

Table S1 Reaction system conditions for genomic DNA removal.

Reagent	Amount
5 ×g DNA Eraser buffer	2.0 µl
gDNA Eraser	1.0 µl
Total RNA	800 ng
RNase-free dH ₂ O	up to 10 µl

Table S2 Reverse transcription reaction system.

Reagent	Amount
Reaction mixture	10.0 µl
PrimeScriptRT enzyme Mix I	1.0 µl
RT primer Mix	1.0 µl
5× PrimeScriptBuffer 2 (for real time)	4.0 µl
Rnase-free dH ₂ O	4.0 µl
Total concentration	20 µl

Table S3 Reverse transcription PCR program.

Temperature (°C)	Time
37	15 min
85	5 s
Save at 4 ° C	

Table S4 Summary of the sequencing data of *H. erectus*.

Group	Control group	HM-group t	MP-group
Number of original sequences (n)	49 556 386	49 404 042	48 043 100
Number of sequences after filtering (n)	47 880 348	47 987 534	46 520 778
Number of bases after filtration (bp)	7 182 052 200	7 198 130 100	6 978 116 700
Q30 (%)	94.03	94.08	94.32

Table S5 Assembly results.

Length range (bp)	Trinity	Unigene
200–600	33 659	22 513
600–1 000	11 264	5 247
1 000–2 000	19 264	6 887
> 2 000	22 525	7 617
Count	86 712	42 264
Minimum length	201	201
Maximum length	18 307	18 307
Mean length	1 443.25	1 122.71
N50	2 485	2 223
N90	646	406

Table S6 Summary of comments in each database.

Database	Number of single gene clusters	Percentage (%)
Non-redundant protein database (Nr)	22 671	53.6
Nucleic acid sequence database (Nt)	18 801	44.5
Tokyo Encyclopedia of Genes and Genomes (KO)	12 046	28.5
Protein sequence database (Swiss-Prot)	18 276	43.2
Protein family domain database (Pfam)	14 248	33.7
Gene Ontology Data (GO)	19 506	46.2
Gene evolutionary lineage: unsupervised orthologous group (eggNOG)	17 228	40.8
Clustering of Eukaryotic Protein Adjacent Classes (KOG)	15 454	36.6
Total single gene cluster (Total unigenes)	42 264	100

Appendix S1. Preparation of MP bait

Attachment of MPs to mysid

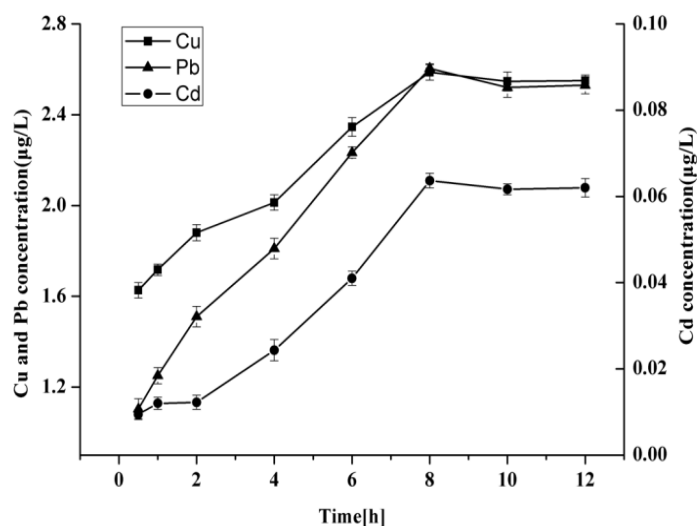
After thawing the frozen mysid shrimp and washing the intact individuals, 10 g of mysid was weighed, and 20 mg of MPs was added and thoroughly mixed. Thereafter, the mixture was poured into a glass jar containing 60 L of seawater, and 1 g of mysid sample was collected at 0, 0.5, 1, 1.5, and 2 h at 25–26 °C. The attachment of MPs to mysid was determined via using ultrasonic cleaning to elute the MPs particles attached to the mysid individuals, and the MPs were counted under the microscope to determine the number of MPs particles attached to a unit mysid.

Heavy metal MPs preparation

MPs (0.10 g) were washed sequentially with 2% HNO₃ and ultrapure water, and placed in a 4 L glass jar containing 3 L of filtered seawater, which was covered with aluminum foil. After 36 h aeration at 25–26 °C when all MPs had been fully infiltrated and even distributed in the seawater, a mixture of 0.05 mg/L Cu, 0.01 mg/L Cd and 0.05 mg/L Pb was added, followed by aeration to facilitate the attachment of the heavy metal ions to MPs. At the same time 50–200 MPs were samples at 0.5, 1, 2, 4, 6, 8, and 10 h, and the MPs within the water samples were collected through a 1- μ m nitrocellulose filter membrane (Shanghai Xinya Purification Material Factory, China). The concentrations of Cu, Cd, and Pb ions on the surface of MPs on the filter membranes was determined using a PE NexION 300X inductively coupled plasma emission mass spectrometer (ICP-MS). The saturated time and concentration of each heavy metal by microplastic attached were measured.

Results

This result indicated that the attachment of heavy metals to the surface of the MPs saturated after 8 h at 25–26 °C. Based on the pre-laboratory basis it is known that there was a gradual increase in the concentrations of heavy metals attached to the surface of MPs with increasing aeration time, peaking 8 h, and plateauing afterwards. The adsorption of centrally located shrimp by MPs leveled off after 1.5 h, when the number of MPs on the bait was 1600/g.



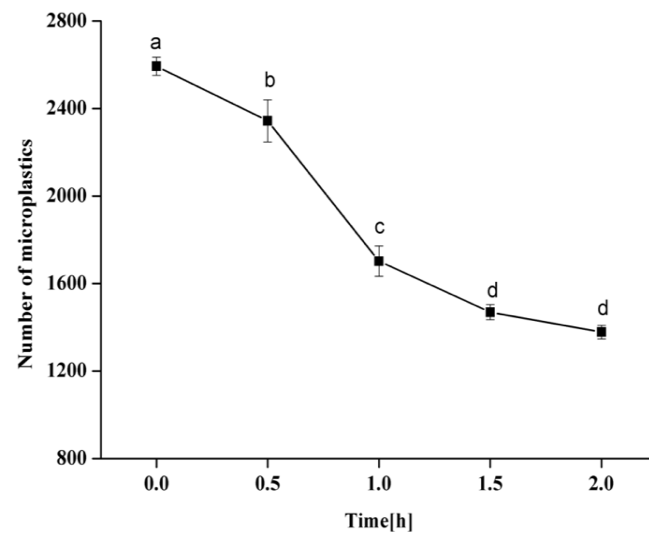


Figure. The attachment dynamics of heavy metal(a) and mysid to PE microplastics(b)
Different letters indicate significant differences at $p < 0.05$.