

Remiern



Physiological Aspects of Milk Somatic Cell Count in Small Ruminants—A Review

Shehadeh Kaskous ¹,*¹, Sabine Farschtschi ², and Michael W. Pfaffl ²

- ¹ Department of Research and Development, Siliconform, Schelmengriesstrasse 1, 86842 Türkheim, Germany
- ² Department of Animal Physiology and Immunology, TUM School of Life Sciences, Technical University of
- Munich, Weihenstephan, Weihenstephaner Berg 3, 85354 Freising-Weihenstephan, Germany
- Correspondence: skaskous@siliconform.com

Abstract: The aim of this review was to focus on the physiological aspects of milk somatic cell count (SCC) in small ruminants (SM). The SCC is an important component naturally present in milk and is generally used as an indicator of milk quality and udder health in milk producing ruminants. SCC contains the following cells: polymorphonuclear neutrophils (PMN), macrophages, lymphocytes, and many milk epithelial (MEC) cells, cell fragments, and cytoplasmic particles/vesicles. PMN (40-80%) represent the major cell type in milk in healthy uninfected goats, whereas the macrophages (45–88%) are the major cell type in sheep's milk. However, dairy goats and sheep have an apocrine secretory system that produces cytoplasmic cellular particles/vesicles and large numbers of cell fragments, resulting in the physiological SCC limit being exceeded. It is obvious that the SCC level in milk of SM can be affected by various influencing factors, such as milk fraction, breed, stage of lactation, parity, type of birth, milking system, and others. An increase in the SCC above the physiological level not only indicates an udder or general health problem but reduces milk production, changes the milk composition, and hence affects milk processing. Moreover, the milking machine plays an important role in maintaining udder health in SM and stable SCC at physiological levels in the milk obtained. So far, there are no healthy or pathological physiological SCC levels defined in SM milk. Furthermore, a differential cell count (DCC) or even a high resolution DCC (HRDCC), which were recently developed for cattle milk, could also help in SM to gain deeper insight into the immunology of the mammary gland and find biomarkers to assess udder health. In conclusion, SCC is an indication of udder health or exposure of the udder to infectious agents or mechanical stress and should therefore always be considered a warning sign.

Keywords: goat; sheep; somatic cell count; mastitis; milk quality; physiology

1. Introduction

Dairy goat and sheep farming are important economically in many countries in the world. Their milk production is a valuable raw material for many milk products. The quality of these products is dependent upon the quality of raw milk used [1]. The somatic cell count (SCC) in SM is an important indicator of udder health and the raw milk quality and also reflects the animal's general health status [2–5], but the predictive values are better in ewes than in goats [6]. However, SCC plays a major role in protecting the milk epithelial cells (MEC) in the udder in general, which appears in the udder in response to invading pathogens. Polymorphonuclear neutrophils (PMNs), especially, invade in a higher number in the alveolar lumen. In addition, the SCCs in the SM are comprised of macrophages, lymphocytes, many cell fragments, and cytoplasmic particles/vesicles. PMNs were the major cell type in milk from healthy uninfected goats and constitute 40 to 80% of the SCC [7–12], whereas the macrophages (45–88%) are the major cell type in sheep's milk, with fewer PMNs (10–35%), lymphocytes (10–17%), and epithelial cells (2–3%) [7,10,13]. Mastitis (subclinical or clinical) increases the PMN content in sheep's milk but not in goat's



Citation: Kaskous, S.; Farschtschi, S.; Pfaffl, M.W. Physiological Aspects of Milk Somatic Cell Count in Small Ruminants—A Review. *Dairy* **2023**, *4*, 26–42. https://doi.org/10.3390/ dairy4010002

Academic Editors: Lotte Bach Larsen and Erminio Trevisi

Received: 23 October 2022 Revised: 8 December 2022 Accepted: 23 December 2022 Published: 30 December 2022



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). milk, in which they still represent the dominant type [9]. Thus, PMNs in goat's milk are the main cellular component in both healthy and in infected glands [14]. It is noteworthy that the function of PMNs in healthy mammary glands does not have the same effect as in mammary gland with mastitis [15]. The histopathological examination of goat udders with high SCCs demonstrated the absence of mastitis [16]. However, a high SCC value is normal in a healthy goat udder [8,17,18]. Various studies have reported that dairy goats and sheep have a different secretory system than other lactating animals. The apocrine system of goats and sheep produces cytoplasmic particles/vesicles (containing DNA) [10,19], and their milk can contain large numbers of cell fragments, resulting in exceeding the limit of SCCs, especially in the late stages of lactation [20]. The cytoplasmic particles/vesicles and cell fragments are counted "false positive" as somatic cells when samples are tested using automated counting methods. As a result, goat's and sheep's milk SCCs are higher than those found in other lactating animals. However, the concentrations of cytoplasmic particles/vesicles in goat milk are much higher than in ewe's milk [10]. The concentration of cytoplasmic particles/vesicles in goat's milk is about 150×10^3 cells/mL [20], whereas sheep's milk has about 15×10^3 cell/mL [7,21]. However, only 10% of SCCs are cytoplasmic particles/vesicles and cell fragments that are normally secreted with goat's milk [7]; the remaining 90% of the somatic cells are blood cells, which contribute to the immune defence of the mammary gland. In any case, good hygiene on the SM farms is a key to a low SCC in the milk produced and minimizes risks to guarantee an optimal consumption quality [3]. Furthermore, low levels of SCC in the milk are desirable to ensure good extraction of protein from raw milk, whereas high levels of SCC depress casein and other component levels in milk [3,22]. It has been shown that goats having high hemoglobin concentrations were characterized by a high milk yield and a lower SCC. In contrast, low hemoglobin concentrations (less than 5.6 mmol) resulted in higher cell counts [20]. Two main factors influence the SCC levels in sheep's and goat's milk—namely, physiological and environmental factors.

In the following, the physiological aspects of milk SCC in SM are summarized and discussed and could help to define the healthy and pathological physiological SCC levels in SM milk. Furthermore, it could help to gain deeper insight into the immunology of the mammary gland and find biomarkers to assess udder health in SM.

2. Physiological Level of SCC in Sheep Milk

So far, there is no clearly defined threshold value for physiological milk SCC in sheep that represents a healthy udder. However, the physiological level of SCC in ewe's milk is still under discussion. Several lines of evidence suggest that the physiological SCC levels in ewe's milk need to be limited in relation to mastitis [23]. Table 1 shows the physiological level of milk SCC in sheep's milk, and the mean value from all publications is 375×10^3 cells/mL. Previous studies have reported that the physiological level of milk SCC in dairy ewes ranged between 10×10^3 and 100×10^3 [24–27]. In a review by Tancin et al. [28], it was observed that SCC occurs at around 150×10^3 cells/mL in bulk milk tanks without mastitis in different breeds of sheep. Other researchers have pointed out that the upper threshold for milk SCC in the udder of a healthy ewe should be 250×10^3 cells/mL [29–31]. It was found in Manchega sheep that SCC 300×10^3 cells/mL was considered as the ideal value for the diagnosis of subclinical mastitis [32]. Zafalon et al. [33] found that the value of SCC above 400×10^3 cells/mL applies to the diagnosis of subclinical mastitis in herds. A similar cutoff value for the Santa Ines and Texel breeds in the lactation season 400×10^3 cells/mL in milk was reported by Kern et al. [34]. On the other hand, some studies clearly showed that the limit values for milk SCC in healthy udders are higher than previously announced and the limit value in different sheep breeds was 500×10^3 cells/mL [35–38]. Tvarožková et al. [39] also found that an SCC of more than 500×10^3 cells/mL of milk could be important for the detection of subclinical mastitis at half udder level in dairy sheep. Similar results reported that the threshold of SCC > 500×10^3 cells/mL was determined as an indicator of a change in milk

quality in ewes [40]. Likewise, it was clearly demonstrated that the negative milk samples from East Friesian ewes showed low somatic cell counts ($734 \pm 3153 \times 10^3$ cells/mL; $5.15 \pm 0.55 \log \text{ cells/mL}$) compared to the positive milk samples with higher somatic cell counts ($4432 \pm 6069 \times 10^3 \text{ cells/mL}$; $5.97 \pm 0.96 \log \text{ cells/mL}$) [41]. In this study, it was shown that approximately 84% of the samples that tested negative had less than $500 \times 10^3 \text{ cells/mL}$, and only 5% had more than $1000 \times 10^3 \text{ cells/mL}$. Conversely, 32% of the positive samples had less than $500 \times 10^3 \text{ cells/mL}$. Furthermore, Tancin et al. [28] reported that an SCC limit of $1000 \times 10^3 \text{ cells/mL}$ was established between healthy and mastitis in sheep. In any case, raw ewe's milk without pathogen bacteria had the lowest average SCC [42].

		Commence (MC ¹¹	600 L 1	
Location	Breed	Source of Milk Sample	SCC Level (×10 ³ Cells/mL)	Authors
Spain	Menchega	Animal	250	[26]
Syria	Awassi	Right-udder half Left-udder half	162 199	[43]
Slovenia	Domestic highland, East Friesland a. Awassi	Udder half	250	[30]
Bulgaria	East Friesian \times 1/4 East Friesian \times Awassi	Animal	1000	[44]
Germany	East Friesian	Animal	174	[41]
Germany	East Friesian black and white	Udder half	61	[45]
Spain	Manchega sheep	*	300	[32]
Brazil	Corriedale and Texel	Animal	317	[46]
Italy	Sarda Sarda × Lacaune	Farm tank Farm tank	206 171	[47]
France	Lacaune Red-face Manech	Farm tank Farm tank	500 1050	[48]
Kosovo	Bar, Sha; Kos, Bal	Animal	500	[49]
Slovak Republic	Different races a. crossbred	Animal	593	[50]
Brazil	Lacaune	Animal	1600	[51]
Poland	Polish a. Polish Lowland	Udder half	200	[42]
Slovak Republic	Lacaune	Farm tank	146	[28]
Spain	Different races	Farm milk	90	[3]
Kashmir	Local breeds	Animal	241	[12]
Slovak Republic	Slovak dairy sheep a. Lacaune	Animal	400	[52]
Iraq	Local ewes	Animal	39	[53]
Italy	Sarda	Half-udder	235	[54]
Slovak Republic	Lacaune	Half udder	200	[55]
Greece	Different races a. crossbred	Farm tank	501	[56]
Mean			375	

 Table 1. Physiological levels of SCC in sheep's milk relative to breeds and geography.

3. Physiological Level of SCC in Goat's Milk

SCC in goat's milk is widely used for evaluating milk quality and as an indicator of udder health [5,57–59]. However, the SCC in goat's milk is generally higher than in sheep's and cow's milk [17], and SCC in milk as an indicator of IMI (Intramammary Infection) may be lower in goats than in sheep and cows since many physiological factors can increase SCC in the uninfected halves [11,17,60]. The average physiological level of SCC in goat milk was 764 \times 10³ cells/mL and fluctuated between 200 \times 10³ and 1500×10^3 cells/mL (Table 2). According to several authors, it is clear that higher SCC in goat's milk is bound with a high proportion of cell fragments. This means that the milk secretion in the goat, in contrast to the cow, leads to greater cell losses of the secretory glandular cells [8,19]. However, it has been confirmed that SCC is a relevant predictor of IMI and hygienic milk quality in goats [5], and SCC measured with a Delaval cell counter was strongly associated with bacterial growth in the half udder milk samples [61]. Rupp et al. [5] reported that goats with repeatedly healthy udders (80% of negative milk samples) had the lowest SCC values with an average of 277×10^3 cells/mL. According to the observations of Csanadi et al. [62], the average milk SCC of five different breeds of goat in Hungary was 664×10^3 cells/mL. Furthermore, milk SCC for goats free of IMI range between 270 and 2000×10^3 cells/mL [7]. The mean SCC for uninfected and infected halves with coagulated negative staphylococci were 272×10^3 and 932×10^3 cells/mL, respectively [22]. Contreras et al. [63] reported that a value of 500×10^3 cells/mL was a useful threshold to distinguish between infected and uninfected halves of the udder. The obtained results by Zeng and Escobar [16] demonstrated that 56% of the milking goats produced milk with 1000×10^3 SCC cells/mL; they concluded that healthy milk from healthy goats showing no signs of mastitis can contain up to 1000×10^3 SCC/mL. Values of 370×10^3 cells/mL milk [14] up to 1000×10^3 cells/mL milk are also given for the healthy goat udder [64]. A review by Koop et al. [65] suggested a cutoff value for SCC in goat's milk of 1500×10^3 cells/mL. According to Min et al. [66], mean SCC values in infected dairy goats ranged between 2000 and 4000×10^3 cells/mL, and they concluded that SCC in goat's milk does not strongly correlate with intramammary infection.

Location	Breed	Source of Milk Sample	SCC Level \times 10 ³ Cells/mL *	Author
Czech Republic	White shorthair	Animal	600	[18]
Czech Republic	White shorthair	Animal	1422	[67]
Greece	Different races a. crossbred	Farm tank	770	[56]
Italy	Alpine a. Saanen	Animal	303	[54]
Kashmir	Local races	Animal	608	[12]
Kosovo	Alpine a. native Red	Animal	1000	[68]
Spain	Different races	Farm tank	660	[3]
Spain	Local breed	Farm tank	1200	[7]
Sweden	Swedish landrace	Animal	481	[61]
Turkey	Saanen	Animal	206	[69]
USA	**	Farm tank	1150 ***	[70]
Mean			764	

Table 2. Physiological levels of SCC in goat's milk relative to breeds and geography.

SCC *: This is geometric SCC; ** not specified; ***: by the fifth parity.

4. Considerations Relative to SCC Levels and Limits in Sheep's and Goat's Milk for Dairy

The legal SCC limit for bulk tank milk in the United States for sheep and goat is 750×10^3 cells/mL and 1000×10^3 cells/mL, respectively [7,62]. It is noteworthy that until now there was no milk SCC legal limit for sheep and goats in the European Union [62,67]. Corrales et al. [71] reported that the limit of 1500×10^3 cells/mL should be acceptable for goat's milk in the European Union. Currently, in the European Community the EU has yet to regulate values of SCC level in ewe's and goat's milk [3]. According to bulk tank milk, the SCC level can be established in both species as follows: 1. Good or acceptable ewe's or goat's milk when bulk tank milk SCC < 750×10^3 cells/mL; 2. Intermediate ewe's or goat's milk when bulk tank milk SCC is between 750 and 1500×10^3 cells/mL; and 3. Bad ewe's or goat's milk when bulk tank milk SCC > 1500×10^3 cells/mL [72,73].

5. Influence of Physiological Factors on SCC in Small Ruminants

It was observed that SCC level in SM can be affected by physiological factors such as milk fraction, breed, stage of lactation, parity, and type of birth [10,62,74–78]. An interesting aspect is that 48% of SCC variance can be attributed to physiological factors [79,80].

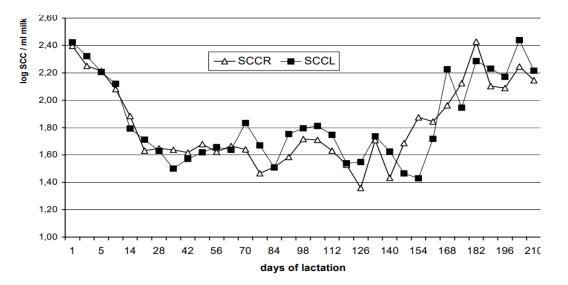
5.1. Effect of Milking Fraction

Usually, the first squirts before milking are the fraction used for bacteriological diagnosis and SCC determination. Several studies have shown that the first milk squirts have a similar SCC as in the main milking fraction. However, the first milk squirts have slightly lower SCC compared to main milking fraction [81], and the average values of SCC were 687×10^3 and 763×10^3 cells/mL, respectively [82]. A review by Martinez [83] found similar results, and the mean values of SCC were 998×10^3 and 1139×10^3 cells/mL in the first milk squirts and the main milking fraction, respectively. Kaskous [41] showed the opposite trend in East Friesian sheep in Germany, and the average SCC was $202.09 \times 10^3 \pm 471.02 \times 10^3$ cells/mL ($5.83 \pm 0.82 \log cells/mL$) from the first milk squirts, whereas the average SCC reached to $174.27 \times 10^3 \pm 247.27 \times 10^3$ cells/mL ($5.44 \pm 0.62 \log cells/mL$) from the total milk produced of each animal. In addition, research by Skapetas et al. [84] showed that SCC was lower in whole machine milk than in hand-stripped milk from Chios ewes.

5.2. Effect of Lactation Stage

Corrales et al. [85] found that SCC was very high at the end of lactation, and it is impossible to distinguish between infected and healthy mammary glands by SCC. It has been thought that the increase in SCC in the milk of SM with advanced lactation is due to a dilution effect [86–88]. This means that the SCC in milk is higher at the end of lactation due to the small amount of milk [27,89]. Paape and Capuco [19] emphasized this statement, and the SCC cannot increase in the late stage of lactation if the milk yield remains high until the end of lactation. Thus, the increase in SCC was associated with the progression of lactation in goats with or without a diagnosis of intramammary infection [86]. Studies by Kaskous [43] showed that normal SCC was higher in early and late stages of lactation (Figure 1).

Figure 1 shows that the available SCC values between 15 and 150 days of lactation were lower than 100×10^3 cells/mL. In other similar studies, SCC was lowest, averaging 200×10^3 cells/mL, at 15 days of lactation and reached a maximum of around 500×10^3 at 285 days [67]. Zafalon et al. [90] clearly show that the milk SCC values in sheep were higher at the end of lactation than in the second week of lactation, although the positive microorganisms in milk samples from half udders were higher in the second week of lactation (117/763, 15.3%) than at the end of lactation (86/694, 12.4%) (Table 3). One explanation for these contradictions is that the sheep produce less milk at the end of lactation, which increases the SCC value in the milk. This low milk yield at the end of lactation influences the lower incidence of subclinical mastitis. This is due to an increase



in macrophages and polymorphonuclear leukocytes in this phase, which are involved in udder defense during the dry period.

Figure 1. SCC level during lactation in Syrian Awassi ewes. SCCR: Somatic cell count in the right half of the udder; SCCL: Somatic cell count in the left half of the udder; log SCC: logarithmic somatic cell count. "Reprinted/adapted with permission from Ref. [43]. Copyright year 2021, copyright owner's name Kaskous". More details on "Copyright and Licensing" are available via the following link: https://www.mdpi.com/ethics#10.

Table 3. Negative and positive results of microorganism tests in samples of milk from mammary halves of different sheep breeds during the second week and at the end of lactation. "Reprinted/adapted with permission from Ref. [90] with some changes. Copyright year 2022, copyright owner's name: Kaskous". More details on "Copyright and Licensing" are available via the following link: https://www.mdpi.com/ethics#10.

	Start of Lactation			End of Lactation			
Breeds	Number of Samples	Negative (% ^a)	Positive (% ^b)	Number of Samples	Negative (% ^a)	Positive (% ^b)	
Santa Ines	402	76.6	23.4	343	81	19	
Texel	120	94.2	5.8	116	95.7	4.3	
IIe de France	120	90	10	104	91.3	8.7	
Dorper	121	96.7	3.3	131	94.7	5.3	
Total	763	84.7	15.3	694	87.6	12.4	

^a: Percentage negative of pathogens; ^b: Percentage positive of pathogens.

On the other hand, goat milk SCC is subject to greater physiological variability than cow milk SCC [91], and this variability is mainly related to lactation stage and goat breed [92]. Wilson et al. [86] have confirmed that more than 90% of SCC variations in goat milk may not be due to bacterial udder infections but to physiological factors such as lactation stage and month of the year or oestrus [93].

5.3. Effect of Parity

The influence of parity on the SCC seems to depend on the state of health of the udder and the pathogens involved [81]. A review by Paape et al. [67] observed an increase in SCC from the first to the fifth lactation. In general, in SM, increased SCC in milk was associated with higher parity [11,19]. The explanation for the fact that the content of somatic cells and especially PMN in the milk increases with the number of lactations is not an age-related change but increased infection rates in older animals [94].

5.4. Effect of Type of Birth

Several reports have shown that type of birth (single, twin, or more birth) influences the SCC in the milk in SM [58,95]. However, the highest SCC values were achieved in animals with multiple births rather than in animals with a single birth [95]. On the contrary, some studies have shown that multiple births have no effect on SCC values [96]. In sheep and goats, some of them are known to have twins or more at birth. Such animals can produce more milk but also higher SCC, since udder health deteriorated in those sheep and goats that suckled two lambs instead of one. Threfore, suckling influenced later udder health during the milking period.

5.5. Effect of Milk Yield

The results from goats indicated that less productive animals without infection lead to higher SCC [83,86,96]. The converse is also correct. This means that goats producing >3 kg milk/day had the lowest SCC (<954 × 10^3 cells/mL) in the milk controls [95]. The results of several experiments support the hypothesis that the reduced milk production in goats and sheep is most likely due to the competent alveolar cells of the mammary gland. Therefore, the SCC in the milk increased significantly. This means that the alveoli have an impaired ability to secrete.

5.6. Effect of Breed

A review by Zafalon et al. [90] observed that the breed of sheep plays a very important role in SCC levels. The established limits of the SCC values for the diagnosis of subclinical mastitis were shown in different sheep breeds during the beginning and the end of lactation (Table 4). However, the SCC values in milk at the end of lactation were higher than those at the beginning of lactation.

Table 4. Cutoff values of SCC in ewe's milk of different breeds in Brazil."Reprinted/adapted with permission from Ref. [90]. Copyright year 2022, copyright owner's name Kaskous". More details on "Copyright and Licensing" are available via the following link: https://www.mdpi.com/ethics#10.

Breeds	Second Week of Lactation (SCC $ imes$ 10 ³ Cells/mL)	End of Lactation (SCC \times 10 ³ Cells/mL)
Santa Ines	487	1171
Texel	419	802
Ile de France	781	554
Dorper	1062	1276

On the other hand, it was found that the SCC values in different dairy goat breeds (Alpine and Nubian) cannot confirm any significant differences [88], and the racial differences are due to differences in health, production levels, and management characteristics between them [97]. Similar results were reported by Csanadi et al. [62] in Hungary, and the SCCs of milk samples do not differ significantly depending on the genotype (Native, Saanen and Alpine × Saanen cross-bred). Based on the observations of Tancin et al. [28], the Lacaune breed of sheep was shown to have a higher percentage of mastitis milk (also higher SCC) compared to other breeds or crossbreeds (purebred Tsigai ewe, crossbred Slovak dairy ewe and crossbred Valachian \times Lacanune ewes).

Finally, based on the observations of Paape et al. [7], it has been shown that physiological factors such as parity, stage of lactation, and milk yield did not have any clear influence on the SCC in milk, whereas intramammary infection has significantly increased the milk SCC in goats and sheep.

5.7. Effect of Stress

A new study in goats has clearly shown that the cumulative effect of various challenges imposed resulted in a change in hormone release and an increase in SCC in the milk produced as well as a reduction in milk yield [98]. In fact, stress in goats and sheep has been reported to increase SCC in the milk. Heat, vaccinations, dietary changes, and changes in milking routine are factors that lead to physiological stress. Since goats and sheep are very sensitive, the resulting decreases in milk yield under stress could explain the increase in SCC [98].

6. Increase of Somatic Cell Counts over Healthy Level in Small Ruminant

The main cause of increased SCC in milk of dairy ruminants is IMI [10,80,93,99–101]. Persson et al. [23] found a significantly higher SCC for udder halves with intramammary infection compared to udder halves without bacterial findings. As a result, high SCC has been suggested as the main reason for culling dairy sheep. But not every increase in the SCC in milk indicated an infection of the udder in SM [102]. Rupp et al. [5] reported that in lactating goats about 50% of the milk samples had a higher number of SCC (1542×10^3 cells/mL) and no udder pathogens were detected, whereas positive udder pathogens with low SCC (855×10^3 cells/mL) were detected in about 32.7% of the milk samples (Table 5). Tvarožková et al. [39] have shown clear results that SCC $\geq 500 \times 10^3$ cells/mL were detected in 92.5% bacteriologically positive milk samples. This means that the presence of pathogens increased the SCC in the milk significantly (p < 0.001) compared to samples that were free of pathogens. This resembles the results of Olechnowicz and Jaskowski [22], which showed that total bacterial count is significantly correlated with the number of somatic cells in bulk milk. Likewise, results from 155 French herds showed that annual geometric means of 750×10^3 , 1000×10^{3} , and 1500×10^3 SCC/mL corresponded to $30 \pm 12\%$, $39 \pm 8\%$, and $51 \pm 8\%$ of infected goat udders, respectively [103,104].

Table 5. Milk bacteriological results for the high-and low SCC in milk samples from healthy goats or goats with nonacute mastitis, with some changes. "Reprinted/adapted with permission from Ref. [5] with some changes. Copyright year 2022, copyright owner's name Kaskous". More details on "Copyright and Licensing" are available via the following link: https://www.mdpi.com/ethics#10.

Item	Total ^a		$\begin{array}{c} \text{Low-SCC} \\ 855 \times 10^3 \end{array}$		$\begin{array}{l} \text{High-SCC} \\ \textbf{1542}\times \textbf{10}^{\textbf{3}} \end{array}$	
	Ν	%	Ν	%	Ν	%
Negative milk samples	1547	57.55	757	67.3	790	50.5
Positive milk samples	1141	42.45	367	32.7	774	49.5
-Staphylococci (total)	922	34.3	281	25	641	41
-Streptococci	75	2.8	27	2.4	48	3.1
-Bacillus	102	3.8	47	4.2	55	3.5
-Micrococci	22	0.8	8	0.7	14	0.9
-Others ^b	20	0.7	4	0.4	16	1.0

^a Results were from 2688 milk samples from 9 sampling time points per udder half during first lactation. ^b Others: Acinetobacter, Aerococcus, Aspergillus, Corynebacterium, Escherichia coli, Pseudomonas and yeast.

7. Influence of the Milking Procedures on the SCC Level in SM

Proper milking procedures are essential to minimizing the risk of a bacterial infection causing mastitis and to reducing the SCC in the milk produced [105]. Therefore, milk SCC level is influenced by machine milking [105–108]. The prevention of udder diseases is mainly based on milking machine management, hygiene, annual milking machine checks, and optimization of milking technology [6]. It is essential that the milking system meet the physiological requirements of sheep and goats in order to increase milk yield and achieve better milk quality. Thus, the udder remains healthy [105,109]. Besides the liner, there

are three operating parameters that regulate mechanical milking: vacuum level, pulsation rate, and pulsation ratios. The milking system needs to provide a stable vacuum, adequate pulsation, and gentle milking action. High vacuum levels (>42 kPa) are often used during machine milking in dairy ewes and goats [110]. However, the use of 44 kPa in the goat milking machine showed an increase in the tissue thickness of the teats over 5%, and the average conductivity of the milk tended to increase [109]. Consequently, higher SCC can be found in the produced milk in SM. Several reports have shown that the vacuum required to open the teat canals in the sheep and goats is between 25 and 35 kPa [111,112]. On the other hand, sheep and goats store the milk in their gland cistern more than in the milk-producing alveoli's udder [113]. However, cisternal milk fraction of SM can range from 50% in sheep [114] to 85% in goats [115]. This, along with the smaller teat size, makes a faster pulsation an acceptable option. In recent years many milking machines for sheep and goats have been developed. However, many farmers are suffering from the performance of these milking machines, as they do not empty the udder completely and many udders will become diseased as well as the SCC increased in the milk produced [84,116,117]. The cause of this problem is the absence of proper milking machines that adapt to the physiology of sheep's and goats' udders. For this reason, Siliconform from Germany has developed a new milking machine for sheep and goats, which includes teat cups with an integrated air inlet (Figure 2).



Figure 2. Goats were milked with the new milking machines from Siliconform-Germany (pictures from Kaskous, 2020). Permission has been obtained and there is no copyright issue.

For SM, it is important during machine milking to monitor mammary gland health through diagnostic testing and teat dipping after milking is complete [2,6]. However, improving hygienic conditions during milking is a key to assessing the microbiological quality of the milk [22]. It is noteworthy that the correlation coefficients between SCC and morphological traits of the udder suggest that some of the morphological traits need to be taken into consideration in the evaluation of the suitability of SM for machine milking [118–120]. It has been estimated that ewes with a more horizontal teat position and larger teats had higher SCC, since they are more prone to develop subclinical mastitis [121]. Due to the above observation, it seems necessary to show the course and consequences of machine milking in SM on milk quality and health of the teat end [22]. In addition, the type of milking had a significant impact on bulk tank SCC in ewes [22].

Bulk tank SCC is also affected by the frequency of milking in dairy goats [122–124]. To improve the udder health status of SM, it is necessary to ensure hygienic conditions of animal maintenance and optimization of milking machine standards and parlour systems.

8. Impact of High Somatic Cell Counts on Milk Quality and Processing

The determination of SCC in sheep's and goat's milk is very important for milk processors, because SCC with other factors plays a big role in determining safety and hygienic quality of the final product [3,62]. A review by Podhorecka et al. [18] observed that low SCC values may significantly affect the technological properties of goat's milk, and SCC should therefore be routinely screened by dairy manufacturers to assure the consumer of high end-product quality. The study by Giaccone et al. [125] clearly showed that an

influence of a higher milk SCC of the Valle del Belice sheep breed influences the properties of cheese production. The same authors found that the fat, protein, casein, and lactose contents in the milk of the higher cell number group (>1000 × 10^3 SCC/mL) were lower than that of the lower cell number group (<1000 × 10^3 SCC/mL) (Table 6).

Table 6. Effect of somatic cell count on milk quality in sheep, "Reprinted/adapted with permission from Ref. [125] with some changes. Copyright year 2022, copyright owner's name Kaskous". More details on "Copyright and Licensing" are available via the following link: https://www.mdpi.com/ethics#10.

Parameters	High Level of SCC (>1000 \times 10 ³)	Low Level of SCC (<1000 \times 10 ³)
Somatic cell count (log ₁₀)	6.40 ^A	5.56 ^B
Fat (%)	6.29 ^A	6.92 ^B
Protein (%)	5.27	5.32
Casein (%)	4.25	4.39
Lactose (%)	4.38 ^a	4.71 ^b
Whey protein (%)	1.08	1.03
Urea (mg/dl)	31.69 ^A	33.07 ^B
pH	6.79 ^A	6.68 ^B
Calcium (g/l)	1.89	1.93
Phosphorus (g/l)	1.42	1.35

Within row, different letters are significant at p < 0.05 (small letters) and at p < 0.01 (capital letters).

Furthermore, the casein fractions (α S1-casein and β -casein) and fat content in fresh feta cheese were significantly lower when the bulk tank of SCC was high, whereas pH and fatty acid content were increased [22]. It is known that an increase of SCC causes a decrease in milk yield and affects milk composition [16,28,126–128], which leads to reduced cheese-making potential [129]. Moreover, it has been estimated that milk with three levels of SCC content in sheep showed differences (p < 0.01) in whey protein, lactose, pH, and total Na content. Furthermore, protein recovery rate was higher in cheese from low SCC milk, whereas adjusted cheese yield did not show significant differences [130]. Consequently, Csanadi et al. [62] noted that goat's milk having very high SCC is not suitable for making any goat milk products.

On the contrary, Chen et al. [131] found that milk composition did not change when milk SCC varied from 214×10^3 to 1450×10^3 cells/mL in Alpine goats without evidence of clinical mastitis, and no significant differences in the yield of semisoft goat cheeses were detected. However, total sensory scores and body and texture scores (hardness, springiness) for cheeses made from the high SCC milk were lower than those for cheeses made from the low and medium SCC milk. Moreover, it is also noted that individual and total free fatty acid (FFA) significantly increased ripening, regardless of the SCC levels.

9. Potential Use of Differential Cell Count in Milk of Small Ruminants

Since milk leukocytes comprise different cell types, each with specific roles in the immune defense of the mammary gland [132], it could be valuable to perform a differential cell count (DCC), which has already proven beneficial in dairy cows [133]. Today, there are even automated cytometers available to determine the main populations of immune cells in cow's milk [134,135]. Moreover, using fluorochrome-conjugated antibody to capture a particular cluster of differentiation (CD) molecule on the cell's surface, it is possible to further differentiate subpopulations of immune cells based on their specific wavelength [136]. Winnicka et al. [137], for example, used this method to analyze blood and milk leukocytes of healthy goats to document the percentages of different lymphocyte populations over

the course of lactation. The authors showed that the percentage of T helper cells (CD4+) increased during early lactation until day 14 and decreased in mid-lactation, whereas the amount of cytotoxic T cells (CD8+) increased until day 21 and then remained stable. For dairy cows, the ratio of CD4+ to CD8+ was proposed as a biomarker of low mastitis resistance, with a threshold below 1 [138]. Tatarczuch et al. [139] examined secretions of involuting udders of sheep and found CD8+ T cells to be the predominant lymphocyte population during early involution. In another flow cytometric study, the authors additionally determined the viability of several subpopulations in caprine milk [140]. Blagitz et al. [141] observed a reduced viability of PMN in goat milk samples with a low SCC and suggested a higher susceptibility to intramammary infections in such cases. For dairy cows, several biomarkers have already been introduced to diagnose mastitis (e.g., the logarithmic ratio of polymorphonuclear neutrophilic leukocytes to lymphocytes [142] or the expression of the inflammatory marker β -integrin CD11b on milk leukocytes [143]). In a different approach, Farschtschi et al. [144] used three different vaccines as systemic immune stimuli and could demonstrate an influence on both blood and milk DCCs. Further research in DCC could help to gain a deeper insight into the immunology of the mammary gland of SM and find biomarkers to assess udder health [10] or even evaluate the systemic immune status, as proposed for dairy cows [144].

10. Conclusions

Intramammary infection is the main cause of increased SCC in milk of lactating animals. However, physiological factors such as lactating stage, lactation number, type of birth, milk yield, and breed have also an influence on the SCC in sheep and goats. It is also noted that the physiological SCC levels in milk remain low when housing, feeding, and milking are carried out under ideal conditions. Importantly, the milking system must meet the physiological requirements of sheep and goats in order to increase milk yield, achieve better milk quality, and maintain udder health. So far, there are currently no legal limits for somatic cells in goat's and sheep's milk. However, a maximal limit of 750×10^3 cells/mL for sheep's milk and 1000×10^3 cells/mL for goat's milk has been set in the USA. In relation to this study, the threshold in the US is high and our recommended SCC values in sheep's and goat's milk are 500×10^3 cells/mL and 750×10^3 cells/mL, respectively. It is noteworthy that this proposed high SCC threshold in goat's milk is due to the high concentration of cytoplasmic particles/vesicles.

Author Contributions: S.K., conceptualization and writing the original manuscript, except point number 9, which was written by S.F.; M.W.P., revision of some content and language improvement of the original manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: There is no additional funding for this research other than salary.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: All data used to support the findings of this study are included within.

Conflicts of Interest: Shehadeh Kaskous: As head of the research and development department, I am an employer at Siliconfrom. No company materials or equipment were used as the paper is a review. There was no financial support other than my salary. Shehadeh Kaskous was the senior author involved in the design of the manuscript.

References

- Knuth, R.M.; Stewart, W.C.; Taylor, J.B.; Bisha, B.; Yeoman, C.J.; Van Emon, M.L.; Murphy, T.W. Relationships among intramammary health, udder and teat characteristics, and productivity of extensively managed ewes. J. Anim. Sci. 2021, 99, 1–10. [CrossRef]
 [PubMed]
- 2. Bergonier, D.; Berthold, X. New advances in epizootiology and control of ewe mastitis. Livest. Prod. Sci. 2003, 79, 1–16. [CrossRef]
- 3. Gonzalo, C. Milk hygiene in small ruminants: A review. Span. J. Agric. Res. 2017, 15, e05R02. [CrossRef]

- Albenzio, M.; Figliola, L.; Caroprese, M.; Marino, R.; Sevi, A.; Santillo, A. Somatic cell count in sheep milk. *Small Rumin. Res.* 2019, 176, 24–30. [CrossRef]
- Rupp, R.; Huau, C.; Caillat, H.; Fassier, T.; Bouvier, F.; Pampouille, E.; Clement, V.; Palhiere, I.; Larroque, H.; Tosser-Klopp, G.; et al. Divergent selection on milk somatic cell count in goats improves udder health and milk quality with no effect on nematode resistance. J. Dairy Sci. 2019, 102, 5242–5253. [CrossRef]
- Bergonier, D.; De Cremoux, R.; Ruup, R.; Lagriffoul, G.; Berthold, X. Mastitis of dairy small ruminants. Vet. Res. 2003, 34, 680–716. [CrossRef]
- Paape, M.J.; Poutrl, B.; Contreras, A.; Marco, J.C.; Capuco, A.V. Milk somatic cell count and lactation in small ruminants. *J. Dairy* Sci. 2001, 84, E237–E244. [CrossRef]
- Haenlein, G.F.W. Relationship of somatic cell counts in goat milk to mastitis and productivity. *Small Rumin. Res.* 2002, 45, 163–178. [CrossRef]
- Bagnicka, E.; Winnicka, A.; Jozwik, A.; Rzewuska, M.; Strzalkowska, N.; Kosciuczuk, E.; Prusak, B.; Kaba, J.; Horbanczuk, J.; Krzyzewski, J. Relationship between somatic cell count and bacterial pathogens in goat milk. *Small Rumin. Res.* 2011, 100, 72–77. [CrossRef]
- 10. Souza, F.N.; Blagitz, M.G.; Penna, C.; Della Libera, A.M.M.P.; Heinemann, M.B.; Cerqueira, M.M.O.P. Somatic cell count in small ruminants: Friend or foe? *Small Rumin. Res.* 2012, 107, 65–75. [CrossRef]
- 11. Jimenez-Granado, R.; Sanchez-Rodriguez, M.; Arce, C.; Rodriguez-Estevez, V. Factors affecting somatic cell count in dairy goats: A review. *Span. J. Agric. Res.* **2014**, *12*, 133–150. [CrossRef]
- 12. Shah, A.; Darzi, M.M.; Kamil, S.A.; Mir, M.S.; Maqbool, R.; Ali, R.; Kashani, B.; Wani, H.; Bashir, A.; Dar, A.A.; et al. Somatic cell alteration in healthy and mastitis milk of sheep and goats. *J. Entomol. Zool. Stud.* **2017**, *5*, 27–33.
- 13. Alhussien, M.N.; Dang, A.K. Milk somatic cells, factors influencing their release, future prospects, and practical utility in dairy animals—An overview. *Vet. World* **2018**, *11*, 562–577. [CrossRef] [PubMed]
- Sierra, D.; Sanchez, A.; Corrales, J.C.; Contreras, A. Differential cell counts in goats' milk. In *Milking and Milk Production of Dairy* Sheep and Goats; Wageningen Pres EAAP: Wageningen, The Netherlands, 1999; Volume 95, pp. 178–180.
- 15. Manlongat, N.; Yang, T.J.; Hinckley, L.S.; Bendel, R.B.; Krider, M. Physiologic-Chemoattractant- induced migration pf polymorphonuclear leukocytes in milk. *Clin. Diagn. Lab. Immunol.* **1998**, *5*, 375–381. [CrossRef] [PubMed]
- Zeng, S.S.; Escobar, E.N. Influence of somatic cell count in goat milk on yield and quality of soft cheese. In Proceedings of the IDF/Greek National Committee of IDF/CIRAVAL Seminar on Production and Utilization of Ewe and Goat Milk, Crete, Greece, 19–21 October 1995; pp. 109–113.
- 17. Contreras, A.; Sierra, D.; Sanchez, A.; Corrales, J.C.; Marco, J.C.; Paape, M.J.; Gonzalo, C. Mastitis in small ruminants. *Small Rumin. Res.* 2007, *68*, 145–153. [CrossRef]
- 18. Podhorecka, K.; Borkova, M.; Sulc, M.; Seydlova, R.; Dragounova, H.; Svejcarova, M.; Peroutkova, J.; Elich, O. Somatic cell count in goat milk: An indirect quality indicator. *Food* **2021**, *10*, 1046. [CrossRef]
- 19. Paape, M.J.; Capuco, A.V. Cellular defence mechanisms in the udder and lactation of goats. *J. Anim. Sci.* **1997**, 75, 556–565. [CrossRef]
- 20. Olechnowicz, J.; Jaskowski, J.M. Somatic cells in goat milk. Med. Weter. 2004, 60, 1263–1266.
- Plummer, P.J.; Plummer, C. Diseases of the Mammary Gland in Sheep and Goat. A Text Book of Veterinary Medicine, 2nd ed.; Pugh-Baird, Ed.; Elsevier: Amsterdam, The Netherlands, 2011; Chapter 15; pp. 442–465.
- Olechnowicz, J.; Jaskowski, J.M. Somatic cell counts and total bacterial count in bulk tank milk of small ruminants. *Slov. Vet. Res.* 2012, 49, 13–18.
- 23. Persson, Y.; Nyman, A.K.; Söderquist, L.; Tomic, N.; Persson Waller, K. Intramammary infections and somatic cell count in meat and pelt producing ewes with clinically healthy udders. *Small Rumin. Res.* 2017, 156, 66–72. [CrossRef]
- Albizu, I.; Penades, J.R.; Baselga, R.; Amorena, B.; Marco, J. Prevalence of subclinical mastitis in rasa aragonesa ewes. *Med. Vet.* 1991, 12, 723–728.
- 25. Regi, G.; Honegger, R.; Büchi, S.; Segessemann, V.; Rüsch, P. Somatic cell count and California mastitis test results in milk sheep with normal udder health during a complete lactation period. *Schweizer Arch. Tierheilk.* **1991**, *133*, 75–80.
- De la Cruz, M.; Serrano, E.; Montoro, V.; Marco, J.; Romeo, M.; Baselga, R.; Albizu, I.; Amorena, B. Etiology and prevalence of subclinical mastitis in the Menchega sheep at mid-date lactation. *Small Rumin. Res.* 1994, 14, 175–180. [CrossRef]
- Cuccuru, C.; Moroni, P.; Zecconi, A.; Casu, S.; Caria, A.; Contini, A. Milk differential cell counts in relation to total counts in Sardinian ewes. *Small Rumin. Res.* 1997, 25, 169–173. [CrossRef]
- Tancin, V.; Baranovic, S.; Uhrincat, M.; Macuhova, L.; Vrskova, M.; Oravcova, M. Somatic cell counts in raw ewes' milk in dairy practice: Frequency of distribution and possible effect on milk yield and composition. *MLjekarstvo* 2017, 67, 253–260. [CrossRef]
- 29. Menzies, P.I. Mastitis of sheep-overview of recent literature. In Proceedings of the 6th Great Lakes Dairy Sheep Symposium, Guelph, ON, Canada, 2–4 November 2000; pp. 68–76.
- 30. Pengov, A. The Role of coagulase-negative *staphylococcus* spp. and associated somatic cell counts in the ovine mammary gland. *J. Dairy Sci.* **2001**, *84*, 572–584. [CrossRef]
- Caboni, P.; Manis, C.; Ibba, I.; Contu, M.; Coroneo, V.; Scano, P. Compositional profile of ovine milk with a high somatic cell count: A metabolomics approach. *Int. Dairy J.* 2017, 69, 33–39. [CrossRef]

- Arias, R.; Oliete, B.; Ramón, M.; Arias, C.; Gallego, R.; Montoro, V.; Gonzalo, C.; Pérez-Guzmán, M.D. Long-term study of environmental effects on test- day somatic cell count and milk yield in Manchega sheep. *Small Rumin. Res.* 2012, 106, 92–97. [CrossRef]
- 33. Zafalon, L.F.; Mascarenhas Santana, R.C.; Pilon, L.E.; Fim Júnior, G.A. Diagnosis of subclinical mastitis in Santa Ines and Morada Nova sheep in south eastern Brazil. *Trop. Anim. Health Prod.* **2016**, *48*, 697–972. [CrossRef]
- Kern, G.; Traulsen, I.; Kemper, N.; Krieter, J. Analysis of somatic cell counts and risk factors associated with occurrence of bacteria in ewes of different primary purposes. *Livest. Sci.* 2013, 157, 597–604. [CrossRef]
- 35. Berthelot, X.; Lagriffoul, G.; Concordet, D.; Barillet, F.; Bergonier, D. Physiological and pathological thresholds of somatic cell count in ewe milk. *Small Rumin. Res.* **2006**, *62*, 27–31. [CrossRef]
- 36. Kiossis, E.; Brozos, C.N.; Petridou, E.; Boscos, C. Program for the control of subclinical mastitis in dairy Chios breed ewes during lactation. *Small Rumin. Res.* 2007, 73, 194–199. [CrossRef]
- Nunes, G.R.; Blagitz, M.G.; Freitas, C.B.; Souza, F.N.; Ricciardi, M.; Stricagnold, C.R.; Sanches, B.G.S.; Azedo, M.R.; Sucupira, M.C.A.; Della Libera, A.M.M.P. Avaliacao de Indicadores Inflamatorios no Diagnostico da Mamite ovina; Arquivos do Instituto Biologico: Sao Paulo, Brazil, 2008; Volume 75, pp. 271–278.
- Fragkou, I.A.; Boscos, C.M.; Fthenakis, G.C. Diagnosis of clinical or subclinical mastitis in ewes. Small Rumin. Res. 2014, 118, 86–92. [CrossRef]
- 39. Tvarožková, K.; Tančin, V.; Uhrinčať, M.; Hleba, L.; Mačuhová, L. Mastitis Pathogens and somatic cell count in ewe's milk. *Potravin. Slovak J. Food Sci.* **2020**, *14*, 164–169.
- Sutera, A.M.; Portolano, B.; Di Gerlando, R.; Sardina, M.T.; Mastrangelo, S.; Tolone, M. Determination of milk production losses and variations of fat and protein percentages according to different levels of somatic cell count in Valle del Belice dairy sheep. *Small Rumin. Res.* 2018, 162, 39–42. [CrossRef]
- 41. Kaskous, S. The effect of some factors in electrical conductivity of sheep milk and their relationship with the other milk parameters. *J. Agric. Sci.* **2006**, *22*, 81–96.
- 42. Swiderek, W.P.; Charon, K.M.; Winnicka, A.; Gruszczyriska, J.; Pierzchala, M. Physiological threshold of somatic cell count in milk of polish health sheep and polish lowland sheep. *Ann. Anim. Sci.* **2016**, *16*, 155–170. [CrossRef]
- 43. Kaskous, S. Correlation of somatic cell count to other methods for Detection of Subclinical Mastitis in Syrian Awassi Ewe Milk. *J. Agric. Sci.* 2000, *16*, 21–34.
- 44. Petrova, N. Effect of different somatic cell count on the production, composition, and some properties of sheep milk. *Bulg. J. Agric Sci.* **2002**, *8*, 87–96.
- 45. Barth, K.; Burow, E.; Knappstein, K. EC and CMT detect subclinical mastitis in dairy sheep but less sensitive than in dairy cows. *Landbauforsch.-Vti Agric. For. Res.* **2008**, *58*, 65–69.
- 46. Pradiee, J.; Moraes, C.R.; Goncalves, M.; Vilanova, M.S.; Correa, G.F.; Ferreira, O.; Osorio, M.; Schmidt, V. Somatic cell count and California Mastitis test as a diagnostic tool for subclinical mastitis in ewes. *Acta Sci. Vet.* **2012**, *40*, 1038.
- 47. Salaris, S.; Sechi, S.; Casu, S.; Carta, A. Relationship between somatic cell count and milk yield in the Sarda dairy sheep breed. Proceedings of the 38th Conference ICAR Annual Meeting, Cork, Ireland, 27 May–1 June 2012.
- Lagriffoul, G.; Batut, E.; Astruc, J.M.; de Cremoux, R.; Bergonier, D. Relatinship between somatic cell counts and other udder phenotypes in dairy sheep. In Proceedings of the ICAR Conference 11, Auckland, New Zealand, 7–11 February 2018; pp. 1–4.
- 49. Bytyqi, H.; Mehmeti, H.; Vehapi, I.; Rrustemaj, F.; Mehmeti, I. Effect of Bacterial content, and Somatic cell count on sheep milk quality in Kosovo. *Food Nutr. Sci.* 2013, 4, 414–419. [CrossRef]
- 50. Makovicky, P.; Makovicky, P.; Nagy, M.; Rimarova, K.; Diablekova, J. Genetic parameters for somatic cell count, Log SCC, and somatic cell score of breeds: Improved Valachian, Tsigal, Lacaune and their crossers. *Acta Vet.* **2014**, *64*, 386–396.
- 51. Merlin Junior, I.A.; Santos, J.S.; Costa, L.G.; Costa, R.G.; Ludovico, A.; Rego, F.C.; Santana, E.H. Sheep milk: Physical–chemical characteristics and microbiological quality. *Arch. Latinoam. Nutr.* **2015**, *65*, 193–198.
- 52. Tvarožková, K.; Tančin, V.; Uhrinčať, M.; Mačuhová, L.; Toman, R.; Tunegová, M. Evaluation of somatic cells in milk of ewes as possible physiological level. *Acta Fytotechn Zootechn*. **2018**, *21*, 149–151. [CrossRef]
- 53. Ilyas, N.M. Effect of age on somatic cell count and milk composition in sheep milk. J. Univ. Duhok 2020, 23, 114–117. [CrossRef]
- Puggioni, G.M.G.; Tedde, V.; Uzzau, S.; Dore, S.; Liciardi, M.; Cannas, E.A.; Pollera, C.; Moroni, P.; Bronzo, V.; Addis, M.F. Relationship of late lactation milk somatic cell count and cathelicidin with intramammary infection in small ruminants. *Pathogens* 2020, *9*, 37. [CrossRef]
- 55. Tvarožková, K.; Vasicek, J.; Uhrincat, M.; Macuhova, L.; Helba, L.; Tancin, V. The presence of pathogens in milk of ewes in relation to the somatic cell count and subpopulations of leukocytes. *Czech J. Anim. Sci.* **2021**, *66*, 315–322. [CrossRef]
- Lianou, D.T.; Michael, C.K.; Vasileiou, N.G.C.; Liagka, D.V.; Mavrogianni, V.S.; Caroprese, M.; Fthenakis, G.C. Association of Breed of Sheep or Goats with Somatic Cell Counts and Total Bacterial Counts of Bulk-Tank Milk. *Appl. Sci.* 2021, 11, 7356. [CrossRef]
- 57. Mcdougall, S.; Pankey, W.; Delaney, C.; Barlow, J.; Murdough, P.A.; Scruton, D. Prevalence, and incidence of subclinical mastitis in goats and dairy ewes in Vermont, USA. *Small Rumin. Res.* **2002**, *46*, 115–121. [CrossRef]
- Luengo, C.; Sanchez, A.; Corrales, J.C.; Fernandez, C.; Contreras, A. Influence of intramammary infection and non-infection factors on somatic cell counts in dairy goats. J. Dairy Res. 2004, 71, 169–174. [CrossRef] [PubMed]

- Park, Y. Improving goat milk. In *Improving the Safety and Quality of Milk, Milk Production and Processing*; Woodhead Publishing Series in Food Science, Technology, and Nutrition; Woodhead Publishing Ltd.: Cambridge, UK, 2010; pp. 304–346.
- Boettcher, P.J.; Moroni, P.; Pisoni, G.; Gianola, D. Application of a finite mixture model to somatic cell scores of Italian goats. J. Dairy Sci. 2005, 88, 2209–2216. [CrossRef] [PubMed]
- Persson, Y.; Olofsson, I. Direct and indirect measurement of somatic cell count as indicator of intramammary infection in dairy goats. Acta Vet. Scand. 2011, 53, 1–5. [CrossRef] [PubMed]
- 62. Csanadi, J.; Fenyvessy, J.; Bohata, S. Somatic cell count of milk from different goat breeds. *Acta Univ. Sapientiale Alimentaria* 2015, *8*, 45–54. [CrossRef]
- 63. Contreras, A.; Sierra, D.; Corrales, J.C.; Sanchez, A.; Marco, J. Physiological threshold of somatic cell count and California mastitis test for diagnosis of caprine subclinical mastitis. *Small Rumi. Res.* **1996**, *21*, 259–264. [CrossRef]
- 64. Fahr, R.D.; Suess, R.; Kaskous, S.; Finn, G. Somatic Cell Count and Milk quality in the Small Ruminants. In Proceedings of the Second, VDL-Special Symposium on Research in the Sheep Sector, Halle, Germany, 3–4 November 1999; pp. 13–17.
- 65. Koop, G.; Van-Werven, T.; Toft, N.; Nielen, M. Estimating test characteristics of somatic cell count to detect Staphylococcus aureus-infected dairy goats using latent class analysis. *J. Dairy Sci.* 2011, 94, 2902–2911. [CrossRef]
- Min, B.R.; Tomita, G.; Hart, S.P. Effect of subclinical intramammary infection on somatic cell counts and chemical composition of goats' milk. J. Dairy Res. 2007, 74, 204–2010. [CrossRef]
- 67. Rychtarova, J.; Krupova, Z.; Brzakova, M.; Borkova, M.; Elich, O.; Dragounova, H.; Seydlova, R.; Sztankoova, Z. Milk quality, somatic cell count, and economics of dairy goat's farm in the Czech Republic. In *Goat Science-Environment, Health and Economy*; Kukovics, S., Ed.; Intech Open: London, UK, 2021.
- 68. Gecaj, R.M.; Ajazi, F.; Bytyqi, H.; Mehmedi, B.; Cadraku, H.; Ismaili, M. Somatic cell number, physiochemical and microbiological parameters of raw milk of goats during the end of lactation as compared by breeds and number of lactations. *Front. Vet. Sci.* **2021**, *8*, 694114. [CrossRef]
- 69. Degirmencioglu, T. Effect of high and low somatic cell counts on the milk composition and yield of Saanen goats. *J. Anim. Husb. Dairy Sci.* **2018**, *2*, 41–45.
- Paape, M.J.; Wiggans, G.R.; Bannerman, D.D.; Thomas, D.L.; Sanders, A.H.; Contreras, A.; Moroni, P.; Miller, R.H. Monitoring goat and sheep milk somatic cell counts. *Small Rumin. Res.* 2007, 68, 114–125. [CrossRef]
- 71. Corrales, J.C.; Sanchez, A.; Luengo, C.; Poveda, J.B.; Contreras, A. Effect of clinical contagious agalactia on the bulk tank milk somatic cell count in Murciano-Granadins goat herds. *J. Dairy Sci.* 2004, *87*, 3165–3171. [CrossRef] [PubMed]
- Gonzalo, C.; Tardaguila, A.; Ariznabarreta, A.; Romeo, M.; Montoro, V.; Perez-Guzman, M.D.; Marco, J.C. Somatic cell counts in dairy livestock and control strategies: Situation in Spain. In *Mastitis and Milk Quality*; Contreras, A., Sanchez, A., Romero, J.C., Eds.; Diego Marin Publishing: Murcia, Spain, 2000; pp. 145–151.
- 73. Silanikove, N.; Merin, U.; Leitner, G. On effects of subclinical mastitis and stage of lactation on milk quality in goats. *Small Rumin. Res.* **2014**, *122*, 76–82. [CrossRef]
- 74. Bergonier, D.; Lagriffoul, G.; Berthelot, X.; Barillet, F. *Facteurs de Variation Non Infectieux des Comptages de Cellules Somatiques Chez les Ovins et Caprins Laitiers*; Wageningen Pers EAAP: Wageningen, The Netherlands, 1994; Volume 77, pp. 112–135.
- 75. Anderson, D.E.; Hull, B.L.; Pugh, D.G. Enfermidades da glandula mamaria. In *Clinica de Ovinos e Caprinos*; Pugh, D.G., Ed.; Roca: Sao Paulo, Brazil, 2005; pp. 379–399.
- 76. Gonzalo, C. Somatic cell of sheep and goat milks, analytical, sanitary, productive, and technological aspects. In Proceedings of the International dairy federation (Future of sheep and goats' dairy sector) 0501/part 3(special Issue), Zaragoza, Spain, 28–30 October 2004; pp. 128–133.
- 77. Ceballos, L.S.; Morales, E.R.; Adarve, G.; Castro, J.D.; Martinez, L.P.; Sampelayo, M.R.S. Composition of goat and cow milk production under similar conditions and analysed by identical methodology. *J. Food Compos. Anal.* 2009, 22, 322–329. [CrossRef]
- Goetsch, A.L.; Zeng, S.S.; Gipson, T.A. Factors affecting goat milk production and quality. *Small Rumin. Res.* 2011, 101, 55–63. [CrossRef]
- 79. Gonzalo, C.; Ariznabarreta, A.; Carriedo, J.A.; San Primitive, F. Mammary pathogens and their relationship to somatic cell count and milk yield losses in dairy ewes. *J. Dairy Sci.* 2002, *85*, 1460–1467. [CrossRef]
- 80. Gonzalo, C.; Carriedo, J.A.; Blanco, M.A.; Beneitez, E.; Juarez, M.T.; De La Fuente, L.F.; San Primitivo, F. Factors of variation influencing bulk tank somatic cell count in dairy sheep. *J. Dairy Sci.* 2005, *88*, 969–974. [CrossRef]
- 81. De Cremoux, R.; Pillet, R.; Ducelliez, M.; Heuchel, V.; Puotrel, B. *Influence du Nombre et du Stade de Lactation sur les Numerations Cellulaires du Lait de Chevre*; EAAP: Bella, Italy; Wageningen Pers: Wageningen, The Netherlands, 1996; Volume 77, pp. 61–165.
- 82. Contreras, A.; Sanchez, A.; Corrales, J.C.; Marco, J.C. Concepto e importancia de las mamitis caprinas. Ovis 1997, 58, 11–31.
- 83. Martinez, B. El Recuento de Celulas Somaticas en la Leche de Cabra, Factors de Variacion y Efecto Sobre la Produccion y Composicion de la Leche. Ph.D. Thesis, Universidad Politecnica de Valencia, Valencia, Spain, 2000.
- 84. Skapetas, B.; Bampidis, V.; Christodoulou, V.; Kalaitizidou, M. Fatty acid profile, somatic cell count and microbiological quality of total machine milk and hand stripped milk of Chios ewes. *MLjekarstvo* **2017**, *67*, 146–154. [CrossRef]
- Corrales, J.C.; Sanchez, A.; Sierra, D.; Marco, J.C.; Contreras, A. Relationship between somatic cell counts and intramammary pathogens goats. In *Somatic Cells and Milk Small Ruminants*; Rubino, R., Ed.; Wageningen Pers: Wageningen, The Netherland, 1996; pp. 35–39.

- Wilson, D.J.; Stewart, K.N.; Sears, P.M. Effects of stage of lactation, production, parity, and season on somatic cell counts in infected and uninfected dairy goats. *Small Rumin. Res.* 1995, 16, 165–169. [CrossRef]
- Bergonier, D.; Longo, F.; Lagriffoul, G.; Consalvi, P.J.; Van de Wiele, A.; Berthelot, X. Frequence et persistence des staphylocoques coagulase negative au tarissement et relations avec les numerations cellulaires chez la brebis laitiere. In *Somatic Cells and Milk of Small Ruminants*; Rubino, R., Ed.; Wageningen Pers: Wageningen, The Netherland, 1996; pp. 53–59.
- Zeng, S.S.; Escobar, E.N.; Brown-Crowder, I. Evaluation of screening tests for detection of antibiotic residues in goat milk. *Small Rumin. Res.* 1996, 21, 155–160. [CrossRef]
- 89. Gomes, V.; Libera, A.M.; Paiva, M.; Madureira, K.M.; Araujo, W.P. Effect of the stage of lactation on somatic cell counts in healthy goats (Caprae hircus) breed in Brazil. *Small Rumin. Res.* **2006**, *64*, 30–34. [CrossRef]
- 90. Zafalon, L.F.; Mascarenhas Santana, R.C.; Esteves, S.N.; Fim Júnior, G.A. Somatic cell count in the diagnosis of subclinical mastitis in sheep of different breeds. *Semin. Cienc. Agrar. Londrina* **2018**, *39*, 1555–1564. [CrossRef]
- Kloppert, B.; Ehrenberg, A.; Wolter, W.; Zschök, M. Konzept zur Überwachung und Eigenkontrolle in Schaf- und Ziegenmilcherzeugerbetrieben im Sinne der Milchverordnung. In Proceedings of the 41 Arbeitstagung Lebensmittelhygiene, Garmisch-Partenkirchen, Gießen, Germany, 25–28 September 2000; pp. 213–219.
- 92. Lerondelle, C.; Richard, Y.; Issartial, J. Factors affecting somatic cell counts in goat milk. *Small Rumin. Res.* **1992**, *8*, 129–139. [CrossRef]
- 93. Raynal-Ljutovac, K.; Pirisl, A.; de Cremoux, R.; Gonzalo, C. Somatic cells of goat and sheep milk: Analytical, sanitary, productive, and technological aspects. *Small Rumin.* 2007, *68*, 126–144. [CrossRef]
- 94. Krömker, V. Milchkunde und Milchhygiene; Parey in MVS Medezinverlag: Stuttgart, Germany, 2007; pp. 47–74.
- 95. Jimenez-Granado, R.; Rodriguez-Estevez, V.; Morantes, M.; Arce, C.; Sanchez-Rodriguez, M. Relationship between reproductive parameters and somatic cell count in dairy goats. *Reprod. Domest. Anim.* **2012**, *47*, 90–123.
- 96. Sanchez-Rodriguez, M.; Gomez Castro, A.G.; Mata Moreno, C.; Domenech Garcia, V.; Lopez Baldan, D.; Romero, A.M.; Cabello, A. Resultados productivos del rebano experimental de raza Florida. *Feagas* **2000**, *1*, 105–107.
- 97. Sanchez, A.; Corrales, J.C.; Marco, J.; Contreras, A. Aplicacion del recuento de celulas somaticas para el control de las mamitis caprinas. *Ovis* **1998**, *54*, 37–51.
- De Vasconcelos, M.; Silva, P.; Merighe, G.; Oliveira, S.; Negrao, J. Cumulative effect of different acute stressors on physiological and hormonal responses and milk yield in lactating Sannen goats. *Semin. Cienc. Agrar. Londrina* 2022, 43, 1891–1906. [CrossRef]
- Gonzalo, C.; Carriedo, J.A.; Beneitez, E.; Juarez, M.T.; De La Fuente, L.F.; San Primitivo, F. Bulk tank total bacterial count in dairy sheep: Factors of variation and relationship with somatic cell count: Short communication. J. Dairy Sci. 2006, 89, 549–552. [CrossRef]
- Koop, G.; Dik, N.; Nielen, M.; Lipman, L.J.A. Repeatability of differential goat bulk milk culture and associations with somatic cell count, total bacterial count, and standard plate count: Short communication. J. Dairy Sci. 2010, 93, 2569–2573. [CrossRef]
- 101. Koop, G.; Nielen, M.; van Werven, T. Bulk milk somatic cell counts are related to bulk milk total bacterial counts and several herd-level risk factors in dairy goats. *J. Dairy Sci.* **2009**, *92*, 4355–4364. [CrossRef] [PubMed]
- 102. Park, Y.W.; Humphrey, R.D. Bacterial cell counts in goat milk and their correlation with somatic cell counts, percent fat, and protein. *J. Dairy Sci.* **1986**, *69*, 32–37. [CrossRef]
- De Cremoux, R.; Heuchel, V.; Berny, F. Interpretation des numerations cellulaires de lait de troupeau en elevage caprin: Resultats preliminaires. In Proceedings of the 7th International Conference on Goat, Tours, France, 15–21 May 2000; p. 764.
- 104. De Cremoux, R.; Heuchel, V.; Berny, F. Description et interpretation des comptages de cellules somatiques des laits de troupeaux en elevage caprin. In Proceedings of the 8es Rencontres Autour des Recherches sur les Ruiminata, Institute de Lelevage, Paris, France, 5–6 December 2001; p. 163.
- 105. Kaskous, S. Laboratory tests to optimize the milking machine settings with air inlet teat cups for sheep and goats. *Dairy* **2022**, *3*, 3. [CrossRef]
- Fernandez, G.; Diaz, J.R.; Peris, C.; Rodriguez, M.; Molina, M.P.; Torres, A. Machine milking parameters for the Manchega sheep breed. In Proceedings of the Sixth International Symposium on the Milking of Small Ruminants, Athens, Greece, 26 September–1 October 1999; pp. 65–68.
- 107. Peris, C.; Diaz, J.R.; Balasch, S.; Beltran, M.C. Influence of vacuum level and overmilking on udder health and teat thickness changes in dairy ewes. *J. Dairy Sci.* 2003, *86*, 3891–3898. [CrossRef] [PubMed]
- Peris, C.; Diaz, J.R.; Segura, C.; Marti, A.; Fernandz, N. Influence of pulsation rate on udder health and teat thickness changes in dairy ewes. J. Dairy Sci. 2003, 86, 530–537. [CrossRef]
- 109. Fernandez, N.; Marti, J.V.; Rodriguez, M.; Peris, C.; Balasch, S. Machine milking parameters for Murciano-Granadina breed goats. J. Dairy Sci. 2020, 103, 507–513. [CrossRef]
- 110. Caria, M.; Boselli, C.; Murgia, L.; Rosati, R.; Pazzona, A. Influence of low vacuum levels on milking characteristics of sheep, goat, and buffalo. *J. Agric. Eng.* **2013**, *XLIV*, e43. [CrossRef]
- 111. Marnet, P.G.; Combaud, J.F.; Dano, Y. Relationships between characteristics of the teat and milkability in Lacuune sheep. In Proceedings of the 6th International Symposium in Machine Milking of Small Ruminants: Milking and Milk Production of Dairy Sheep and Goats, Athens, Greece, 26 September–1 October 1999; pp. 41–44.

- 112. Sinapis, E.; Marnet, P.G.; Skapetas, B.; Hatziminaoglou, I. Vacuum level for opening the teat sphincter and the change of the teat end wall thickness during the machine milking of mountainous Greek breed (Boutsiko) ewes. *Small Rumin. Res.* 2007, 69, 136–143. [CrossRef]
- 113. Dzidic, A.; Rovai, M.; Poulet, J.L.; Leclerc, M.; Marnet, P.G. Review: Milking routine and cluster detachment levels in small ruminants. *Animal* **2019**, *13* (Suppl. S1), 86–93. [CrossRef]
- 114. Marnet, P.G.; McKusick, B.C. Regulation of milk ejection and milkability in small ruminants. *Livest. Sci.* 2001, 7, 125–133. [CrossRef]
- 115. Torres, A.; Castro, N.; Hernández-Castellano, L.E.; Argüello, A.; Capote, J. Short communication: Effects of milking frequency on udder morphology, milk partitioning, and milk quality in 3 dairy goat breeds. J. Dairy Sci. 2013, 69, 1071–1074. [CrossRef] [PubMed]
- 116. Laga, V.; Skapetas, B.; Katanos, I.; Sinapis, E.; Hatziminiaoglou, I. Efficiency of milking machines for dairy ewes in central Macedonia, Greece. *Anim. Sci. Rev.* 2007, *36*, 23–40.
- 117. Tölü, C.; Irmak, S.; Acikel, S.; Akbag, H.I.; Savas, T. Comparison of milk yield, milk composition and residual milk of machine and Hand-Milked in Turkish Saanen Goats. J. Agric. Sci. 2016, 22, 462–470.
- Dzidic, A.; Kaps, M.; Bruckmaier, R.M. Machine milking of Istrian dairy crossbreed ewes: Udder morphology and milking characteristics. *Small Rumin. Res.* 2004, 55, 183–189. [CrossRef]
- 119. Legarra, A.; Ugarte, E. Genetic parameters of udder traits, somatic cell score, and milk yield in Latxa sheep. *J. Dairy Sci.* 2005, *88*, 2238–2245. [CrossRef]
- 120. Marie-Etancelin, C.; Astruc, J.M.; Porte, D.; Larroque, H.; Robert-Granie, C. Multiple trait genetic parameters and genetic evaluation of udder-type traits in Lacaune dairy ewes. *Livest. Prod. Sci.* 2005, 97, 211–218. [CrossRef]
- Margetin, M.; Spanik, J.; Milerski, M.; Capistrak, A.; Apolen, D. Relationships between morphological and functional udder traits and somatic cell count in milk of ewes. In Proceedings of the International Conference Physiological and Technical Aspects of Machine milking, Nitra, Slovak Republic, 26–28 April 2005.
- 122. Sinapis, E.; Hatziminaoglu, I.; Marnet, P.G.; Abas, Z.; Bolou, A. Influence of vacuum level, pulsation rate and pulsator ratio on machine milking efficiency in local Greek goats. *Livest. Prod. Sci.* **2000**, *64*, 175–181. [CrossRef]
- 123. Salama, A.A.K.; Such, X.; Caja, G.; Rovai, M.; Casals, R.; Albanell, E.; Marin, M.P.; Marti, A. Effects of once versus twice daily milking throughout lactation on milk yield and milk composition in dairy goats. J. Dairy Sci. 2003, 86, 1673–1680. [CrossRef]
- 124. Capote, J.; Castro, N.; Caja, G.; Fernandez, G.; Briggs, H.; Argüello, A. Effect of the frequency of milking and lactation stage on milk fractions and milk composition in Tinerfena dairy goats. *Small Rumin. Res.* **2008**, *75*, 252–255. [CrossRef]
- 125. Giaccone, P.; Scatassa, M.L.; Todaro, M. The influence of somatic cell count on sheep milk composition and cheese-making properties. *Ital. J. Anim. Sci.* 2005, 4 (Suppl. S2), 345–347. [CrossRef]
- Margetin, M.; Milerski, M.; Apolen, D.; Capistak, A.; Oravcova, M.; Debreceni, O. Relationship between production, quality of milk and udder health status of ewes during machine milking. J. Cent. Eur. Agric. 2013, 14, 328–340. [CrossRef]
- 127. Baranovic, S.; Tancin, V.; Tvarozkova, K.; Uhrincat, M.; Macuhova, L.; Palkovic, J. Impact of somatic cell count and lameness on the production and composition of ewe's milk. *Potravinarstvo* **2018**, *12*, 116–121. [CrossRef] [PubMed]
- 128. Tvarožková, k.; Tančin, V.; Uhrinčať, M.; Hleba, L.; Mačuhová, L.; Miklas, S. The impact of somatic cell count on milk yield and composition. *Sci. J. Phytotech. Zootech.* 2021, 24, 49–52. [CrossRef]
- 129. Barbano, D.M.; Rasmussen, R.R.; Lynch, J.M. Influence of milk somatic cell count and milk age on cheese yield. *J. Dairy Res.* **1991**, 74, 369–388. [CrossRef]
- Pirisi, A.; Piredda, G.; Corona, M.; Pes, M.; Pintus, S.; Ledda, A. Influence of somatic cell count on ewe's milk composition, cheese yield and cheese quality. In Proceedings of the 6th Great Lakes Dairy Sheep Symposium, Guelph, ON, Canada, 2–4 November 2000; pp. 47–59.
- 131. Chen, S.X.; Wang, J.Z.; Van Kessel, J.S.; Ren, F.Z.; Zeng, S.S. Effect of somatic cell count in goat milk on yield, sensory quality, and fatty acid profile of semisoft cheese. *J. Dairy Sci.* 2010, *93*, 1345–1354. [CrossRef]
- 132. Sordillo, L.M. Mammary Gland Immunobiology and Resistance to Mastitis. *Vet. Clin. North Am. Food Anim. Pract.* 2018, 34, 507–523. [CrossRef]
- 133. Farschtschi, S.; Mattes, M.; Pfaffl, M.W. Advantages and Challenges of Differential Immune Cell Count Determination in Blood and Milk for Monitoring the Health and Well-Being of Dairy Cows. *Vet. Sci.* **2022**, *9*, 255. [CrossRef]
- Godden, S.M.; Royster, E.; Timmerman, J.; Rapnicki, P.; Green, H. Evaluation of an automated milk leukocyte differential test and the California Mastitis Test for detecting intramammary infection in early- and late-lactation quarters and cows. J. Dairy Sci. 2017, 100, 6527–6544. [CrossRef]
- 135. Schwarz, D.; Kleinhans, S.; Reimann, G.; Stuckler, P.; Reith, F.; Ilves, K.; Pedastsaar, K.; Yan, L.; Zhang, Z.; Valdivieso, M.; et al. Investigation of dairy cow performance in different udder health groups defined based on a combination of somatic cell count and differential somatic cell count. *Prev. Vet. Med.* 2020, 183, 105123. [CrossRef]
- 136. Cossarizza, A.; Chang, H.-D.; Radbruch, A.; Akdis, M.; Andrä, I.; Annunziato, F.; Bacher, P.; Barnaba, V.; Battistini, L.; Bauer, W.M.; et al. Guidelines for the use of flow cytometry and cell sorting in immunological studies. *Eur. J. Immunol.* 2017, 47, 1584–1797. [CrossRef]
- 137. Winnicka, A.; Klucinski, W.; Hoser, G.; Sikora, J.; Kawiak, J. Flow Cytometry analysis of milk and peripheral blood cells from goats during lactation. *J. Vet. Med.* **1999**, *46*, 459–464. [CrossRef] [PubMed]

- Park, Y.H.; Joo, Y.S.; Park, J.Y.; Moon, J.S.; Kim, S.H.; Kwon, N.H.; Ahn, J.S.; Davis, W.C.; Davies, C.J. Characterization of lymphocyte subpopulations and major histocompatibility complex haplotypes of mastitis-resistant and susceptible cows. *J. Vet. Sci.* 2004, *5*, 29–39. [CrossRef] [PubMed]
- 139. Tatarczuch, L.; Philip, C.; Bischof, R.J.; Lee, C.S. Leukocyte pheno- types in involuting and fully involuted mammary glandular tissues and secretions of sheep. *J. Anat.* 2000, *196*, 313–326. [CrossRef] [PubMed]
- 140. Boulaaba, A.; Grabowski, N.; Klein, G. Differential cell count of caprine milk by flow cytometry and microscopy. *Small Rumin. Res.* **2011**, *97*, 117–123. [CrossRef]
- 141. Blagitz, M.G.; Souza, F.N.; Gomes, V.; Della Libera, A.M.M.P. Apoptosis and necrosis of polymorphonuclear leukocytes in goat milk with high and low somatic cell counts. *Small Rumin. Res.* **2011**, *100*, 67–71. [CrossRef]
- 142. Pilla, R.; Malvisi, M.; Snel, G.G.; Schwarz, D.; Konig, S.; Czerny, C.P.; Piccinini, R. Differential cell count as an alternative method to diagnose dairy cow mastitis. *J. Dairy Sci.* 2013, *96*, 1653–1660. [CrossRef]
- 143. De Matteis, G.; Grandoni, F.; Scata, M.C.; Catillo, G.; Moioli, B.; Buttazzoni, L. Flow Cytometry-Detected Immunological Markers and on Farm Recorded Parameters in Composite Cow Milk as Related to Udder Health Status. *Vet. Sci.* 2020, 7, 114. [CrossRef]
- 144. Farschtschi, S.; Mattes, M.; Hildebrandt, A.; Chiang, D.; Kirchner, B.; Kliem, H.; Pfaffl, M.W. Development of an advanced flow cytometry based high-resolution immunophenotyping method to benchmark early immune response in dairy cows. *Sci. Rep.* 2021, *11*, 22896. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.