

## Supplementary Data

### BSEM equations and parameters

The general form of equations governing the biogeochemical model is expressed by

$$\frac{\partial A}{\partial t} + \nabla(\vec{u}.A) = F_A + R(A) \quad (S1)$$

where  $A$  denotes each of the state variables,  $\vec{u}$  is the three dimensional fluid velocity vector,  $F_A$  denotes the sum of horizontal and vertical diffusion terms,  $R(A)$  refers the local rates of change of the 12 model compartments. Except  $R(A)$ , the mathematical forms of all terms in the differential equation above and their numerical solution procedures are similar to those of the temperature and salinity transport equations in the GETM/GOTM [1]. GETM and GOTM modules solve 3D transport equations for 12 non-conservative substances that BSEM involves. Detailed representation of source-sink terms is described below for each model compartment.

#### S.1. The autotrophs

The biological source-sink term  $R(P)$  for the phytoplankton include the phytoplankton growth that is reduced by overall limitation function,  $\Psi_X$ , grazing by different zooplankton groups, and physiological mortality (Eq. S2). Temporal variations of the large ( $P_L$ ) and small ( $P_S$ ) phytoplankton biomass are governed by the biological source-sink terms and settling velocity of the form

$$R(P_X) = \sigma_X \cdot \Psi_X \cdot P_X - \sum GR(P_X) \cdot Z_Y - m_{PX} \cdot P_X - w_p \frac{\partial P_X}{\partial z}, \quad (S2)$$

where

$$\Psi_X = f_X(N_n, N_a, PO) \cdot f_X(I) \cdot f_X(T) \quad (S3)$$

represents the total limitation function of the primary production, and the subscript  $X$  denotes either  $L$  for the large or  $S$  for the small size group, while  $Y$  denotes  $S$ ,  $L$  or  $N$ , for small-, large zooplankton or *Noctiluca*, respectively;  $w_p$  is the settling velocity of the phytoplankton ( $w_p = 0$  for the small phytoplankton and  $w_p = w_{pL}$  for large); and  $z$  is the depth. The phytoplankton growth is modelled as

the product of the maximum specific growth rate,  $\sigma_x$ , total limitation function,  $\Psi_x$ , and phytoplankton biomass  $P_x$ . The function  $f_x(N_n, N_a, PO)$  is parameterised assuming that either nitrogen or phosphorus controls phytoplankton growth [2]. This function represents the nutrient uptake expressed as the minimum of total nitrogen (sum of nitrate and ammonium) limitations  $f_x(N_n, N_a) = f_x(N_n) + f_x(N_a)$  and phosphorus limitation  $f_x(PO)$ .

Thus the function  $\Psi_x$ , that limits the primary production is defined by

$$\Psi_x = f_x(I) \cdot f_x(T) \cdot \min\{f_x(N_n) + f_x(N_a), f_x(PO)\}. \quad (S4)$$

The limiting functions in equation (S4) are expressed by the Monod-type hyperbolic functions involving a saturation response at high resource concentrations.

$$f_x(N_a) = \left[ \frac{N_a}{K_{AX} + N_a} \right], \quad f_x(N_n) = \left[ \frac{N_n}{K_{NX} + N_n} \right] \cdot e^{-\lambda \cdot N_a}, \quad f_x(PO) = \left[ \frac{PO}{K_{POX} + PO} \right], \quad (S5)$$

where

$$K_{NX} = a + bP_x, \quad K_{AX} = 0.1 K_{NX} \quad \text{and} \quad K_{POX} = K_{NX}/np \quad (S6)$$

are the corresponding half saturation functions of nitrate, ammonium and phosphate uptakes. They are parameterised following [3]. Values of the constants  $a$ ,  $b$  and  $\lambda$  are given in Table S2, and the denominator  $np$  represents the  $N:P$  ratio. The nitrogen limitation function  $f_x(N_n)$  (Eq. S5) is based on [4] and accounts for the inhibition of nitrate uptake in the presence of ammonium. Following Liebig's law of the minimum, the nutrient uptake rate is thus limited by either total nitrogen or phosphorus.

The growth is also subject to simultaneous limitations by the photosynthetically available radiation  $f_x(I)$ , and temperature  $f_x(T)$ . Using a spectrally unresolved model, the light limitation is parameterized by [5]

$$f_x(I) = [\tanh(\alpha_x I)] \cdot e^{-\beta_x I}, \quad (S7)$$

where  $\alpha_x$  is a parameter controlling slope of the photosynthesis-irradiance curve at low values of the photosynthetically available irradiance (PAR) whose intensity  $I$  is calculated by the hydrodynamic model,  $\beta_x$  is the photoinhibition parameter to reduce the growth at high irradiance conditions.

The silicate control on the diatom growth is neglected as the available data does not yield evidence for the prevailing role of silicate limitation although its input from major rivers tends to decline during the last two decades [6]. The temperature controls of the growth,  $f_x(T)$  for the large and small phytoplankton are assumed to be

$$f_L(T) = Q10^{(12-T)/12} \quad \text{and} \quad f_S(T) = Q10^{(T-20)/10}. \quad (S8)$$

In this way the large phytoplankton group can grow faster at low temperatures, while the small phytoplankton grows faster above 20° C. Weak temperature control is imposed for the small phytoplankton group, however the lower growth rate of the large phytoplankton group gives indirectly the small phytoplankton group a growth advantage.

Phytoplankton biomass decreases due to grazing by the zooplankton ( $Z_S$  and  $Z_L$ ) and heterotrophic dinoflagellate species *Noctiluca* ( $Z_N$ ), and phytoplankton mortality (second and third terms in the right-hand side of Eq. S2). The expressions  $GR(P_X)$ , used to model the grazing term is given in the next paragraph, where the ingestion of the heterotrophs and carnivores are presented. Coefficients of the phytoplankton mortality rates,  $m_{PX}$ , are given in Table S2.

## S.2. The heterotrophs and carnivores

Changes in the zooplankton biomass are controlled by ingestion, predation, excretion, and mortality which are expressed by

$$R(Z_Y) = \varphi \left[ \sum_i GR_j(A_i) \cdot Z_Y \right] - \sum_W GR_W(Z_Y) \cdot Z_W - \mu_Y \cdot Z_Y - m_{ZY} \cdot Z_Y^2, \quad (S9)$$

where  $Z_Y$  and  $Z_W \neq Z_Y$  ( $Y$  or  $W$  denote either  $S$ ,  $L$ ,  $N$  or  $G$ );  $\varphi$  is the coefficient of assimilation efficiency,  $\mu_Y$  and  $m_{ZY}$  are, respectively, the excretion rate and the natural mortality rate expressed in the quadratic form.

The ingestion terms  $GR_j(A_i)$  in Eq. (S9) are represented by the Michaelis-Menten (the so-called Holling type II) functional form for  $j$ th predator on  $i$ th prey, in terms of the maximum rate  $g_j$ , the temperature limitation function  $f_j(T)$ , and the food capture efficiency coefficient  $b_{j,i}$  for the food item  $A_i$ .

$$GR_j(A_i) = g_j \cdot f_j(T) \cdot \frac{b_{j,i} A_i}{K_j + [\sum_i b_{j,i} A_i]}, \quad b_{j,i} = \frac{a_{j,i} A_i}{\sum_i a_{j,i} A_i}, \quad (S10)$$

where the terms within the square bracket in the denominator refer to the total food available for the consumption of any zooplankton group;  $K_j$  denotes its half-saturation value;  $a_{j,i}$  denotes the constant food preference coefficient specified externally as in Table S1. According to the above equations, when a food type declines, its grazing preference decreases [7]. In this case, zooplankton select an alternative food type having higher biomass. Thus, grazing preferences may switch from one prey to another depending on local conditions and the predator may select temporally and spatially most favourable food types.

The temperature control of the growth,  $f_j(T)$ , is introduced in the form

$$f_S(T) = 1 \quad (S11a)$$

$$f_L(T) = 1 \quad (S11b)$$

$$f_N(T) = Q10^{(T-12)/8} \quad (S11c)$$

$$f_G(T) = 4.5 - 1/(T - 15) \quad \text{for } T > 16 \quad \text{and } f_G(T) = 1 \quad \text{otherwise.} \quad (S11d)$$

According to the observations, *Noctiluca* can maintain its growth at a wide temperature range of 12-30°C [8]. Eq. S11c suppresses the *Noctiluca* growth at low temperatures but favours it in spring and summer months when the surface mixed layer starts warming up. Eq. S11d imposes the growth advantage of *Mnemiopsis* and *Beroe Ovata* population at high temperatures during July-August and strong decline at the end of the year [9]. The mortality rate of  $Z_G$  is increased four times in November-

December in order to maintain  $Z_G$  peaks until October. The mortality rate of the large zooplankton,  $m_{ZL}$ , is controlled by the temperature, so,  $m_{ZL} = m_{0ZL}Q10^{(T-20)/10}$  and  $m_{0ZL}$  is a constant listed in Table S2.

### S.3. Particulate organic nitrogen and phosphorus

It is assumed that the dissolved organic nitrogen and phosphorus pool is controlled by the egestion and sloppy feeding (i.e. unassimilated part of the food grazed), phytoplankton and zooplankton mortalities, its consumption by zooplankton groups within the water column (the third term), the transformation into the dissolved organic nitrogen and phosphorus pool at a rate  $\varepsilon(DO)$  and the sinking rate of detritus,  $w_D$  (Eq. S12). As in [10], 70% of particulate organic nitrogen is assumed to decompose directly into ammonium and phosphorus.

$$R(D) = (1 - \varphi) \cdot Graz + Mort - \varphi \cdot Cons - \varepsilon_n \cdot D - w_D \frac{\partial D}{\partial Z}, \quad (S12)$$

$$Graz = GR_S(P_S) \cdot Z_S + [GR_L(P_L) + GR_L(Z_S) + GR_L(Z_N)] \cdot Z_L + [GR_N(P_L) + GR_N(Z_S)] \cdot Z_N + [GR_G(Z_S) + GR_G(Z_L) + GR_G(Z_N)] \cdot Z_G, \quad (S13)$$

$$Mort = \sum_X m_{PX} \cdot P_X + \sum_Y m_{ZY} \cdot Z_Y^2, \quad (S14)$$

$$Cons = GR_L(D) \cdot Z_L + GR_N(D) \cdot Z_N. \quad (S15)$$

$$\varepsilon_n = \varepsilon(DO) \cdot f_D(T), \text{ where } f_D(T) = e^{d_o T}, \quad (S16)$$

$$\varepsilon(DO) = \begin{cases} 1 & \text{for } DO > 250 \text{ mmol O}_2 \text{ m}^{-3} \\ \left\{ 1 + \frac{K_{DO}}{K_{DO} + DO} \right\} & \text{for } 250 \text{ mmol O}_2 \text{ m}^{-3} \geq DO \geq DO_l \\ 0.25 & \text{for } DO < DO_l \end{cases} \quad (S17)$$

The detritus decomposition rate depends on oxygen and temperature (Eq. S16). Eq. S17 parameterises the higher decomposition rate in the oxygen deficient part of the water column due to more active bacterial processes with respect to the surface aerobic layer. The decomposition processes continue even in an absence of oxygen due to bacterial activities employing electron acceptors other than oxygen [11].

#### S.4. Dissolved inorganic nitrogen and phosphorus

The changes in ammonium, nitrate and phosphate concentrations are expressed by

$$R(N_a) = \frac{1-np}{np} \{ \lambda \cdot \varepsilon_n \cdot D + \sum_Y \mu_{ZY} \cdot Z_Y \} - \sum_X [ \sigma_X \cdot f_X(N_a) \cdot f_X(I) \cdot f_X(T) \cdot P_X ] - r_n \cdot \frac{I_n}{I_n+I} \cdot N_a - r_a \cdot N_n \cdot N_a, \quad (S18)$$

$$R(N_n) = r_n \cdot \frac{I_n}{I_n+I} \cdot N_a - \sum_X [ \sigma_X \cdot f_X(N_n) \cdot f_X(I) \cdot f_X(T) \cdot P_X ] - \frac{3}{5} r_a \cdot N_n \cdot N_a - \frac{4}{53} r_s \cdot HS \cdot N_n, \quad (S19)$$

$$R(PO) = \{ \varepsilon_n \cdot D + \sum_Y \mu_{ZY} \cdot Z_Y - \sum_X [ \sigma_X \cdot f_X(PO) \cdot f_X(I) \cdot f_X(T) \cdot P_X ] \} / np \quad (S20)$$

In Eq. S18, the first and second terms represent ammonium sources due to decomposition of detritus (remineralisation) and zooplankton excretion, respectively. The source terms are multiplied by the expression of  $np$ , which represents the phytoplankton and zooplankton  $N:P$  stoichiometry. This is because the detritus represents both dissolved organic nitrogen and phosphorus, and zooplankton excretions contain both nitrogen and phosphorus, as well [12].  $1/np$  part of detritus remineralisation and zooplankton excretion is assumed to be source of PO and the rest is the source of  $N_a$ . The third term represents its uptake during the primary production and the last two terms are the ammonium oxidation by oxygen (nitrification) and by nitrate in the absence of oxygen. The light dependent nitrification rate is applied.

Nitrification is the only internal (reaction) source of nitrate (Eq. S19). According to Eq. S19 nitrate is consumed due to its uptake by the phytoplankton (the second term), anaerobic particulate matter remineralization (the third term) that applies at oxygen concentrations less than  $10 \text{ mmol O}_2 \text{ m}^{-3}$ , the oxidations of ammonium and hydrogen sulphide taking place at oxygen concentrations less than  $10 \text{ mmol O}_2 \text{ m}^{-3}$  following the reaction equations in [13].

The phosphate sources (Eq. S20) are the same as for the ammonium divided by the parameter  $np$ , which represents the phytoplankton and zooplankton  $N:P$  stoichiometry. The third term represents its uptake during the primary production

#### S.5. Dissolved oxygen and hydrogen sulphide

Dissolved oxygen concentration is altered by a balance between its photosynthetic production by the autotrophs and the consumption due to the pelagic decomposition of organic matter (Eq. S12). The excretion of zooplankton as well as the oxidation of ammonium within the oxygenated parts of the water column ( $DO > DO_l$ ) and oxidation of hydrogen sulphide near the anoxic interface is given by

$$R(DO) = 8.125[\sum_X \Psi_X \cdot P_X] - 6.625[\varepsilon_n \cdot D + \sum_Y \mu_{ZY} \cdot Z_Y] - 2 r_a \cdot N_a - \frac{1}{2} r_o \cdot HS \cdot DO \quad (S21)$$

The air-sea exchanges of surface dissolved oxygen concentration is given by the model of [14] (see also in [15]).

The reaction kinetics governing temporal changes of hydrogen sulphide concentration are given by

$$R(HS) = 0.5 \varepsilon_n \cdot D - r_o \cdot HS \cdot DO - r_s \cdot HS \cdot N_n, \quad (S22)$$

where the first term represents hydrogen sulphide production by the process of sulphate based anaerobic organic matter decomposition, the second and third terms express the oxidation reactions of H<sub>2</sub>S by nitrate and oxygen (Eqs. S19 and S21). The coefficients are given in Table S2.

**Table S1.** Food preference coefficients of the predator groups on the prey groups.

	$Z_S$	$Z_L$	$Z_N$	$Z_G$
$P_L$	--	1.0	0.5	--
$P_S$	1.0	--	0.5	--
$D$	--	0.5	1.0	--
$Z_S$	--	0.5	0.25	1.0
$Z_L$	--	--	--	0.5
$Z_N$	--	0.25	--	0.3

**Table S2.** BSEM input parameters.

Parameter	Value	Unit	Definition
$\alpha_L$	0.18	$W^{-1} m^2$	Initial slope of P-I curve for $P_L$
$\alpha_S$	0.12	$W^{-1} m^2$	Initial slope of P-I curve for $P_S$
$\beta_L$	0.001	$W^{-1} m^2$	Photo inhibition parameter for $P_L$
$\beta_S$	0.035	$W^{-1} m^2$	Photo inhibition parameter for $P_S$
$\gamma_{ZL}$	0.5	$d^{-1}$	Maximum grazing rate of $Z_L$
$\gamma_{ZS}$	0.8	$d^{-1}$	Maximum grazing rate of $Z_S$
$\gamma_{ZN}$	0.5	$d^{-1}$	Maximum grazing rate of $Z_N$
$\gamma_{ZG}$	0.15	$d^{-1}$	Maximum grazing rate of $Z_G$
$\varepsilon_n$	0.2	$d^{-1}$	Default remineralisation rate of $D$
$\lambda$	3.0	$(mmol N)^{-1}$	$N_a$ inhibition parameter

Parameter	Value	Unit	Definition
$\mu_{ZL}$	0.05	$d^{-1}$	Excretion rate of $Z_L$
$\mu_{ZS}$	0.06	$d^{-1}$	Excretion rate of $Z_S$
$\mu_{ZN}$	0.06	$d^{-1}$	Excretion rate of $Z_N$
$\mu_{ZG}$	0.08	$d^{-1}$	Excretion rate of $Z_G$
$\sigma_L$	2.5	$d^{-1}$	Maximum growth rate $P_L$
$\sigma_S$	2.0	$d^{-1}$	Maximum growth rate $P_S$
$\varphi$	0.7	-	Assimilation efficiency
$a$	0.5	$\text{mmol N m}^{-3}$	Half-saturation constant in Eq. S3
$b$	0.5	-	Half-saturation constant in Eq. S3
$d_o$	0.15	$^{\circ}\text{C}^{-1}$	Temperature factor of mineralisation
$DO_l$	10	$\text{mmol O}_2 \text{ m}^{-3}$	Minimum oxygen concentration Eqs. S6c and d
$K_{DO}$	150	$\text{mmol O}_2 \text{ m}^{-3}$	Oxygen half-saturation constant for $D$ remineralisation
$K_{ZL}$	0.5	$\text{mmol N m}^{-3}$	Half-saturation constant for grazing of $Z_L$
$K_{ZS}$	0.4	$\text{mmol N m}^{-3}$	Half-saturation constant for grazing of $Z_S$
$K_{ZN}$	0.4	$\text{mmol N m}^{-3}$	Half-saturation constant for grazing of $Z_N$
$K_{ZG}$	0.25	$\text{mmol N m}^{-3}$	Half-saturation constant for grazing of $Z_G$
$I_n$	10	$\text{W m}^{-2}$	Limiting value of light attenuation (Eq. S7a)
$m_{PL}$	0.04	$d^{-1}$	Mortality rate of $P_L$
$m_{PS}$	0.02	$d^{-1}$	Mortality rate of $P_S$
$m_{0ZL}$	0.25	$d^{-1}$	Default mortality rate of $Z_L$
$m_{ZS}$	0.1	$d^{-1}$	Mortality rate of $Z_S$
$m_{ZN}$	0.15	$d^{-1}$	Mortality rate of $Z_N$
$m_{ZG}$	0.04	$d^{-1}$	Mortality rate of $Z_G$
$np$	16	-	Redfield ratio
$Q_{10}$	2.0	-	Factor of temperature control
$r_a$	0.01	$d^{-1}$	Default oxidation rate of $N_a$ by $N_n$
$r_n$	0.1	$d^{-1}$	Default nitrification rate
$r_o$	0.01	$d^{-1}$	Default oxidation rate of $HS$ by $DO$
$r_s$	0.01	$d^{-1}$	Default oxidation rate of $HS$ by $N_n$
$w_D$	5.0	$\text{m d}^{-1}$	Detritus sedimentation rate
$w_{PL}$	0.5	$\text{m d}^{-1}$	$P_L$ sedimentation rate

## References

1. GETM, 3D General Estuarine Transport Model Available online: <http://www.getm.eu/> (accessed on 1 October 2020).
2. Oguz, T.; Merico, A. Factors controlling the summer *Emiliana huxleyi* bloom in the Black Sea: A modeling study. *J. Mar. Syst.* **2006**, *59*, 173–188, doi:10.1016/j.jmarsys.2005.08.002.
3. Tsiaras, K.P.; Kourafalou, V.H.; Davidov, A.; Staneva, J. A three-dimensional coupled model of the Western Black Sea plankton dynamics: Seasonal variability and comparison to Sea WiFS data. *J. Geophys. Res. Ocean.* **2008**, *113*, doi:10.1029/2006JC003959.
4. Wroblewski, J. A model of phytoplankton plume formation during variable Oregon upwelling. *J. Mar. Res.* **1977**, *35*, 357–394.
5. Jassby, A.D.; Platt, T. Mathematical formulation of the relationship between photosynthesis and light for phytoplankton. *Limnol. Oceanogr.* **1976**, *21*, 540–547, doi:10.4319/lo.1976.21.4.0540.
6. Ludwig, W.; Dumont, E.; Meybeck, M.; Heussner, S. River discharges of water and nutrients to the Mediterranean and Black Sea: Major drivers for ecosystem changes during past and future decades? *Prog. Oceanogr.* **2009**, *80*, 199–217, doi:10.1016/j.pocean.2009.02.001.
7. Gentleman, W.; Leising, A.; Frost, B.; Strom, S.; Murray, J. Functional responses for zooplankton

- feeding on multiple resources: A review of assumptions and biological dynamics. *Deep. Res. Part II Top. Stud. Oceanogr.* **2003**, *50*, 2847–2875, doi:10.1016/j.dsr2.2003.07.001.
8. Kopuz, U.; Feyzioglu, A.M.; Valente, A. An Unusual Red-Tide Event of *Noctiluca Scintillans* (Macartney) in the Southeastern Black Sea. *Turkish J. Fish. Aquat. Sci.* **2014**, *14*, 261–268, doi:10.4194/1303-2712-v14\_1\_28.
  9. A., S.T.; Alekseenko, E.; Lidia, M.; Paul, N. Modelling assessment of interactions in the Black Sea of the invasive ctenophores *Mnemiopsis leidyi* and *Beroe ovata*. *Ecol. Modell.* **2018**, *376*, 1–14, doi:10.1016/j.ecolmodel.2018.02.008.
  10. Druon, J.N.; Mannino, A.; Signorini, S.; McClain, C.; Friedrichs, M.; Wilkin, J.; Fennel, K. Modeling the dynamics and export of dissolved organic matter in the Northeastern U.S. continental shelf. *Estuar. Coast. Shelf Sci.* **2010**, *88*, 488–507, doi:10.1016/j.ecss.2010.05.010.
  11. Fenchel, T.; Blackburn, H.; King, G. *Bacterial biogeo-chemistry: the ecophysiology of mineral cycling*; 3rd Ed.; Academic Press: San Diego, 2012; ISBN 9780124159747.
  12. Hargrave, B.T.; Geen, G.H. Phosphorus excretion by zooplankton. *Limnol. Oceanogr.* **1968**, *13*, 332–342, doi:10.4319/lo.1968.13.2.0332.
  13. Oguz, T. Modeling aggregate dynamics of transparent exopolymer particles (TEP) and their interactions with a pelagic food web. *Mar. Ecol. Prog. Ser.* **2017**, *582*, 15–31, doi:10.3354/meps12330.
  14. Liss, P.S.; Merlivat, L. Air-Sea Gas Exchange Rates: Introduction and Synthesis. In *The Role of Air-Sea Exchange in Geochemical Cycling*; Springer Netherlands, 1986; pp. 113–127.
  15. Miladinova, S.; Stips, A. Sensitivity of oxygen dynamics in the water column of the Baltic Sea to external forcing. *Ocean Sci.* **2010**, *6*, 461–474, doi:10.5194/os-6-461-2010.