

Electroactive Nanocarbon as Novel Label for DNA Analysis[†]

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[†] Presented at the 7th International Symposium on Sensor Science, Napoli, Italy, 9–11 May 2019.

Published: 31 July 2019

Abstract: We use here a carbon nanomaterial as the electroactive label for the development of a low-cost and easy-to-miniaturize non-optical polymerase chain reaction (PCR). To this aim, graphene oxide nanocolloids (GONCs) were first conjugated to the DNA primers either by physical adsorption or by the formation of a covalent bond. Subsequently, GONC-modified primers were hybridized to their complementary counterpart to obtain GONC-modified double stranded DNA (GONC-dsDNA). The electrochemical signal, due to the reduction of the electrochemically reducible oxygen functionalities on GONC surface was then measured and correlated to the concentration of GONC-dsDNA.

Keywords: electroactive nanocarbon; nano-graphene oxide; DNA analysis; electrochemistry

1. Introduction

Among the several chemically modified graphene materials, nano-graphene oxide has attracted vast attention from many research fields owing to its exceptional properties (eg. good conductivity, high surface area, possibility of functionalization, intrinsic electroactivity and fluorescence properties) that it possesses [1–6]. Nano-graphene oxide material consists of highly oxidized two-dimensional monoatomic sheets of large surface area, with different oxygen functionalities namely epoxy, peroxy, hydroxyl, carbonyl, and carboxyl groups [1,5]. Existing studies show that graphene oxide possesses an intrinsic electroactivity due to the presence of electrochemically reducible oxygen functionalities on its surface [6]. Specifically, a significant reduction peak can be observed at around −1.2 V when the potential is cathodically scanned. In this work, we employ graphene oxide nanocolloid (GONC) materials as a label for DNA detection. The abundance of electrochemically reducible oxygen containing groups (OCGs) on the material surface, together with the nanoscale size of the colloids make GONC the ideal material for cost-effective and sensitive labelling.

2. Materials and Methods

Graphene Oxide Nanocolloids (GONC), sodium chloride, sodium diphosphate basic monohydrate, *N*-(3-dimethylaminopropyl)-*N*'-ethylcarbodiimide hydrochloride (EDC), 2-(*N*-Morpholino) ethanesulfonic acid (MES), *N*-hydroxysulfosuccinimide sodium (sulfo-NHS), were obtained from Sigma Aldrich (Singapore). DNA oligonucleotides used in this work were purchased from Integrated DNA Technologies, Inc. (Singapore) and their sequences were the following: 5'-GCTCC TACAAATGCCATCA-3' (probe); 5'-TGATGGCATTGTAGGAGC-3' (target); for the covalent conjugation with GONCs, amino-modification of DNA sequences 5'-end has been carried out. Covalent modification of DNA sequences with GONC has been carried out according to previous

literature [7]. A similar protocol has been followed for the physical adsorption protocol by skipping the covalent conjugation step. Calibration curves of increasing concentrations of GONC (0.01 mg/mL–1 mg/mL) and GONC-DNA conjugates (0.0125 mg/mL–0.2 mg/mL) were performed by dropcasting 3 μ L of the sample on the electrode surface and letting it dry before measuring the electrochemical signal. Hybridisation of DNA primer sequences was conducted by incubating the GONC modified DNA sequences with the complementary target at a concentration of 0.48 μ M, at 42°C for 40 minutes. After centrifugation, three washing steps were carried out in PBS buffer (pH 7.4).

3. Results

As part of material characterization, cyclic voltammetry was performed to investigate the presence of OCGs on GONC surface and the intrinsic electroactivity of the material. As shown in Figure 1, a significant reduction peak at about 1.15 V was observed during the cathodic scan of GONC, which arose from the reduction of electrochemically active OCGs present on GONC surface. The obtained reduction signal confirmed the suitability of the material for the current study.

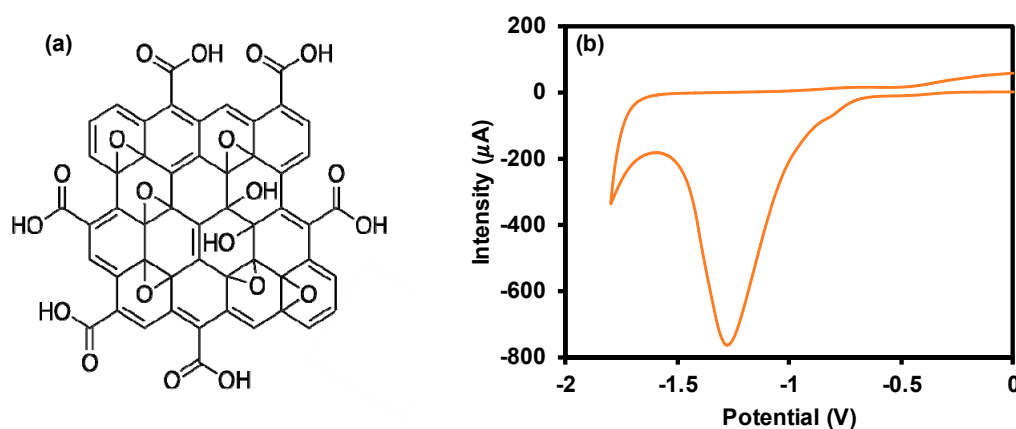


Figure 1. Graphene oxide nanocolloid (GONC) structure (a) and electrochemical reduction peak (b). Conditions: scan rate 50 mV; 0.1 M PBS, pH 7.0.

Moving on, GONC material was conjugated to specific DNA sequences following two different protocols namely physical adsorption and covalent conjugation. After which, the GONC modified DNA probes were hybridized with the corresponding complementary target sequences in order to obtain a GONC-dsDNA conjugate. The electrochemical signal obtained from different concentrations of the GONC-dsDNA conjugates synthesized by following the two conjugation protocols was then measured and compared with that obtained from the bare GONCs. Results from this study are shown in Figure 2. Figure 2a depicts the calibration curve that shows the peak heights obtained by differential pulse voltammetry (DPV) measurements for increasing concentrations of GONCs. Figure 2b shows the peak heights from the reduction peaks for increasing concentrations of GONC-dsDNA conjugate obtained with the physical adsorption protocol, while Figure 2c shows the calibration curve obtained by increasing concentrations of GONC-dsDNA conjugate from the covalent conjugation.

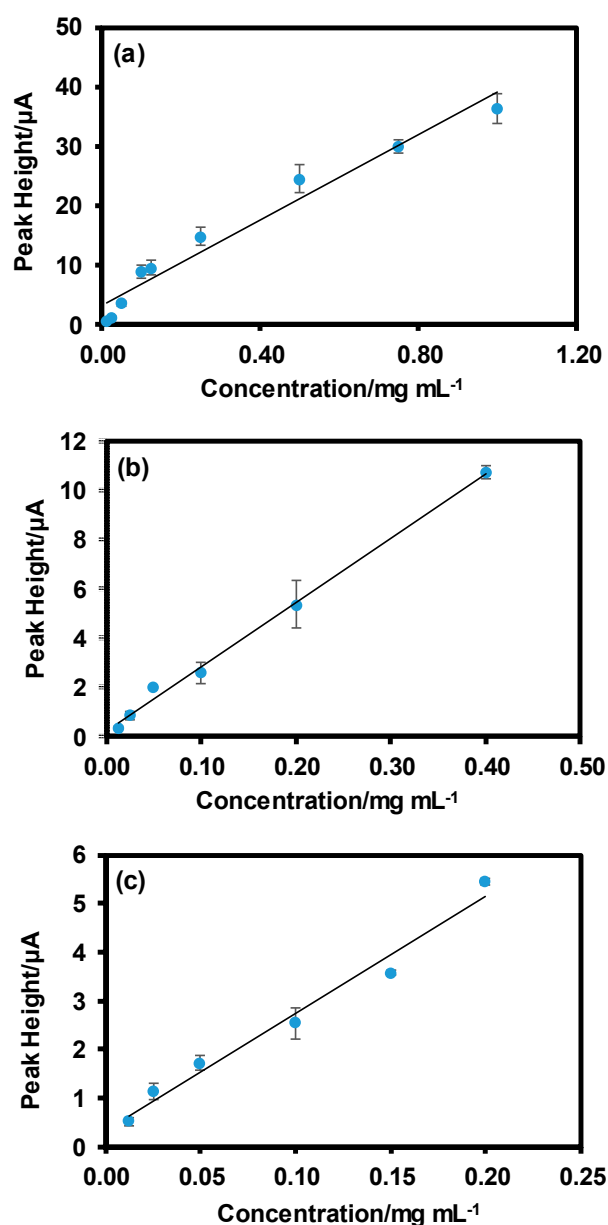


Figure 2. Calibration curve depicting the signal intensity versus concentration for: (a) Bare GONC material; (b) GONC-dsDNA conjugate obtained by physical adsorption; (c) GONC-dsDNA conjugate obtained by covalent conjugation.

4. Discussion

In this preliminary study we showed how electrochemically active nano-graphene oxide material could be successfully employed for DNA labelling. As shown from the calibration curves depicted in Figure 2, GONC material maintains a significant electroactivity after the conjugation with the DNA probes and hybridization with complementary targets, allowing the final quantitative analysis of DNA. This system could be eventually integrated in a PCR protocol, with the use of GONC-modified DNA primers that would allow an electrochemical detection and provide a cost-effective alternative to the current optical PCR.

Acknowledgments: The authors gratefully acknowledge a Ministry of Education (MOE), AcRF Tier 1 grant (Reference No: RG18/17) for the funding of this research.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

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