

Article

Abscisic Acid in Coordination with Nitrogen Alleviates Salinity-Inhibited Photosynthetic Potential in Mustard by Improving Proline Accumulation and Antioxidant Activity

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Abstract: This investigation was done to assess the role of abscisic acid (ABA; 25 μ M) and/or nitrogen (N; 10 mM) in the alleviation of salinity (NaCl; 100 mM)-induced reduction in photosynthetic activity and growth, N and sulfur (S) assimilation of mustard (*Brassica juncea* L.) cv. RH0-749. Salinity treatment caused oxidative stress and significantly elevated the content of both H₂O₂ and thiobarbituric acid reactive substances (TBARS), and impaired photosynthetic activity and growth, but increased the content of nitrogenous osmolyte proline and the activity of antioxidant enzymes involved in the metabolism of reactive oxygen species. The application of 25 μ M ABA under a controlled condition negatively affected photosynthesis and growth. However, ABA, when combined with N, minimized oxidative stress and mitigated the salinity-inhibited effects by increasing the activity of antioxidant enzymes (superoxide dismutase, SOD; glutathione reductase, GR; ascorbate peroxidase, APX) and proline content. Overall, the supplementation of 10 mM N combined with 25 μ M ABA provides an important strategy for enhancing the photosynthetic potential of *B. juncea* under saline conditions.

Keywords: abscisic acid; *Brassica juncea*; nitrogen; salinity; oxidative stress; photosynthesis

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1. Introduction

Oilseed crops play a prominent role in agricultural industries and trade throughout the world [1]. The edible oilseed crop, Indian mustard (*Brassica juncea* L.), stands second to peanuts; accounts for 27.8% of the country's oilseed economy; and is cultivated throughout the northern Indian plains [2]. Unfortunately, most of its cultivation depends on groundwater sources varying from medium to high salinity levels [3]. Salinity is a serious soil constraint that negatively affects agriculture production, particularly in arid and semi-arid regions of the world [4]. Salinity affects about 2% (6.74 Mha) of Indian land [5,6].

In plants, salt stress mainly causes ionic stress by increasing Na⁺ and Cl⁻ ions concentrations and thereby prevents essential elements attainment, homeostasis and metabolism [7,8]. In addition to causing cellular water imbalance, osmotic stress and abscission, salinity stress may also significantly impact all phases of photosynthesis and other gas exchange traits [9]. Notably, the accumulation of Na⁺ and Cl⁻ ions (ionic toxicity) causes oxidative stress due to the physiological imbalance between generation and antioxidants-mediated scavenging of reactive oxygen species (ROS), such as H₂O₂, O₂⁻ and ·OH [10]. Elevated or non-metabolized ROS may cause the oxidation of proteins and membrane lipids and also impair cellular redox homeostasis [11].

Plants tolerance to salinity stress mainly involve the maintenance of ionic

homeostasis; metabolic adjustments that lead to the accumulation of compounds containing nitrogen (N; such as an organic solute proline) or sulfur (S; such as cysteine, Cys; glutathione, GSH); induction of phytohormones (such as abscisic acid, ABA) and/or the scavenging of ROS through antioxidant enzymes (such as superoxide dismutase, SOD; glutathione reductase, GR; ascorbate peroxidase, APX) and antioxidant metabolites/non-enzymes (such as ascorbate, AsA; glutathione, GSH) [12–14]. Osmotic adjustment can be maintained by the endogenous production of ABA and/or nitrogenous compatible solutes, such as proline. Interestingly, both ABA and proline are closely linked, where exogenously applied ABA-mediated salinity tolerance has been linked to the increased production of proline [15,16]). On the one hand, ABA can effectively reduce Na^+ and Cl^- content and Na^+/K^+ ratio, increase K^+ and Ca^{2+} content and the accumulation of proline [17]. On the other hand, in addition to osmotic adjustment, proline scavenges varied ROS and thereby contributes to the strengthening of the antioxidant defense system and the eventual alleviation of stress impacts in plants [16]. Proline metabolism in salinity-exposed plants is also closely linked with N nutrition, where N-availability regulates proline production and plant salinity tolerance [14,18]. Notably, S-assimilatory products in plant salinity tolerance are also of great significance, where one of the S-assimilation products and S-containing amino acid Cys acts as a precursor or donor of reduced S for a range of S-compounds including GSH [19]. Interestingly, coordination occurs between the assimilatory pathways of N and S, where nitrate reductase (NR) is involved in NO_3^- reduction and ATP-sulfurylase (ATP-S) activates the metabolically inert SO_4^{2-} ; and makes it usable in Cys and GSH [14,18,19]. In turn, besides acting as ROS-scavengers, GSH (and also proline) can modulate the photosynthesis functions by influencing the activity of ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco) [20–22]. It has also been shown that exogenous supplementation of ABA or N enhances salinity tolerance in plants [23–25]. Additionally, N-supply was reported to improve salinity tolerance by maintaining GSH production [26].

Despite the above facts, the study on the interaction of ABA and N in the regulation of photosynthetic activity via the control on the antioxidant defense system under salt stress has not been done. Additionally, the projected increase in salinity-affected areas to 16.2 Mha by 2050 due to climate change can further aggravate the situation of the cultivation of oilseed crops [27]. Hence, it is hypothesized that a coordination between N-containing osmolyte (proline) and phytohormone (ABA) can help to maintain the cellular ionic (Na^+ and Cl^-) homeostasis; reduced S-containing compounds (Cys and GSH); and a fine-tuning among antioxidant enzymes, and thereby can combat elevated ROS-accrued oxidative stress and improve cellular redox homeostasis, and eventually improve growth and stomatal behavior; photosynthesis in salinity-exposed plants supplied with optimum concentrations of ABA and N. To test the highlighted hypothesis, the present work investigated the interactive effects of ABA and N in the regulation of photosynthetic activity, growth and N and S-assimilation, antioxidant activity and proline in *B. juncea* plants under NaCl stressed conditions.

2. Results

In the first experiment, where the screening of ABA concentrations was performed, exogenously supplied ABA ($>5 \mu\text{M}$) significantly decreased photosynthetic and growth attributes in the absence of NaCl but did not influence the status of oxidative stress parameters (H_2O_2 and TBARS contents) in comparison to the control. However, in the presence of NaCl (100 mM), photosynthetic and growth parameters were favorably influenced by exogenously supplied ABA when compared to the NaCl-stressed plants. Under 100 mM NaCl stress, a lesser effect of $50 \mu\text{M}$ ABA on the studied parameters was observed when compared to $25 \mu\text{M}$ ABA. However, among the ABA concentrations screened, $25 \mu\text{M}$ ABA most efficiently increased chlorophyll content (+56.6%), net photosynthesis (+108.1%), stomatal conductance (+55.6%), intercellular CO_2 concentration (+59%), leaf area (+50.2%), plant dry mass (+60.3%) and reduced the

contents of H₂O₂ (−59.3%) and TBARS (−54.0%) in comparison to the NaCl-stressed plants (Table 1).

Table 1. Selection of ABA concentration, Chlorophyll content (SPAD value), net photosynthesis ($\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$), stomatal conductance ($\text{mmol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$), intercellular CO₂ concentration ($\mu\text{mol CO}_2 \text{ mol}^{-1}$), leaf area ($\text{cm}^2 \text{ plant}^{-1}$), plant dry mass (g plant^{-1}), Na⁺ and Cl[−] content (mg g^{-1} leaf dry mass) and H₂O₂ and TBARS content (nmol g^{-1} fresh weight) of Indian mustard (*Brassica juncea* L.) at 30 days after sowing. Plants were grown individually with 0, 5, 10, 25 and 50 μM ABA in the presence or absence of 100 mM NaCl. Data are presented as treatments mean \pm SE ($n = 4$). The same letter in columns indicates that data did not differ significantly by Duncan’s multiple range test at $p < 0.05$. ABA, Abscisic acid.

Treatments/Parameters	Control	NaCl (100 mM)	ABA Concentration (μM)				5 μM ABA + 10 μM ABA 25 μM ABA 50 μM ABA			
			5	10	25	50	100 mM NaCl	+100 mM NaCl	+100 mM NaCl	+100 mM NaCl
Chlorophyll content	36.2 \pm 1.15a	18.2 \pm 0.58f	34.2 \pm 1.08a	31.1 \pm 0.98b	24.5 \pm 0.78d	19.3 \pm 0.61f	20.0 \pm 0.63ef	22.0 \pm 0.71e	28.5 \pm 0.91c	25.0 \pm 0.79d
Net photosynthesis	13.7 \pm 0.44a	6.2 \pm 0.19f	13.4 \pm 0.43a	12.3 \pm 0.39b	10.13 \pm 0.33c	6.9 \pm 0.22f	7.9 \pm 0.25e	9.14 \pm 0.29d	12.9 \pm 0.41ab	9.1 \pm 0.29d
Stomatal conductance	238.5 \pm 6.23a	139.4 \pm 6.04e	230.2 \pm 6.01ab	219.6 \pm 5.74b	195 \pm 5.09b	142.6 \pm 6.18c	170 \pm 4.44e	182 \pm 4.75cd	217 \pm 5.67b	193 \pm 5.04c
Intercellular CO ₂ concentration	294.9 \pm 9.40a	161.6 \pm 7.01f	288.5 \pm 9.19a	278.2 \pm 8.87b	242.8 \pm 6.34c	166 \pm 7.20f	180.2 \pm 7.82e	192.2 \pm 8.34d	257 \pm 6.72ab	201 \pm 9.88d
Leaf area	116.64 \pm 3.05a	65 \pm 2.28gh	114.4 \pm 2.99a	106.02 \pm 2.77b	77.8 \pm 2.48ef	61.4 \pm 2.86h	82.6 \pm 2.63de	88.8 \pm 2.83d	97.64 \pm 3.11c	72.14 \pm 2.3fg
Plant dry mass	2.2 \pm 0.07a	1.31 \pm 0.05e	1.78 \pm 0.06cd	1.8 \pm 0.05cd	1.94 \pm 0.05bc	1.39 \pm 0.06e	1.63 \pm 0.07d	1.71 \pm 0.07d	2.101 \pm 0.05ab	1.33 \pm 0.06e
H ₂ O ₂ content	89.92 \pm 4.42c	157.8 \pm 4.03a	87.2 \pm 4.28c	86.4 \pm 4.24c	85.2 \pm 4.18c	84.7 \pm 2.57c	119.6 \pm 3.62b	110.2 \pm 4.78b	64.23 \pm 3.15d	87.32 \pm 4.29c
TBARS content	17.5 \pm 0.86c	28.2 \pm 0.72a	16.2 \pm 0.79c	15.9 \pm 0.78c	16.9 \pm 0.83c	15.7 \pm 0.47c	21.43 \pm 0.65b	20.35 \pm 0.88b	13.0 \pm 0.63d	17.2 \pm 0.84c

2.1. Impact of ABA and/or N on Accumulation of Na⁺ and Cl[−] Ions and Oxidative Stress

The content of Na⁺ and Cl[−] in both the leaves and roots of plants was analyzed to determine the potential of 25 μM ABA along with 10 mM N in modulating the ion accumulation NaCl (100 mM)-exposed *B. juncea* plants. The content of Na⁺ and Cl[−] significantly increased in NaCl-treated *B. juncea* roots and leaves; however, higher contents of Na⁺ and Cl[−] were observed in roots than in leaves (Figure 1A–D). Exogenously applied ABA or N individually lowered Na⁺ and Cl[−] ion accumulation in comparison with both the control and salt-stressed plants. Additionally, the combined treatment of ABA and N further reduced the accumulation of these ions in both roots and leaves when compared to the control plants. In particular, under salt stress conditions, N reduced leaf and root Na⁺ accumulation by 33.1% and 35.2% and Cl[−] accumulation by 38.0% and 28.0%, respectively; whereas ABA reduced leaf and root Na⁺ accumulation by 26.6% and 21.1% and Cl[−] accumulation by 32.1% and 20.9%, respectively, in comparison to the NaCl-stressed plants. Moreover, the application of ABA together with N reduced root Na⁺ by 43.7%, root Cl[−] by 36.4%, leaf Na⁺ by 43.1% and leaf Cl[−] by 46.2% in the presence of salt stress compared to the NaCl alone treated plants (Figure 1A–D).

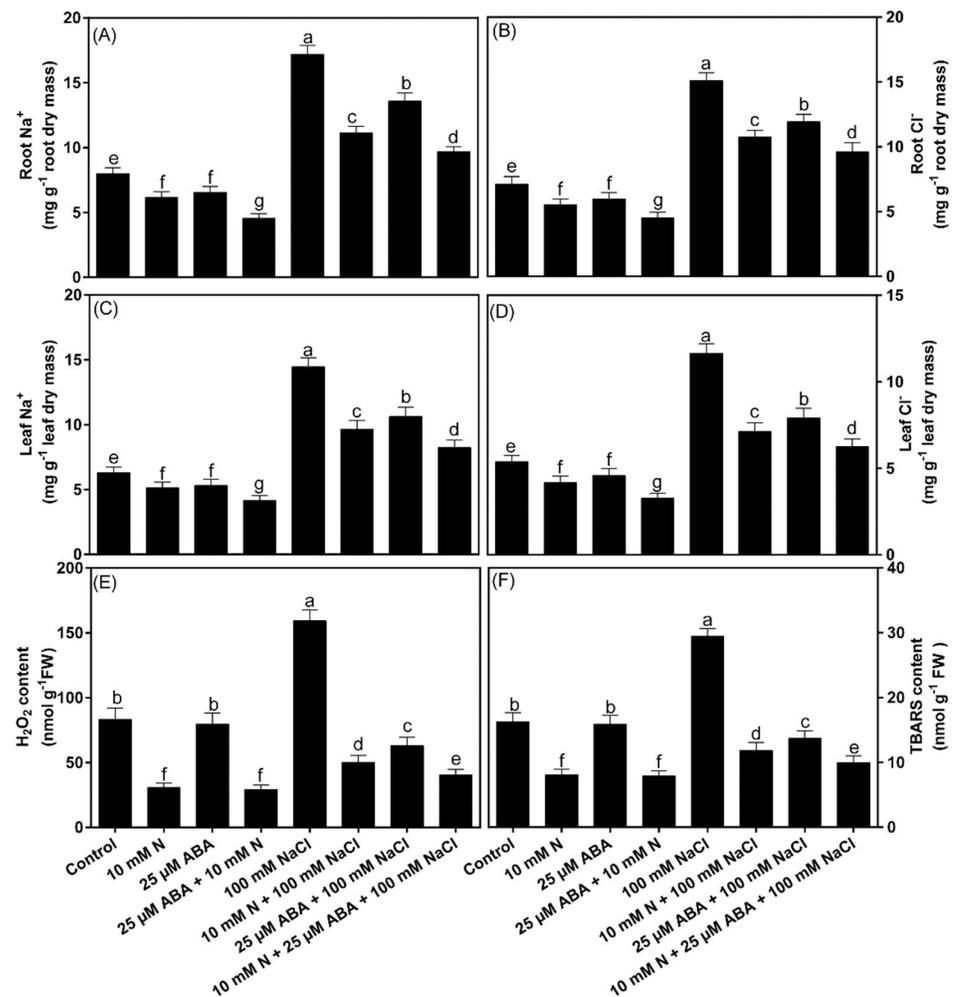


Figure 1. Contents of leaf and root Na⁺ and Cl⁻ (A–D), H₂O₂ (E), thiobarbituric acid reactive substances (TBARS; (F)) in Indian mustard (*Brassica juncea* L.) at 30 days after sowing. Plants were grown individually with 0 or 10 mM N and treated with 25 μM ABA in the presence or absence of 100 mM NaCl. Data are presented as treatments mean ± SE (*n* = 4). The same letter above bars indicates that data did not differ significantly by Duncan’s multiple range test at *p* < 0.05. ABA, Abscisic acid; N, Nitrogen.

Contents of H₂O₂ and TBARS were quantified in order to evaluate the role of ABA and/or N in the mitigation of oxidative stress. NaCl (100 mM) stress alone increased the content of H₂O₂ (+99.5%) and TBARS (+81.1%) in comparison to the control. Plants treated with ABA or N without NaCl showed N-mediated reduction in H₂O₂ and TBARS content compared to control; whereas ABA treated plants had values equivalent to control plants. In the presence of NaCl, both ABA and N independently reduced H₂O₂ by 69.1% and 59.1% and TBARS by 62.0% and 53.1%, respectively. However, ABA combined with N reduced oxidative stress more conspicuously and reduced the content of H₂O₂ by 75.6% and TBARS by 66.0% in comparison to NaCl-stressed plants (Figure 1E,F).

2.2. Visualization of the Status of O₂⁻ and H₂O₂ in Leaves Using Histochemical Staining and Confocal Laser Scanning Microscopy

Histochemical staining method was employed to visualize the status of oxidative stress parameters, such as O₂⁻ generation (as shown by blue staining of leaves) and H₂O₂ (as shown by brown staining of leaves) using NBT and DAB staining methods,

respectively. The staining spots were more pronounced in NaCl treated leaf discs compared to the control. However, restricted staining spots were observed under salt stress in the leaves of plants that received ABA or N alone in comparison with NaCl treated leaves. Moreover, ABA, together with N, more prominently reduced the staining spots in the presence of NaCl stress (Figure 2A–J).

Further, the analysis with 2',7'-dichlorofluorescein (H₂DCFDA) fluorescence revealed the status of the accumulation of H₂O₂ in roots. The roots grown with NaCl alone showed a higher intensity of green fluorescence. In contrast, the roots of plants supplemented with ABA or N individually exhibited a lesser intensity of green fluorescence, though the result was more conspicuous with N than ABA supply. The combined supplementation of ABA and N under NaCl stress most effectively reduced the H₂O₂ content, which was revealed through a lesser green fluorescence, almost similar to the green fluorescence observed in the roots of the control plants (Figure 3A–E).

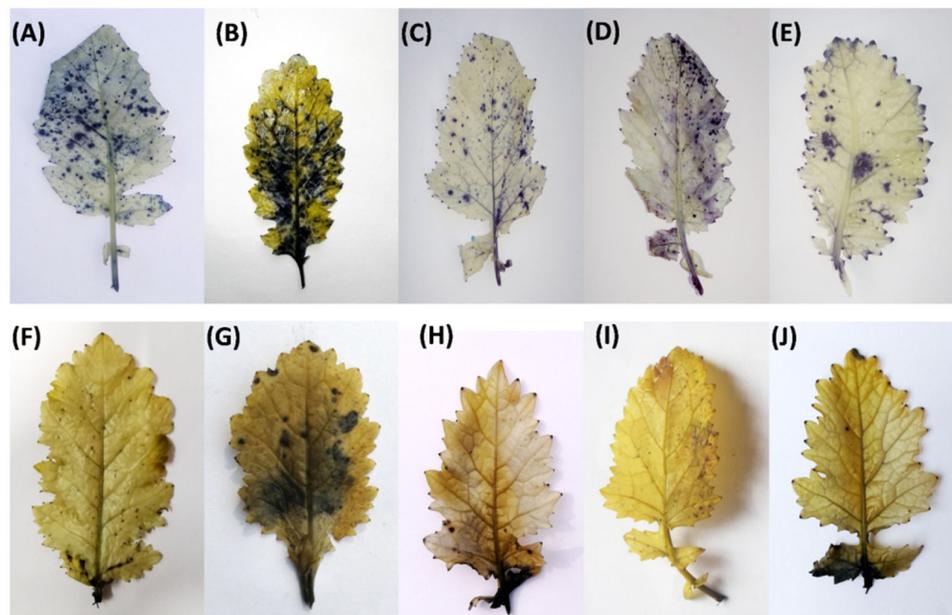


Figure 2. Representative images showing the level of in situ generation of superoxide ions (O_2^-) by nitro blue tetrazolium (NBT) staining of the leaves (A–E) and generation of H_2O_2 by 3,3'-diaminobenzidine (DAB) staining of the leaves (F–J) in Indian mustard (*Brassica juncea* L.) at 30 days after sowing. Plants were grown with 0 (A,F), 100 mM NaCl (B,G), 10 mM N + 100 mM NaCl (C,H), 25 μ M ABA + 100 mM NaCl (D,I) and 25 μ M ABA + 10 mM N + 100 mM NaCl (E,J). NaCl, Sodium chloride; ABA, Abscisic acid; N, Nitrogen.

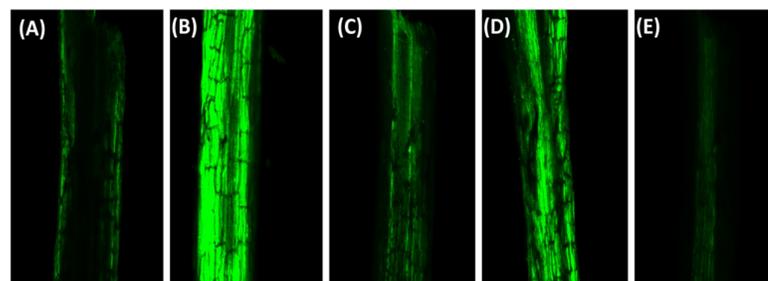


Figure 3. Representative confocal microscopic images of H_2O_2 formation in roots of Indian mustard (*Brassica juncea* L.) using H₂DCFDA staining (A–E) at 30 days after sowing. Plants were grown with 0 (A), 100 mM NaCl (B), 10 mM N + 100 mM NaCl (C), 25 μ M ABA + 100 mM NaCl (D) and 25 μ M ABA + 10 mM N + 100 mM NaCl (E). NaCl, Sodium chloride; ABA, Abscisic acid; N, Nitrogen.

2.3. Influence of ABA and/or N on Photosynthetic Characteristics under NaCl Stress

Salt stress severely inhibited photosynthetic characteristics (P_N , g_s , C_i and chlorophyll). Plants receiving N showed higher values of the photosynthetic characteristics in comparison to the control, both under stress and non-stress conditions. Under control conditions (without stress), ABA treatment decreased the values of photosynthetic parameters. However, in NaCl stressed plants, supplementation of ABA or N independently improved P_N by 109% and 181%, g_s by 63% and 101%, C_i by 52% and 98% and Chl content by 55% and 96%, respectively, in comparison to NaCl alone treated plants. The combined supplementation of ABA and N resulted in prominent increases in P_N by 238%, g_s by 136%, C_i by 133% and Chl content by 137% in comparison to NaCl-stressed plants (Table 2). NaCl stress severely inhibited Rubisco activity in comparison to the control plants. The plants supplemented with N showed higher Rubisco activity both under control and stressed conditions. Under non-stress conditions, plants treated with ABA showed reduced Rubisco activity. However, under stress conditions, plants supplemented with ABA or N individually resulted in enhanced Rubisco activity by 173.1% and 247.0%, respectively, in comparison to the NaCl-treated plants. Further, supplementation of ABA and N together maximally increased Rubisco activity by about three-fold in comparison to the NaCl-treated plants (Table 2).

Table 2. Net photosynthetic rate ($\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$), stomatal conductance ($\text{mmol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$), intercellular CO_2 ($\mu\text{mol CO}_2 \text{ mol}^{-1}$), chlorophyll content (SPAD value), Rubisco activity ($\mu\text{mol CO}_2 \text{ mg}^{-1} \text{ protein m}^{-1}$), proline content (mg g^{-1} fresh mass), leaf area ($\text{cm}^2 \text{ plant}^{-1}$) and plant dry mass (g plant^{-1}) in Indian mustard (*Brassica juncea* L.) at 30 days after sowing. Plants were grown individually with 0 or 10 mM N, treated with 25 μM ABA in the presence or absence of 100 mM NaCl. Data are presented as treatments mean \pm SE ($n = 4$). The same letter in rows indicates that data did not differ significantly by Duncan's multiple range test at $p < 0.05$. ABA, Abscisic acid; N, Nitrogen.

Treatments/ Parameters	Net Photosynthesis	Stomatal Conductance	Intercellular CO_2 Concentration	Chlorophyll Content	Rubisco Activity	Proline Content	Leaf Area	Plant Dry Mass
Control	13.72 \pm 0.59e	241.32 \pm 7.69e	278.23 \pm 8.87e	28.8 \pm 1.2498e	49.11 \pm 1.56e	7.32 \pm 0.32e	113.36 \pm 5.27e	2.5 \pm 0.08e
100 mM NaCl	6.3 \pm 0.27h	147.42 \pm 6.39h	169.72 \pm 7.36h	16.4 \pm 0.7117h	16.14 \pm 1.02h	9.56 \pm 0.41d	61.7 \pm 3.74h	1.12 \pm 0.07h
10 mM N	21.2 \pm 0.67a	339.62 \pm 8.87a	388.21 \pm 10.14a	42.3 \pm 1.3487a	79.20 \pm 2.07a	12.85 \pm 0.41c	182.24 \pm 5.81a	4.15 \pm 0.08a
25 μM ABA	10.54 \pm 0.33g	194.52 \pm 6.20g	223.23 \pm 7.12g	22.3 \pm 0.7110g	33.94 \pm 1.24g	13.60 \pm 0.41bc	78.92 \pm 3.67g	1.78 \pm 0.08g
25 μM ABA + 10 mM N	17.9 \pm 0.57c	292.8 \pm 7.65c	333.25 \pm 8.71c	35.5 \pm 1.1319c	62.60 \pm 1.64c	13.09 \pm 0.42c	147.8 \pm 6.03c	3.32 \pm 0.10c
10 mM N + 100 mM NaCl	16.3 \pm 0.51d	269.4 \pm 8.58d	305.73 \pm 9.74d	32.2 \pm 1.0267d	55.96 \pm 1.78d	15.74 \pm 0.50ab	130.58 \pm 6.07d	2.98 \pm 0.09d
25 μM ABA + 100 mM NaCl	12.13 \pm 0.38f	217.92 \pm 6.94f	250.77 \pm 7.99f	25.5 \pm 0.8130f	44.02 \pm 1.40f	14.32 \pm 0.46b	96.14 \pm 4.47f	2.21 \pm 0.10f
25 μM ABA + 10 mM N + 100 mM NaCl	19.6 \pm 0.62b	316.2 \pm 8.26b	360.69 \pm 9.42b	38.93 \pm 1.2412b	71.21 \pm 1.86b	17.21 \pm 0.55a	165.02 \pm 5.26b	3.61 \pm 0.11b

2.4. Impact of ABA and/or N on Growth under NaCl Stress

Salt stress significantly decreased leaf area and plant dry mass. In contrast, N supplementation increased leaf area by 52.0% and plant dry mass by 66.2% under non-stressed conditions in comparison to control plants. ABA application in the absence of NaCl resulted in reduced leaf area by 30.2% and plant dry mass by 28.8% in comparison to the control plants. Under NaCl stress conditions, leaf area and plant dry mass were improved by 55.8% and 97.1% with ABA and by 112.0% and 166.1% with N application. A higher increase of 167.2% in leaf area and 222.1% in plant dry mass was

noted with the combined application of ABA and N in comparison to salt-stressed plants (Table 2).

2.5. Impact of ABA and/or N on Proline Content under NaCl Stress

NaCl treatment induced proline accumulation in plants. Exogenous supplementation of either N or ABA individually increased proline content compared to the control plants. However, ABA supplementation with N increased proline accumulation by 80% under salt stress and 36.9% under non-stress compared to salt-stressed plants (Table 2).

2.6. Impact of ABA and/or N on Stomatal Behavior under NaCl Stress

The leaf samples of NaCl treated plants showed closed stomata, while stomata were open in the leaf samples of control plants. The treatment of ABA in the presence of NaCl resulted in stomatal closure. Plants treated with N alone or N + ABA in the presence of NaCl showed that stomata were partially opened (Figure 4).

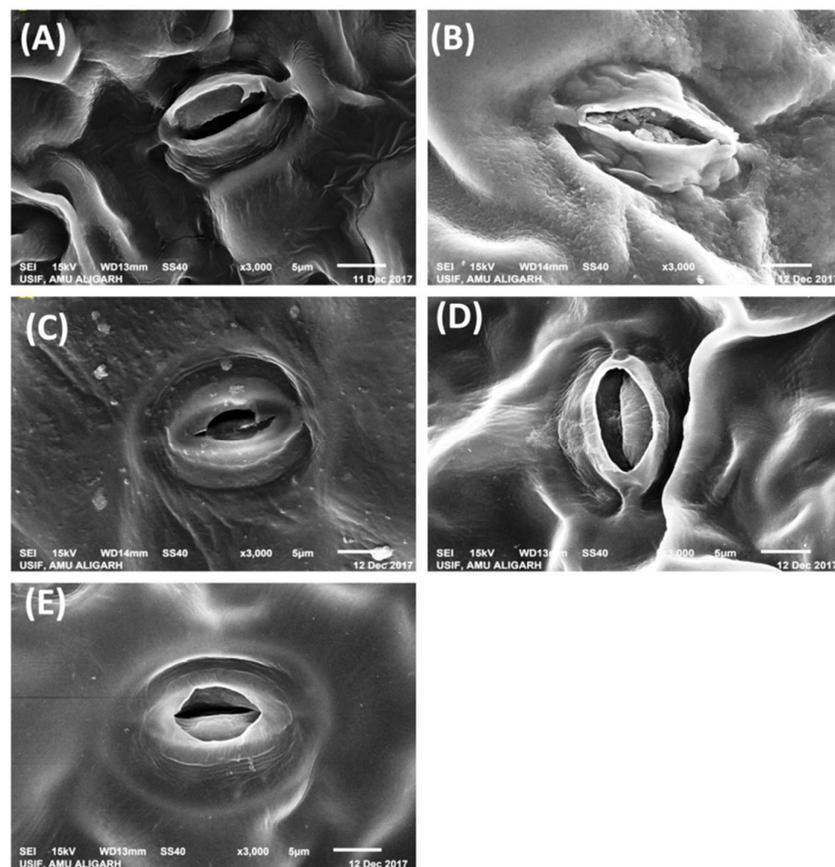


Figure 4. Representative images showing the leaf stomatal behavior in Indian mustard (*Brassica juncea* L.) under 0 (A), 100 mM NaCl (B), 10 mM N + 100 mM NaCl (C), 25 μ M ABA + 100 mM NaCl (D) and 25 μ M ABA + 10 mM N + 100 mM NaCl (E). The opening and closing of stomata were studied under the scanning electron microscope at 3000 \times magnifications at 30 days after sowing. NaCl, Sodium chloride; ABA, Abscisic acid; N, Nitrogen.

2.7. Impact of ABA and/or N on Antioxidant Metabolism under NaCl Stress

The activity of APX, GR and SOD was measured for assessing the involvement of these enzymes in the antioxidant defense system when treated with ABA and/or N under NaCl-induced oxidative stress. Under NaCl stress, the activity of APX, GR, and SOD

increased by 33.3%, 53.6% and 42.2%, respectively, in comparison to the control plants. Individual supplementation of N increased the activity of these enzymes both under stress and control conditions. On the contrary individual supplementation of ABA under control conditions did not show any significant change in the activity of these enzymes. However, under stress conditions, plants supplemented with ABA exhibited an increase in the activity of APX by 120.3%, GR by 117.0% and SOD by 74.2%; whereas, plants receiving N exhibited an increase in APX by 120.2%, GR by 117.5% and SOD by 78.2% in comparison to the control plants. A comparatively more pronounced increase in the activity of these enzymes was observed when ABA + N was supplemented together under NaCl stress conditions. The combined application of ABA + N increased activity of APX, GR and SOD by 182.0%, 175.1% and 92.2%, respectively, in comparison to the plants exposed to NaCl stress (Table 3).

Table 3. Activity of ascorbate peroxidase (APX), glutathione reductase (GR) and superoxide dismutase (SOD) in Indian mustard (*Brassica juncea* L.) at 30 days after sowing. Plants were grown individually with 0 or 10 mM N and treated with 25 μ M ABA in the presence or absence of 100 mM NaCl. Data are presented as treatments mean \pm SE ($n = 4$). The same letter in rows indicates that data did not differ significantly by Duncan's multiple range test at $p < 0.05$. ABA, Abscisic acid; N, Nitrogen.

Treatments/ Parameters	APX	GR	SOD
	(U mg ⁻¹ Protein min ⁻¹)		
Control	0.634 \pm 0.018e	0.178 \pm 0.005e	4.54 \pm 0.108e
100 mM NaCl	0.845 \pm 0.024d	0.273 \pm 0.008d	6.46 \pm 0.153d
10 mM N	1.146 \pm 0.027c	0.345 \pm 0.008c	7.06 \pm 0.168c
25 μ M ABA	0.676 \pm 0.019e	0.195 \pm 0.005e	4.74 \pm 0.113e
25 μ M ABA + 10 mM N	1.188 \pm 0.028c	0.358 \pm 0.008c	7.29 \pm 0.173c
10 mM N + 100 mM NaCl	1.435 \pm 0.034b	0.405 \pm 0.009b	8.12 \pm 0.193b
25 μ M ABA + 100 mM NaCl	1.401 \pm 0.033b	0.386 \pm 0.009b	7.89 \pm 0.188b
25 μ M ABA + 10 mM N + 100 mM NaCl	1.790 \pm 0.036a	0.482 \pm 0.009a	8.72 \pm 0.207a

2.8. Impact of ABA and/or N on Cysteine and GSH Content under NaCl Stress

Under NaCl stress, the contents of Cys and GSH increased by 14.6% and 39.2%, respectively, in comparison to the control plants. Individual supplementation of N showed increased Cys and GSH content both under stress and in control conditions. ABA alone could not induce Cys and GSH content under control conditions, while it showed a significant increase in the contents of these parameters in N-supplemented plants. Under control conditions, N increased GSH content by 27.3% and Cys content by 128.1% in comparison to the control. Under stress conditions, plants supplemented with ABA exhibited an increase in GSH content by 26.1% and Cys content by 98.0%, while plants receiving N exhibited an increase in GSH content by 34.0% and Cys content by 109.2% in comparison to the NaCl-treated plants. A comparatively more pronounced increase was found under stress conditions when ABA was supplemented together with N. The combined application of ABA plus N under salt stress increased GSH by 45.2% and Cys by 126.1% in comparison to the NaCl-treated plants (Figure 5).

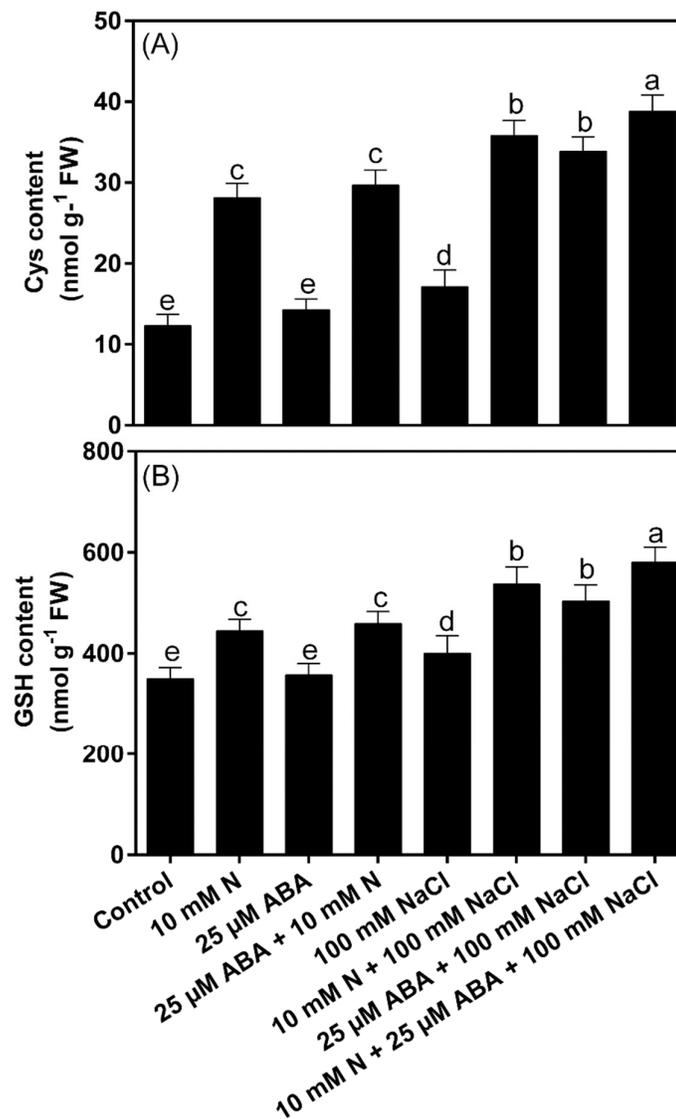


Figure 5. Content of cysteine (Cys; **(A)**) and reduced glutathione (GSH; **(B)**) in Indian mustard (*Brassica juncea* L.) at 30 days after sowing. Plants were grown individually with 0 or 10 mM N and treated with 25 μ M ABA in the presence or absence of 100 mM NaCl. Data are presented as treatments mean \pm SE ($n = 4$). The same letter above bars indicates that data did not differ significantly by Duncan's multiple range test at $p < 0.05$. ABA, Abscisic acid; N, Nitrogen; FW, Fresh weight.

2.9. Effect of ABA and/or N on Contents of N, S and Activity of NR and ATP-S under NaCl

NaCl treatment reduced N content by 48.1% and NR activity by 23.6% in comparison to the control plants. Under control conditions, the supplementation of N alone or N together with ABA increased N content and NR activity equally by about 62.2% and 46.3%, respectively, while individual supplementation of ABA did not result in any significant change in both the parameters. However, under NaCl stress, plants supplemented with ABA or N alone showed an increase in the N content (25.2% or 49.1%) and NR activity (49.2% or 57.1%), respectively. The combined treatment of ABA and N showed a more prominent increase in N content and NR activity by 130.5% and 116.0%, respectively, in comparison to the NaCl-treated plants (Figure 6A,B). Under NaCl stress, S content decreased by 44.0%, while ATP-S activity increased by 33.4% in comparison to the control plants. Under non-stress conditions, N supplementation

increased S content by 43.1% and ATP-S activity by 76.0%, while ABA supplementation alone had no significant effect in comparison to the control plants. Plants grown with salt stress and N showed increased S content and ATP-S activity by 50.1% and 48.2%, respectively, while plants receiving ABA in the presence of salt stress increased S content and ATP-S activity by 26.3% and 8.2%, respectively, in comparison to the NaCl-treated plants. However, a more prominent increase was noted with combined treatment of ABA and N; where the increase of 122.2% and 60.0% in S content and ATP-S activity, respectively, was observed in comparison to the NaCl-treated plants (Figure 6C,D).

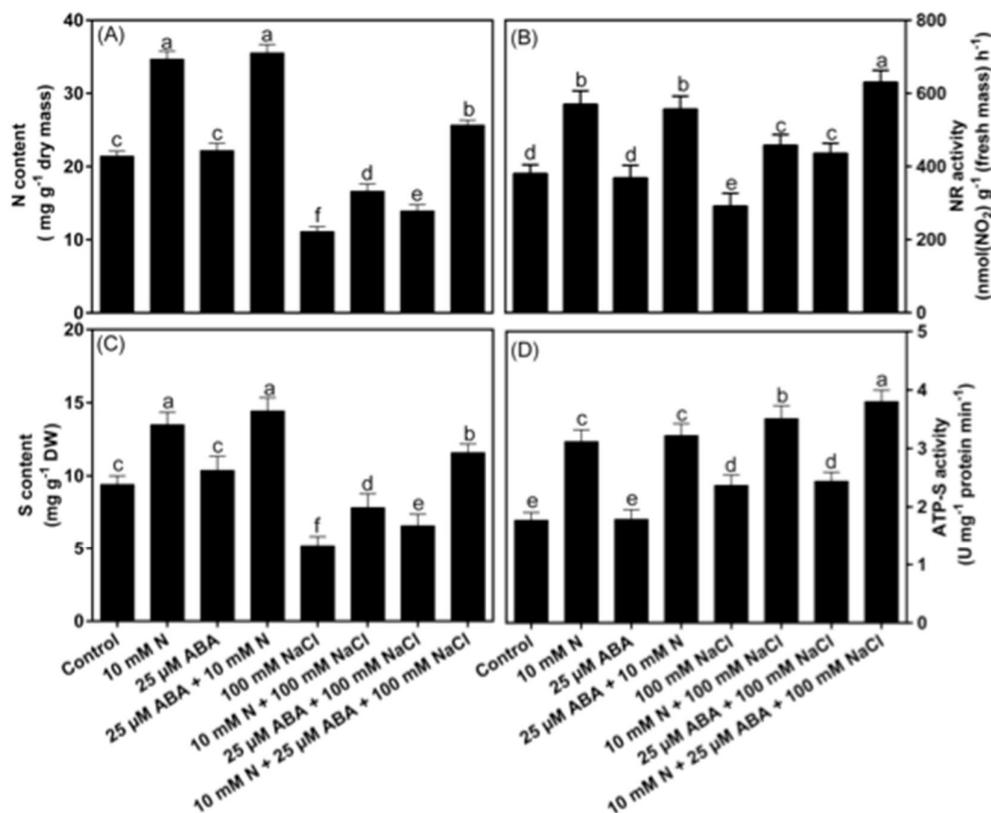


Figure 6. Content of nitrogen (N; (A)), sulfur (S; (C)) and the activity of nitrate reductase (NR; (B)) and ATP-sulfurylase (ATP-S; (D)) in Indian mustard (*Brassica juncea* L.) at 30 days after sowing. Plants were grown individually with 0 or 10 mM N and treated with 25 µM ABA in the presence or absence of 100 mM NaCl. Data are presented as treatments mean ± SE ($n = 4$). The same letter above bars indicates that data did not differ significantly by Duncan's multiple range test at $p < 0.05$. ABA, Abscisic acid; N, Nitrogen; DW, Dry weight.

3. Discussion

To date, most studies on salinity, ABA and N have focused either on salinity as a limiting factor for plant health and productivity [10] or on the ABA [28] or N [29] influence on plant health and productivity, while their interactive effects are still not fully understood. Hence, understanding the interaction between salinity, ABA and N supply could be of great agronomic importance. Given this, the effort was made herein to interpret the results and discuss the same in detail the ABA, N and their interactive significance taking into account three major aspects: (i) modulation of ionic toxicity (Na^+ and Cl^-) and mineral elements (N and S)-accumulation, and the status of oxidative stress and antioxidant metabolism; (ii) control of the contents of N and S; N-containing osmolyte proline and S-containing compounds (Cys and GSH), and activity of NR and ATP-S; and (iii) the significance of the cumulative outcome of the aspects on the overall improvement of the photosynthetic and growth parameters.

3.1. Modulation of Ionic Toxicity (Na^+ and Cl^-) and Mineral Elements (N and S)-Accumulation, and the Status of Oxidative Stress and Antioxidant Metabolism

As observed in the present study, the elevated accumulation of Na^+ and Cl^- ions in plant tissues was considered an obvious response in plants exposed to soils with high NaCl concentrations [30]. Upon entry into the cells, these Na^+ and Cl^- may cause osmotic stress (causing cellular water deficit), imbalance of the uptake of essential nutrients (such as N and S) and/or elevated or non-metabolized ROS-accrued oxidative stress [4]. Plants tend to acquire nutrients from the environment surrounding their root system. However, the presence of soil salinity and excess Na^+ and Cl^- ions can disrupt the uptake of most nutrients in glycophytes (such as *B. juncea*) through competitive interactions or by affecting the membrane selectivity for ions [13]. In context with the salinity impact on N and S contents, salinity (50 mM NaCl)-mediated decreases in the contents of both N and S were reported in *B. juncea* [31]. In particular, enhancing the supply of N or its sources (such as NO_3^-) in salinity-exposed crop plants was reported to significantly reduce Na^+ and Cl^- contents, increase N level in leaves and thereby alleviate crop salt injury or enhance its salt tolerance [26,32]. On the other hand, the occurrence of the positive relationship between ABA accumulation and salinity tolerance, and the ABA-mediated accumulation of K^+ , Ca^{2+} and compatible solutes, such as proline (and also sugars), in root vacuoles were argued to counteract the uptake of Na^+ and Cl^- [17,33].

Elevated ions (Na^+ and Cl^-) in plant organs, including roots and leaves, can cause oxidative stress by hampering the balance between the generation and scavenging of ROS, such as studied herein O_2^- and H_2O_2 [34]. Excess or non-metabolized ROS may cause the oxidation of macromolecules, including membrane lipids [11]. Measured herein in terms of TBARS content, the highest lipid peroxidation was observed in NaCl (100 mM)-alone treated *B. juncea*, which coincides with recent reports [34,35]. However, the contents of both oxidative stress parameters (O_2^- and H_2O_2) and lipid peroxidation indicator (TBARS) were least in NaCl (100 mM)-exposed *B. juncea* supplied with 25 μM ABA and 10 mM N, which could be attributed to ABA- and/or N-mediated decreases in the toxicity of ions (Na^+ and Cl^-) and eventual minimization of oxidative stress and its consequences.

As one of the defense strategies, the increased oxidative stress induced the antioxidant defense system in order to protect *B. juncea* against salt stress-caused damages. This study revealed ABA (25 μM) and N (10 mM)-mediated maximal enhancements in the activity of SOD, an enzyme involved in O_2^- -dismutation to H_2O_2 ; APX, an enzyme involved in H_2O_2 -metabolism; and also the activity of GR, an enzyme involved in the regeneration of reduced GSH that as a major component of the Halliwell–Asada pathway is directly or indirectly involved in ROS-metabolism [36–40]. Earlier, the supply of N [41] and ABA [42–45] has been reported to enhance ROS-metabolizing enzymes in different stressed test plants.

3.2. Control of the Status of N-containing Osmolyte Proline and S-containing Compounds (Cys and GSH), and Activity of NR and ATP-S

Plants tend to metabolically adapt to salinity stress by mainly enhancing the synthesis of S-rich compounds (such as Cys and GSH) and N-containing proteogenic amino acids/osmolytes (such as proline) [22]. Therefore, in relation to ABA and N supplies in NaCl-exposed *B. juncea*, it is imperative to discuss the status of tuning between N- and S-assimilatory enzymes, NR and ATP-S, respectively; and N- and S-assimilatory products, proline, and Cys and GSH, respectively. On the one hand, exogenously supplied N triggers the synthesis of compatible solutes, such as proline (a nitrogenous osmolyte), and the solute played a vital role in osmotic adjustment in NaCl-exposed plants [46]. In the study presented here, the maximal increase in proline was observed when 10 mM N and 25 μM ABA were supplemented together under both stressed and non-stressed conditions. Notably, besides acting as a compatible solute,

proline also played an important role in scavenging free radicals and protecting redox potential under NaCl stress conditions [47]. The plant status of proline and other compatible solutes was regulated by phytohormones (including ABA) and mineral nutrients (including N) [16]. Further, exogenously supplied proline was also reported to mediate the upregulation of genes associated with the antioxidant defense system in salinity-stressed plants [48]. Interestingly, proline biosynthesis is regulated by the crosstalk of ABA signaling and phosphate homeostasis through activation of the P5CS1 gene [49]. On the other hand, the assimilatory pathways of N and S are well coordinated in plants; and the S metabolism is strongly affected by the plant's N status [50]. Hence, N (10 mM)-supply mediated enhancements in the contents of N and S, the activity of NR and APT-S and increases in Cys and GSH contents in NaCl (100 mM)-exposed *B. juncea* are obvious. NR is a rate-limiting enzyme for NO_3^- reduction, where its sustained activity is crucial for N assimilation; whereas, ATP-S activates the metabolically inert SO_4^{2-} and makes it usable in Cys and GSH during S assimilation [18,19]. Earlier, exogenous supplementation of N was reported to restore the ATP-S activity in N-deficient mediums [51]. Additionally, an exogenous supply of S enhanced the ATP-sulfurylase and NR activity when compared with plants grown without S [52]. In earlier studies, the exogenous supplementation of GSH improved salt stress tolerance and thereby growth and development in plants under NaCl stress [53–55]. With an improved S assimilation pathway, the plants' potential to survive under oxidative stress conditions has been reported in various studies [56,57].

3.3. Overall Improvement in the Photosynthetic and Growth Parameters

Photosynthetic and growth parameters are among the major parameters at the whole plant level that can reflect the cell-level metabolic processes in plants growing under varying conditions. The reduction in growth and photosynthesis under salt stress may be due to the overproduction of ROS, as increased TBARS and H_2O_2 content were observed. ROS interferes with the proper functioning of cell membrane lipids, proteins and other important enzymes of metabolic pathways, and thus, resulted in reduced growth and photosynthetic attributes. Several studies showed a decrease in growth under NaCl stress [58–60]. The reduction in leaf area under salt stress might be due to the toxicity of Na^+ and Cl^- in the shoot cells. If finely coordinated, various components of the antioxidant defense system (such as GSH, proline, SOD, APX and GR) can efficiently metabolize varied ROS, such as H_2O_2 and O_2^- and $\cdot\text{OH}$; and thereby protect photosynthetic and growth parameters. As discussed above in Sections 4.1 and 4.2, the combined supply of ABA (25 μM) and N (10 mM) significantly improved primarily: (i) NR activity that contributed to an increased cellular level of proline, which in turn maintained osmotic adjustment; and secondarily (ii) enhanced ATP-S activity contributed in yielding first Cys and thereafter the reduced pool of GSH; and thereby cumulatively scavenged varied ROS and protected photosynthetic and growth parameters of *B. juncea* against NaCl (100 mM)-impacts. The increased leaf area and plant dry mass with N has been attributed to the increased GSH level [61]. This is in confirmation of the earlier findings that exogenously supplied N improves growth and development by increasing photosynthesis, chlorophyll content, proline production and nitrogen metabolism [62,63]. Fatma et al. [64] reported that increased ATP-S activity in *B. juncea* indicated its higher sulfate accumulation capacity with increased photosynthetic attributes and plant dry mass. Notably, GSH and proline can influence the activity of Rubisco and modulate the photosynthetic functions and eventually growth parameters [20–22].

Notably, the scanning electron microscopic study revealed the potential of ABA and N on the stomatal behavioral response of plants. In our study, comparatively closed stomata were found in NaCl treated plants compared to the control plants. Treatment with N reduced the closing effect of NaCl stress on stomata, and combined treatment of ABA and N maximally reduced the effect of NaCl on stomatal width aperture. Generally, it has been shown that ABA induces stomatal closure, but it has also been found that an increase in intracellular GSH suppresses stomatal closure [65]. The combined treatment

of ABA and N prominently influenced GSH production, which resulted in the opening of stomata when compared with salt-treated plants. It is interesting to note that stomatal closure occurs on the reduction of intracellular GSH and that intracellular GSH regulates stomatal behavior. Hence, this study confirmed the role of intracellular GSH in N and ABA-mediated regulation of NaCl-induced ROS production and stomatal closure.

4. Methodology

4.1. Experimental Design and Growth Conditions

Seeds of Indian mustard (*Brassica juncea* L. cv. RH0-749) obtained from the Indian Agricultural Research Institute (IARI), New Delhi, were sterilized using HgCl₂ solution (0.01%) and were washed repeatedly with double distilled water (DDW). Sterilized seeds were sown in clay pots filled with 5 kg of acid-washed sand, purified according to the method given by Hewitt [66]. The pots were kept in the greenhouse of the Department of Botany, Aligarh Muslim University, Aligarh, India, with average day/night temperatures of 23/14 ± 3 °C, 620 μmol m⁻² s⁻¹ photosynthetically active radiation (PAR) and the relative humidity was 62 ± 3%. After germination, three plants per pot were maintained. Two experiments were performed. In the first experiment, a concentration of ABA exhibiting the highest alleviation of salinity stress-impact was worked out by applying 0, 5, 10, 25 and 50 μM ABA to plants grown with 0 or 100 mM NaCl and studying photosynthesis, growth and oxidative stress. ABA was dissolved in ethanol to prepare the stock solution (100 mM), which was then diluted with DDW to obtain the desired concentrations of the solution. The ABA solution was sprayed on the foliage evenly at 15 days after germination using a hand sprayer. Based on the results obtained from this first experiment, 25 μM ABA was found to maximally alleviate the impact of 100 mM NaCl on photosynthesis, growth and oxidative stress. Additionally, based on an earlier report, 10 mM N was considered as the optimal concentration for N in alleviating the salinity impact on *B. juncea* [18]. Hence, 25 μM ABA and 10 mM N were considered in the second experiment, where 0 or 25 μM ABA was applied to the leaf of plants grown with 10 mM N in the absence or presence of 100 mM NaCl at 20 days after sowing (DAS). Salt and N treatments were given at 10 DAS. KNO₃ was used as the source of N, K⁺ concentration was maintained in all treatments by the addition of KCl. The control set of plants was supplemented with 250 mL of Hoagland nutrient solution at alternate days and 250 mL of distilled water daily. Sampling was done at 30 DAS. The design of the experiment was a randomized complete block design, and the number of replicates for each treatment was four.

4.2. Analyses of Na⁺ and Cl⁻ Content

Content of Na⁺ and Cl⁻ was measured in roots and leaves. One gram of oven-dried plant tissue was dissolved in 4 mL concentrated HNO₃ (68%) in a 100 mL glass beaker. The beaker containing the digested sample was heated on a water bath until brown effervescence was observed. Afterward, 38 mL of tri acid mixture solution was added drop-wise till a clear solution was obtained. The solution containing plant samples was dried on the hot plate, and after that, dried samples were diluted with DDW to make a final volume of 100 mL. The Na⁺ content was measured using a flame photometer (Khera-391: Khera Instruments, New Delhi), and Cl⁻ content by titration against the 0.02 N silver nitrate using 5% potassium chromate solution as an indicator.

4.3. Content of H₂O₂ and TBARS

The H₂O₂ content was measured in leaves by following the method of Okuda et al. [67]. Leaves (1.0 g) were grounded using 200 mM perchloric acid (ice-cold) and centrifuged at 1200×g for 10 min, and the supernatant was neutralized by adding 4 M potassium hydroxide. For measuring optical density (OD) at 590 nm, 2 mL of the eluate was mixed with 1 mL solution containing 160 μL of 3-methyl-2-benzothiazoline hydrazine, 40 μL of peroxidase and 800 μL of 12.5 mM 3-(dimethylamino) benzoic acid.

The TBARS content was measured in leaves by following the method of Dhindsa et al. [68]. Leaves (1.0 g) grounded in 0.25% thiobarbituric acid (TBA) were heated for 30 min in a water bath and rapidly cooled in cold water, followed by centrifugation (10,000× g) for 15 min. For measuring OD at 532 nm, 2 mL of supernatant was mixed with 8 mL 20% TCA containing 0.5% TBA. The content of TBARS was calculated using an extinction coefficient of 155 mM⁻¹ cm⁻¹.

4.4. Superoxide Ion (O₂⁻) and H₂O₂ by a Histochemical Staining Method

In situ determination of the level of superoxide ion (O₂⁻) and H₂O₂ generation were visually detected by following the method of Kumar et al. [69] with slight modification. Freshly prepared nitro blue tetrazolium (NBT) solution and 3,3-diaminobenzidine (DAB) solution were used for detecting O₂⁻ and H₂O₂, respectively. NBT solution was formed by dissolving 0.2 g of NBT in 100 mL of sodium phosphate buffer (50 mM; pH 7.5). The DAB solution (pH 3.8) was formed by dissolving 100 mg of DAB in DDW in an amber-colored bottle. For NBT and DAB staining, leaf samples were soaked in NBT solution and DAB solutions, respectively, and incubated overnight at room temperature. The samples were then boiled for 20 min in absolute ethanol, and then photographs were taken.

4.5. Analysis of H₂O₂ in Roots by Confocal Laser Scanning Microscopy

The content, as well as visualization of H₂O₂, was done in leaves. In order to know the status of ROS (such as H₂O₂) at the root level, confocal microscopy was employed to visualize H₂O₂ formation in roots of *B. juncea* using 2',7' dichlorofluorescein (H₂DCFDA) staining. In brief, root samples were immersed for 15 min in a freshly prepared 12.5 μM H₂DCFDA solution. After repeated washing with DDW, temporary slides of stained samples were prepared, and fluorescence was monitored using a confocal laser scanning microscope (Model LSM 780; Carl Zeiss, Oberkochen, Germany) at excitation 400–490 nm and emission ≥ 520 nm.

4.6. Photosynthetic Parameters and Rubisco Activity

Net photosynthetic rate (P_N), intercellular CO₂ concentration (C_i) and stomatal conductance (g_s) were measured using an Infrared Gas Analyzer (IRGA, Model CID-340). The parameters were measured between 11.00 a.m. and 1.00 p.m. at light saturating intensity (PAR: 760 μmol m⁻² s⁻¹), temperature 24 ± 2 °C and at 385 ± 15 μmol mol⁻¹ atmospheric CO₂ concentration. Chlorophyll content was measured in fully expanded young leaves using the SPAD chlorophyll meter (Model 502 DL PLUS, Konica Minolta, Inc., Tokyo, Japan). The Rubisco activity was measured in leaves by following the method of Usuda [70].

4.7. Growth Parameters

The plants were dried in the oven at 80 °C, and the dried material was weighed on an electrical balance. Leaf area (LA) was calculated with a leaf area meter (Model LA 211, Systronics, New Delhi, India).

4.8. Assay of Antioxidant Enzymes

Fresh leaves (0.2 g) were homogenized with an extraction buffer using a chilled mortar and pestle. The extraction buffer was prepared by dissolving 0.05% Triton X-100 and 1% polyvinylpyrrolidone in 100 mM potassium phosphate buffer (pH 7.0). The homogenate was centrifuged (15,000× g) at 4 °C for 20 min, and the supernatant was used for the assay of superoxide dismutase (SOD) and glutathione reductase (GR). For the ascorbate peroxidase (APX) assay, 2 mM ascorbate was added to the extraction buffer. The activity of APX and SOD was calculated by following the method of Asada [71] and Beyer and Fridovich [72], respectively. The activity of GR was measured according to the method of Foyer and Halliwell [73].

4.9. Determination of Nitrate Reductase (NR) Activity and N Content

The NR activity was measured according to the method of Kuo et al. [74]. One gram of leaves was frozen in liquid nitrogen, ground to a powder and then homogenized in 250 mM Tris-HCl buffer (pH 8.5) using a chilled mortar and pestle. The buffer was prepared by dissolving 1 mM EDTA, 10 mM Cys, 1 mM DTT and 20 μ M FAD in 10% glycerol. The homogenate was centrifuged (10,000 \times g) at 4 °C for 30 min. NR activity was measured as the rate of nitrite production at 28 °C following the method of Nakagawa et al. [75]. In the reaction mixture, NADH was used for initiating the reaction. After 20 min, the reaction was ended by adding 1 N HCl containing 1 mL of 1% sulphanilamide solution. Subsequently, 1 mL of 0.02% aqueous NED was added. The reaction mixture (1.5 mL) containing enzyme extract, 10 mM KNO₃, 0.065 M HEPES (pH 7.0) and 0.5 mM NADH in 0.04 mM phosphate buffer (pH 7.2) was used for measuring absorbance at 540 nm using a spectrophotometer after 10 min.

The content of N was measured by the Kjeldahl digestion method, as described by Lindner [76]. A 20 mL aliquot of the digested leaf sample was taken in a 100 mL volumetric flask. To this flask, 10% sodium silicate (2 mL) and 4 mL of 2.5 N sodium hydroxide solutions were added to prevent turbidity and neutralize the excess of acid, respectively. The volume was made up to 100 mL with DDW. In a 20 mL test tube, 10 mL aliquot was taken, and 1 mL Nessler's reagent was added. The final volume was maintained with DDW. The OD was recorded on a spectrophotometer at 525 nm.

4.10. Determination of Proline Content

The proline content was calculated according to the method of Bates et al. [77]. Fresh leaf tissues (1.0 g) were homogenized in 10 mL of 3% sulphosalicylic acid, and 4 mL each of acid ninhydrin and glacial acetic acid was added to the filtrate. The test tubes containing the homogenate filtrate were heated in a water bath for one hour, followed by immediate cooling of the test tubes in ice-cold water. Afterward, the mixture was extracted with toluene, and the OD was measured on a spectrophotometer at 520 nm using L-proline as a standard.

4.11. Analyses of Content of S, Cysteine, GSH, and ATP-S Activity

For measuring S content, 0.2 g oven-dried leaves were ground and dissolved in a solution containing 70% HNO₃ and 60% HClO₄ (85:15, v/v). The sulfur content was measured following the turbidimetric method. For turbidity development, 2.5 mL gum acacia solution (0.25%) and 1.0 g BaCl₂ were added to a 5 mL aliquot, and the final volume was made using DDW. Within 10 min of the turbidity development, the OD was measured at 415 nm.

The content of Cys and GSH was measured according to the method of Gaitonde [78] and Anderson [79], respectively. In fresh leaves, the activity of ATP-sulfurylase (ATP-S) was measured according to the method of Lappartient and Touraine [80].

4.12. Analysis of Stomatal Behavior

Fresh leaves were fixed with 2.5% glutaraldehyde solution for 4–5 h at room temperature. Following repeated washing steps using phosphate buffer (15 min at each step), the samples were dehydrated through a graded series of ethanol solutions (60%, 70%, 80% and 95%) for about 20 min at each step. After that, the samples were placed in absolute ethanol. The small sections of dehydrated samples were coated with gold-palladium and observed under the scanning electron microscope (SEM; Carl Zeiss EVO 40, Carl Zeiss AG, Oberkochen, Germany) at a magnification of 3000 \times g. The stomata were visualized using SEM images.

4.13. Statistical Analysis

The data were subjected to statistical analysis using analysis of variance (ANOVA), and Duncan's multiple range test was used to compare means of different treatments by IBM SPSS software (version 22.0). The number of replicates for each treatment was four, and data are presented as a mean \pm SE. The least significant difference (LSD) at $p < 0.05$ was considered as significant.

5. Conclusions

Overall, the results indicated that NaCl stress severely induced oxidative stress through overproducing ROS and eventually adversely affected the photosynthesis and plant growth of *B. juncea*. Exogenously supplied N enhanced photosynthesis and growth parameters under no stress and protected these parameters against salt stress. However, ABA supplementation improved these parameters only under stressed conditions. Compared to the individual influence of N and ABA, their combined application proved to be the most effective in combating NaCl-induced toxic effects on photosynthesis and growth. The more positive influence of the combined application of ABA and N was through their effect on nitrogenous osmolyte (proline), antioxidant enzymes (SOD, APX and GR), N and S-assimilation and production of Cys and GSH. Therefore, the supplementation of 10 mM N in combination with 25 μ M ABA prominently enhanced the photosynthetic potential of oleiferous brassica under saline conditions. However, the precise regulatory molecular mechanism of ABA and/or N-induced NaCl stress tolerance opens the avenue for future research.

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