



Editorial

New Strategies to Increase Fish Larval Physiological Responses against Changes in Water Temperature

Carmen Navarro-Guillén

Instituto de Ciencias Marinas de Andalucía (ICMAN-CSIC), 11519 Puerto Real, Spain; carmen.navarro@csic.es

Aquaculture is probably the fastest-growing food-producing sector, accounting for nearly 50 percent of the world's food fish. However, efforts are needed to improve efficiency and to assure the sustainability of the aquaculture sector. Climate change is now considered a challenge to the global food chain and a major threat to fish production [1]. The increase in global temperature will affect offshore and inland aquaculture production, testing the resilience of domesticated fish species. Temperature plays a critical role in the growth and development of aquatic animals [2]. As ectotherms, fish may particularly be sensitive to temperature variations. Increasing temperatures affect fish physiology and behavior through chronic stress and increases in metabolic demands, in addition to having an evident impact on nutrient utilization and growth [3,4].

Aquaculture production output relies, to a high degree, on larviculture efficiency. In marine hatcheries, high mortalities are normally observed in larval stages in optimized farming conditions (70–80%), thus affecting fish production. The consequences and implications of variations in those conditions, such as an increase in temperature, are an important challenge for the future. Thus, understanding the mechanisms that control early development and growth and their relationship with abiotic factors is critical for the identification of strategies promoting climate-resilient adaptations from early stages.

In hearing larvae (*Clupea harengus*), elevated temperatures significantly affected the larval growth and swimming kinematics. Larvae reared at the highest temperature showed the lowest growth. This is probably because increasing temperature reduces the conversion of energy to somatic growth by increasing metabolic demand. Concomitantly, swimming behavior was also affected by temperature owing to changes in growth rates (i.e., larger larvae swim faster) [5]. For common sole (*Solea solea*), a flatfish species with a strong metamorphosis during the larval stage, temperature promoted growth, since metamorphosis at 19 °C started earlier than at 16 °C. However, by the end of metamorphosis, larvae from the 19 °C treatment were shorter and presented incidences of metamorphosis defects compared with those reared at lower temperatures [6]. Similarly, for gilthead seabream (*Sparus aurata*), exposure to high temperatures during early development led to lower fish swimming performance and promoted upper-jaw abnormalities [7].

Since all the results described above present potential threats to aquaculture efficiency and product quality, exploring how to promote climate-resilient adaptations in farmed fish is a key strategy for helping to attain food security under changing environmental conditions. The dietary inclusion of additives with high biological activity may enhance the growth performance and welfare of farmed fish species. The inclusion of grape seed extract in diets for Senegalese sole (*Solea senegalensis*) promoted the activity of antioxidant enzymes and decreased oxidative damage after thermal acute stress [8]. A protective role against thermal stress has been also described for dietary taurine. For heat stress, taurine supplementation led to increased HSP70 gene expression levels and C4 complement levels in tiger puffer (*Takifugu rubripes*) [9], while improved growth performance, oxidative system, and innate immunity of olive flounder (*Paralichthys olivaceus*) and pufferfish (*Takifugu obscurus*) were observed under low-temperature stress [10,11]. Other dietary interventions, such as modification in the concentration of specific macronutrients, may play an important



Citation: Navarro-Guillén, C. New Strategies to Increase Fish Larval Physiological Responses against Changes in Water Temperature. Fishes 2023, 8, 299. https://doi.org/ 10.3390/fishes8060299

Received: 24 May 2023 Accepted: 31 May 2023 Published: 3 June 2023



Copyright: © 2023 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

Fishes 2023, 8, 299 2 of 2

protective role and improve the thermal resilience of aquaculture species. For example, in barramundi (*Lates calcarifer*), a high-fat diet increases juvenile growth, while, by contrast, it reduces fish tolerance to extreme water temperatures [12]. On the other hand, in sea bass (*Dicentrarchus labrax*), feeding a high-fat diet to fish acclimatized to warmer waters has been established as good practice for improving growth [13]. Although the abovementioned studies boost aquaculture resilience to cope with challenging temperatures, most of them were performed on juveniles. To improve production output, studies are also needed on larvae.

New effective strategies promoting fish robustness and resilience from early developmental stages are needed to enable the development of resilient food production systems and ensure sustainable food security in the face of climate change.

Funding: CN-G was supported by the researcher contract DOC_01203 funded by the Andalusian Plan for Research Development and Innovation (PAIDI 2020).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data sharing not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Maulu, S.; Hasimuna, O.J.; Haambiya, L.H.; Monde, C.; Musuka, C.G.; Makorwa, T.H.; Munganga, B.P.; Phiri, K.J.; Nsekanabo, J.D. Climate Change Effects on Aquaculture Production: Sustainability Implications, Mitigation, and Adaptations. *Front. Sustain. Food Syst.* **2021**, *5*, 609097. [CrossRef]

- 2. Ngoan, L.D. Effects of climate change in aquaculture: Case study in Thua Thien Hue Province, Vietnam. *Biomed. J. Sci. Tech. Res.* **2018**, *10*, 2018. [CrossRef]
- Stewart, H.A.; Aboagye, D.I.; Ramee, S.W.; Allen, P.J. Effects of acute thermal stress on acid-base regulation, haematology, ion-osmoregulation and aerobic metabolism in Channel Catfish (*Ictalurus punctatus*). Aquac. Res. 2019, 50, 2133–2141. [CrossRef]
- 4. Navarro-Guillén, C.; Gilannejad, N.; Pérez-Hilario, D.; Martínez-Rodríguez, G.; Yúfera, M. Gut transit of daily consecutive meals in greater amberjack juveniles reared at different temperatures. *Aquaculture* **2023**, *567*, 739244. [CrossRef]
- 5. Allan, B.J.M.; Browman, H.I.; Shema, S.; Skiftesvik, A.E.; Folkvord, A.; Durif, C.M.F.; Kjesbu, O.S. Increasing temperature and prey availability affect the growth and swimming kinematics of Atlantic herring (*Clupea harengus*) larvae. *J. Plankton Res.* **2022**, 44, 401–413. [CrossRef] [PubMed]
- 6. Sardi, A.E.; Bégout, M.L.; Lalles, A.L.; Cousin, X.; Budzinski, H. Temperature and feeding frequency impact the survival, growth, and metamorphosis success of *Solea solea* larvae. *PLoS ONE* **2023**, *18*, e0281193. [CrossRef] [PubMed]
- 7. Kourkouta, C.; Printzi, A.; Geladakis, G.; Mitrizakis, N.; Papandroulakis, N.; Koumoundouros, G. Long lasting effects of early temperature exposure on the swimming performance and skeleton development of metamorphosing gilthead seabream (*Sparus aurata* L.) larvae. *Sci. Rep.* **2021**, *11*, 8787. [CrossRef] [PubMed]
- 8. Xavier, M.J.; Conceição, L.E.C.; Valente, L.M.P.; Colen, R.; Rodrigues, A.C.M.; Rocha, R.J.M.; Custódio, L.; Carballo, C.; Manchado, M.; Engrola, S. Dietary Natural Plant Extracts Can Promote Growth and Modulate Oxidative Status of Senegalese Sole Postlarvae under Standard/Challenge Conditions. *Animals* **2021**, *11*, 1398. [CrossRef] [PubMed]
- 9. Shi, L.; Zhao, Y.; Zhou, J.; Zhai, H.; Wei, L.; Liu, Z.; Wang, W.; Jiang, Z.; Ren, T.; Han, Y. Dietary taurine impacts the growth, amino acid profile and resistance against heat stress of tiger puffer (*Takifugu rubripes*). Aquacult. Nutr. 2020, 26, 1691–1701. [CrossRef]
- 10. Cheng, C.H.; Guo, Z.X.; Wang, A.L. The protective effects of taurine on oxidative stress, cytoplasmic free-Ca(2+) and apoptosis of pufferfish (*Takifugu obscurus*) under low temperature stress. *Fish Shellfish Immun.* **2018**, 77, 457–464. [CrossRef] [PubMed]
- 11. Kim, J.-M.; Malintha, G.H.T.; Gunathilaka, G.L.B.E.; Lee, C.; Kim, M.-G.; Lee, B.-J.; Kim, J.-D.; Lee, K.-J. Taurine supplementation in diet for olive flounder at low water temperature. *Fish. Aquatic Sci.* **2017**, 20, 20. [CrossRef]
- 12. Gomez Isaza, D.F.; Cramp, R.L.; Smullen, R.; Glencross, B.D.; Franklin, C.E. Coping with climatic extremes: Dietary fat content decreased the thermal resilience of barramundi (*Lates calcarifer*). *Comp. Biochem. Phys. A* **2019**, 230, 64–70. [CrossRef] [PubMed]
- 13. Katsika, L.; Huesca Flores, M.; Kotzamanis, Y.; Estevez, A.; Chatzifotis, S. Understanding the Interaction Effects between Dietary Lipid Content and Rearing Temperature on Growth Performance, Feed Utilization, and Fat Deposition of Sea Bass (*Dicentrarchus labrax*). *Animals* **2021**, *11*, 392. [CrossRef] [PubMed]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.