

Article

Hox Gene Collinearity May Be Related to Noether Theory on Symmetry and Its Linked Conserved Quantity

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Abstract: Hox Gene Collinearity (HGC) is a fundamental property that controls the development of many animal species, including vertebrates. In the Hox gene clusters, the genes are located in a sequential order Hox1, Hox2, Hox3, etc., along the 3′ to 5′ direction of the cluster in the chromosome. During Hox cluster activation, the Hox genes are expressed sequentially in the ontogenetic units D1, D2, D3, etc., along the anterior–posterior axis (A-P) of the early embryo. This collinearity, first observed by E.B. Lewis, is surprising because the spatial collinearity of these structures (Hox clusters and embryos) correlates entities that differ by about four orders of magnitude. Biomolecular mechanisms alone cannot explain such correlations. Long-range physical interactions, such as diffusion or electric attractions, should be involved. A biophysical model (BM) was formulated, which, in alignment with the biomolecular processes, successfully describes the existing vertebrate genetic engineering data. One hundred years ago, Emmy Noether made a fundamental discovery in mathematics and physics. She proved, rigorously, that a physical system obeying a symmetry law (e.g., rotations or self-similarity) is followed by a conserved physical quantity. It is argued here that HGC obeys a ‘primitive’ self-similarity symmetry. In this case, the associated primitive conserved quantity is the irreversibly increasing ‘ratchet’-like Hoxgene ordering where some genes may be missing. The genes of a vertebrate Hox cluster are located along a finite straight line. The same order follows the ontogenetic units of the vertebrate embryo. Therefore, HGC is a manifestation of a primitive Noether Theory (NT). NT may be applied to other than the vertebrate case, for instance, to animals with a circular topological symmetry. For example, the observed abnormal Hox gene ordering of the echinoderm Hox clusters may be reproduced by a double-strand break of the circular Hox gene ordering and its subsequent incorporation in the flanking chromosome.

Keywords: Hox gene collinearity; vertebrate Hox cluster; anterior–posterior axis; embryonic rotational symmetry; sea urchin

1. Introduction

In 2018, the anniversaries of two major scientific discoveries were celebrated. One in biology for the forty years of Hox Gene Collinearity (HGC) that was formulated by E.B. Lewis in 1978 [1]. The other was the centenary of the far-reaching theory in physics and mathematics, as introduced by Emmy Noether [2–4]. In the present study, it is argued that these two groundbreaking discoveries may be linked.

In 1918, Emmy Noether proposed a rigorous and elegant correlation between two fundamental concepts in physics and mathematics: symmetry and conservation laws [2]. In simple terms, Noether proved that, under some well-founded conditions in classical mechanics, a physical system obeying a symmetry law is necessarily followed by a conserved physical quantity [3,4]. For instance, the physical

laws that are symmetrical under translations in space (freedom to choose the origin of the coordinate system) lead to the conservation of momentum. Similarly, energy conservation during physical processes is a consequence of the symmetry of the physical laws under time translations [4,5]. (See the Appendix A). Symmetry is a very broad concept with many facets covering aesthetic, philosophical, physical, and mathematical aspects. Hermann Weyl gave a broad definition as follows: A body, a spatial configuration, is symmetric with respect to a given plane E if it is carried into itself by reflection (mirror image) in E [6]. Similarly, the operation of rotations on several physical bodies may define rotational symmetry. For example, if a rotation of a body around a given axis, say by thirty degrees or (30°), carries every point of the body to another point that belongs to the body. Thus, according to Pythagoras, the circle and the sphere are completely rotationally symmetric (invariant) in two- and three-dimensional spaces, respectively [6].

Many operations (not only geometric) can be applied to physical bodies or entities, which may lead to an invariance (symmetry) related to this particular operation. Noether’s theory (NT) proved very helpful in the solution of physical problems even in the most modern fields of elementary particles and quarks. Given a symmetry, the search is directed to discover the corresponding conserved quantity (particle) and vice versa. For instance, testing the validity of some supersymmetry theories applied to elementary particles, a corresponding conserved quantity (particle) is expected [4,7]. The lasting search for this particle led to the discovery of the Higgs boson in the CERN Laboratory in 2012 [7].

2. Hox Gene Collinearity on a Straight Line

Hox genes are a group of genes playing a fundamental role in the development of most animals, including vertebrates (hence, also the humans). In many cases, some of these genes are grouped in clusters and denoted Hox1, Hox2, Hox3, etc. Numbering starts at the 3’ end of the telomeric side of the cluster, and it is directed toward the 5’ end of the centromeric side of the DNA sequence. In Figure 1a, a (theoretical) common ancestor Hox cluster is shown together with a prototypic animal clade, the mouse Hox A cluster (Figure 1b) [8]. In Figure 1a,b the distances between the Hox genes are not faithfully represented.

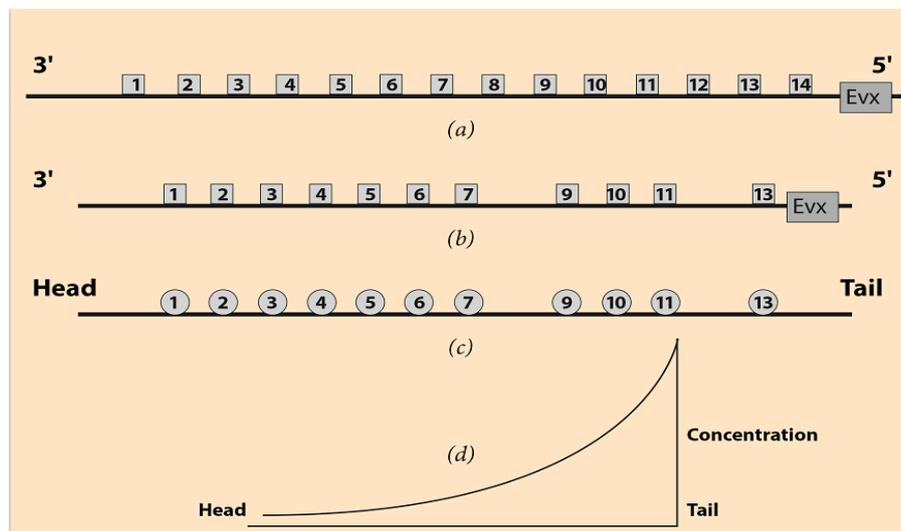


Figure 1. Ordering of Hox genes and the series of the ontogenetic units. (a) Hox gene ordering of a (theoretical) common ancestor. Gene *Evx* is located next to the 5’ end of the Hox cluster. (b) Ordering of the mouse HoxA cluster. Hox8 and Hox12 are missing. (c) The corresponding ontogenetic units of the mouse along the A-P axis. (d) The steady-state monotonic concentration gradient of a morphogen. The peak is at the tail region.

The Even Skipped Gene (*Evx*) lies close to the centromeric side of the mouse *HoxA* cluster. It is believed that the tandem duplication of an ancestral *ur-Hox* gene, followed by several evolutionary modifications, may be the origin of these organized *Hox* gene arrays [9]. Many other hypotheses have been proposed based on new data on gene duplication and gene ordering [10]. The speculative common ancestor *Hox* cluster is complete, and the gene ordering is normal from the first *Hox1* to the last *Hox* gene (Figure 1a) [8–10]. In vertebrates, there are four paralogue *Hox* clusters (*HoxA*, *HoxB*, *HoxC*, and *HoxD*). These paralogue *Hox* clusters are complementary to each other, and their combinatorial cooperation guides their evolutionary modifications. In each paralogue *Hox* cluster, some *Hox* genes are missing as, for example, in the mouse *HoxA* in Figure 1b. The origin and crucial role of such ‘holes’ in the vertebrate *Hox* clusters are not well understood (see Section 6).

In the early stages of embryogenesis, one can ontogenetically divide the embryo into body Domains D1, D2, D3, etc., following the anterior–posterior axis from head to tail (Figure 1c). E.B. Lewis noticed that some *Drosophila* *Hox* genes are activated in a collinear manner [1]. (See the Appendix A). Schematically, *Hox1* is activated in D1, *Hox2* in D2, *Hox3* in D3, etc. For instance, in the cells of D1, only the *Hox1* gene is transcribed. In the cells of D2, the *Hox2* gene is transcribed, etc. The clear cut expression regions of the *Hox* genes in the ontogenetic domains D1, D2, etc., of the vertebrates are only a simplification. In reality, these expressions are partially overlapping and nested gradually toward the tail end of the embryo or the limb bud tip, e.g., [11,12]. This nested pattern, denoted as *Hox* Gene Collinearity (HGC), can be easily incorporated in the present simplifying representation. In addition, the spatial and temporal features of HGC are still intensively studied in several animal species [13]. The above particular local transcription of the *Hox* genes is a very surprising event that proved later to be a universal property widespread to most animal species where the *Hox* genes are grouped in clusters [8].

The linear size of the *Hox* clusters extends up to some hundreds of nanometers (nm), whereas the linear size of an early embryo (head to tail) is about 1 millimeter (mm) (Figure 1). The sizes of these two entities (gene cluster and early embryo) differ by more than four orders of magnitude. In HGC, the unexpected one-to-one correspondence of the two sequences constitutes an enigma of developmental biology. Several explanatory models were proposed which can correctly reproduce many of the performed genetic engineering experimental results, e.g., [11,14]. All these models are based solely on well-established biomolecular processes responsible for gene activation or inhibition. Nevertheless, the collinearity phenomenon is still an enigma (see Section 6). Notice that the multiscale nature of HGC hints at the action of long-range forces which may cooperate with the biomolecular mechanisms. Physical processes, such as diffusion or electric attraction, can trigger such forces [15].

3. The Macro- and Micro-Influences in the Biophysical Model

As early as 2001, a biophysical model (BM) was proposed, which combines long-range gradients based mainly on diffusion, causing pulling forces at the level of the *Hox* clusters [15–18]. For many years this model was poorly endorsed because supporting experimental evidence was lacking. However, in recent years novel experimental techniques were developed that enabled the observation of physical–geometric deformations of the *Hox* clusters during *Hox* gene activation in mice [19–21]. These novel techniques, mainly based on super-resolution imaging (STORM), made it possible to measure gradual elongations of the *Hox* clusters. These elongations could be explained by the action of physical forces in the BM [16].

A one-dimensional mathematical model was introduced to explain how a morphogen monotonic gradient may be formed. The model assumes that a morphogen is produced at one end of a morphogenetic field, and it spreads mainly via passive diffusion. It has been verified that several molecules play the role of morphogens, e.g., the fibroblast growth factor (FGF), or the Sonic hedgehog [22,23]. According to this diffusion mechanism, the morphogen is produced at the tail of the embryo, where its concentration C is kept constant (C_0). C diffuses along the anterior–posterior

axis (Figure 1d). At the same time, the morphogen is decomposed by first-order chemical kinetics. The diffusion-plus-degradation equation in space x and time t of $C(x,t)$ is [16,17]

$$\frac{\partial C}{\partial t} = D \frac{\partial^2 C}{\partial x^2} - kC \tag{1}$$

D is the diffusion constant, and k is the chemical decomposition constant.

Following the method of Carslow and Jaeger, the asymptotic steady-state solution $C(x)$ of Equation (1) is

$$C(x) = C_0 e^{-A(x_0-x)} \tag{2}$$

where $A = \sqrt{k/D}$, x_0 is the tail position and x varies between 0 and x_0 ($0 \leq x \leq x_0$) [24].

In an experiment testing the time dependence of the morphogen (FGF) spreading in the chick limb bud, it is confirmed that diffusion is the responsible main mechanism [25].

In the following, the morphogen signals are transduced inside the cells and produce specific molecules that are allocated in particular positions inside the cell nucleus. The existence and action of such molecules (e.g., molecule proteins called SMAD) are thoroughly studied [26,27]. In analogy, it is here assumed that similar molecules P are produced and anchored facing the Hox cluster (Figure 2). It is further assumed that the P molecules are positively charged, while the Hox gene clusters are negatively charged (N). An attractive force acting on the Hox cluster is created resembling a truncated Coulomb force [16,17]. A simple heuristic form for the attractive force F can be introduced:

$$F = P \times N \tag{3}$$

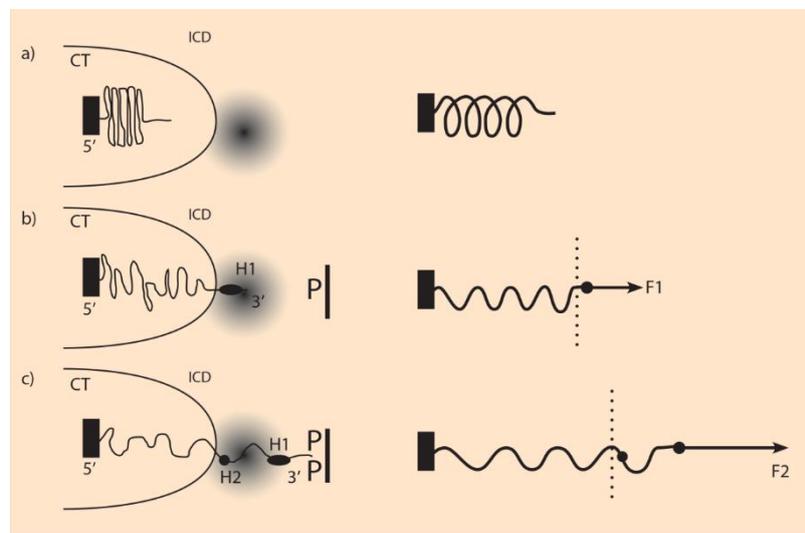


Figure 2. Hox cluster elongation and its mechanical analog (adapted from [17]). (a) (Left) Before activation, the Hox cluster is condensed inside the chromatin territory (CT).The transcription factory region outside CT is shadowed. (Right) Mechanical analog of an uncharged elastic spring fastened at its left end. (b) (Left) The fixed pole P pulls Hox1 (H1) outside the cluster in the intra chromosomal domain (ICD) towards the transcription factory. (Right) A small force F_1 expands the spring. (c) (Left) The attracting force of pole P increases (PP), and besides Hox1, Hox2 is also extruded. (Right) The attracting force F_2 is stronger than F_1 , and the spring is further elongated.

The factor P in Equation (3) varies according to the cell position along the morphogen gradient in the anterior–posterior axis. P is high at the tail and low at the head of the embryo. The factor N represents the Hox cluster, which is negatively charged, as mentioned above. The physical force F pulls the Hox genes gradually to a region where gene activation is possible (Figure 2). These regions

(transcription factories) are studied in depth by Cook and collaborators [28]. In this model, the force F of Equation (3) depends on both macro (P) and micro (N) influences, and the Hox cluster behaves like an expanding elastic spring [17,18].

The intermediate dimensions are not included in formulating Equation (3), although they are important for the transition from single-cell expression to tissue development. For instance, Acemel et al. analyzed the bipartite chromatin structure of jawed vertebrate HoxD clusters. This architecture allows Hox genes to pattern disparate parts of the body, including limbs [29]. In the evolutionary process, each Hox gene of the vertebrate Hox cluster is endowed with specific features and a functional role. In this course, some of these genes are lost (deleted) (see Figure 1b). Even in the case of missing genes, the ordering remains ever-increasing like a ‘ratchet’. For instance, the Hox ordering (1, 2, . . . , 4, 5, . . .) may be observed, but the ordering (1, 2, . . . , 5, 4, . . .) is not allowed since it represents a gene reversal (See the Appendix A). This Hox gene ordering and the ontogenetic units of the embryo are located on a finite straight line with discrete ends (Figure 1b,c).

A phenomenon or object is self-similar if the image of the part is similar to the image of the whole. For example, the image of a coastline is the same, independent of the scale of the image. Similarly, the branching picture of a neuron is the same, independent of the magnification of the drawing. These phenomena or objects were coined ‘fractals’ by Mandelbrot, who was the first to study them consistently [30]. In this analysis, self-similarity is a continuous symmetry applying to all dimensional sizes (scale invariance). The next step is to incorporate the fractals in NT: self-similarity is the symmetry, and the conserved quantity is the complete Hox gene ordering (Figure 1a).

The genes of a Hox cluster and the embryonic body elements of the vertebrates are somehow ‘self-similar’ objects embedded in the same space, namely the one-dimensional finite straight line with distinct ends. The Hox gene collinearity is a ‘primitive’ self-similarity on a straight line since it applies to only two discrete dimensional scales (genetic and embryonic). In contrast, the proper self-similarity is a continuous symmetry applying to all scale dimensions. Therefore NT applies to the vertebrate Hox clusters where the primitive symmetry is linked to a partially conserved Hox gene ordering as in an irreversibly increasing ratchet with holes (Figure 1b). An inspection of the existing data on vertebrate Hox clusters confirms the structure of these orderings [8,31].

A fundamental question may be raised: Are the physical (and chemical) laws invariant when the scale changes? The answer is no [5]. The reason is that the basic constituents of matter (electrons, protons, and other elementary particles) have concrete and fixed material properties (mass, electric charge, spin, etc.). This ‘atomic’ nature of matter is not arbitrary. Therefore, it imposes its specific scale, so it affects the properties of macroscopic matter, such as the strength, elasticity, etc., of different materials.

The above property is equally true in biological phenomena. For instance, in the Reaction–Diffusion theory of Turing, patterns in space and time can be spontaneously created [32]. These patterns are scale-dependent since both Diffusion and Chemical kinetics have their inherent ‘atomic’ scales. Therefore the original genius theory of Turing cannot describe pattern regulation in Morphogenesis—the ability of embryos to adjust their pattern to variable sizes. To achieve pattern regulation, a supplementary mechanism to the Reaction–Diffusion theory must be introduced, see, e.g., [33].

4. Hox Gene Ordering in Rotationally Symmetric Animals

The above primitive collinearity applies to ‘directly’ developing organisms, such as arthropods or vertebrates. These organisms, in their very early embryogenesis, look like a miniature version of an adult animal. The major body elements are formed at once during embryogenesis along the A-P axis [34]. However, besides these directly developing animals, there is a large variety of invertebrates that, at the very early stages of embryogenesis, are ‘indirectly’ developing a larva, where the adult body shape is formed. This body plan differs from the initial A-P organization [34,35]. The powerful mechanism of evolution drives the creation of a new species by imposing a novel symmetry at the larva stage. For instance, twenty-nine hours after fertilization in the larva stage of Echinoderms, a rotational

symmetry is superimposed on the linear symmetry (A-P) of the initial embryo [35]. In Figure 3a (left), the larva of *Holopneustes purpuratus* is drawn with the morphology of five podia A-E located on a closed contour on top of the initial A-P axis [36].

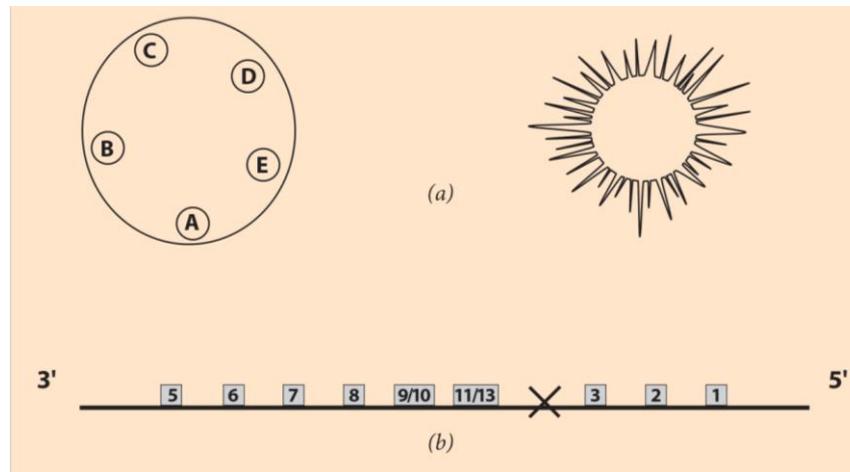


Figure 3. Developmental stages of the echinoderms. (a) (Left) *Holopneustes purpuratus* larva with A, B, C, D, E the podia locations (adapted from [36]). (Right) Juvenile sea urchin. (b) The Hox gene cluster ordering of the sea urchin. Hox4 is missing (hole), and the normal gene ordering is reshuffled.

The finite line with distinct ends is substituted by this rotationally symmetric contour [36]. In Figure 3a (left and right), a larva and an adult echinoderm are, respectively, depicted.

After the powerful new sequencing techniques developed in the last decade or so, many unexpected data have been discovered concerning the Hox gene ordering of the rotationally symmetric animals. For instance, the Hox gene cluster of the sea urchin was surprising (Figure 3b). The normally anterior class of genes (Hox1, Hox2, Hox3) are 'translocated and inverted' to the 5' end of the cluster, as shown in Figure 3b [37,38]. This reorganization is coined TAI, and many hypotheses have been put forward to explain its origin [31,38].

Vertebrates and the major deuterostome clades (Echinoderms included) have a supposedly common ancestor [8,31]. The vertebrates retain the ancestor's Hox gene ordering partially while the Echinoderms deviate radically from this common ancestor's Hox gene ordering [31,38]. Below, as a typical example, the case of the vertebrates will be compared to the Echinoderms Hox orderings.

According to the self-similarity hypothesis proposed here, without breaking the common ancestor's ordering of Hox genes, a bending of the linear arrangement brings Hox1 close to Hox13 (Figure 4, left).

In this figure, the 13 Hox genes are depicted in a circular ordering like a clock face where the clock 'hours' and the sequencing is self-similar to the rotationally symmetric larva body elements [37]. How this symmetry change is imposed remains to be clarified. After the establishment of the new symmetry, the circular contour has to be incorporated in the DNA sequence. The DNA fiber undergoes a double-strand break, shown in Figure 4 (middle). Such events are spontaneous, and they constitute the innovative force of evolution. This break enables two switching combinations to the endings Hox1 and Hox13 genes of the Hox cluster. Note that the number of Hox genes remains constant as in the common ancestor case (see, however, Section 6).

If Hox1 and Hox13 are attached to the 3' and 5' chromosome ends, respectively, no novel DNA sequence is created. This case could explain the Asteroid *planci* Hox cluster ordering. Although this Asteroid is rotationally symmetric, its Hox gene cluster ordering is comparable to the normal vertebrate ordering [37]. In contrast, a novel Hox gene sequence will be created if Hox1 and Hox13 are attached to the 5' and 3' ends of the flanking chromosome, respectively (Figure 4, right). If a subsequent break occurs at the level of Hox4, the reshuffling indicated in Figure 4 (right) leads to the Hox gene linear ordering of Figure 3b. At the location of the break, Hox4 is deleted. This resulting gene sequence is

the observed ordering of Hox genes in the sea urchin Hox cluster. According to NT, the linear Hox gene ordering of Figure 3b is the partially conserved quantity linked to the primitive self-symmetry of rotationally symmetric animals. Every Hox gene of the cluster has been endowed with a specific role at the larva stage, where the circular structure was led down.

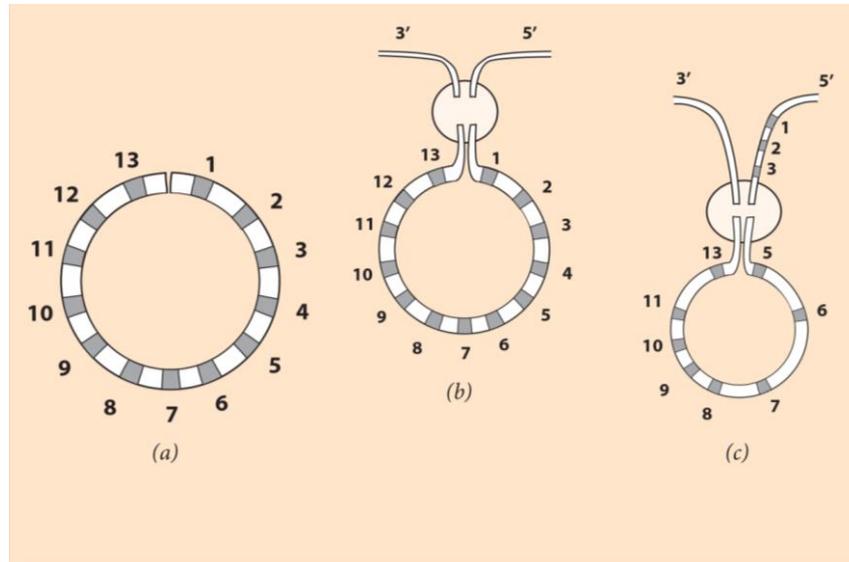


Figure 4. Circular ordering of Hoxgenes. (Adapted from S. Papageorgiou (2016) *Current Genomics* 17: 444–449) [37]. (a) The linear DNA is bent, and the two ends of the Hox cluster come close together. (b) In the encircled domain, the ends Hox1 and Hox13 of the cluster are connected to the 3' and 5' end of the flanking chromosome. If Hox1 is attached to the 3' end and Hox13 to the 5' end, the produced linear arrangement is the normal one and could represent the observed *A. planici* gene ordering. (c) If Hox5 is connected to the 3' end and Hox13 to Hox3 on the flanking chromosome, the linear ordering is the sea urchin Hox cluster arrangement, as shown in Figure 3.

5. Equivalence of Biophysical Model and Primitive Noether Theory

In vertebrates, both spatial and temporal collinearities are confirmed. According to BM, spatial collinearity (SC) proceeds gradually in coordination and entanglement with temporal collinearity (TC) [17,18]. In particular, several experiments were proposed to test further the space–time entanglement inherent in BM. Below, a BM prediction is compared to the corresponding NT prediction.

(A) According to BM and its elastic spring approximation for a Hox cluster, the centromeric end of the cluster is fastened at its neighboring DNA area. If the fastening domain is cut off, the fastening ability is reduced, and a weak force applied at the telomeric end of the cluster could oblige the cluster to slide abnormally toward the 3' direction [39]. It is, therefore, predicted that the elimination of only the centromeric domain neighboring the cluster would cause an abnormally premature activation of the cluster [39]. This experimental prediction is a variation of the Kondo and Duboule experiment, which had an unexpected outcome [40].

(B) According to NT, a primitive symmetry is endowed to the Hox cluster. This symmetry conveys to the cluster an invariant property, which consists of the irreversible ordering of the Hox genes where some genes are missing. This gene ordering remains the same even after the excision of the centromeric domain. However, other physical quantities, such as timing, may change in relation to the excision. It is expected that a weaker or tighter fastening of the Hox cluster will cause its premature or retarded activation, respectively. This result is consistent with the above case A of BM. The compatibility of A and B hints at a possible generalized equivalence of BM and the primitive NT. Additional experimental evidence is necessary to confirm further this equivalence.

When the fastening of a Hox cluster is very loose, a minimal pulling force suffices to make the whole cluster to slide in the 3' direction. In this spirit, one could explain the observed absence of temporal collinearity in *Drosophila* (see Appendix A.2). If the *Drosophila* Hox cluster is really loosely fastened, from the start of its activation, the whole cluster is translocated inside the transcription factory domain, and no TC is observed (Figure 2). Therefore, during Hox gene activation, only spatial collinearity is taking place. This means Hox genes are activated at the same time in all ontogenetic domains Ds. This daring assumption has to be confirmed. In contrast, vertebrates follow a quite different pathway of activation where both TC and SC are entangled.

6. Discussion and Conclusions

The evolutionary pathways differ in directly and indirectly developing animals. In the first case of direct development (e.g., vertebrates), paralogue Hox clusters with missing Hox genes cooperate to organize the different species. In indirect development, at some early embryonic stage, the larva develops so that the initial embryonic organization along the A-P axis switches to a circular form. The corresponding circular Hox cluster breaks at a certain gene location so that the cluster is incorporated in the flanking DNA sequence. It has been observed in another context that Hox gene loss is possible if 'the gene's role in development is lost or significantly modified during evolution [41].

According to NT and in analogy to the irreversible Hox gene ordering along a finite line, another conserved ordering should be expected for the ontogenetic units along the embryonic A-P axis. The Antennapedia mutation in *Drosophila* is a case of ordering violation between Hox genes and the resulting ectopic growth of middle legs instead of antennae [42] (see also Appendix A.2).

In rotationally symmetric animals, no paralog Hox clusters are formed, and the number of genes is complete except the gene at the breaking point. Several spatial and temporal features of the Hox gene expressions in echinoderms have already been reported [43]. A similar analysis for other rotationally symmetric animals with differing morphologies and Hox gene cluster orderings could be illuminating. It should be interesting to clarify if the Hox4 loss is the cause or the effect of a partially conserved gene ordering in the sea urchin.

A hypothesis was proposed according to which, during Hox gene activation, physical forces precede and organize the geometry of the Hox clusters, followed by the biomolecular interactions [44]. This prediction is in agreement with the conclusion that 'the data suggest the structural organization of the HoxD gene cluster may predate transcriptional activation' [20].

Krumlauf rightly noticed that, although significant progress has been made in exploring Hox gene collinearity, 'collinearity remains as intriguing today as it did when Ed Lewis first described this unique feature . . . ' [12]. The reason for this failure is probably due to the fact that all approaches up to now ignore the role of Noether's theory. As it is indicated here, the inclusion of NT in explaining HGC leads to the observed gene ordering in the finite (with two ends) linear vertebrate Hox clusters. Furthermore, NT can also explain Hox gene ordering in animals with different endless topologies and symmetries, as in the case of circular embryos (e.g., echinoderms).

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Conflicts of Interest: The authors declare no conflict of interest.

Appendix A. Some Epistemological and Philosophical Considerations

Appendix A.1. Noether Theory

Frank Wilczek, in his thought provoking book 'A beautiful question', defines symmetry in a compact form: an operation acting on a material body (or mechanism) causes a change. If this change

leaves the body invariant, the body is symmetrical to this change: “Change without change” [4]. Of course, many well-known experts have given their own definitions (see, e.g., [6]), but I think Wilczek’s proposition in the form of a short aphorism is more suitable to deal with Noether’s Theory.

Energy conservation is a dogma that was raised in the 19th century and is still not completely settled. It has never been tested accurately since all movements of material bodies are accompanied by friction (however small) and other small forces [4]. Therefore this dogma remains as an idealization of reality. The accurate NT is based only on the principles of uniformity of space- and time-invariance of physical laws in spatial translations and different times. All existing evidence (from the quanta realm to remote astrophysical observations) is consistent with the validity of this uniformity principle. Therefore, Wilczek argues, NT with its mathematical accuracy, guarantees energy conservation (See Section 1).

Appendix A.2. Hox Gene Collinearity

Comparing the orderings in Figure 1b,c, it is noticed that they are similar, and they differ only in the scale. It was concluded that this is a primitive self-similarity (Section 3). It is legitimate to proceed to the next step and assume that a primitive NT is applicable during Hox gene activation (transcription and expression). By inspection of all determined (up to now) orderings of Hox gene clusters of directly developing animals, the irreversibly increasing ratchet ordering is the semi-conserved quantity of the primitive NT, which differs from the complete ordering of Figure 1a. In this ordering, no gene reversal is allowed.

Considering the Antennapedia gene (a *Drosophila* Hox gene), it was noticed that the spontaneous mutation of this gene leads to the development of middle legs in the head instead of antennae. Such a mutation is a ‘gain of function’ [42]. The reverse mutation is also possible transforming middle legs to antennae. The latter case (coined ‘loss of function’) is recessive and lethal [42]. The above abnormal spontaneous mutations are caused by gene reversals, which are not allowed in normal development. The proposed partially conserved gene orderings are part of a primitive NT with the HGC being a manifestation of this theory, as applied to complex biosystems.

Appendix A.3. A ‘Light’ Philosophical Divertimento

An extension of NT to universal ontogeny is depicted in the following diagram.

Sunlight→Materialobjects→Shadow

This universal applicability of NT is related to an aesthetic metaphor: sunlight hits matter conveying a symmetry to the material bodies. Subsequently, this symmetry creates an indispensable shadow linked to these bodies. Light and shadow are metaphorically the symmetry and conserved quantity of NT.

The above metaphor is reminiscent of Plato’s *Politeia* (Republic) where Socrates narrates an allegory of the human bondage: human beings are chained like slaves in a cave and a fire behind them (a substitute of the sun) projects on a wall the shadows of people moving around in the cave. These shadows constitute for the chained prisoners the sole experiences of the world. This Platonic allegory has strongly influenced western philosophical thinking up to recent times [45]. It is as if this cognition is imprinted in the human intellectual faculty, and Noether Theory reflects a side of its mathematical content.

References

1. Lewis, E.B. A gene complex controlling segmentation in *Drosophila*. *Nature* **1978**, *276*, 565–570. [[CrossRef](#)] [[PubMed](#)]
2. Noether, E. Invariante Variationsprobleme. *Nachr. Ges. Wiss. Gött. Math. Phys. Klasse* **1918**, *1918*, 235–257.
3. Marinho, R.M. Noether’s theorem in Classical Mechanics revisited. *arXiv* **2006**, arXiv:physics/0608264v13. [[CrossRef](#)]

4. Wilczek, F. *A Beautiful Question*; PenguinBooks: New York, NY, USA, 2015.
5. Feynman, R.P. *Six Not-So-Easy Pieces*; Basics Books Perseus Books Group: New York, NY, USA, 2015.
6. Weyl, H. *Symmetry*; Princeton University Press: Princeton, NJ, USA, 1952.
7. Ellis, J.; Gaillard, M.K.; Nanopoulos, D.V. A Historical Profile of the Higgs Boson. *arXiv* **2012**, arXiv:1201.6045v01hep-ph.
8. Duboule, D. Rise and fall of Hox gene cluster. *Development* **2007**, *134*, 2549–2560. [[CrossRef](#)]
9. Gehring, W.J.; Kloter, U.; Suga, H. Evolution of the Hox gene complex from an evolutionary ground state. *Curr. Top. Dev. Biol.* **2009**, *88*, 35–61.
10. Ferrier, D.E. Evolution of Hox complexes. *Adv. Exp. Med. Biol.* **2010**, *689*, 91–100.
11. Tarchini, B.; Duboule, D. Control of Hoxd genes' collinearity during early limb development. *Dev. Cell* **2006**, *10*, 93–103. [[CrossRef](#)]
12. Krumlauf, R. Hox genes, Clusters and Collinearity. *Int. J. Dev. Biol.* **2018**, *62*, 659–663. [[CrossRef](#)]
13. Durston, A.J. Two-tier Hox gene collinearity mediates vertebrate axial patterning. *Front. Cell Dev. Biol.* **2018**, *4*, 102. [[CrossRef](#)]
14. Tschopp, P.; Tarchini, B.; Spitz, F.; Zakany, J.; Duboule, D. Uncoupling time and space in the collinear regulation of Hox genes. *PLoS Genet.* **2009**, *5*, e1000398. [[CrossRef](#)] [[PubMed](#)]
15. Papageorgiou, S. A physical force may expose Hox genes to express in a morphogenetic density gradient. *Bull. Math. Biol.* **2001**, *63*, 185–200. [[CrossRef](#)] [[PubMed](#)]
16. Papageorgiou, S. Pulling forces acting on Hox gene clusters cause expression collinearity. *Int. J. Dev. Biol.* **2006**, *50*, 301–308. [[CrossRef](#)] [[PubMed](#)]
17. Papageorgiou, S. Comparison of models for the collinearity of Hox genes in the developmental axes of Vertebrates. *Curr. Genom.* **2012**, *13*, 245–251. [[CrossRef](#)]
18. Papageorgiou, S. Physical Forces may cause the HoxD gene cluster elongation. *Biology* **2017**, *6*, 32. [[CrossRef](#)]
19. Noordermeer, D.; Leleu, M.; Schorderet, P.; Joye, E.; Chabaud, F.; Duboule, D. Temporal dynamics and developmental memory of 3D chromatin architecture at Hox gene loci. *eLife* **2014**, *3*, e02557. [[CrossRef](#)]
20. Fabre, P.J.; Benke, A.; Joye, E.; Huynh, T.H.N.; Manley, S.; Duboule, D. Nanoscale spatial organization of the HoxD gene cluster in distinct transcriptional states. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 13964–13969. [[CrossRef](#)]
21. Fabre, P.J.; Benke, A.; Manley, S.; Duboule, D. Visualizing the HoxD Gene Cluster at the Nanoscale Level. *Cold Spring Harb. Symp. Quant. Biol.* **2015**, *80*, 9–16. [[CrossRef](#)]
22. Towers, M.; Wolpert, L.; Tickle, C. Gradients of signalling in the developing limb. *Curr. Opin. Cell Biol.* **2012**, *24*, 181–187. [[CrossRef](#)]
23. Bökel, C.; Brand, M. Generation and interpretation of FGF morphogen gradients in vertebrates. *Curr. Opin. Genet Dev.* **2013**, *23*, 415–422. [[CrossRef](#)]
24. Carslaw, H.S.; Jaeger, J.C. *Conduction of Heat in Solids*; Oxford University Press: Oxford, UK, 1959.
25. Vargesson, N.; Kostakopoulou, K.; Drossopoulou, G.; Papageorgiou, S.; Tickle, C. Characterisation of Hoxa gene expression in the chick limb bud in response to FGF. *Dev. Dyn.* **2001**, *220*, 87–90. [[CrossRef](#)]
26. Shimizu, K.; Gurdon, J.B. A quantitative analysis of signal transduction from activin receptor to nucleus and its relevance to morphogen gradient interpretation. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 6791–6796. [[CrossRef](#)] [[PubMed](#)]
27. Simeoni, I.; Gurdon, J.B. Interpretation of BMP signaling in early Xenopus development. *Dev. Biol.* **2007**, *308*, 82–92. [[CrossRef](#)] [[PubMed](#)]
28. Papantonis, A.; Cook, P.R. Fixing the model for transcription: The DNA moves, not the Polymerase. *Transcription* **2011**, *2*, 41–44. [[CrossRef](#)]
29. Acemel, R.D.; Tena, J.J.; Irastorza-Azcarate, I.; Marlétaz, F.; Gómez-Marín, C.; de la Calle-Mustienes, E.; Bertrand, S.; Diaz, S.G.; Aldea, D.; Aury, J.M.; et al. A single three-dimensional chromatin compartment in amphioxus indicates a stepwise evolution of vertebrate Hox bimodal regulation. *Nat. Genet.* **2016**, *48*, 336–341. [[CrossRef](#)]
30. Mandelbrot, B.B. *The Fractal Geometry of Nature*; W.H. Freeman & Company: New York, NY, USA, 1982.
31. David, B.; Mooi, R. How Hox genes can shed light on the place of echinoderms among the deuterostomes. *Evol. Dev. Biol.* **2014**, *5*, 22–41. [[CrossRef](#)]
32. Turing, A.M. The chemical basis of Morphogenesis. *Philos. Trans. R. Soc. B* **1952**, *237*, 37–72.

33. Papageorgiou, S. A morphogen gradient model for pattern regulation: I. Formation of non-repetitive and repetitive structures. *Biophys. Chem.* **1980**, *11*, 183–190. [[CrossRef](#)]
34. Arenas-Mena, C.; Martinez, P.; Cameron, R.A.; Davidson, E.H. Expression of the Hox gene complex in the indirect development of a sea urchin. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 13062–13067. [[CrossRef](#)]
35. Mooi, R.; David, B. Radial symmetry, the anterior/posterior axis and echinoderm Hox genes. *Annu. Rev. Ecol. Evol.* **2008**, *39*, 43–62. [[CrossRef](#)]
36. Morris, V.B. Origins of radial symmetry identified in an echinoderm during adult development and the inferred axes of ancestral bilateral symmetry. *Proc. R. Soc. B* **2007**, *274*, 1511–1516. [[CrossRef](#)] [[PubMed](#)]
37. Papageorgiou, S. Hox Gene Collinearity: From A-P patterning to radially symmetric animals. *Curr. Genomics* **2016**, *17*, 444–449. [[CrossRef](#)] [[PubMed](#)]
38. Cameron, R.A.; Rowen, L.; Nesbitt, R.; Bloom, S.; Rast, J.P.; Berney, K.; Arenas-Mena, C.; Martinez, P.; Lucas, S.; Richardson, P.M.; et al. Unusual gene order and organization of the sea urchin Hox cluster. *J. Exp. Zool. B. Mol. Dev. Evol.* **2006**, *306*, 45–58. [[CrossRef](#)] [[PubMed](#)]
39. Papageorgiou, S. Abnormal elongation of Hox gene clusters may cause Cancer. *Front. Cell Dev. Biol.* **2018**, *6*, 25. [[CrossRef](#)]
40. Kondo, T.; Duboule, D. Breaking collinearity in the mouse HoxD complex. *Cell* **1999**, *97*, 407–417. [[CrossRef](#)]
41. Butts, T.; Holland, P.W.; Ferrier, D.E. Ancient homeobox gene loss and the evolution of chordate brain and pharynx development: Deductions from amphioxus gene expression. *Proc. Biol. Sci.* **2010**, *277*, 3381–3389. [[CrossRef](#)]
42. Gehring, W.J. *Master Control Genes in Development and Evolution*; Yale University Press: New Haven, CT, USA, 1999.
43. Byrne, M.; Martinez, P.; Morris, V. Evolution of a pentamer body plan was not linked to translocation of anterior Hox genes: The echinoderm HOX cluster revisited. *Evol. Dev.* **2016**, *18*, 137–143. [[CrossRef](#)]
44. Papageorgiou, S. Biophysics precedes Biochemistry in Hox Gene Collinearity. Available online: http://webmedcentralplus.com/article_view/405 (accessed on 23 July 2014).
45. Heisenberg, W. *Physics and Philosophy*; George Allen & Unwin Ltd.: London, UK, 1958.



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