



Proceedings

Preliminary Study for Detection of Hydrogen Peroxide Using a Hydroxyethyl Cellulose Membrane †

Helena Vasconcelos ^{1,2}, José M. M. M. de Almeida ^{2,4}, Cristina Saraiva ¹, Pedro A. S. Jorge ^{2,3} and Luis Coelho ^{2,*}

- School of Agrarian and Veterinary Sciences, CECAV-Centro de Ciência Animal e Veterinária, University of Trás-os-Montes e Alto Douro, 5001-801 Vila Real, Portugal
- ² INESC TEC—Technology and Science and Faculty of Sciences, University of Porto, Rua do Campo Alegre, 4169-007 Porto, Portugal
- ³ Department of Physics and Astronomy of Faculty of Sciences, University of Porto, 4169-007 Porto, Portugal
- Department of Physics, School of Science and Technology, University of Trás-os-Montes e Alto Douro, 5001-801 Vila Real, Portugal
- * Correspondence: lcoelho@inesctec.pt
- † Presented at the 7th International Symposium on Sensor Science, Napoli, Italy, 9-11 May 2019.

Published: 5 July 2019

Abstract: High concentration of biogenic amines (BA) is an indicator of deterioration of food and the determination of their concentration is an important method of food control. The hydrogen peroxide (H₂O₂) is a side product of the degradation of BAs by certain enzymes. It is presented an experimental technique grounded on chemiluminescence to measure small quantities of H₂O₂ with concentrations as low as 0.01%w/w up to 0.08%w/w. Luminol and cobalt hydroxide are added to hydroxyethyl cellulose to obtain an active membrane which will react with the sampling solution and the amount of total light emission is directly related to the H₂O₂ concentration.

Keywords: Biogenic amines; Chemiluminescence; Food storage

1. Introduction

Providing safe food to consumers has never been more important and more challenging for the food industry. In addition of being nutritious and appealing, food must be safe [1]. Deterioration is a complex process characterized by changes in food product that makes it unacceptable to be consumed [2]. However, microbial activity, the most common cause of deterioration, manifest as visible growth of slime, colonies and texture changes, due to polymer degradation, and development of off-odors and off-flavors [3]. High concentrations of biogenic amines (BAs) can be found as a consequence of microbial activity in foods such as wine, fermented meat and fish products, cheeses and fermented vegetables.

BAs are nitrogenous organic polar or semi-polar bases of low molecular weight. On the basis of their chemical structure these amines can be aliphatic (cadaverine, putrescine, spermine and spermidine), aromatic (phenylethylamine, tyramine), or heterocyclic (pyrrolidine, histamine) [4], and according to their number of amine groups they can be divided into monoamines (tyramine and phenylethylamine) and diamines (histamine, putrescine and cadaverine) [5]. Usually, these nitrogen compounds are formed mainly by microbial decarboxylation of amino acids and by amination and transamination of aldehydes and ketones.

Since the formation of BAs is used as an indication of food spoilage [6], the determination of their concentration in foods is an important method of food control [7]. As hydrogen peroxide (H₂O₂) is a side product of the degradation of BA by the enzyme diamine oxidase and produces

Proceedings 2019, 15, 7

luminescence upon reaction with 5-amino-2,3-dihydrophthalazine-1,4-dione (C₈H₇N₃O₂), known as luminol [8], it was used chemiluminescence as an indirect method for detection of H₂O₂.

In this study we present the preliminary development of a small membrane based on hydroxyethyl cellulose combined with luminol and a catalyst which glows in the presence of H₂O₂ with concentrations as low as 0.01%w/w.

2. Materials and Methods

The luminol-based membrane was developed with luminol, sodium phosphate, cobalt (II) chloride hexahydrate, sodium lauryl sulphate, hydrogen peroxide (30%) and hydroxyethyl cellulose (Sigma Aldrich, Darmstadt, Germany). The procedure established by Miklicanin and Valzacchi [8] was refined to establish the experiment protocol.

In a first stage luminol (0.2 mg), sodium phosphate (8.6 mg), sodium lauryl sulphate (60 μ L, 34.36 mmol/L) and hydroxyethyl cellulose (150 mg) was added to 10 mL of Milli-Q® water. The mixture was placed on a magnetic stirrer for 30 min and 1 mL was dispensed into home-made carrier, made of Teflon to decrease adhesion and facilitate removal of the membrane. The membranes were dried in an oven at 70 °C for 4 h then cooled and stored in a desiccator with vacuum.

A set of sample solutions with a concentration of 0.01 to 0.07%w/w of H_2O_2 were prepared by diluting a standard 30%w/w solution of H_2O_2 . To all sample solutions was added cobalt hydroxide (200 μ L, 5.0 mmol/L).

After drying, the membrane was carefully split into small pieces of the same size to increase the area of contact with the sample solution and placed inside a cuvette. The cuvette was placed on a small and portable acquisition system specially developed for this work containing a spectrometer (C12880MA from Hamamatsu) that collects part of the light emitted. The sample solution (250 $\mu L)$ was added and the emission spectra were acquired being recording at every 100 ms and the total light detected was integrated up to 50 min. The same procedure was repeated to all $\rm H_2O_2$ concentrations.

3. Results and Discussion

The light intensity variation emitted with a sample solution concentration of 0.01%w/w is represented in Figure 1a with a maximum peak of intensity at approximately 426 nm as expected [9]. A normalization of the data was carried out to eliminate fluctuations due to noise. The light emission spectra that characterize the reaction of peroxide with luminol over time, illustrated in Figure 1b, showed a clear difference between a concentration of 0.01% and 0.08% of H₂O₂.

It is evident that for a higher concentration, the peak of intensity is higher, and the reaction stays longer time. When the H_2O_2 concentration increases, it is observed a quasi-linear behavior of the total integral peak intensity, as shown in Figure 1c.

Although the data acquisition conditions are kept the same, some variability has been obtained in particular for higher concentrations. This can be justified because the light emission is not uniform, due to the unevenly rehydration of the membrane.

To verify the stability of the detection system the detection process was repeated four times for the same concentration. Figure 1d shows the relative data for three steps of different concentrations of H₂O₂. To check the stability and repeatability of the sensing scheme, similar samples were used to obtain several acquisitions. The system resolution, R, can be estimated considering the values obtained from two measurement linked with two different values of the concentration [10]

$$R = -\frac{\sigma}{S} = -\frac{\delta c}{\delta I}\sigma,\tag{1}$$

where the δc is the H₂O₂ concentration, S is the sensitivity to concentrations of H₂O₂ variations, δl is the difference in the total intensity of light detected and σ is the highest standard deviation associated with the light detection associated to a step variation of H₂O₂ concentration.

The resolutions obtained between levels 0.01 and 0.02%w/w; 0.02 and 0.03%w/w and 0.03 and 0.04%w/w were 0.0025, 0.00022 and 0.0008%w/w respectively.

Proceedings **2019**, 15, 7 3 of 4

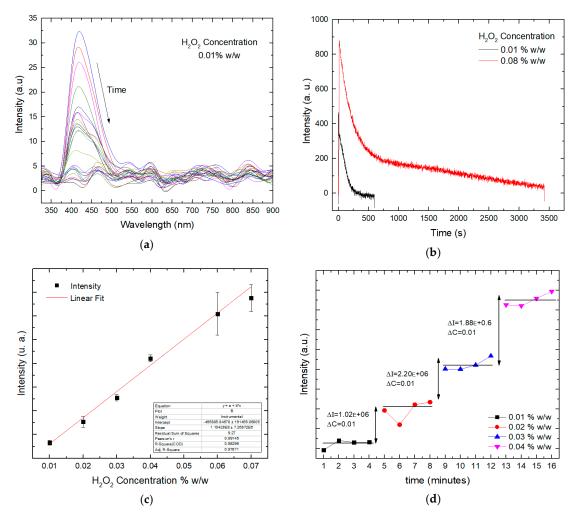


Figure 1. (a) Spectra showing the variation of the intensity of the light emission to the concentration 0.01%w/w as a function of the reaction time; (b) decay time of the spectra integral as a function of the chemical reaction; (c) integral of the decay time for each H_2O_2 concentration; (d) measured intensity when the H_2O_2 undertakes step increases.

4. Conclusions

A preliminary study for the detection of hydrogen peroxide using a hydroxyethyl cellulose membrane was presented. The results have shown that with this method it is possible to detect very low concentrations of H_2O_2 down to 0.01%w/w with resolutions better than 0.0025%w/w in the range up to 0.04%w/w. As the H_2O_2 concentration increased, an increase in light-emitting intensity and reaction time was observed following a quasi-linear behavior.

Funding: This work is financed by the North Portugal Regional Operational Programme (NORTE 2020), under the PORTUGAL 2020 Partnership Agreement, and through the European Regional Development Fund (ERDF). within projects «POCI-01-0145-FEDER-006961» and «Coral - Sustainable Ocean Exploitation: Tools and Sensors/NORTE-01-0145-FEDER-000036».

 $\label{lem:acknowledges} \textbf{Acknowledges} \ \ \text{financial support from FCT: SFRH/BD/120064/2016} \ \ \text{and from POCH program.}$

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Bohrer, B.M. Trends in Food Science & Technology Review: Nutrient density and nutritional value of meat products and non-meat foods high in protein. *Trends Food Sci. Technol.* **2017**, *65*, 103–112.

Proceedings 2019, 15, 7 4 of 4

2. Mariutti, L.R.B.; Bragagnolo, N. Influence of salt on lipid oxidation in meat and seafood products: A review. *Food Res. Int.* **2017**, *94*, 90–100.

- 3. Gram, L.; Ravn, L.; Rasch, M.; Bruhn, J.B.; Christensen, A.B.; Givskov, M. Food spoilage—Interactions between food spoilage bacteria. *Int. J. Food Microbiol.* **2002**, *78*, 79–97.
- 4. Cunha, S.C.; Lopes, R.; Fernandes, J.O. Biogenic amines in liqueurs: Influence of processing and composition. *J. Food Compos. Anal.* **2017**, *56*, 147–155.
- 5. Loizzo, M.R.; Menichini, F.; Picci, N.; Puoci, F.; Spizzirri, U.G.; Restuccia, D. Technological aspects and analytical determination of biogenic amines in cheese. *Trends Food Sci. Technol.* **2013**, *30*, 38–55.
- 6. Sørensen, K.M.; Aru, V.; Khakimov, B.; Aunskjær, U.; Engelsen, S.B. Biogenic amines: a key freshness parameter of animal protein products in the coming circular economy. *Curr. Opin. Food Sci.* **2018**, *22*, 167–173.
- 7. Erim, F.B. Recent analytical approaches to the analysis of biogenic amines in food samples. *TrAC Trends Anal. Chem.* **2013**, *52*, 239–247.
- 8. Omanovic-Miklicanin, E.; Valzacchi, S. Development of new chemiluminescence biosensors for determination of biogenic amines in meat. *Food Chem.* **2017**, *235*, 98–103.
- 9. Lee, J.; Seliger, H.H. Spectral Characteristics of the Excited States of the Product of the Chemiluminescence of Luminol. *Photochem. Photobiol.* **1970**, *11*, 247–258.
- 10. Díaz-Herrera, N.; González-Cano, A.; Viegas, D.; Santos, J.L.; Navarrete, M.C. Refractive index sensing of aqueous media based on plasmonic resonance in tapered optical fibres operating in the 1.5 μm region. *Sens. Actuators B Chem.* **2010**, *146*, 195–198.



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).