-P. Opportunistic bacteria use quorum sensing to disturb coral symbiotic communities and mediate the occurrence of coral bleaching. *Environ. Microbiol.* 2020, 22, 1944–1962. [CrossRef] [PubMed]
- 107. Frère, L.; Maignien, L.; Chalopin, M. Microplastic bacterial communities in the Bay of Brest: Influence of polymer type and size. *Environ. Pollut.* **2018**, 242 *Pt A*, 614–625. [CrossRef]
- 108. Bryant, J.A.; Clemente, T.M.; Viviani, D.A.; Fong, A.A.; Thomas, K.A.; Kemp, P.; Karl, D.M.; White, A.E.; DeLong, E. Diversity and activity of communities inhabiting plastic debris in the North Pacific Gyre. *mSystems* 2016, 1, e00016–e00024. [CrossRef] [PubMed]
- Goldstein, M.C.; Carson, H.S.; Eriksen, M. Relationship of diversity and habitat area in North Pacific plastic-associated rafting communities. *Mar. Biol.* 2014, 161, 1441–1453. [CrossRef]
- 110. Soares, M.O.; de Lima Xavier, F.R.; Dias, N.M.; da Silva, M.Q.M.; Lima, J.P.; Barroso, C.X.; Vieira, L.M.; Paiva, S.V.; Matthews-Cascon, H.; Bezerra, L.E.A.; et al. Alien hotspot: Benthic marine species introduced in the Brazilian semiarid coast. *Mar. Pollut. Bull.* 2022, 174, 113250. [CrossRef] [PubMed]
- Castro, M.C.T.; Fileman, T.W.; Hall-Spencer, J.M. Invasive species in the Northeastern and Southwestern Atlantic Ocean: A review. *Mar. Pollut. Bull.* 2017, 116, 41–47. [CrossRef]
- 112. Hughes, K.A.; Pescott, O.L.; Peyton, J.; Adriaens, T.; Cottier-Cook, E.J.; Key, G.; Rabbitsch, W.; Tricarico, E.; Barnes, D.K.A.; Baxter, N.; et al. Invasive non-native species likely to threaten biodiversity and ecosystems in the Antarctic Peninsula region. *Glob. Change Biol.* 2020, 26, 2702–2716. [CrossRef]
- Richard, H.; Carpenter, E.J.; Komada, T.; Palmer, P.T.; Rochman, C.M. Biofilm facilitates metal accumulation onto microplastics in estuarine waters. *Sci. Total Environ.* 2019, 683, 600–608. [CrossRef]
- 114. Freese, H.M.; Methner, A.; Overmann, J. Adaptation of surface-associated bacteria to the open ocean: A genomically distinct subpopulation of *Phaeobacter gallaeciensis* colonizes Pacific mesozooplankton. *Front. Microbiol.* **2017**, *8*, 1659. [CrossRef]

- 115. Debroas, D.; Mone, A.; Halle, A.T. Plastics in the North Atlantic garbage patch: A boat-microbe for hitchhikers and plastic degraders. *Sci. Total Environ.* **2017**, 599–600, 1222–1232. [CrossRef]
- 116. Delacuvellerie, A.; Cyriaque, V.; Gobert, S.; Benali, S.; Wattiez, R. The plastisphere in marine ecosystem hosts potential specific microbial degraders including *Alcanivorax borkumensis* as a key player for the low-density polyethylene degradation. *J. Hazard. Mat.* 2019, 280, 120899. [CrossRef] [PubMed]
- 117. Ghosh, S.K.; Pal, S.; Ray, S. Study of microbes having potentiality for biodegradation of plastics. *Environ. Sci. Pollut. Res. Int.* **2013**, 20, 4339–4355. [CrossRef] [PubMed]
- 118. Baptista Neto, J.A.B.; Gaylarde, C.; Beech, I.; Bastos, A.; Quaresma, V.S.; de Carvalho, D.G. Microplastics and attached microorganisms in sediments of the Vitória Bay estuarine system in SE Brazil. *Ocean. Coast. Manag.* 2019, 169, 247–253. [CrossRef]
- De Tender, C.; Devriese, L.I.; Haegeman, A.; Maes, S.; Vangeyte, J.; Cattrijsse, A.; Dawyndt, P.; Ruttink, T. Temporal dynamics of bacterial and fungal colonization on plastic debris in the North Sea. *Environ. Sci. Technol.* 2017, *51*, 7350–7360. [CrossRef] [PubMed]
- 120. Kowalczyk, A.; Chyc, M.; Ryszka, P.; Latowski, D. *Achromobacter xylosoxidans* as a new microorganism strain colonizing highdensity polyethylene as a key step to its biodegradation. *Environ. Sci. Pollut. Res.* **2016**, *23*, 11349–11356. [CrossRef]
- 121. Gilan, O.; Hadar, Y.; Sivan, A. Colonization, biofilm formation and biodegradation of polyethylene by a strain of *Rhodococcus ruber*. *Appl. Microbiol. Biotechnol.* **2004**, *65*, 97–104. [CrossRef]
- Hadad, D.; Geresh, S.; Sivan, A. Biodegradation of polyethylene by the thermophilic bacterium *Brevibacillus borstelensis*. J. Appl. Microbiol. 2005, 98, 1093–1100. [CrossRef]
- 123. Harshvardhan, K.; Jha, B. Biodegradation of low-density polyethylene by marine bacteria from pelagic waters, Arabian Sea, India. *Mar. Pollut. Bull.* **2013**, *77*, 100–106. [CrossRef]
- 124. Huang, X.; Cao, L.; Qin, Z.; Li, S.; Kong, W.; Liu, Y. Tat-independent secretion of polyethylene terephthalate hydrolase PETase in *Bacillus subtilis* 168 mediated by its native signal peptide. *J. Agric. Food Chem.* **2018**, *66*, 13217–13227. [CrossRef]
- 125. Sudhakar, M.; Doble, M.; Murthy, P.S.; Venkatesan, R. Marine microbe-mediated biodegradation of low- and high-density polyethylenes. *Int. Biodegrad.* 2008, *61*, 203–213. [CrossRef]
- 126. Roth, C.; Wei, R.; Oeser, T.; Then, J.; Follner, C.; Zimmermann, W.; Strater, N. Structural and functional studies on a thermostable polyethylene terephthalate degrading hydrolase from *Thermobifida fusca*. *Appl. Microbiol. Biotechnol.* **2014**, *98*, 7815–7823. [CrossRef] [PubMed]
- 127. Ribitsch, D.; Herrero Acero, E.; Greimel, K.; Dellacher, A.; Zitzenbacher, S.; Marold, A.; Rodriguez, R.D.; Steinkellner, G.; Gruber, K.; Schwab, H.; et al. A new esterase from *Thermobifida halotolerans* hydrolyses polyethylene terephthalate (PET) and polylactic acid (PLA). *Polymers* **2021**, *4*, 617. [CrossRef]
- 128. Oberbeckmann, S.; Loeder, M.G.J.; Gerdts, G.; Osborn, A.M. Spatial and seasonal variation in diversity and structure of microbial biofilms on marine plastics in Northern European waters. *FEMS Microb. Ecol.* **2014**, *90*, 478–492. [CrossRef] [PubMed]
- 129. Yan, F.; Wei, R.; Cui, Q.; Bornscheuer, U.T.; Liu, Y.-J. Thermophilic whole-cell degradation of polyethylene terephthalate using engineered *Clostridium thermocellum*. *Microb. Biotechnol.* **2021**, *14*, 374–385. [CrossRef]
- Kawai, F.; Oda, M.; Tamashiro, T.; Waku, T.; Tanaka, N.; Yamamoto, M.; Mizushima, H.; Miyakawa, T.; Tanokura, M. A novel Ca²⁺-activated, thermostabilized polyesterase capable of hydrolyzing polyethylene terephthalate from *Saccharomonospora viridis* AHK190. *Appl. Microbiol. Biotechnol.* 2014, 98, 10053–10064. [CrossRef]
- 131. Yamada-Onodera, K.; Mukumoto, H.; Katsuyaya, Y.; Saiganji, A.; Tani, Y. Degradation of polyethylene by a fungus *Penicillium* simplicissimus YK. *Polym. Degrad. Stab.* **2001**, *72*, 323–327. [CrossRef]
- 132. Almeida, E.L.; Carrillo, R.A.F.; Jackson, S.A.; Dobson, A.D.W. In silico screening and heterologous expression of a polyethylene terephthalate hydrolase (PETase)-like enzyme (SM14est) with polycaprolactone (PCL)-degrading activity, from the marine sponge-derived strain *Streptomyces* sp. SM14. *Front. Microbiol* **2019**, *10*, 2187. [CrossRef]
- 133. Pometto, A.L.; Lee, B.; Johnson, K.E. Production of an extracellular polyethylene-degrading enzyme by *Streptomyces* sp. *Appl. Environ. Microbiol.* **1992**, *58*, 731–733. [CrossRef]
- 134. Shao, H.; Chen, M.; Fei, X.; Zhang, R.; Zhong, Y.-X.; Ni, W.; Tao, X.; He, X.; Zhang, E.; Yong, B.; et al. Complete genome sequence and characterization of a polyethylene biodegradation strain, *Streptomyces albogriseolus* LBX-2. *Microorganisms* **2019**, *7*, 379. [CrossRef]
- 135. Devi, R.S.; Kannan, V.R.; Nivas, D. Biodegradation of HDPE by *Aspergillus* spp. from marine ecosystem of Gulf of Mannar, India. *Mar. Pollut. Bull.* **2015**, *96*, 32–40. [CrossRef]
- 136. Sowmya, H.V.; Ramalingappa, M.; Krishnappa, M.; Thippeswamy, B. Degradation of polyethylene by *Penicillium simplicissimum* isolated from local dumpsite of Shivamogga district. *Environ. Dev. Sustain.* **2014**, *17*, 731–745. [CrossRef]
- 137. Pattnaik, P.; Dangayach, G.S.; Bhardwaj, A.K. A review on the sustainability of textile industries wastewater with and without treatment methodologies. *Rev. Environ. Health* **2018**, *33*, 163–203. [CrossRef] [PubMed]
- 138. Kvale, K.F.; Friederike, P.A.E.; Oschlies, A. A critical examination of the role of marine snow and zooplankton fecal pellets in removing ocean surface microplastic. *Front. Mar. Sci.* 2020, *6*, 808. [CrossRef]
- 139. Gigault, J.; Ter Halle, A.; Baudrimont, M.; Pascal, P.-Y.; Gauffre, F.; Phi, T.-L.; El Hadri, H.; Grassi, B.; Reynaud, S. Current opinion: What is a nanoplastic? *Environ. Pollut.* **2018**, 235, 1030–1034. [CrossRef] [PubMed]
- 140. Shimao, M. Biodegradation of plastics. Curr. Opin. Biotechnol. 2001, 12, 242–247. [CrossRef]

- 141. Mabrouk, M.M.; Sabry, S.A. Degradation of poly (3hydroxybutyrate) and its copolymer poly (3-hydroxybutyrate-co3-hydroxyvalerate) by a marine *Streptomyces* sp. SNG9. *Microbiol. Res.* **2001**, *156*, 323–335. [CrossRef]
- Paço, A.; Duarte, K.; da Costa, J.P.; Santos, P.S.M.; Pereira, R.; Freitas, A.C.; Duarte, A.C.; Rocha-Santos, T.A.P. Biodegradation of polyethylene microplastics by the marine fungus *Zalerion maritimum*. *Sci. Total Environ.* 2017, *586*, 10–15. [CrossRef]
- 143. Sekiguchi, T.; Saika, A.; Nomura, K.; Watanabe, T.; Watanabe, T.; Fujimoto, Y.; Enoki, M.; Sato, T.; Kato, C.; Kanehiro, H. Biodegradation of aliphatic polyesters soaked in deep seawaters and isolation of poly(ε-caprolactone)-degrading bacteria. *Polym. Degrad. Stab.* 2011, 96, 1397–1403. [CrossRef]
- 144. Suzuki, M.; Tachibana, Y.; Oba, K.; Takizawa, R.; Kasuya, K.-i. Microbial degradation of poly(ε-caprolactone) in a coastal environment. *Polym. Degrad. Stab.* **2018**, 149, 1–8. [CrossRef]
- 145. Chua, T.-K.; Tseng, M.; Yang, M.-K. Degradation of poly (ε-caprolactone) by thermophilic *Streptomyces thermoviolaceus* subsp. *thermoviolaceus* 76T-2. *AMB Express* **2013**, *3*, 8. [CrossRef]
- 146. Oberbeckmann, S.; Labrenz, M. Marine microbial assemblages on microplastics: Diversity, adaptation, and role in degradation. *Ann. Rev. Mar. Sci.* 2020, 12, 209–232. [CrossRef] [PubMed]
- 147. Tomita, K.; Kuroki, Y.; Nagai, K. Isolation of thermophiles degrading poly (*L*-lactic acid). *J. Biosci. Bioeng.* **1999**, *87*, 752–755. [CrossRef]
- 148. Kim, D.Y.; Rhee, Y.H. Biodegradation of microbial and synthetic polyesters by fungi. *Appl. Microbiol. Biotechnol.* **2003**, *61*, 300–308. [CrossRef] [PubMed]
- 149. Akutsu, Y.; Nakajima-Kambe, T.; Nomura, N.; Nakahara, T. Purification and Properties of a Polyester Polyurethane-Degrading Enzyme from *Comamonas acidovorans* TB-35. *Appl. Environ. Microbiol.* **1998**, *64*, 62–67. [CrossRef]
- 150. Howard, G.T.; Ruiz, C.; Hillard, N.P. Growth of *Pseudomonas chlororaphis* on a polyester-polyurethane and the purification and characterization of a polyurethane-esterase enzyme. *Int. Biodeter. Biodegrad.* **1999**, *43*, 7–12. [CrossRef]
- Peng, Y.H.; Shih, Y.H.; Lai, Y.C.; Liu, Y.-Z.; Liu, Y.-T.; Lin, N.-C. Degradation of polyurethane by bacterium isolated from soil and assessment of polyurethanolytic activity of a *Pseudomonas putida* strain. *Environ. Sci. Pollut. Res.* 2014, 21, 9529–9537. [CrossRef]
- 152. Howard, G.T.; Norton, W.N.; Burks, T. Growth of *Acinetobacter gerneri* P7 on polyurethane and the purification and characterization of a polyurethanase enzyme. *Biodegradation* **2012**, *23*, 561–573. [CrossRef]
- 153. Nawaz, A.; Hasan, F.; Shah, A.A. Degradation of poly (ε-caprolactone) (PCL) by a newly isolated *Brevundimonas* sp. strain MRL-AN1 from soil. FEMS Microbiol. Lett. 2015, 362, 1–7. [CrossRef]
- 154. Matsumiya, Y.; Murata, N.; Tanabe, E.; Kubota, K.; Kubo, M. Isolation and characterization of an ether-type polyurethanedegrading micro-organism and analysis of degradation mechanism by *Alternaria* sp. *J. Appl. Microb.* **2010**, *108*, 1946–1953. [CrossRef]
- 155. Danko, A.S.; Luo, M.; Bagwell, C.E.; Brigman, R.I.; Freedman, D.L. Involvement of linear plasmids in aerobic biodegradation of vinyl chloride. *Appl. Environ. Microbiol.* **2014**, *70*, 6092–6097. [CrossRef]
- 156. Pereira, O.; Hochart, C.; Auguet, J.C.; Debroas, D.; Galand, P.E. Genomic ecology of Marine Group II, the most common marine planktonic Archaea across the surface ocean. *MicrobiologyOpen* **2019**, *8*, e852. [CrossRef] [PubMed]
- 157. Webb, J.S.; Nixon, M.; Eastwood, I.M.; Greenhalgh, M.; Robson, G.D.; Handley, P.S. Fungal colonization and biodeterioration of plasticized polyvinyl chloride. *Appl. Environ. Microbiol.* **2000**, *66*, 3194–3200. [CrossRef] [PubMed]
- 158. Mor, R.; Sivan, A. Biofilm formation and partial biodegradation of polystyrene by the actinomycete *Rhodococcus ruber*. *Biodegradation* **2008**, *19*, 851–858. [CrossRef] [PubMed]
- 159. Nakamiya, K.; Sakasita, G.; Ooi, T.; Kinoshita, S. Enzymatic degradation of polystyrene by hydroquinone peroxidase of *Azotobacter* beijerinckii HM121. J. Ferm. Bioeng. **1997**, 84, 480–482. [CrossRef]
- 160. Yoshida, S.; Hiraga, K.; Takehana, T.; Taniguchi, I.; Yamaji, H.; Maeda, Y.; Toyohara, K.; Miyamoto, K.; Kimura, Y.; Oda, K. A bacterium that degrades and assimilates poly(ethylene terephthalate). *Science* **2016**, *351*, 1196–1199. [CrossRef]
- 161. Tournier, V.; Topham, C.M.; Gilles, A.; David, B.; Folgoas, C.; Moya-Leclair, E.; Kamionka, E.; Desrousseaux, M.-L.; Texier, H.; Gavalda, S.; et al. An engineered PET depolymerase to break down and recycle plastic bottles. *Nature* 2020, 580, 216–219. [CrossRef]
- 162. Brandon, J.A.; Jones, W.; Ohman, M.D. Multidecadal increase in plastic particles in coastal ocean sediments. *Sci. Adv.* **2019**, *5*, eaax0587. [CrossRef]
- 163. Choy, C.A.; Robison, B.H.; Gagne, T.O.; Erwin, B.; Firl, E.; Halden, R.U.; Hamiltin, J.P.; Katija, K.; Lisin, S.E.; Rolsky, C.; et al. The vertical distribution and biological transport of marine microplastics across the epipelagic and mesopelagic water column. *Sci. Rep.* 2019, *9*, 7843. [CrossRef]
- Fazey, F.M.; Ryan, P.G. Biofouling on buoyant marine plastics: An experimental study into the effect of size on surface longevity. *Environ. Pollut.* 2016, 210, 354–360. [CrossRef]
- 165. Kane, I.A.; Clare, M.A.; Miramontes, E.; Wogelius, R.; Rothwell, J.J.; Garreau, P.; Pohl, F. Seafloor microplastic hotspots controlled by deep-sea circulation. *Science* **2020**, *368*, 1140–1145. [CrossRef]
- 166. Jamieson, A.J.; Brooks, L.S.R.; Reid, W.D.K.; Piertney, S.B.; Narayanaswamy, B.E.; Linley, T.D. Microplastics and synthetic particles ingested by deep-sea amphipods in six of the deepest marine ecosystems on Earth. *R. Soc. Open Sci.* 2019, *6*, 180667. [CrossRef] [PubMed]
- Zhang, D.; Liu, X.; Huang, W.; Li, J.; Wang, C.; Zhang, D.; Zhang, C. Microplastic pollution in deep-sea sediments and organisms of the Western Pacific Ocean. *Environ. Poll.* 2020, 259, 113948. [CrossRef] [PubMed]

- 168. Urbanek, A.K.; Rymowicz, W.; Mirończuk, A.M. Degradation of plastics and plastic-degrading bacteria in cold marine habitats. *Appl. Microbiol. Biotechnol.* **2018**, *102*, 7669–7678. [CrossRef] [PubMed]
- Tagg, A.S.; Oberbeckmann, S.; Fischer, D.; Kreikemeyer, B. Paint particles are distinct and variable substrate for marine bacteria. *Mar. Pollut. Bull.* 2019, 146, 117–124. [CrossRef]
- Gangadoo, S.; Owen, S.; Rajapaksha, P.; Plaisted, K.; Cheeseman, S.; Haddara, H.; Truong, V.K.; Ngo, S.T.; Vu, V.; Cozzolino, D.; et al. Nano-plastics and their analytical characterisation and fate in the marine environment: From source to sea. *Sci. Total Environ.* 2020, 732, 138792. [CrossRef]
- 171. Dedman, C.J.; King, A.M.; Christie-Oleza, J.A.; Davies, G.L. Environmentally relevant concentrations of titanium dioxide nanoparticles pose negligible risk to marine microbes. *Environ. Sci. Nano* **2021**, *8*, 1236–1255. [CrossRef]
- 172. Abioye, O.P.; Loto, C.A.; Fayomi, O.S.I. Evaluation of Anti-biofouling Progresses in Marine Application. J. Bio-Tribo-Corros. 2019, 5, 22. [CrossRef]
- 173. Al-Naamani, L.; Dobretsov, S.; Dutta, J.; Burgess, J.G. Chitosan-zinc oxide nanocomposite coatings for the prevention of marine biofouling. *Chemosphere* **2017**, *168*, 408–417. [CrossRef]
- 174. Wang, D.; Xu, J.; Yang, J.; Zhou, S. Preparation and synergistic antifouling effect of self-renewable coatings containing quaternary ammonium-functionalized SiO₂ nanoparticles. J. Colloid. Interface Sci. 2020, 563, 261–271. [CrossRef]
- 175. Reyes-Estebanez, M.; Ortega-Morales, B.O.; Chan-Bacab, M.; Granados-Echegoyer, C.; Camacho-Chab, J.C.; Pereanez-Sacarias, J.E.; Gaylarde, C. Antimicrobial engineered nanoparticles in the built cultural heritage context and their ecotoxicological impact on animals and plants: A brief review. *Herit. Sci.* 2018, 6, 52. [CrossRef]
- 176. Falfushynska, H.; Sokolova, I.; Stoika, R. Uptake, biodistribution, and mechanisms of toxicity of metal-containing nanoparticles in aquatic invertebrates and vertebrates. In *Biomedical Nanomaterials*; Springer: Cham, Switzerland, 2022; pp. 227–263. [CrossRef]
- 177. Zha, S.J.; Rong, J.H.; Guan, X.F.; Tang, Y.; Han, Y.; Liu, G. Immunotoxicity of four nanoparticles to a marine bivalve species, *Tegillarca granosa. J. Hazard. Mater.* **2019**, 377, 237–248. [CrossRef] [PubMed]
- 178. Solano, R.; Patiño-Ruiz, D.; Tejeda-Benitez, L.; Herrera, A. Metal- and metal/oxide-based engineered nanoparticles and nanostructures: A review on the applications, nanotoxicological effects, and risk control strategies. *Environ. Sci. Pollut. Res.* 2021, 28, 16962–16981. [CrossRef] [PubMed]
- 179. Han, L.; Zhai, Y.; Liu, Y.; Hao, L.; Guo, H. Comparison of the in vitro and in vivo toxic effects of three sizes of zinc oxide (ZnO) particles using flounder gill (FG) cells and zebrafish embryos. *J. Ocean Univ. China* 2017, *16*, 93–106. [CrossRef]
- 180. Ferry, J.; Craig, P.; Hexel, C.; Sisco, P.; Frey, R.; Pennington, P.L.; Fulton, M.H.; Scott, I.G.; Decho, A.W.; Kashiwada, S.; et al. Transfer of gold nanoparticles from the water column to the estuarine food web. *Nat. Nanotechnol.* **2009**, *4*, 441–444. [CrossRef]
- 181. Garner, K.L.; Suh, S.; Keller, A.A. Assessing the risk of engineered nanomaterials in the environment: Development and application of the nanoFate model. *Environ. Sci. Technol.* **2017**, *51*, 5541–5551. [CrossRef] [PubMed]
- 182. Ciacci, C.; Grimmelpont, M.V.; Corsi, I.; Bergami, E.; Curzi, D.; Burini, D.; Bouchet, V.M.P.; Ambrogini, P.; Gobbi, P.; Ujiie, Y.; et al. Nanoparticle-biological interactions in a marine benthic foraminifer. *Sci. Rep.* **2019**, *9*, 19441. [CrossRef]
- 183. Peijnenburg, W.J.G.M.; Baalousha, M.; Chen, J.; Chaudry, Q.; von der Kammer, F.; Kuhlbusch, T.A.J.; Nickel, C.; Quick, J.T.K.; Renkerg, M.; Koelmans, A.A. A Review of the properties and processes determining the fate of engineered nanomaterials in the aquatic environment. *Crit. Rev. Environ. Sci. Technol.* 2015, 45, 2084–2134. [CrossRef]
- Chiu, M.; Khan, Z.A.; Garcia, S.G. Effect of engineered nanoparticles on exopolymeric substances release from marine phytoplankton. *Nanoscale Res. Lett.* 2017, 12, 620. [CrossRef]
- Sendra, M.; Moreno, I.; Blasco, J. Toxicity of metal and metal oxide engineered nanoparticles to phytoplankton. In *Ecotoxicity of Nanoparticles in Aquatic Systems*; Blasco, J., Corsi, I., Eds.; CRC Press: Boca Raton, FL, USA, 2019; ISBN 1351657550/9781351657556.
- 186. Tsiola, A.; Toncelli, C.; Fodelianakis, S.; Michaud, G.; Bucheli, T.; Gavriilidou, A.; Kagiorgi, M.; Kalantzi, I.; Knauer, K.; Kotulas, G.; et al. Low-dose addition of silver nanoparticles stresses marine plankton communities. *Environ. Sci. Nano* 2018, *5*, 1965–1980. [CrossRef]
- 187. Zhao, J.; Lin, M.; Wang, Z.; Cao, X.; Xing, B. Engineered nanomaterials in the environment: Are they safe? *Crit. Rev. Environ. Sci. Technol.* **2021**, *51*, 1443–1478. [CrossRef]
- 188. Gillard, B.; Chatzievangelou, D.; Thomsen, L.; Ullrich, M.S. Heavy-metal-resistant microorganisms in deep-sea sediments disturbed by mining activity: An application toward the development of experimental in vitro systems. *Front. Mar. Sci.* 2019, *6*, 432. [CrossRef]
- 189. Cho, H.; Kim, K.; Son, S.K.; Le, A.D.; Kagiri, A.; Ramos, J.; Tsai, S.M.; Drobenaire, H.W.; Santschi, P.H.; Quigg, A. Fine-scale microbial communities associated with manganese nodules in deep-sea sediment of the Korea Deep Ocean Study Area in the Northeast Equatorial Pacific. Ocean Sci. J. 2018, 53, 337–353. [CrossRef]
- 190. Lemaitre, N.; de Souza, G.F.; Archer, C.; Wang, R.-M.; Planquette, H.; Sarthou, G.; Vance, D. Pervasive sources of isotopically light zinc in the North Atlantic Ocean. *Earth Planet Sci. Lett.* **2020**, *539*, 116216. [CrossRef]
- Conway, T.M.; Hamilton, D.S.; Shelley, R.U.; Aguilar-Islas, A.M.; Landing, W.M.; Mahowald, M.N.; John, S.G. Tracing and constraining anthropogenic aerosol iron fluxes to the North Atlantic Ocean using iron isotopes. *Nat. Commun.* 2019, 10, 2628. [CrossRef] [PubMed]
- 192. Hamilton, D.S.; Moore, J.K.; Arneth, A.; Bond, T.C.; Carslaw, K.S.; Hantson, S.; Ito, A.; Kaplan, J.O.; Lindsay, K.; Nieradzik, L.P.; et al. Impact of changes to the atmospheric soluble iron deposition flux on ocean biogeochemical cycles in the Anthropocene. *Glob. Biogeochem. Cycles* 2020, 34, e2019GB006448. [CrossRef]

- 193. Chuang, C.-Y.; Santschi, P.H.; Ho, Y.-F.; Conte, M.; Guo, L.; Schumann, D.; Ayranov, M.; Li, Y.-h. Biopolymers as major carrier phases and redox regulators of Th, Pa, Pb, Po, and Be in settling particles from the Atlantic Ocean. *Mar. Chem.* 2013, 15, 131–143. [CrossRef]
- 194. Boyd, P.; Ellwood, M.; Tagliabue, A.; Twining, B.S. Biotic and abiotic retention, recycling and remineralization of metals in the ocean. *Nat. Geosci.* 2017, *10*, 167–173. [CrossRef]
- 195. Hoffman, C.L.; Nicholas, S.L.; Ohnemus, D.C.; Fitzsimmons, J.; Sherrell, R.; German, C.; Heller, M.; Lee, J.-M.; Lam, P.; Toner, B.M.; et al. Near-field iron and carbon chemistry of non-buoyant hydrothermal plume particles, Southern East Pacific Rise 15°S. *Mar. Chem.* 2018, 201, 183–197. [CrossRef]
- 196. Sundseth, K.; Pacyna, J.M.; Pacyna, E.G.; Pirrone, N.; Thorne, R.J. Global sources and pathways of mercury in the context of human health. *Int. J. Environ. Res. Public Health* **2017**, *14*, 105. [CrossRef]
- 197. Arctic Monitoring and Assessment Programme (AMAP). United Nations Environment Programme (UNEP) Global Mercury Assessment: Sources, Emissions, Releases and Environmental Transport; UNEP Chemicals Branch: Geneva, Switzerland, 2013.
- 198. Gworek, B.; Bemowska-Kałabun, O.; Kijeńska, M.; Wrzosek-Jakubowska, J. Mercury in marine and oceanic waters—A review. *Water Air Soil Pollut.* **2016**, 227, 371. [CrossRef]
- Zhang, L.; Wu, S.; Zhao, L.; Liu, X.; Pierce, E.M.; Gu, B. Mercury sorption and desorption on organo-mineral particulates as a source for microbial methylation. *Environ. Sci. Technol.* 2019, 53, 2426–2433. [CrossRef] [PubMed]
- Martinez-Ruiz, F.; Paytan, A.; Gonzalez-Muñoz, M.T.; Jroundi, F.; Abad, M.M.; Lam, P.J.; Kastner, M. Barite formation in the ocean: Origin of amorphous and crystalline precipitates. *Chem. Geol.* 2019, *511*, 441–451. [CrossRef]
- 201. Zamanillo, M.; Ortega-Retuerta, E.; Nunes, S.; Rodriguez-Ros, P.; Dall'Osto, M.; Estrada, M.; Sala, M.M.; Simo, R. Main drivers of transparent exopolymer particle distribution across the surface Atlantic Ocean. *Biogeosciences* **2019**, *16*, 733–749. [CrossRef]
- 202. Lundgreen, R.B.C.; Jaspers, C.; Traving, S.J.; Ayala, D.J.; Lombard, F.; Grossart, H.-P.; Nielsen, T.G.; Munk, P.; Riemann, L. Eukaryotic and cyanobacterial communities associated with marine snow particles in the oligotrophic Sargasso Sea. *Sci. Rep.* 2019, *9*, 8891. [CrossRef]
- Arnosti, C.; Ziervogel, K.; Yang, T.; Teske, A. Oil-derived marine 817 aggregates–hot spots of polysaccharide degradation by specialized bacterial 818 communities. *Deep-Sea Res. PT II* 2016, 129, 179–186. [CrossRef]
- Duran Suja, L.; Summers, S.; Gutierrez, T. Role of EPS, Dispersant and Nutrients on the Microbial Response and MOS Formation in the Subarctic Northeast Atlantic. *Front. Microbiol.* 2017, *8*, 676. [CrossRef] [PubMed]
- 205. Capo, E.; Bravo, A.G.; Soerensen, A.L.; Bertilsson, S.; Pinhassi, J.; Feng, C.; Andersson, A.F.; Buck, M.; Bjorn, E. Marine snow as a habitat for microbial mercury methylators in the Baltic Sea. *bioRxiv* 2020, *3*, 975987. [CrossRef]
- 206. Achberger, A.M.; Doyle, S.M.; Mills, M.I.; Holmes II, C.P.; Quigg, A.; Sylvan, J.B. Bacteria-Oil Microaggregates Are an Important Mechanism for Hydrocarbon Degradation in the Marine Water Column. *mSystems* **2021**, *6*, e01105-21. [CrossRef]
- Duret, M.T.; Lampitt, R.S.; Lam, P. Prokaryotic niche partitioning between suspended and sinking marine particles. *Environ. Microbiol. Rep.* 2019, 11, 386–400. [CrossRef]
- Datta, M.; Sliwerska, E.; Gore, J.; Polz, M.F.; Cordero, O.X. Microbial interactions lead to rapid micro-scale successions on model marine particles. *Nat. Commun.* 2016, 7, 11965. [CrossRef]
- Liu, Y.; Fang, J.; Jia, Z.; Chen, S.; Zhang, L.; Gao, W. DNA stable-isotope probing reveals potential key players for microbial decomposition and degradation of diatom-derived marine particulate matter. *MicrobiologyOpen* 2020, 9, e1013. [CrossRef] [PubMed]
- Alcolombri, U.; Peaudecerf, F.J.; Fernandez, V.I.; Behrendt, L.; Lee, K.S.; Stocker, R. Sinking enhances the degradation of organic particles by marine bacteria. *Nat. Geosci.* 2021, 14, 775–780. [CrossRef]
- Bochdansky, A.B.; Clouse, M.A.; Herndl, G.J. Eukaryotic microbes, principally fungi and labyrinthulomycetes, dominate biomass on bathypelagic marine snow. *ISME J.* 2017, 11, 362–373. [CrossRef] [PubMed]
- Schultz, D.; Zühlke, D.; Bernhardt, J.; Francis, T.B.; Albrecht, D.; Hirschfeld, C.; Markert, S.; Riedel, K. An optimized metaproteomics protocol for a holistic taxonomic and functional characterization of microbial communities from marine particles. *Environ. Microbiol. Rep.* 2020, *12*, 367–376. [CrossRef]
- López-Pérez, M.; Kimes, N.E.; Haro-Moreno, J.M.; Rodriguez-Valera, F. Not all particles are equal: The selective enrichment of particle-associated bacteria from the Mediterranean sea. *Front. Microbiol.* 2016, 7, 996. [CrossRef]
- 214. Gregson, B.H.; McKew, B.A.; Holland, R.D.; Nedwed, T.; Prince, R.; McGenity, T. Marine oil snow, a microbial perspective. *Front. Mar. Sci.* 2021, 28, 11. [CrossRef]