



Article

Growing Degree Day Targets for Fruit Development of Australian Mango Cultivars

Marcelo H. Amaral ^{1,*}, Cameron McConchie ¹, Geoffrey Dickinson ² and Kerry B. Walsh ^{1,*}

¹ Institute for Future Farming Systems, Central Queensland University, Rockhampton, QLD 4701, Australia; cmaxmail@bigpond.com

² Department of Agriculture and Fisheries, Mareeba, QLD 4880, Australia; g.dickinson@cqu.edu.au

* Correspondence: m.m.amaral@cquemail.com (M.H.A.); k.walsh@cqu.edu.au (K.B.W.)

Abstract: A forward estimate of mango (*Mangifera indica* L.) harvest timing is required for farm management (e.g., for organization of harvest labour and marketing). This forward estimate can be based on accumulated growing degree days (GDD) from an early stage of flowering to fruit harvest maturity, with fruit maturity judged on a destructive assessment of flesh colour and dry matter content. The current study was undertaken to improve GDD targets for Australian mango cultivars, to improve estimation of harvest maturity, and to document a methodology recommended for future work characterizing fruit maturation GDD for other mango cultivars. An alternate algorithm on GDD calculation involving use of a function that penalizes high temperatures as well as low temperatures was demonstrated to better predict harvest maturity in warmer climates. Across multiple locations and seasons, the required heat units (GDD, $T_b = 12\text{ }^\circ\text{C}$, $T_B = 32\text{ }^\circ\text{C}$; where T_B is upper base temperature of $32\text{ }^\circ\text{C}$ and T_b is lower base temperature of $12\text{ }^\circ\text{C}$) to achieve maturity from asparagus stage of flowering was documented as 2185, 1728, and 1740 for the cultivars Keitt, Calypso and Honey Gold, respectively. GDD difference between the asparagus and two-thirds floral opening stages of flowering was 188 ± 18 for Calypso, 184 ± 12 for Honey Gold, 238 ± 21 for Keitt and 175 ± 10 for KP. Colour specifications for a colour card set suitable for maturity assessment of all cultivars was also proposed. A flesh colour harvest maturity card specification of 9 was proposed for the cultivar Honey Gold and 13 for the cultivar Keitt.

Keywords: growing degree days; flesh colour; temperature monitoring



Citation: Amaral, M.H.; McConchie, C.; Dickinson, G.; Walsh, K.B.

Growing Degree Day Targets for Fruit Development of Australian Mango Cultivars. *Horticulturae* **2023**, *9*, 489. <https://doi.org/10.3390/horticulturae9040489>

Academic Editors: Riccardo Lo Bianco, Roberto Massenti and Antonino Pisciotta

Received: 22 March 2023

Revised: 7 April 2023

Accepted: 9 April 2023

Published: 13 April 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

1.1. GDD

A forward estimate of mango (*Mangifera indica* L.) harvest timing is required for farm management. For example, a forward estimate of harvest date is required several months before harvest for organization of harvest resourcing, including hire of harvest labor, order of packing materials, and transport. Harvest time forecast is also essential to market planning, with longer lead time required for longer supply chains (e.g., export of Australian mango requires booking of fruit fly treatment facilities and shipping).

The first use of temperature in forecasts of mango fruit harvest maturity involved a recommendation of 1000 h above $17.9\text{ }^\circ\text{C}$ (Oppenheimer, 1947; as cited in Diczbalis et al. [1]). Subsequently, cultivar specific growing degree day (GDD) requirements have been established for mango reproductive development from a given stage of flowering to harvest maturity (Table 1). The GDD calculation involves summation across days of the average of daily minimum and maximum temperature minus a ‘minimum base temperature’ (T_b), which assumes fruit development halts below this temperature, and can involve penalty for temperatures above a ‘maximum base temperature (T_B), which assumes fruit development slows at high temperatures (see calculations in Section 1.4). A forecast of harvest

date from a given date of flowering is typically based on a historical record of daily minimum and maximum temperatures, updated with current season temperature data as the season progresses.

Table 1. Required heat units for fruit maturation, including upper (TB) and lower base temperature (Tb) utilized for calculation of GDD in each study, from the reproductive stages of asparagus, Christmas tree stage (2/3 flowers open, with 1/3 still not open on panicle tip) and fruit set. n/a is not applicable. The sensor location ‘adjacent’ refers to sensor placement adjacent to the orchard block.

Location	Cultivar	Tb (°C)	TB (°C)	Reproductive Stage	Heat Units	Temperature Sensor Location
Australia [1]	Kensington Pride	12	n/a	asparagus	1600	inside canopy
Australia [2]	Calypso	12	n/a	asparagus	1680	on farm
Australia [3]	Calypso	10	n/a	Christmas tree	1640	adjacent
Australia [2]	Honey Gold	12	n/a	asparagus	1800	on farm
Australia [4]	Honey Gold	12	n/a	Christmas tree	1500	adjacent
Australia [2]	R2E2	12	n/a	asparagus	1800	on farm
Brazil [5]	Tommy Atkins	13	32	Christmas tree	1428	adjacent
Brazil [6]	Tommy Atkins	13	32	fruit set	1158	adjacent
Brazil [7]	Alfa	10	n/a	Christmas tree	2117	1 km from farm
Brazil [8]	Roxa	10.6	n/a	pea size fruit	1710	n/a
Brazil [9]	Uba	10	n/a	bud swelling	2399	n/a
Mexico [10]	Tommy Atkins	10	n/a	Christmas tree	1600	inside canopy
Mexico [10]	Keitt	10	n/a	Christmas tree	2100	inside canopy
Mexico [10]	Kent	10	n/a	Christmas tree	1800	inside canopy
Mexico [10]	Ataulfo	10	n/a	Christmas tree	1600	inside canopy
India [11]	Alphonso	10	n/a	fruit set	1867	n/a
India [11]	Alphonso	17.9	n/a	fruit set	919	n/a
India [12]	Kesar	17.9	n/a	fruit set	1020	n/a

References: Diczbalis et al. [1], Moore [2], Hofman et al. [3], Winston et al. [4], Castro et al. [5], Rodrigues et al. [6], Barros et al. [7], Callejas et al. [8], Lemos et al. [9], Osuna-Garcia [10], Zagade et al. [11], Halepotara et al. [12].

GDD recommendations made to the Australian industry diverge in both the flowering stage used and the Tb value, causing some confusion in grower usage. For example, for cultivar Calypso, Moore [2] recommended 1680 GDD using a base temperature (Tb) of 12 °C from asparagus stage, while Hofman et al. [3] suggested the use of 1640 GDD on Tb of 10 °C from Christmas tree stage. Limitations to this previous work are discussed in the following sections.

1.2. Estimating Harvest Time from Flowering

Most mango GDD recommendations have been based on an ‘eyeball’ estimation that most panicles in the orchard are at a given reproductive stage, and that the fruit is ready for commercial harvest. The former estimate has a qualitative element, and the latter estimate is subject to variation on commercial and agronomic grounds. Further, existing recommendations on GDD requirements for mango maturation have generally involved work in a single season, without validation across seasons and growing conditions. As such there is a level of uncertainty in these recommendations.

For example, of the Australian work, Moore [2] based GDD recommendations on work involving tagging of trees at ‘early’ and ‘late’ flowering. Hofman et al. [3] relied on an orchard wide estimate of flowering stage in setting GDD for cultivar Calypso, with values from 1300 to 1820 units recorded at different sites and seasons. Values of the Northern Territory (NT) Australia sites only were averaged to achieve what is now an industry accepted GDD target for this cultivar (1640 units from asparagus stage on a Tb = 10 °C). Similarly, the Winston et al. [4] recommendation of 1500 units for cultivar Honey Gold fruit development was based on grower estimates of when flowering across the orchard was, on average, at Christmas tree stage.

The use of whole tree or orchard assessments of flowering and commercial harvests is convenient, but a more accurate estimate of the GDD target should be achieved by tracking of individual fruit from flowering to harvest maturity. Panicles can be tagged at a reproductive stage that has a short duration, commonly asparagus stage (Figure 1). Fruit from these panicles can be destructively harvested in the weeks before and after the date of anticipated harvest, with assessment of internal attributes used in establishing the date of optimal harvest maturity. For example, Osuna-Garcia [10] tagged individual panicles on trees with a temperature logger within the canopy in a study set in Mexico that recommended 2100–2200 GDD on a Tb of 10 °C for Keitt to reach harvest maturity from the Christmas tree stage.



Figure 1. Stages of development of flowering illustrated by images of cultivar Calypso: (a) asparagus stage, (b) elongation phase, (c) Christmas tree stage (two thirds of flowers on panicle open), (d) fruit set stage.

The stages of flower development vary in duration (Figure 1). Lemos et al. [9] reported 182 to 276 GDD (equivalent to 14–21 days) from bud swelling to flower initiation (a), then 623 GDD from bud swelling to flower opening (equivalent to 49 days) (c) using Equation (1), with a Tb of 10 for mango cv. Uba. Ideally GDD estimates should be based on use of the floral stage with the shortest duration, i.e., asparagus stage (a). However, the proportion of terminals on a tree in asparagus stage is difficult to assess visually when driving through a row, and Christmas tree stage (c) is therefore usually assessed in commercial practice.

1.3. Estimating Harvest Maturity

The determination of when fruit is at ‘harvest maturity’ can be more problematic than determination of panicle development stage. ‘Harvest maturity’ is a commercial target which will vary by market and shelf-life needs (e.g., from a distant market served by sea-freight requiring a maximum storage potential to a local restaurant market seeking tree ripened fruit).

A range of fruit attributes change as fruit matures on tree, including skin colour, fruit shape, flesh colour, dry matter content (DMC), juice soluble solids content (SSC), titratable acidity (TA), SSC: TA and flesh firmness [13]. However, the levels of these attributes associated with a given stage of maturity can be cultivar and growing condition dependent.

Within Australian supply chains, flesh colour targets have been set for fruit harvest maturity for Kensington Pride (KP) [14] and Calypso [15]. In both cases, colour cards have been produced to assist growers in assessment of flesh colour. DMC targets have also been set for harvest of fruit in Australia (e.g., Table 2). The US National Mango Board [16] has promoted use of flesh colour as a maturity standard and has provided cultivar specific target flesh colours, and, more recently, minimum DMC values. These values are used by growers exporting to the USA and Europe from Mexico, Peru, Brazil, Costa Rica, Guatemala, and other central and South American countries (pers. comm., Agrodan, Brazil).

However, the DMC targets have been established in context of SSC and eating quality of ripened fruit, and not maturity *per se*. Fruit DMC increases with time on tree, but the absolute level of DMC can vary with growing condition, e.g., with water status [17]. Walsh et al. [18] recommend that the level of DMC associated with harvest maturity as indexed by flesh colour should be established for a given growing condition.

Table 2. Specifications on harvest minimum Dry Matter Content (DMC) and flesh colour for some relevant Australian grown mangoes. Flesh colours refer to colour cards produced by the named source. Kensington Pride is abbreviated to KP.

Cultivar	DMC (%)	Source	Target Flesh Colour Card	Source
Calypso	14	Whiley and Hofman [19]	7	DAF [15]
Honey Gold	15	Henriod [20]	none	
KP	15	Henriod et al. [21]	single “mature” colour card	NT Farmers Association [14]
R2E2	13	Henriod et al. [21]	none	
Keitt	16	Silva Neta [22]	2	National Mango Board, Orlando, FL, USA [18]

The first Australian study to suggest GDD targets dealt with cultivar Kensington Pride. It was based on eating quality of ripened fruit, which is associated with DMC [1]. Moore [2] based GDD recommendations for all major Australian cultivars on a fruit DMC maturity specification of 14.0% (*w/w*). In the most comprehensive work undertaken on the setting of harvest maturity standards for an Australian cultivar (Calypso), Hofman et al. [3] recommended a harvest specification of a minimum DMC (14% *w/w*), flesh colour of 7 on colour score cards, SSC of 7% (*w/v*) and GDD of 1640 (from Christmas tree stage, $T_b = 10\text{ }^\circ\text{C}$), with DMC, flesh colour and GDD promoted as the three most reliable attributes. Winston et al. [4] reported an attempt to use paint colour charts as references for flesh colour in Honey Gold maturity evaluation, however ‘the method was discontinued in year 2 due to inconsistencies in the methods and the time involved’, with preference given to the use of a GDD target established using a target DMC.

Henriod and Sole [23] established development of minimum mango harvest maturity standards for ‘1243’, a cultivar recently released from the Australian National Mango Plant Breeding Program. Fruit were harvested at intervals around time of expected maturation on the tree and assessed for quality once ripened. It was concluded that at-harvest flesh colour (hue), SSC, TA, DMC and GDD (T_b of $12\text{ }^\circ\text{C}$, from Christmas tree stage) were all suitable maturity indicators, with minimum values for these attributes of 102 (hue), 7% *w/v*, 2.3% *w/v*, 13% *w/w* and 1040 GDD, respectively. Unfortunately, the report was not clear on the method used to record flowering (e.g., eyeball of orchard average or tagging of panicles). The study also involved a single growing location and season.

1.4. GDD Calculation and Temperature Measurement

Most GDD estimates for mango fruit maturation have been based on the Arnold [24] algorithm (Equation (1)), with variation in the base temperature (T_b) between 10.0 and 17.9 $^\circ\text{C}$ and the stage of flowering stage used (Table 1). However, two studies (Table 1) have adopted use of an upper temperature threshold (T_B), as proposed by Ometto [25] (Equation (2); referred to as the ‘Upper T’ method in the current study). However, there is no published justification of the choice of T_b or T_B values.

$$\text{GDD} = \frac{T_{\text{max}} + T_{\text{min}}}{2} - T_b \quad (1)$$

If $T_B > T_b > T_M > T_m$; then $\text{GDD} = 0$,

If $T_B > T_M > T_m > T_b$; then $\text{GDD} = \left(\frac{T_{\text{Max}} - T_{\text{min}}}{2} \right) + (T_{\text{min}} - T_b)$,

If $T_B > T_M > T_b > T_m$; then $\text{GDD} = \frac{(T_{\text{Max}} - T_b)^2}{2 * (T_{\text{Max}} - T_{\text{min}})}$,

$$\text{If } TM > TB > Tm > Tb; \text{ then GDD} = \frac{2*(TMax-Tmin)*(Tmin-Tb) + (TMax-Tmin)^2 - (TMax-Tb)^2}{2*(TMax-Tmin)},$$

$$\text{If } TM > TB > Tb > Tm; \text{ then GDD} = \frac{1}{2} * \left[\frac{((TMax - Tb)^2 - (TMax - TB)^2)}{TMax - Tmin} \right] \quad (2)$$

where TB is Upper base temperature, Tb is Lower base temperature, $Tmax$ is maximum daily temperature and $Tmin$ is minimum daily temperature.

Another limitation to previous work is the location of the temperature sensor used in calculation of GDD. Data has been used from sensors located within or outside the mango tree canopy and located adjacent to the monitored orchard or not reported assuming is from a government recording station many kilometers distant to the farm (Table 1). With branch terminals largely positioned in the outer tree canopy, use of a Bureau of Meteorology standard [26] for sensor location is recommended (i.e., placement of the temperature sensor within a white coloured weather screen with ventilated sides, positioned 1.2 m above a ground surface covered with vegetation or mulch, in an open area away from other structures by at least four times the height of those objects). Given potential variation in temperatures across a farm, placement of a sensor in the near vicinity of each monitored orchard is also recommended.

1.5. Cultivars

The Australian mango industry is based on the domestically developed Kensington Pride (43% of production volume), Calypso™ 'B74' (25%), R2E2 (19%) and Honey Gold™ (8%) cultivars, with minor production of Asian and Florida bred cultivars, including Keitt (information from Australian Mango Industry Association website, accessed on 1 December 2022)). Kensington, also known as KP or Bowen, was selected from a poly-embryonic line brought India to Bowen, Australia, in the late 1880s. The mono-embryonic cultivar B74 originated as a cross of Kensington Pride and of the mono-embryonic Florida variety, Sensation. Sensation is a late season cultivar that has Florida cultivars Haden and Brooks parentage. The poly-embryonic Honey Gold was selected from a Kensington Pride mother tree pollinated by an unknown cultivar in Rockhampton, QLD. The mono embryonic Keitt is a late season cultivar originating in Florida that is usually grown to extend the end of Australian mango season to late March.

1.6. Research Aims and Objectives

The aim of the current study is to improve existing GDD recommendations for mango reproductive development for four Australian grown cultivars (Kensington Pride, Calypso, Honey Gold and Keitt), and to provide a methodology for estimation of GDD targets for maturation of mango fruit of any cultivar, with optimization of Tb and TB values. Additionally, a comparison of a GDD calculation employing a minimum base temperature only [24] and a calculation using both a minimum (Tb) and maximum (TB) base temperature [25] is undertaken.

To improve GDD estimates over previous studies, several procedures were adopted: (i) tagging of individual panicles at an early development stage of short duration, such as asparagus stage; (ii) a time series of destructive measurements of fruit internal attributes to establish harvest timing; (iii) use of data of multiple 'calibration' sites varying in region and season; (iv) testing of recommendations at several 'validation' sites across different seasons; and (vi) use of on-farm temperature sensors positioned within six meters of an orchard block as opposed to use of more remote weather stations. In addition, given variation in colour by printers and in screen display, attention was also given to better documentation of the process of producing colour comparison cards for estimation of flesh colour.

2. Materials and Methods

2.1. Temperature Assessment

An orchard block as defined as a management unit with consistent tree cultivar, age, management history, irrigation infrastructure and harvest. Blocks typically have an area of 1 to 5 ha with more than 312 trees/ha, given average density (8×4) planting. For each orchard block with tagged fruit, temperature was monitored using a temperature sensor (Sensor Host, Rockhampton, Australia) in a ventilated shade screen mounted 1.2 m above covered ground, outside of the tree canopy, with temperature logged at 15 min intervals. The exceptions were the Darwin and Bungundarra site in 2018 and 2019, when the farm temperature record was used. These records were based on Hobo Onset (USA) temperature loggers within screens placed inside the tree canopy. Daily minimum and maximum temperatures were used in calculation of daily GDD.

2.2. Sites and Panicle Tagging Exercises

Flowering events in the 2018, 2019, and 2020 seasons were tagged, generally at asparagus stage, within 9 orchards across Australia and in Brazil (Table 3). These sites involved the cultivars dominating commercial production in Australia (KP, Calypso, R2E2 and Honey Gold), and a cultivar common to production in both Australia and Brazil (Keitt). In total, 22 populations were selected, where each population is specific in cultivar, location and date (involving 38 tagging events, given tagging of multiple flowering events in some locations) (Appendix A Table A1). The two populations of cultivar R2E2 that were tagged did not hold any fruit for two consecutive seasons. Populations from the 2018–2021 season were used in ‘calibration’ of GDD targets, while the 13 populations of the 2021/22 season were used in validation of proposed targets. Additional fruit from these exercises also measured non-destructively for a sizing exercise reported in Amaral and Walsh [27].

Table 3. Site locations.

Region	Latitude	Longitude	Cultivars	Seasons
Darwin, NT	−12.754125°	131.167722°	Calypso	2018/19/20/21
Darwin, NT	−12.548013°	131.259296°	Honey Gold	2021
Katherine, NT	−14.615475°	132.205328°	KP	2020/21
Katherine, NT	−14.583944°	131.995526°	Calypso	2020/21
Katherine, NT	−14.544315°	132.471902°	Honey Gold	2020/21
Dimbullah, QLD	−17.136831°	145.088776°	Calypso, Honey Gold	2020/21
Bungundara, QLD	−23.025202°	150.641147°	Honey Gold, Keitt, KP	2018/19/20/21
Belem do Sao Francisco, PE, Brazil	−8.678973°	−39.165941°	Keitt	2020/21
Curaca, BA, Brazil	−9.038435°	−39.930138°	Keitt	2021/22

Panicles at several developmental stages were tagged on a single date and harvested on a single date in 2018 at Darwin site, while at the Bungundarra site, panicles at asparagus stage were recorded weekly, with all fruit harvested at a single date (Table A1). In other years, panicles were tagged at asparagus stage on a single date, with panicles monitored weekly for the achievement of Christmas tree stage, and resulting fruit harvested over several weeks ($n = 20$ per week) around the expected (GDD forecast) date of harvest maturation. Asparagus stage terminals were marked on the subtending vegetative stem, with the tag later moved onto the panicle when fruit set was successful for that terminal. If asparagus stage terminals were tagged on different dates (i.e., different flowering ‘events’), a different colour of flagging tape was used for the different date. Asparagus stage was missed at some locations, necessitating tagging at elongation or Christmas tree stage (Table A1). At each site, typically 10 panicles were tagged on each of 10 trees, with panicles selected from around the tree canopy (Table A1). The 100 panicles typically resulted in >20 fruit, but in some cases fewer fruit were retained to harvest maturity (Table A1).

Fruit were destructively sampled at weekly intervals around the anticipated harvest maturity date anticipated from the currently recommended GDD for a given cultivar. Fruit

flesh colour (CIE LAB and hue) and oven-DM was assessed in all populations, while SSC, TA and carotenoid content (mg/kg) was assessed in some populations. These values were used to estimate the date and GDD at which harvest maturity was achieved, from the published commercial colour specifications for cultivars Calypso and Keitt. A flesh colour specification for Honey Gold was established using the existing GDD specification of 1800 units [2].

2021–2022 populations were used as validation sets for Calypso, Honey Gold and Keitt cultivars. The thirteen 2021 season populations (Table A1) were used to validate the recommended target GDD units between the stages of asparagus, Christmas tree and harvest maturity for each of the four cultivars. At each site, resulting fruit ($n = 20$) were destructively assessed for maturity attributes when the fruit reached the target GDD established in the 2018–2020 calibration exercise.

2.3. Estimate of Lower and Upper Base Temperature

An exercise was undertaken to optimize the T_b and T_B values used in the GDD calculations. T_b from 1 to 20 °C at intervals of 1 °C were used in equation 1, and T_B from 25 to 37 °C at intervals of 1 °C were used in equation 2. The T_b (°C) used while varying T_B in the calculation of equation 2 was set to 12 °C. For the T_b exercise, temperature data from four different flowering events at a southern location (Bugundarra, QLD, Australia) was used. For the T_B exercise, data of flowering events of northern sites (two at Darwin, NT, and one event at Katherine, NT) was used (Table 4). The sites and periods were chosen for low temperatures in assessment of T_b and high temperatures in assessment of T_B .

Table 4. Farm locations and range of temperature values used for T_b or T_B in T_b/T_B optimization method. n/a is not applicable. Population # refers to numbering in Table A1.

Region/Population #	T_b (°C)	T_B (°C)	Method	Period
Darwin, NT/10	12	23 to 37	Ometto, 1981	15/06/2021–20/10/2021
Darwin, NT/11	12	23 to 37	Ometto, 1981	15/06/2021–20/10/2021
Katherine, NT/15	12	23 to 37	Ometto, 1981	15/06/2021–20/10/2021
Bugundarra, QLD/2	1 to 20	n/a	Arnold, 1960	07/07/2020–23/12/2020
Bugundarra, QLD/4	1 to 20	n/a	Arnold, 1960	07/07/2020–23/12/2020
Bugundarra, QLD/21	1 to 20	n/a	Arnold, 1960	07/07/2020–23/12/2020

The method of Yang et al. [28], as adapted by Rodrigues et al. [6], was used, with the Coefficient of Variation (CV) (Equation (3)) calculated for GDD values estimated across three sites for each of T_b or T_B values. The T_b or T_B value with the lowest CV was chosen as the most reliable T_b or T_B .

$$CV = \frac{\sigma}{\mu} * 100 \quad (3)$$

2.4. Assessment of Maturity Attributes

Spectra were acquired from a mid-equatorial position on both sides of the fruit using a handheld near infrared spectrometer (F750, Felix Instruments, Camas, WA, USA). Spectra were acquired of each side of 1126 fruit ($n = 2252$ spectra). Fruit were then sliced to remove cheeks on both sides and a core of 20 mm diameter taken from the center of each slice. The skin was removed from the core, and the core then trimmed to a length of 10 mm. The flesh colour of the inside cut was assessed visually by comparison with a colour chart and by use of a Chroma Meter (CR-400, Konica Minolta, Japan) calibrated with a factory standard ceramic white tile c and set to the illumination method 'D65'. Readings were taken in the CIE LAB colour scheme. Hue angle was calculated as \tan^{-1} (CIE B/CIE A) for samples with an A value above 0 and $180 \pm \arctan$ (CIE B/CIE A) for samples with A value below 0 (Ford and Roberts, 1998). Flesh colour was thus assessed at a depth of 10 mm from the skin, rather than a set distance from the stone.

One half of each core (approximately 5 g fresh weight) was used for oven-DMC analysis while the other half was diced and stored at $-20\text{ }^{\circ}\text{C}$ awaiting carotenoid analysis. For oven-DMC assessment, samples were placed on aluminum foil cups and dried in a fan forced home dehydrator EzidryFD2000 (Ezidry, Adelaide, Australia) at $60\text{ }^{\circ}\text{C}$ for 48 h [29], with weight recorded before and after using a scale of 0.001 g resolution. DMC was calculated as $\text{Dry Weight}/\text{Fresh Weight} \times 100$. The rest of the fruit was blended and then filtered. SSC of filtrate was measured with a Bellingham and Stanley RFM320 digital refractometer and TA assessed of a 10 mL sample of juice using 0.1 N NaOH as a titrant and 1% *w/v* citric acid as a reference. TA results were expressed as citric acid equivalents.

Samples frozen at $-20\text{ }^{\circ}\text{C}$ were freeze dried ($-45\text{ }^{\circ}\text{C}$, 200 mT) (Flexidry MP freeze drier, FTS Systems, USA) for approximately 36 h, then crushed in a ceramic mortar and pestle. Approximately 0.1 g of subsample was placed into 15 mL of acetone (99.5%, AR grade, Chem Supply, Australia), then sonicated for 30 min (Soniclean 160TD ultrasonic cleaner; Dudley Park, South Australia) and centrifuged (Heraeus Multifuge, Thermo Fisher Scientific; Sydney, Australia; $1000\times g$ for 5 min) with no volume loss reported. Total carotenoids were assessed using the method of Tomlins et al. [30], with supernatant absorbance at 450 nm measured using a UV-Vis spectrophotometer (Genesys 10S UV-Vis, Thermo Scientific, Australia). The total carotenoid concentration of the extracts (C_e , in mg/L) was calculated using the Beer-Lambert law:

$$C_e = A/\epsilon b \times MW \times 1000 \quad (4)$$

where A is absorbance at 450 nm, ϵ is molar absorptivity (137,400 L/mol/cm), b is path length (1 cm), MW is molecular weight of β -carotene (536.8726 g/mol). The carotenoid content of tissue (C_t , in mg/g dry weight) was calculated as:

$$C_t = C_e \times V/W \quad (5)$$

where V is volume of extract (15 mL), and W is dry weight of tissue.

2.5. Colour Cards

Colour cards are available to assist growers in judging flesh colour. The use of colour cards to assess sample colour can be compromised by variation in ambient lighting, issues with the users' vision (e.g., at extreme, colour blindness), and the printing process used to produce the cards and ageing of the cards. An attempt was made to quantify the colour space values of the cards, as CIE LAB value from the original pdf file as sent to printer (when available) and/or a colorimeter reading of the printed card.

Several card sets were accessed. Calypso colour cards were a product of the work of Whiley and Hofman [26]. Colour space values were accessed from the pdf files. There have been two printing runs producing card sets for grower use, and a card set from each printing run was accessed. A cultivar Kensington Pride 'business card' with harvest maturity colour was produced by NT Farmers association [14] (Darwin, Australia). Keitt colour swatches as electronically published by the US National Mango Board [18] were also accessed.

2.6. Statistical Analysis

Linear correlations between parameters and one-way ANOVA statistical analysis were undertaken using the Rstudio 4.1.2 (Boston, MA, USA). A significance p -value < 0.05 was adopted. Population results were expressed as mean \pm SD., or mean \pm SE for all parameters (DMC, TSS, CIE-Lab, hue, TA and SSC: TA ratio).

3. Results and Discussion

3.1. GDD Algorithm-Choice of T_b and T_B

The use of a different T_b merely creates a daily offset in the GDD increment when the daily average of T_{max} and T_{min} exceeds T_b . For example, if T_b is decreased from

12 to 10 °C, then 2 extra units will be accumulated every day. In this scenario, the use of different T_b values requires a revised value for the heat units associated with harvest maturity, but the date the GDD target is achieved is not affected. However, if the daily average of T_{max} and T_{min} is less than T_b , differences in the estimate of the date of harvest maturity emerge.

The CV on calculated GDD across three populations from a cooler growing area was minimal at a T_b of 12 or 13 °C (Figure 2). A T_b of 12 °C is therefore recommended based on low CV and its current common use as a base temperature in most Australian GDD calculations. For TB, minimal CV occurred at 32 °C (Figure 3). In comparison, Rodrigues et al. [6] recommended a T_{min} of 13 °C and a TB of 32 °C for mango cv. Tommy Atkins in Brazil.

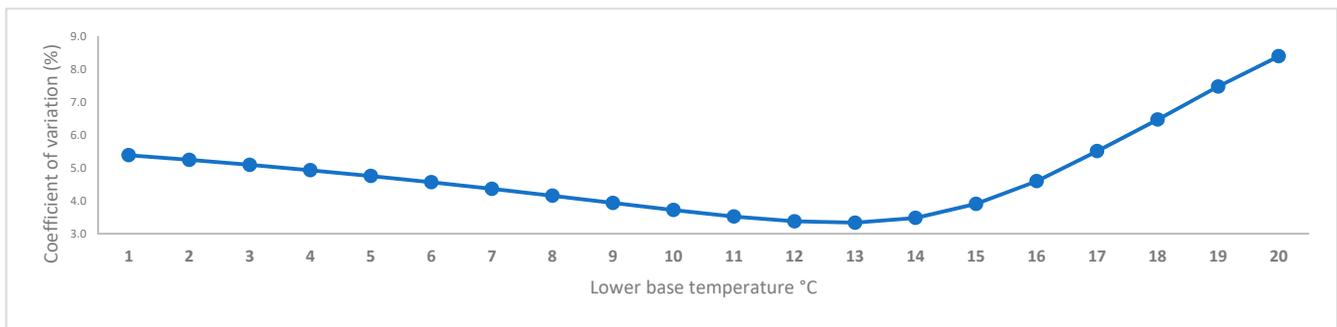


Figure 2. Coefficient of variation for GDD calculated using different lower base temperatures (T_b). Data of three flowering events of the southern-most farm of this study, Bungundarra, QLD (Table A1).

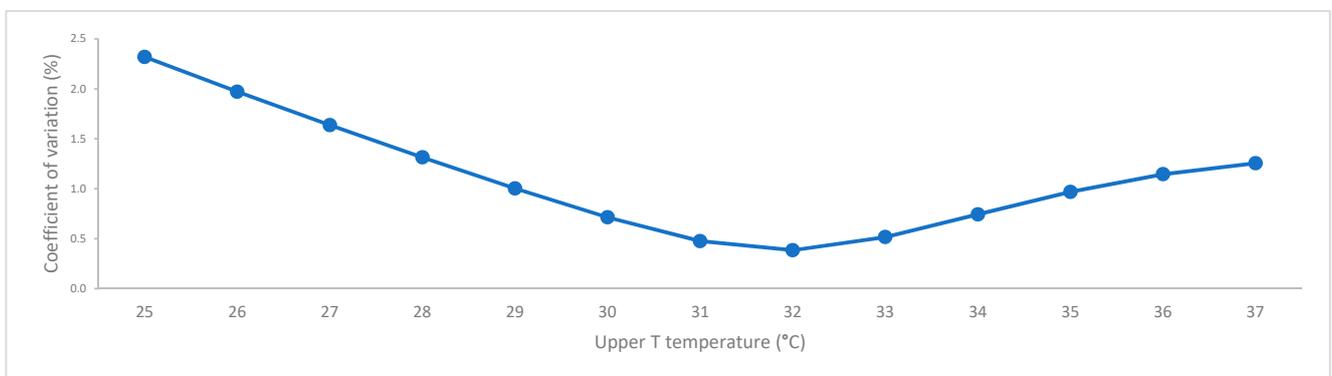


Figure 3. Coefficient of variation for GDD calculated using different upper base temperatures (TB) and a T_b of 12°C. Data of three flowering events of the northern-most farms of this study, in Darwin and Katherine, NT (Table A1).

3.2. GDD Algorithm Implementing TB

The use of a TB in a GDD calculation (Equation (2)) provided target harvest dates up to 13 days later than the standard GDD calculation (Equation (1)) for NT sites, up to 5 days later in Far North Queensland sites and up to 3 days later in Central Queensland sites (Table 5). This difference between sites mirrors differences in the proportion of days with temperatures above 32 °C.

Table 5. Data showing 2021/22 harvest date estimated from flowering date using the standard and the Upper T methods of calculating GDD for 9 populations, varying in location and cultivar. GDD (from asparagus stage, Tb = 12 °C, TB = 32 °C) targets of 1680 for Calypso, 1800 for Honey Gold (HG) and 1600 for Kensington Pride (KP) were used.

Region-Cultivar-Population #	Flowering Date	Harvest Date (Standard GDD)	Harvest Date (Upper T GDD)	Difference in Days
Darwin NT-Calypso-10	4-Jun	18-Sep	1-Oct	13
Darwin NT-HG-12	8-Jul	25-Oct	4-Nov	10
Katherine NT-Calypso-14	15-Jun	14-Oct	22-Oct	8
Katherine NT-KP-15	15-Jun	2-Oct	9-Oct	7
Katherine NT-HG-13	2-Jul	23-Oct	4-Nov	12
Dimbulah FNQ-Calypso-16	30-Jun	22-Nov	26-Nov	4
Dimbulah FNQ-HG-17	30-Jun	29-Nov	4-Dec	5
Bungundarra CQ-KP-20	5-Jun	26-Nov	29-Nov	3
Bungundarra CQ-HG-21	5-Jun	16-Dec	18-Dec	2

refers to Table A1 population numbers.

While the use of the Upper T method impacts the predicted harvest date, it remains to be demonstrated that the physiological premise is correct (i.e., that maturation slows at temperatures above 32 °C). This demonstration is attempted in the following section.

3.3. GDD between Reproductive Stages

A sample of 15 panicles tagged at asparagus stage of a range of Honey Gold, Calypso, Keitt, and Kensington Pride populations were observed either weekly or every three days and the date that Christmas tree stage was reached was recorded (Table 6). The mean and standard deviation, across locations and seasons, on the GDD difference between the two flowering stages was 188 ± 18 for Calypso, 184 ± 12 for Honey Gold, 238 ± 21 for Keitt and 175 ± 10 for KP (Table 6). These estimates should be more accurate than the recommendation of 300 units for all cultivars, as given by Moore [2], given that the latter estimate was based on grower estimates of the date of ‘early’ and ‘late’ flowering stages, rather than from tracking of individual panicles. The difference between the GDD requirement of the three Australian cultivars, which share parentage, and the Florida cultivar, Keitt, is consistent with a genetic component to the GDD requirement.

Table 6. GDD (Tb 12 °C, TB 32 °C) between asparagus and Christmas tree stages ($n = 15$), with average and SD for each cultivar. Population numbers refer to Table A1.

Population #	Cultivar	Date of Christmas Tree Stage	GDD
2018			
1	Calypso	13-Jun	164
2020			
5A	Honey Gold	5-Aug	180
5B	Honey Gold	29-Aug	171
6A	Honey Gold	5-Aug	180
6B	Honey Gold	9-Sep	183
7	Keitt	1-Sep	237
8	Keitt	16-Sep	231
9	Keitt	29-Aug	297
2021			
10	Calypso	18-Jun	180
12	Honey Gold	24-Jun	186
13	Honey Gold	17-Jul	176
14	Calypso	2-Jul	193
15	KP	30-Jun	184
16	Calypso	23-Jul	214

Table 6. Cont.

Population #	Cultivar	Date of Christmas Tree Stage	GDD
17	Honey Gold	23-Jul	214
20	KP	28-Jul	165
21	Honey Gold	21-Jul	184
22	Keitt	4-Aug	235
Average \pm SD			
	Calypso		188 \pm 18
	Honey Gold		184 \pm 12
	Keitt		238 \pm 21
	KP		175 \pm 10

3.4. Colour Cards for Flesh Colour Assessment

A strong correlation between fruit GDD and CIE B existed across cv. Calypso ($R = 0.96$, $n = 513$), cv. HoneyGold ($R = 0.90$, $n = 611$) and cv. Keitt ($R = 0.94$, $n = 240$) populations. In addition, CIE B was strongly correlated to organoleptic parameters such as SSC:TA ratio ($R^2 = 0.89$), TA ($R^2 = 0.84$), DMC (%w/w) ($R^2 = 0.86$), and total carotenoids content ($R^2 = 0.86$), although poorly correlated to SSC (%w/w) ($R^2 = 0.58$) (Appendix B). Therefore, flesh colour is recommended as the primary index in assessment of harvest maturity.

The flesh colour of cut fruit was matched by visual comparison to colour cards. Calypso and KP fruit judged as matching Calypso colour card 7 (CIE B = 32) had a mean CIE B value of 32.9 (with range 30.2 to 35.6) and 32.2 (range 29.4 to 35.0), respectively. Keitt fruit judged as matching Keitt colour card 2 (B = 51) had a mean CIE B value of 51.0 (range 45.9 to 55.0) (Table A2). Human sorting was thus successful in matching fruit to colour cards.

CIE LAB values varied between the pdf associated values and readings taken of cards from different print runs using different printers, although values for a given maturity value were consistent for different prints from the one printer (Table A2).

A set of swatches with colour values spanning the range associated with harvest maturity of all cultivars involved in this study was proposed, with indication of the swatch associated with maturity of each cultivar (Table A3). CIE LAB values are given as expected readings of fruit flesh using a calibrated colorimeter such as the Minolta CR400 (from Tables 5–7). A second CIE L value is given in Table A3, being the value in a pdf electronic file to achieve desired CIE LAB values in a print made by an office printer (Bizhub C4000i, Konica Minolta, Japan). These values were determined by trial and error. A similar optimization is recommended when using other printers.

3.5. Cultivar Specifications on Maturity

3.5.1. Time Course of Maturity Attributes

Fruit attributes were destructively assessed for fruit from fruit stone-hardening stage to past commercial harvest for several populations (Table A1). One example each of Calypso, Honey Gold and Keitt populations is given graphically in Figure 4 (other data is presented in Amaral [31]). In all cultivars, SSC showed little change with time on trees (Figure 4). DMC, TA, and flesh colour, as indexed by CIE B value or hue, changed as fruit matured, while CIE A value changed only in Keitt fruit (Figure 4). Change was not linear, with perturbations likely due to changes in growing conditions, particularly water status (e.g., Anderson et al. [16]).

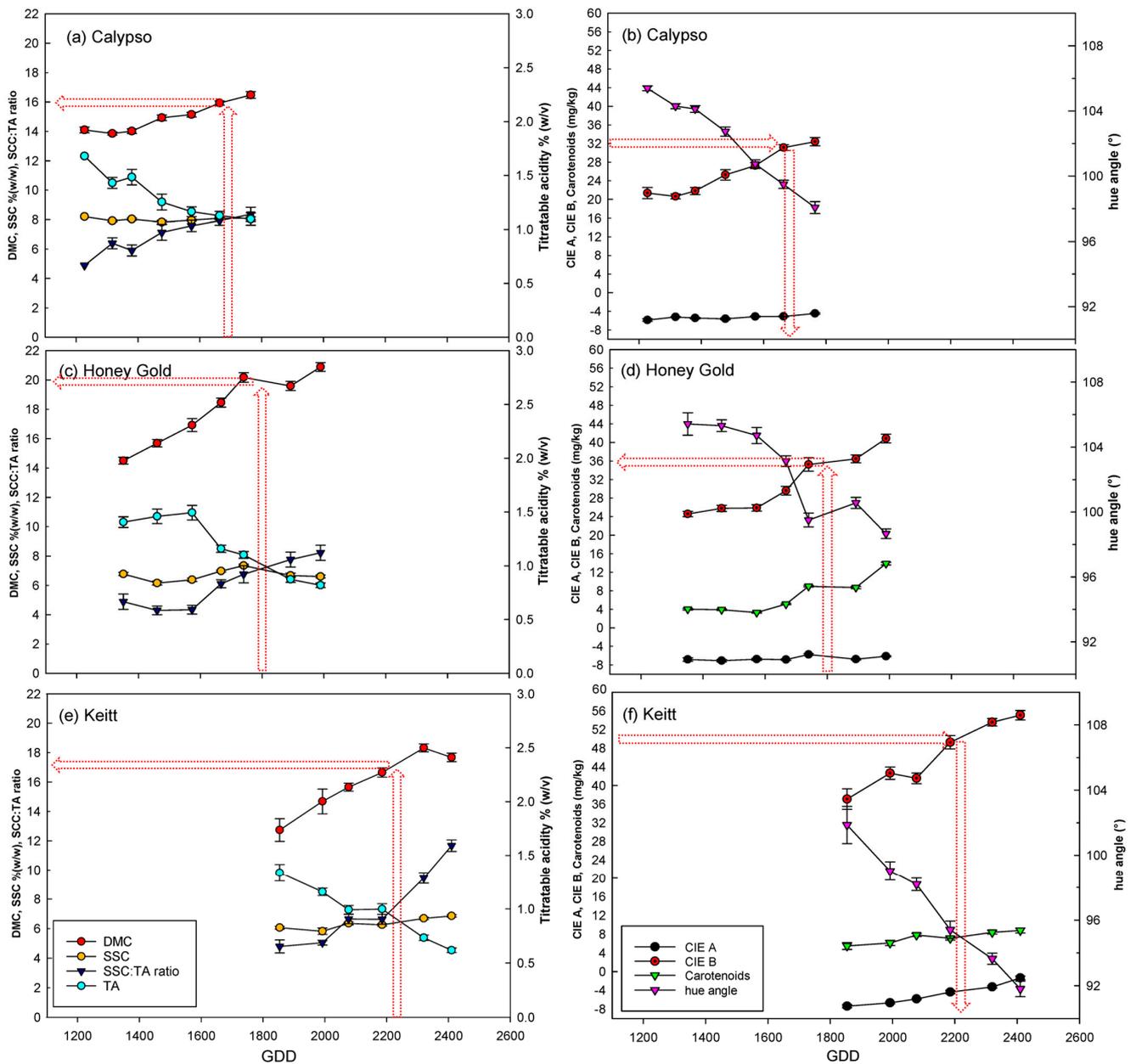


Figure 4. Time course from stone hardening of fruit attributes (for populations 3, 5 and 7 of Table A1, shown in top (a,b), middle (c,d) and bottom (e,f) panels, respectively). Left panels: DMC, SSC, SSC:TA ratio and TA. Right panels: flesh colour CIE AB values and total carotenoids (mg/kg); Data presented as mean with associated standard error ($n = 10$ to 20 fruit for Calypso, 5 to 10 fruit for Honey Gold and Keitt). For Calypso and Kensington Pride, red lines are drawn from the CIE B value associated with harvest maturity, i.e., 32 for Calypso and Kensington Pride and 51 for Keitt, to the associated GDD (right panel), and from this GDD value to the associated DMC value (left panel). For Honey Gold, the red line is drawn from the recommended GDD of 1800 to the associated B and DMC values. X axis is given in heat units (GDD) estimated from asparagus stage of flowering using $T_b 12\text{ }^{\circ}\text{C}$ and $T_B 32\text{ }^{\circ}\text{C}$.

In the example Calypso population, CIE B value increased at about 2.3 units per week while DMC increased at approximately 0.52% w/w per week (Figure 4). The B value harvest maturity target (32 units) was reached at 1664 GDD from asparagus stage and a DMC of 16.0% w/w (see red arrows on Figure 4). For a Keitt population, B value increased at about 4 units per week and DMC increased at approximately 1.0% w/w per week. The B value

target of 51 units was reached at 2210 GDD from asparagus stage with a fruit DMC of 17.0% *w/w*, as estimated by linear interpolation of adjacent values.

The commercially accepted maturity target for Honey Gold is a GDD of 1500 from the Christmas tree stage [4] or 1800 from the asparagus stage [2]. For a Honey Gold population, B value increased at about 2.7 units per week and DMC increased at approximately 1.40% *w/w* per week (Figure 4). The harvest maturity GDD target of 1800 was achieved when fruit flesh B value was 36 and fruit DMC was 20.0% *w/w* (see red arrows on Figure 4).

This procedure was followed with all calibration data sets, to associate attribute levels to the available harvest maturity targets of flesh colour in Calypso, KP and Keitt, and to the GDD target for Honey Gold, as described in the following sections. While the focus has been given to GDD and CIE B, levels of DMC, TA, and SSC:TA ratio are also reported in the following sections.

3.5.2. Calypso

For a 2018 Calypso population, CIE B = 32 was reached at 1720 GDD and a DMC of 16.5% *w/w* (Table 7), based on interpolation between measurements made at 1692 and 1792 GDD. For the same orchard in 2019, CIE B = 32 was reached at GDD 1735 with a DMC of 16.3% *w/w*. From the average of data of these two populations, a GDD of 1728 from asparagus stage is recommended to achieve a flesh B value of 32, with an associated DMC of 16.4% *w/w*, TA of 0.86% *w/w* and SSC:TA ratio of 10.6. This represents an increase of approximately 3 (summer) calendar days over the currently recommended GDD of 1640 on Tb 10 °C [3] or 1680 on Tb 12 °C [2].

Table 7. Observed values of Calypso fruit attributes at the two harvest dates with CIE B value bracketing the target value of 32 (in italics), and values at the target value, in italics, as estimated by interpolation. GDD are calculated from asparagus stage with Tb = 12 °C, and TB = 32 °C for fruit from the same orchard in 2018 and 2019 seasons. Panicles were tagged on different dates and harvested on one date. Population numbers refer to Table 4.

	CIE B	Hue	DMC	TA	SSC:TA Ratio
2018 (pop # 1)					
GDD 1692	28.0 ± 0.6	99.6 ± 0.2	15.4 ± 0.2	0.90 ± 0.03	8.4 ± 0.2
<i>GDD 1720</i>	<i>32</i>	<i>96.5</i>	<i>16.5</i>	<i>0.8</i>	<i>13</i>
GDD 1792	38.3 ± 1.1	93.6 ± 0.6	18.0 ± 0.3	0.40 ± 0.10	20.0 ± 1.4
2019 (pop # 3)					
GDD 1664	31.1 ± 0.6	99.5 ± 0.3	15.9 ± 0.1	1.12 ± 0.10	7.9 ± 0.3
<i>GDD 1735</i>	<i>32</i>	<i>98.7</i>	<i>16.3</i>	<i>1.1</i>	<i>8.2</i>
GDD 1765	32.4 ± 0.9	98.1 ± 0.4	16.5 ± 0.2	1.10 ± 0.10	8.3 ± 0.5

Refers to population numbers in Table A1.

3.5.3. Keitt

In Keitt fruit population #7 (Table 6), the target maturity CIE B value of 51 was achieved at a DMC of 17.0% *w/w*, a TA of 0.85% *w/w*, an SSC:TA ratio of 7 and a total carotenoids content of 7.5 mg/kg at 2210 GDD from asparagus stage (Table 6). From the data of three populations (Table 8), a GDD of 2185 was chosen as a minimum value recommended to achieve a minimum flesh CIE B value of 51.0.

Table 8. Observed values of Keitt fruit attributes at two or three harvest dates with CIE B value bracketing the target value of 51, and values at the target value, in italics, as estimated by interpolation. GDD are calculated from asparagus stage with $T_b = 12\text{ }^\circ\text{C}$, and $T_B = 32\text{ }^\circ\text{C}$ for fruit of three populations. Panicles were tagged on one date and harvested on a range of dates. Population numbers refer to Table 4.

	CIE B	Hue	DMC (%w/w)	TA (%w/v)	SSC:TA Ratio	Carotenoid (mg/kg)	Flesh Card Colour
Pop # 7							
GDD 2185	49.3 ± 1.5	95.4 ± 0.5	16.7 ± 0.3	1.00 ± 0.04	6.6 ± 0.3	7.1 ± 0.4	1.9 ± 0.1
<i>GDD 2210</i>	<i>51</i>	<i>94.8</i>	<i>17</i>	<i>0.85</i>	<i>7</i>	<i>7.5</i>	<i>2</i>
GDD 2320	53.6 ± 0.8	93.6 ± 0.3	18.3 ± 0.3	0.73 ± 0.02	9.5 ± 0.4	8.4 ± 0.4	2.3 ± 0.1
GDD 2410	55.1 ± 1.0	91.8 ± 0.5	17.7 ± 0.3	0.62 ± 0.01	11.6 ± 0.4	8.8 ± 0.5	2.3 ± 0.1
Pop # 8							
GDD 2007	43.2 ± 1.9	98.5 ± 0.6	16.0 ± 0.6	1.12 ± 0.13	6.0 ± 0.5	4.7 ± 0.6	1.9 ± 0.2
GDD 2142	51.0 ± 1.2	94.4 ± 0.6	16.7 ± 0.2	1.05 ± 0.04	6.6 ± 0.3	8.4 ± 0.6	2.4 ± 0.2
GDD 2233	51.1 ± 2.0	92.7 ± 0.7	16.4 ± 0.4	1.05 ± 0.04	7.5 ± 0.2	7.5 ± 0.6	2.7 ± 0.2
Pop # 9							
GDD 2297	55.0 ± 1	95.1 ± 0.4	15.9 ± 0.1	0.77 ± 0.02	10.6 ± 0.4	10.2 ± 0.5	2.1 ± 0.1
GDD 2350	54.5 ± 1.1	95.1 ± 0.4	16.2 ± 0.2	0.81 ± 0.02	10.8 ± 0.3	9.7 ± 0.6	2.1 ± 0.1
GDD 2452	57.0 ± 0.7	94.8 ± 0.4	16.5 ± 0.2	0.70 ± 0.03	11.7 ± 0.5	10.7 ± 0.7	2.6 ± 0.1

Refers to population numbers in Table A1.

3.5.4. Honey Gold

For Honey Gold, a GDD of 1800 from asparagus stage (on $T_b = 12\text{ }^\circ\text{C}$) is a recommended maturity specification [2]. At 1800 GDD, fruit of population 2 reached a CIE B of 36 (interpolation using 4 data points) and a DMC of 17.6% w/w (Table 9). For the same orchard in the next season, fruit at 1800 GDD possessed a CIE B of 40.5 (extrapolated from 3 data points) and a DMC of 24.4% w/w. The difference in values was associated with unusual growing conditions, as the orchard was in a declared bushfire disaster area in November and was subject to high temperatures and dry conditions. In 2020, tagging occurred on two flowering events (FE) on each of two orchards on the one farm (populations 5 and 6). In FE1 of orchard 1, a GDD of 1800 was associated with a CIE B of 34 and a DMC of 18.5% w/w, while in FE2, it was associated with a CIE B of 35.5 and a DMC of 19.5% w/w. In FE1 of orchard 2, a GDD of 1800 was associated with a CIE B of 34 and a DMC of 18.5% w/w, while in FE2, it was associated with a CIE B of 34 and DMC of 19.2% w/w. Averaged across these populations, 1800 GDD was associated with CIE B of 36 ± 2.1 (mean \pm SD) and DMC of 19.5 ± 2.2 . The CIE B value of 36 is therefore recommended as a flesh colour standard for Honey Gold. This value is equivalent to the DAF 2019 print of Calypso colour card 9.

The preceding discussion is based on the Moore (2010) recommendation of a GDD of 1800 from asparagus stage or a GDD of 1500 from Christmas tree stage (on $T_b = 12\text{ }^\circ\text{C}$) as a maturity specification for Honey Gold. Winston et al. [4] confirmed a GDD (on $T_b = 12\text{ }^\circ\text{C}$) of 1500 from the Christmas tree stage. However, the 300 GDD asparagus to Christmas stage difference used by Moore [2] was an approximation used across all cultivars. A GDD of 184 was established in the current study for development between the two flowering stages. An average GDD of 1560 was estimated from Christmas tree stage to harvest maturity, suggesting a GDD of $1560 + 184 = 1744$ should be used for forecasting of Honey Gold harvest maturity from asparagus stage.

Table 9. Observed values of Honey Gold fruit attributes at several harvest dates bracketing the GDD (Tb = 12 °C) target of 1800 units from asparagus stage, and values at the target value, in italics, as estimated by interpolation or extrapolation. Panicles were tagged at asparagus stage on different dates. Population numbers refer to Table 4. ‘n/a’ is not available.

	CIE B	Hue	DMC (% w/w)	TA (% w/w)	SSC:TA Ratio	Carotenoid (mg/kg)
2018 CQ pop 2						
GDD 1658	34.1 ± 0.8	101.8 ± 0.3	16.3 ± 0.2	1.00 ± 0.02	5.7 ± 0.1	n/a
GDD 1791	38.2 ± 3.1	99.4 ± 0.9	18.0 ± 0.4	0.87 ± 0.08	7.4 ± 0.7	n/a
<i>GDD 1800</i>	<i>36</i>	<i>100.3</i>	<i>17.6</i>	<i>0.95</i>	<i>6.8</i>	<i>n/a</i>
GDD 1816	34.7 ± 0.7	100.6 ± 0.2	17.3 ± 0.2	0.95 ± 0.02	6.3 ± 0.1	n/a
GDD 1850	38.7 ± 0.7	99.8 ± 0.2	18.1 ± 0.1	0.85 ± 0.02	7.3 ± 0.2	n/a
2019 CQ pop 4						n/a
GDD 1638	34.8 ± 0.7	96.8 ± 0.4	22.6 ± 0.3	1.49 ± 0.04	6.3 ± 0.2	n/a
GDD 1691	35.7 ± 0.7	96.6 ± 0.4	22.7 ± 0.2	1.45 ± 0.05	6.2 ± 0.1	n/a
GDD 1756	39.2 ± 0.6	96.2 ± 0.2	24.1 ± 0.2	1.33 ± 0.03	6.8 ± 0.1	n/a
<i>GDD 1800</i>	<i>40.5</i>	<i>96</i>	<i>24.4</i>	<i>1.2</i>	<i>6.8</i>	<i>n/a</i>
2020 CQ pop 5						
Orchard 1						
* GDD 1747	32 ± 1.3	101 ± 0.3	17.2 ± 0.3	1.15 ± 0.06	6.2 ± 0.3	5.0 ± 0.5
* <i>GDD 1800</i>	<i>34</i>	<i>101</i>	<i>18.5</i>	<i>1.15</i>	<i>6.2</i>	<i>6.1</i>
* GDD 1850	35.6 ± 1.2	100.5 ± 0.4	19.4 ± 0.4	1.15 ± 0.02	6.3 ± 0.2	6.9 ± 0.5
** GDD 1740	35.3 ± 1.5	99.5 ± 0.4	20.2 ± 0.3	1.1 ± 0.02	6.8 ± 0.2	9.0 ± 0.6
** <i>GDD 1800</i>	<i>35.5</i>	<i>100.5</i>	<i>19.5</i>	<i>1</i>	<i>7</i>	<i>8.5</i>
** GDD 1892	36.5 ± 0.9	100.6 ± 0.3	19.6 ± 0.3	0.87 ± 0.02	7.8 ± 0.2	8.7 ± 0.5
2020 CQ pop 6						
Orchard 2						
* GDD 1747	32.9 ± 0.6	103 ± 0.3	17.1 ± 0.2	1.17 ± 0.03	5.9 ± 0.3	3.3 ± 0.3
* <i>GDD 1800</i>	<i>36.5</i>	<i>101</i>	<i>18.2</i>	<i>1.13</i>	<i>6.3</i>	<i>6.5</i>
* GDD 1850	39.9 ± 1.1	99.8 ± 0.3	19.1 ± 0.2	1.09 ± 0.02	6.6 ± 0.1	5.1 ± 0.3
** GDD 1687	31.4 ± 1.1	101.2 ± 0.3	19.1 ± 0.3	1.15 ± 0.04	6.0 ± 0.2	7.7 ± 0.5
** <i>GDD 1800</i>	<i>34</i>	<i>100.5</i>	<i>19.2</i>	<i>1.18</i>	<i>5.6</i>	<i>8</i>
** GDD 1839	34.3 ± 1	100.3 ± 0.3	19.2 ± 0.5	1.19 ± 0.04	5.5 ± 0.2	8.5 ± 0.6

* Represents the first flower event one for the given orchard; ** Represents the second flower event for the given orchard.

3.6. GDD Validation

The GDD targets established in the preceding section for Calypso and Keitt (1728 and 2185, respectively), associated with flesh CIE B values of 32 and 51, respectively, were trialed in 2021 validation exercises. The Calypso target was also used for KP. Across multiple populations harvested at these GDD values, flesh CIE B value met the colour specification of 32 for all six Calypso populations, and the specification of 51 in five of the six Keitt populations (Table 10). DMC met the specification minimum of 14.0% w/w in all cases, varying from 15.4 to 16.7% w/w across the six Calypso populations, and 14.0 to 17.4% w/w across the six Keitt populations.

Table 10. GDD validation exercise: Maturity attributes of fruit 2021/22 season harvested close to a GDD of 1728, 1740, 2185, and 1600 for Calypso, Honey Gold, Keitt and KP populations, respectively. Population numbers refer to Table A1.

Pop #	Season	Fruit	CIE B	Hue	DMC (%)	Colour Cards	GDD
Calypso							
1 *	2018	95	32.0	96.5	16.5	7.0	1720
3 *	2019	209	32.0	98.7	16.3	7.0	1735
10	2021	88	31.0	100.0	16.0	6.0	1757
11	2021	54	30.0	101.0	16.0	6.6	1700
14	2021	18	31.0	97.9	16.7	6.1	1741
16	2021	48	31.0	100.0	15.4	6.0	1757
Mean ± SE			31.2 ± 0.3	99 ± 0.6	16.2 ± 0.2	6.2 ± 0.1	1735 ± 10.1
Honey Gold							
2 *	2018	186	35.0	100.0	17.5	8.0	1740
4 *	2019	96	38.3	96.5	22.9	9.0	1740
5a	2020	110	32.0	101.0	17.2	7.2	1747
5b	2020	114	35.3	99.5	20.2	8.5	1740
6a	2020	116	32.9	103.0	17.1	7.3	1747
6b	2020	94	33.5	101.0	18.8	8.0	1740
12	2021	26	34.0	98.7	18.9	8.0	1732
13	2021	80	37.0	98.5	17.2	8.7	1733
17	2021	30	34.9	100.0	16.2	8.0	1757
21	2021	30	29.0	103.2	17.9	6.1	1740
Mean ± SE			34.2 ± 0.8	100.1 ± 0.6	18.4 ± 0.6	7.7 ± 0.2	1741.6 ± 2.2
Keitt							
7	2020	142	51.0	95.4	17.0	2.0	2230
8	2020	98	51.0	94.4	16.8	2.0	2142, 2233
9	2020	162	54.9	96.1	15.9	2.1	2297
18	2021	30	55.0	101.9	14.0	1.9	2185
19	2021	30	55.1	96.8	15.3	2.1	2188
22	2021	10	48.4	96.2	17.4	2.0	2185
Mean ± SE			52.5 ± 1.2	96.5 ± 0.8	16.1 ± 0.3	2.0 ± 0.0	2209 ± 8.0
KP							
15	2021	26	32.0	99.4	18.2	6.7	1602
20	2021	4	26.0	104.8	14.7	5.0	1638
Mean ± SE			29.0 ± 1	102.1 ± 1.9	16.5 ± 1.2	6.0 ± 0.6	1620 ± 12.7

* Populations marked with an asterisk were not used in the validation set as results were based on interpolated data for Calypso based on CIE B 32 and for Honey Gold on GDD of 1740. # Refers to population codes in Table A1.

Of the two KP populations, one failed to meet the B value specification of 32 (Table 8). DMC met the specification minimum of 14.0% *w/w* in both cases, at 14.7 and 18.2% *w/w*. The failure of one KP population to achieve CIE B specification at GDD 1600 is attributed to a sampling issue. Only two fruits remained from an initial tagging of 100 panicles for each event, and those fruits were located inside the canopy. Further work is required to confirm the recommended KP GDD.

The recommended Calypso GDD target (on Tb 12 °C, from asparagus) of 1728 is 48 units greater than the Moore [2] specification of 1680. This difference will be achieved in four days in the hotter temperatures prevailing near harvest.

The recommended Keitt GDD target (on Tb 12 °C, from asparagus) of 2185 is much greater than the Moore [2] specification of 1680, but it is consistent with the recommendation of Osuna-Garcia [10] of between 2100 and 2200 GDD (average 2150) on a Tb of 10 °C for Keitt to reach harvest maturity from Christmas tree stage. At an average GDD accumulation rate of 18 units per day, 2150 units is achieved in 107 days. Assuming *T_{min}* is always >12 °C, the equivalent GDD on a Tb = 12 °C is 2150 – (107 × 2) = 1936. Adjusting for the asparagus to Christmas tree development time yields a GDD requirement of

1936 + 240 = 2176 on Tb of 12 °C from asparagus stage. This value is consistent with the recommendation of the current study (2185).

Honey Gold fruit harvest targeted a GDD of 1744. In practice, harvests of the four validation populations occurred at GDD values ranging from 1732 to 1757, with an average of 1740 (Table 8). The flesh colour target of CIE B = 36 for Honey Gold, as established in the calibration exercise, was validated on the 2021 data. B values between 34 and 37 were achieved across four 2021 validation populations, however one population (pop 21) achieved a value of only 29. This result is attributed to the low number of samples in this population ($n = 5$) which resulted from a chilling event injuring 95% of the tagged panicles. The few remaining panicles were positioned inside the canopy and can be expected to experience a cooler microclimate and delayed maturation. Fruit DMC met the specification minimum of 14% *w/w* in all populations, varying from 16.2 to 22.9% *w/w* across the eight populations. The GDD target of 1744 is therefore recommended for use with Honey Gold, being associated with a B value of 36 (ranging from 34 to 37). This is a relatively small change on the previously recommended GDD of 1800, being equivalent to a three (summer) calendar days earlier harvest.

4. Conclusions

A recommendation of a methodology to follow in establishing the GDD of fruit development of mango cultivars has been presented. The Upper T temperature method (Equation (2)), using a $T_b = 12$ °C and $T_B = 32$ °C, is recommended over the standard method for estimation of GDD between flowering and fruit harvest maturity, particularly for lower latitude sites, although further validation is warranted. The availability of GDD values collected using a common methodology will facilitate cultivar comparisons (e.g., for selection of cultivars to achieve a desired market window) and in evaluation of the heritability of the trait.

The current study improves on existing GDD recommendations for mango reproductive development for four Australian grown cultivars (Kensington Pride, Calypso, Honey Gold and Keitt), with optimization of T_b and T_B values. Recommendations on cultivar specific minimum maturity specifications are given in Table 11. The GDD from Christmas tree to harvest maturity was calculated by subtraction of 180 from the asparagus target for Australian cultivars KP, Honey Gold and Calypso, and 240 for Keitt. The Honey Gold recommendation is approximately 60 units higher, or approx. five calendar days, to the 1500 units recommendation of Winston et al. [4]. The Calypso recommendation is 48 units higher, or approx. four calendar days, to the 1680 units recommendation of Moore et al. [2].

Table 11. Recommended minimum harvest maturity specifications by cultivar, based on use of the Upper T calculation of GDD.

Cultivar	CIE B	Colour Card Equivalent-(Table A3)	DMC (% <i>w/w</i>)	SSC: TA	GDD (from Asparagus Stage)	GDD (from Christmas Tree Stage)
KP	32 (29–34)	7	14.7	-	1600 *	1420 **
Calypso	32 (29–34)	7	16.0	6.5	1728	1540
Honey Gold	36 (33–39)	9	18.0	6.5	1740	1560
Keitt	51 (46–55)	13	16.0	6.5	2185	1936

* Diczbalis et al. [1] recommendation; ** Values extrapolated from GDD to reach Christmas tree stage.

The hardware and UpperT method recommended in the current study was implemented in major mango growing areas in Australia, with results viewable on-line given user entered flowering dates (<http://fruitronics.com/>, accessed on 1 November, 2022). Further work is required to confirm the KP recommendation, and additional work could be carried out for other mango cultivars such as R2E2, NamDocMai and the National Mango Breeding Program cultivars in Australia. Further work could also be carried out to establish a GDD range, involving documentation of loss of storage life with increased harvest GDD.

A single colour card set is recommended for assessment of flesh colour across all cultivars, including Keitt. Cards with scores of 11, 13, and 15 added to the existing ‘Calypso’ card set (DAF 2019 print), with CIE B values of 43.0, 51.0, and 58.0 (as illustrated in Table A3).

Further study could be carried out to confirm that the delay in maturation of within canopy fruit compared to external canopy fruit is due to a temperature difference. Also, there is variation in flower opening within a panicle, with flower opening typically begins at the base of the panicle and proceeding towards the tip over a period of a week or so, with consequent variation on pollination and fruit set on a given panicle. Future studies could quantify this variation, which adds uncertainty in the GDD forecast of harvest maturity.

There has been some debate within the mango industry on the relative merits of use of flesh colour and DMC in estimation of fruit maturation. As a non-destructive technique, more fruit can be sampled for NIR-DMC than can be destructively assessed for flesh colour. As harvest GDD approaches, it is recommended that growers use the non-destructive measure of NIR-DMC to select fruit of a range of DMC values, and thus maturities, from an orchard. This fruit can then be cut to assess flesh colour as a confirmation of harvest maturity status. The NIR-DMC of fruit at the harvest maturity flesh colour, as evaluated by comparison to colour charts or by use of a chromameter to measure CIE B value, can then be established. That NIR-DMC value can be used in non-destructive assessment of fruit harvest maturity for orchards with similar growing conditions.

Author Contributions: Conceptualization, M.H.A. and K.B.W.; methodology, M.H.A. and K.B.W.; investigation, M.H.A.; writing—original draft preparation, M.H.A. and K.B.W.; writing—review and editing, M.H.A., C.M., G.D. and K.B.W.; supervision, K.B.W.; project administration, K.B.W.; funding acquisition, K.B.W. All authors have read and agreed to the published version of the manuscript.

Funding: Funding for this project was provided by Hort Innovation from the Australian Government Department of Agriculture, Fisheries and Forestry as part of its Rural R&D for Profit program with Central Queensland University, UNE, Mangoes Australia, NT DITT, NSW DPI and DAF Qld, and by Perfection Fresh, Manbulloo and Pinata. Hort Innovation is the grower-owned, not-for-profit research and development corporation for Australian horticulture. MA acknowledges receipt of a CQU International tuition fee waiver and a living allowance scholarship through Hort Innovation project MG22000.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Acknowledgments: The work formed part of the Master’s Thesis of M.A. M.A. acknowledges the contribution in data collection and support given from Maira Aparecida and Paulo Dantas, Agrodan, Brazil, Martina Matzner from Acacia Hills Farms Ltd., Australia. The contribution of Nicholas Anderson in collection of data in 2018 and 2019 and in training for destructive methodologies is acknowledged.

Conflicts of Interest: The authors declare no competing interests in the undertaking of this work.

Appendix A. Panicle Tagging Exercises

Table A1. Populations and tagging exercises across different sites and seasons. All panicles were tagged at asparagus stage except as noted.

Pop (#)	Cultivar	Region	Tagging Date	Panicle Number	Fruit Number	Retention (%)	Harvest Dates	Comments
2018								
1	Calypso	Darwin, NT	29-May	269	95	35	20/09	40 panicles tagged at asparagus, 20 at elongation, 30 at Christmas tree, 5 at fruit set, GDD between asparagus and each stage estimated
2A	Honey Gold	Bugundarra, QLD	10-Jul	201	31	15	18/12, 27/12, 31/12	
2B	Honey Gold	Bugundarra, QLD	17-Jul	291	73	25	18/12, 27/12, 31/12	
2C	Honey Gold	Bugundarra, QLD	26-Jul	92	47	51	18/12, 27/12, 31/12	
2D	Honey Gold	Bugundarra, QLD	4-Aug	280	30	11	18/12, 27/12, 31/12	
2E	Honey Gold	Bugundarra, QLD	18-Aug	138	28	20	18/12, 27/12, 31/12	
2019								
3A	Calypso	Darwin, NT	22-May	50	12	24	4/10	
3B	Calypso	Darwin, NT	29-May	50	24	48	4/10	
3C	Calypso	Darwin, NT	5-Jun	50	15	30	4/10	
3D	Calypso	Darwin, NT	12-Jun	50	11	22	4/10	
3E	Calypso	Darwin, NT	19-Jun	50	9	18	4/10	
3F	Calypso	Darwin, NT	26-Jun	50	21	42	4/10	
3G	Calypso	Darwin, NT	3-Jul	50	1	2	4/10	
4A	Honey Gold	Bungundarra, QLD	16-Jul	113	16	14	20/12	
4B	Honey Gold	Bungundarra, QLD	25-Jul	60	10	17	20/12	
4C	Honey Gold	Bungundarra, QLD	2-Aug	18	7	39	20/12	
4D	Honey Gold	Bungundarra, QLD	14-Aug	30	13	43	20/12	
4E	Honey Gold	Bungundarra, QLD	30-Aug	24	2	8	20/12	

Table A1. Cont.

Pop (#)	Cultivar	Region	Tagging Date	Panicle Number	Fruit Number	Retention (%)	Harvest Dates	Comments
2020								
5A	Honey Gold	Bungundarra, QLD	5-Jul	600	55	9	03/12, 10/12, 17/12, 23/12, 28/12/2020, 07/01, 15/01/2021	
5B	Honey Gold	Bungundarra, QLD	5-Aug	600	57	10	03/12, 10/12, 17/12, 23/12, 28/12/2020, 07/01, 15/01/2021	
6A	Honey Gold	Bungundarra, QLD	5-Jul	600	58	10	03/12, 10/12, 17/12, 23/12, 28/12/2020, 07/01, 15/01/2021	
6B	Honey Gold	Bungundarra, QLD	13-Aug	600	47	8	03/12, 10/12, 17/12, 23/12, 28/12/2020, 07/01, 15/01/2021	
7	Keitt	Bungundarra, QLD	5-Aug	300	71	24	22/12, 29/12/2020, 07/01, 13/01, 20/01, 29/01, 04/02/2021	
8	Keitt	Bungundarra, QLD	25-Aug	200	49	25	20/01, 29/01, 04/02, 11/02, 10/03/2021	
9	Keitt	Belem do Sao Francisco, Brazil	6-Aug	300	81	27	28/12, 31/12/2020, 05/01/2021	Tagged by farm staff, first samples were cool stored for a week before assessing
2021								
10	Calypso	Darwin, NT	4-Jun	100	44	44	28/09, 01/10, 04/10	
11	Calypso	Darwin, NT	4-Jun	100	27	27	23/09	tagged at Christmas tree stage
12	Honey Gold	Darwin, NT	8-Jul	50	13	26	27/10	
13	Honey Gold	Katherine	2-Jul	100	40	40	26/10	
14	Calypso	Katherine, NT	15-Jun	100	9	9	20/10	
15	KP	Katherine, NT	15-Jun	50	13	26	7/10	
16	Calypso	Dimbulah, QLD	30-Jun	100	29	29	25/11, 29/11	5 fruit were not destroyed at harvest and used in a ripening exercise
17	Honey Gold	Dimbulah, QLD	30-Jun	100	20	20	29/11	5 fruit were not destroyed at harvest and used in a ripening exercise

Table A1. Cont.

Pop (#)	Cultivar	Region	Tagging Date	Panicle Number	Fruit Number	Retention (%)	Harvest Dates	Comments
18	Keitt	Belem do Sao Francisco, PE, Brazil	16-Jun	100	32	32	8/11	17 fruit do not have CIE LAB readings
19	Keitt	Curaca, BA, Brazil	18-Jun	100	24	24	15/11	9 fruit do not have CIE LAB readings
20	KP	Bungundarra, QLD	5-Jul	50	2	4	1/12	all internal fruit (external fruit loss from chilling injury)
21	Honey Gold	Bungundarra, QLD	24-Jun	100	5	5	8/12	all internal fruit (external fruit loss from chilling injury)
22	Keitt	Bungundarra, QLD	5-Jul	100	5	5	5/01	suffered loss from bacterial black spot

Appendix B. Flesh Colour as Maturity Targets

Table A2. CIE B value of mango harvest maturity colour cards. Data is presented for a Calypso card set of the original (2010) and of a second printing (2019), and values from the pdf of the file sent to the printer for the original printing. However, the cards of the second printing had been heavily used in field and were ‘aged’. Data for Keitt is from the pdf file of card swatches and from print. Data is presented of three original Kensington Pride ‘business cards’. Mean and SE of three replicate readings is presented.

DAF Calypso Picking Guide	Card Set 1 (2019 Version)– Second Printing	Card Set 2 (2010 Version)–Original Printing	Card Set 3 (2010 Version)-Digital
3	19.1 ± 0.2	17.5 ± 0.3	17.0 ± 0.0
5	25.7 ± 0.1	22.5 ± 0.3	26.0 ± 0.0
7 *	32.4 ± 0.1	26.2 ± 0.5	34.0 ± 0.0
9	35.8 ± 0.1	34.6 ± 0.5	43.0 ± 0.0
11	41.6 ± 0.1	43.1 ± 0.6	not available
KP ‘business’ card	Card set 1	Card set 2	Card set 3
Mature mango *	31.0 ± 0.0	30.9 ± 0.0	31.0 ± 0.0
Keitt, US Mango Board Maturity and Ripeness guide	Digital version	Printed 1	Printed 2
1	43.0 ± 0.1	34.4 ± 0.2	34.3 ± 0.2
2 *	51.0 ± 0.1	45.8 ± 0.2	45.8 ± 0.2
3	58.0 ± 0.1	48.5 ± 0.2	48.6 ± 0.2
4	66.0 ± 0.1	50.5 ± 0.2	50.5 ± 0.2
5	75.0 ± 0.1	53.0 ± 0.2	52.9 ± 0.2

* The colour recommended as denoting maturity is denoted by an asterisk.

Table A3. Proposed fruit maturity colour swatches and associated CIE LAB values, with maturity targets specific to cultivar. CIE L have two proposed values separated by slash (/), the first for printing purposes based on use of a PDF document, the second being the expected CIE L reading of fruit flesh.

3 CIE L = 97.00/85.00 CIE A = -4.40 CIE B = 18.00	5 CIE L = 97.00/85.00 CIE A = -5.20 CIE B = 26.00	7 (Calypso/KP) CIE L = 97.00/85.00 CIE A = -5.70 CIE B = 32.00	9 (Honey Gold) CIE L = 95.00/84.00 CIE A = -5.90 CIE B = 36.00
11 CIE L = 95.00/84.00 CIE A = -4.70 CIE B = 43.00	13 (Keitt) CIE L = 92.00/83.00 CIE A = -3.80 CIE B = 51.00	15 CIE L = 92.00/83.00 CIE A = -1.08 CIE B = 57.00	17 CIE L = 88.00/80.00 CIE A = 1.60 CIE B = 62.00

References

1. Diczbalis, Y.; Landrigan, M.; Wicks, C. Heat Sums to Predict Fruit Maturity in Mango (cv. Kensington Pride). A Report for Acacia Hills Farm Pty. Ltd. and the Horticultural Research and Development Corporation. HRDC Ref Number: FR605; 1997. Available online: <https://catalogue.nla.gov.au/Record/2936748/Holdings> (accessed on 1 March 2022).
2. Moore, C. Developing a Crop Forecasting System for the Australian Mango Industry. Regional Development, Primary Industry, Fisheries and Resources, Project Number: MG05004. 2010. Available online: <https://www.horticulture.com.au/globalassets/hort-innovation/historic-reports/developing-a-crop-forecasting-system-for-the-australian-mango-industry-mg05004.pdf> (accessed on 1 March 2022).
3. Hofman, P. Development of Best Practice Pre and Postharvest Protocols for Production of Calypso Mango: Phase 2. Sunshine Horticultural Services Pty Ltd., Project Number: MG06005. 2011. Available online: <https://www.horticulture.com.au/globalassets/hort-innovation/historic-reports/development-of-best-practice-pre-and-post-harvest-protocols-for-production-of-calypso-mango---phase-2-mg06005.pdf> (accessed on 1 February 2022).
4. Winston, E.C.; Hofman, P.; Macnish, A.; Marques, R.; Scurr, R. *Improving Fruit Quality and Profitability of 'Honey Gold'™ Mango*; Tropical Horticultural Consulting Pty Ltd., Project Number: MG10009.; Horticulture Australia Ltd.: Sydney, Australia, 2014; Available online: <https://www.horticulture.com.au/globalassets/laserfiche/assets/project-reports/mg10009/mg10009-final-report-complete.pdf> (accessed on 1 February 2022).
5. Castro, W.C.; Rodrigues, J.C.; de Sousa, A.M.L. Thermal necessity of mango trees from northeast Para's state, Brazil. *Rev. Bras. De Agric. Irrig.* **2017**, *11*, 1116–1126. [[CrossRef](#)]
6. Rodrigues, J.C.; Souza, P.J.O.; Lima, R.T. Estimation of basal temperatures and thermal requirement in mango production at the Northeast of Para state, Brazil. *Rev. Bras. Frutic. Jaboticabal SP* **2013**, *35*, 143–150. [[CrossRef](#)]
7. Barros, M.; Zanetti, V.; Fraga, C.; Nince, P.; Campelo Júnior, J.; Lobo, F. Photothermal units and lower base temperatures in fruit of mango tree variety "Alfa", in Baixada Cuiabana. *Rev. Bras. De Frutic.* **2010**, *32*, 479–485. [[CrossRef](#)]
8. Callejas, I.J.A.; Neves, G.A.R.; Tavares, A.D.S.; de Moura, I.B.; de Lima, E.A. Determination of the cardinal temperatures of mango cultivar 'Roxa' through computer simulation using a nonlinear model. *Ambiência* **2014**, *10*, 97–110. [[CrossRef](#)]
9. Lemos, L.M.; Salomão, L.C.; Siqueira, D.L.; Pereira, O.L.; Cecon, P.R. Heat unit accumulation and inflorescence and fruit development in 'ubá' mango trees grown in Visconde do rio branco-MG. *Rev. Bras. De Frutic.* **2018**, *12*, 40. [[CrossRef](#)]
10. Osuna-Garcia, J.A. *Validation of the Heat Unit's Technique to Determine the Optimum Harvest Time on Main Exporting Mango Varieties*; Agreement Instituto Nacional de Investigaciones Forestales, Agrícolas y Pecuarias and National Mango Board, USA; Burleigh Dodds Science Publishing: Santiago Ixcuintla, Mexico, 2019.
11. Zagade, V.V.; Pujari, K.H. Effect of period of maturity on physical characters and heat units required of mango (*Mangifera indica* L.) cv Alphonso. *Plant Arch.* **2014**, *14*, 835–840.
12. Halepotara, F.H.; Kanzaria, D.R.; Rajatiya, J.H.; Solanki, M.B.; Dodiya, K. Effect of heat unit and time duration required for maturation of mango (*Mangifera indica* L.) CV. Kesar. *J. Pharmacogn. Phytochem.* **2019**, *8*, 537–541.
13. De Freitas, S.T.; Guimarães I, T.; Vilvert, J.C.; Amaral, M.H.; Brecht, J.K.; Marques, A.T. Mango dry matter content at harvest to achieve high consumer quality of different cultivars in different growing seasons. *Postharvest Biol. Technol.* **2022**, *189*, 111917. [[CrossRef](#)]
14. NT. Farmers Association 2021. Available online: <https://ntfarmers.org.au/> (accessed on 1 February 2022).
15. The State of Queensland Department of Agriculture and Fisheries. Calypso Mango Picking Guide. 2019. Available online: <https://calypsomango.com.au/> (accessed on 31 January 2021).
16. National Mango Board. Mango Maturity and Ripening Guide. 2010. Available online: https://www.mango.org/wpcontent/uploads/2017/10/Mango_Maturity_And_Ripeness_Guide.pdf (accessed on 10 July 2020).
17. Anderson, N.; Subedi, P.; Walsh, K. Manipulation of mango fruit dry matter content to improve eating quality. *Sci. Hortic.* **2017**, *226*, 316–321. [[CrossRef](#)]
18. Walsh, K.B.; McGlone, V.A.; Han, D.H. The uses of near infra-red spectroscopy in postharvest decision support: A Review. *Postharvest Biol. Technol.* **2020**, *163*, 111139. Available online: <https://doi.org/10.1016/j.postharvbio.2020.111139> (accessed on 20 February 2022). [[CrossRef](#)]
19. Whiley, A.; Hofman, P. *Development of Best Practice Protocols for Production of Calypso™ Mango*; Final Report. Sunshine Horticultural Services Ltd. Project Number: FR02049; Horticulture Australia Ltd.: Nambour, QLD, Australia, 2006.
20. Henriod, R. Industry Moves on Quality Standards. *Mango Matters*, 2015; Volume 20, p. 10. Available online: <https://australian-mangoes.squarespace.com/resource-collection/2015/7/20/mango-matters-winter-2015?rq=Mango%20Matters%20-%20Winter%202015> (accessed on 1 December 2022).
21. Henriod, R.; Sole, D.; Wright, C.; Campbell, T. Determination of Eating Quality Standards for Mango Varieties 'Kensington Pride' and 'R2E2'. MG14504 Final Report, Department of Agriculture, and Fisheries, Queensland Government; 2015. Available online: <https://era.daf.qld.gov.au/id/eprint/6544/1/MG15002%20final%20report-543.pdf> (accessed on 1 December 2022).
22. Silva Neta, M.L. Recommendation of Portable Spectrometers for Monitoring "Keitt" Mango Quality Produced in 'Submedido Vale do São Francisco'. Master's Dissertation, Universidade Federal de Sergipe, São Cristóvão, SE, Brazil, 2019. Available online: <https://ainfo.cnptia.embrapa.br/digital/bitstream/item/209174/1/RECOMENDACAO-DE-ESPECTROMETROS-NIR-2019.pdf> (accessed on 30 September 2022).

23. Henriod, R.; Sole, D. Development of maturity standards for a new Australian mango cultivar. NMBP-1243. *Acta Hortic.* **2017**, *1183*, 17–22. [[CrossRef](#)]
24. Arnold, C.Y. Maximum-minimum temperatures as a basis for computing heat units. *Proc. Am. Soc. Hortic. Sci.* **1960**, *76*, 682–692.
25. Ometto, J. Bioclimatologia vegetal. In *Plant Bioclimatology*; Agronomica Ceres: São Paulo, Brazil, 1981; pp. 129–155.
26. BOM. The Australian Bureau of Meteorology. 2018. Available online: <http://www.bom.gov.au/climate/data/acorn-sat/> (accessed on 31 January 2021).
27. Amaral, M.H.; Walsh, K.B. In-orchard sizing of mango fruit: 2. Forward estimation of size at harvest. *Horticulturae* **2023**, *9*, 54. [[CrossRef](#)]
28. Yang, S.; Logan, J.; Coffey, D. Mathematical formulae for calculating the base temperature for growing degree days. *Agric. For. Meteorol.* **1995**, *74*, 61–74. [[CrossRef](#)]
29. Subedi, P.P.; Walsh, K.B.; Owens, G. Prediction of mango eating quality at harvest using short-wave near infrared spectrometry. *Postharvest Biol. Technol.* **2007**, *43*, 326–334. [[CrossRef](#)]
30. Tomlins, K.; Otori, C.; Bechoff, A.; Menya, G.; Westby, A. Relationship among the carotenoid content, dry matter content and sensory attributes of sweet potato. *Food Chem.* **2012**, *131*, 14–21. [[CrossRef](#)]
31. Amaral, M.H.P. Benchmarking New Methods for Estimation of Quantity and Harvest Timing of the Mango Crop. Master's Thesis, CQ University, Norman Gardens, Australia, 2022; pp. 174–178. Available online: https://figshare.com/articles/thesis/Benchmarking_new_methods_for_estimation_of_quantity_and_harvest_timing_of_the_mango_crop/21708602 (accessed on 10 April 2023).

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.