

Abstract

Clastogenic Effects of Thyme Essential Oil on *Vicia faba* [†]

Victoria Bînzari ¹, Denisa-Ioana Gheorghe ^{1,2}, Carmen Lupu ^{1,*}, Diana Constantinescu-Aruxandei ¹ 
and Florin Oancea ^{1,*} 

¹ National Institute for Research and Development in Chemistry and Petrochemistry Bucharest, 202 Splaiul Independentei, 6th District, 060021 Bucharest, Romania; victoria.binzari@icechim.ro (V.B.); denisa.gheorghe@icechim.ro (D.-I.G.); diana.constantinescu@icechim.ro (D.C.-A.)

² Faculty of Biology, University of Bucharest, 91-95 Splaiul Independentei, 6th District, 050095 Bucharest, Romania

* Correspondence: carmen.lupu@icechim.ro (C.L.); florin.oancea@icechim.ro (F.O.)

[†] Presented at the 17th International Symposium “Priorities of Chemistry for a Sustainable Development” PRIOCHEM, Bucharest, Romania, 27–29 October 2021.

Abstract: Indiscriminate use of pesticides in agriculture has many negative implications on both abiotic and biotic components of the environment [1]. One of the alternative methods to maintaining productivity and quality of life is using compounds produced by the secondary metabolism of plants, such as essential oils [2]. Due to their rapid efficacy and degradability, essential oils are used as bioherbicides, biostimulants, anti-microbial agents, insect repellents, etc. Evaluations of essential oils toxic, cytotoxic, and genotoxic potentials employing ecotoxicological bioassays are of great importance in determining possible risks [3]. To determine genotoxicity and clastogenic effects of various factors, mitotic divisions are used, with the evaluated parameters being the mitotic index (MI) and the frequency of micronuclei [4]. This study aims to analyze the potential phytotoxic effect of thyme essential oil given its potential use as a plant biostimulant. This phytotoxic assay was done by investigating the clastogenic effect on *Vicia faba* root meristems. After sterilization, seeds were left to hydrate for 24 h in sterile water. Sterile deionized water was used for the control variant and thyme essential oil at a 0.1% concentration for the sample. The seed plates were placed at 23 °C under dark conditions until the length of rootlets reached 2–3 cm. For cytological analysis of the mitotic index (MI) and micronucleus (MN) tests, 1–2 cm of rootlets were subjected to Carnoy fixation solution for 24 h. The samples were then rinsed with distilled water and hydrolyzed with 1N HCl at 60 °C for 6 min. Schiff’s reagent was used for staining. The mitotic index was calculated as the number of cells in mitosis divided by the total number of cells, x 100, per 1000 scored cells/sample resulting from 10 separate roots for each group. The mitotic index of *Vicia faba* in the 0.1% essential oil sample did not show significant differences compared to the control sample. The mean values of MI were 31.4% for the control and 31.2% for the sample with thyme essential oil, indicating a similar cell division ratio. Additionally, this essential oil concentration did not significantly lead to micronuclei formation at root meristems relative to the control. Various types of physiological (C-metaphase, stickiness, bridge, laggard, etc.) and clastogenic chromosomal aberrations (chromosomal breaks, fragments, etc.) were not observed when analyzing the cell division phases. Following the study performed on *Vicia faba*, it was noted that 0.1% thyme essential oil has no cytotoxic effect, as no chromosomal aberrations were observed in the samples, and it did not induce the inhibition of cell proliferation in root meristems. The relative frequencies of the various mitotic phases were not affected by thyme essential oil.

Keywords: essential oil; ecotoxicology; mitotic index



Citation: Bînzari, V.; Gheorghe, D.-I.; Lupu, C.; Constantinescu-Aruxandei, D.; Oancea, F. Clastogenic Effects of Thyme Essential Oil on *Vicia faba*. *Chem. Proc.* **2022**, *7*, 30. <https://doi.org/10.3390/chemproc2022007030>

Academic Editors: Mihaela Doni, Zina Vuluga and Radu Claudiu Fierăscu

Published: 8 March 2022

Publisher’s Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Author Contributions: Conceptualization, V.B. and F.O.; methodology, C.L.; software, F.O.; validation, D.-I.G., D.C.-A. and C.L.; formal analysis, F.O.; investigation, V.B.; resources, F.O.; data curation,

D.C.-A.; writing—original draft preparation, V.B.; writing—review and editing, D.C.-A.; visualization, C.L.; supervision, F.O.; project administration, C.L.; funding acquisition, F.O. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by project POC-A1-A1.2.3-G-2015-P_40_352-SECVENT, My_SMIS 105684, “Sequential processes of closing the side streams from bioeconomy and innovative (bio) products resulting from it, subsidiary project SECVENT 81/2016”. The SECVENT project was cofunded by the European Regional Development Fund (ERDF), The Competitiveness Operational Programme (POC), Axis 1.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Issa, M.; Chandel, S.; Singh, H.P.; Batish, D.R.; Kohli, R.K.; Yadav, S.S.; Kumari, A. Appraisal of phytotoxic, cytotoxic and genotoxic potential of essential oil of a medicinal plant *Vitex negundo*. *Ind. Crops Prod.* **2020**, *145*, 112083. [[CrossRef](#)]
2. de Assis Alves, T.; de Assis Alves, T.; da Cunha Henrique, M.K.; de Paula Azevedo, A.F.N.; Carvalho, J.A.M.; Pinheiro, P.F.; Menini, L.; Praça-Fontes, M.M. Phytotoxic and cyto-genotoxic activity of essential oil from leaf residues of *Eucalyptus urophylla* and the hybrid *E. urophylla* × *E. camaldulensis* on *Lactuca sativa* and *Sorghum bicolor*. *Res. Soc. Dev.* **2021**, *10*, e242101119646. [[CrossRef](#)]
3. de Sousa Silva, T.; e Silva, A.P.S.; de Almeida Santos, A.; Ribeiro, K.G.; de Souza, D.C.; Bueno, P.A.A.; Marques, M.M.M.; de Almeida, P.M.; Peron, A.P. Cytotoxicity, genotoxicity, and toxicity of plant biostimulants produced in brazil: Subsidies for determining environmental risk to non-target species. *Water Air Soil Pollut.* **2020**, *231*, 233. [[CrossRef](#)]
4. Andronic, L.I.; Jacota, A.G.; Bujoreanu, V.V.; Grigorov, T.B. Genotoxicity of barley stripe mosaic virus in infected host plants. *Cent. Eur. J. Biol.* **2010**, *5*, 633–640. [[CrossRef](#)]