

Table S1. Primers used for cloning of *PhENDOI*, gene expression analysis by qPCR, and making constructs for VIGS experiments.

Primer Function	Direction	Sequence
RT-PCR clone	Forward	5'- TGGAGCAAAGARGGNCA -3'
	Reverse	5'- TTTCNCCRGCTTCRACACCTT -3'
RACE	Forward	5'- TGGCGACTTATCGGCCCTCTGCGTGTGG -3'
	Reverse	5'- ACTTGGTCCGGCCACACGCAGAGGGC -3'
Full length sequence	Forward	5'- TCATTGTGCTTCTTTGAGCATGTGGG -3'
	Reverse	5'- GACTTGTCATCTCGTGTTACATGGAGAGC -3'
<i>PhENDOI</i> qPCR	Forward	5'- CCACTTGCAGCAACTTGATCAACCA -3'
	Reverse	5'- TCGACGACGACGAACGATACTTGT -3'
<i>PhCP10</i> qPCR	Forward	5'- ACTTTGTGGACTTGCAACGGAAGC -3'
	Reverse	5'- CCAAGCCTATCTCAATCCCATACA -3'
<i>PhACTIN</i> qPCR	Forward	5'- AGCCAACAGAGAGAAGATGACCCA -3'
	Reverse	5'- ACACCATCACCAGAGTCCAACACA -3'
<i>PhRPS13</i> qPCR	Forward	5'- CAGGCAGGTTAAGGCAAAGC -3'
	Reverse	5'- CTAGCAAGGTACAGAAACGGC -3'
VIGS, Exp 1	Forward	5'- catctccatgTCAAGCCCTCTCCACTTCAT -3'
	Reverse	5'- catgtggtaccGTGCAAGTTTGACTTGTGCC -3'
VIGS, Exp 2	Forward	5'- tagtttgatccTGTCTTGCTTTCATTACCCAAC -3'
	Reverse	5'- cataattctagaTGCAAGCTTCATCTGGTGTAT -3'

1 ATGTTGGGGTTAACTTCATTAAGCATTAGTTTCTTTCTGTCTT
 M L G L T S L S I S F F L C L

 46 GCTTTCATTACCCAACATGATGTTGAAGCATGGAGTAAAGAGGGT
 A F I T Q H D V E A **↑** W S K E G

 91 CATATGATGACTTGTCGAATCGCCCAGGAGCTCTTGAATGATGAG
 H M M T C R I A Q E L L N D E

 136 GCAGCTCATGCGGTTAAGATGTTGTTACCGGATTATGTTAATGGC
A A H A V K M L L P D Y V N G

 181 GACTTATCGGCCCTCTGCGTGTGGCCGACCAAGTCCGGCATTGG
D L S A L C V W P D Q V R H W

 226 TACAAGTATAGGTGGTCAAGCCCTCTCCACTTCATTGATACACCA
 Y K Y R W S S P L H F I D T P

 271 GATGAAGCTTGCAACTTTGATTATGATAGGGATTGTCATGATGAA
 D E A C N F D Y D R D C H D E

 316 CATGGAGTGAAGGATATGTGTGTTGCTGGTGCCATCCAAAACTTT
 H G V K D M C V A G A I Q **N** F

 361 ACCACTCAGCTTTCCCATTACCCAGAGGGAAGTCCGATCGCCGA
 T T Q L S H Y P E G T S D R R

 406 TATAATATGACAGAGGCCTTGTTATTCTTGTCACATTTTCATGGGA
 Y **N** M T E A L L F L S H F M G

 451 GATATCCATCAACCGTTGCATGTTGGATTACAAAGTGATGAAGGA
 D I H Q P L H V G F T S D E G

 496 GGAAATACAATTAATCTGCGCTGGTTTAGGCACAAGTCAAACCTG
 G N T I N L R W F R H K S N L

 541 CACCATGTATGGGATAGAGAGATGATTCTACAAGCTGCAAAAGAC
H H V W D R E M I L Q A A K D

 586 TATTATGCAAAGGATGTAAACCTCCTTGAAGCAGACATTGAAGGA
 Y Y A K D V N L L E A D I E G

 631 AACTTCACTGATGGAATCTGGACTGATGATCTTTCTTCTTGAGG
N F T D G I W T D D L S S W R

 676 GAATGTGGAAATCTTCTCCTGTGTGAATAAGTTTGCTGCGGAA
E C G N L F S C V N K F A A E

 721 AGTATAAGTATAGCTTGCAAATGGGGATACAAAGGTGTTGAGGCT
S I S I A C K W G Y K G V E A

 766 GCGGAAACATTATCAGATGATTACTTCAATTCAAGACTCCCAATT
G E T L S D D Y F N S R L P I

 811 GTGATGAAAAGAATAGCTCAAGGTGGAGTACGATTAGCTATGCTT
V M K R I A Q G G V R L A M L

 856 CTAAATCGGGTTTTTGGAGATTCTCAAGAAGATCCACTTGCAGCA
 L N R V F G D S Q E D P L A A

 901 ACTTGA
 T *

Figure S1. The sequences of the *PhENDO1* coding region and the predicted protein. The *PhENDO* cDNA encodes a protein of 301 amino acids. The first 25 amino acids encode a signal peptide, and the arrow indicates the cleavage site of the signal peptide. The solid lines are the 14-12 endonuclease peptides that were sequenced from 2-DE [31]. Three predicted glycosylation sites are located at 119, 137, and 211 (gray boxes).

Figure S2. Alignment of the deduced amino acid sequences of endonuclease enzymes.

Endonuclease *PhENDO1* from petunia is compared with the amino acid sequences from *Nicotiana attenuata* NaEndonuclease1 (XP_019234897), *Solanum tuberosum* StEN1 (AAT79582), *Solanum lycopersicum* SlENDO1 (BAL03523), *Arabidopsis thaliana* ENDO1/BNF1, ENDO2, ENDO3, ENDO4 and ENDO5 (NP_172585, NP_176996, NP_001078420, NP_001328107 and NP_567631), *Zinnia elegans* ZEN1, ZEN2, and ZEN3 (BAA28948, AAD00694 and AAD00695), and *Hemerocallis* hybrid SA6 (AAC34856). Dark gray shading are residues that are identical in all sequences and light gray shading are residues that are functionally identical. Dashes indicate gaps introduced to produce the alignment, asterisks (*) are residues involved in the binding of zinc atoms, plus signs (+) identify residues involved in forming disulfide bonds; and number symbols (#) are structurally important glycosylation sites. Active sites for RNase and DNase activities in nucleases (His residues at positions 85 and 157, respectively, in PhENDO1) are also indicated under the alignment [17].

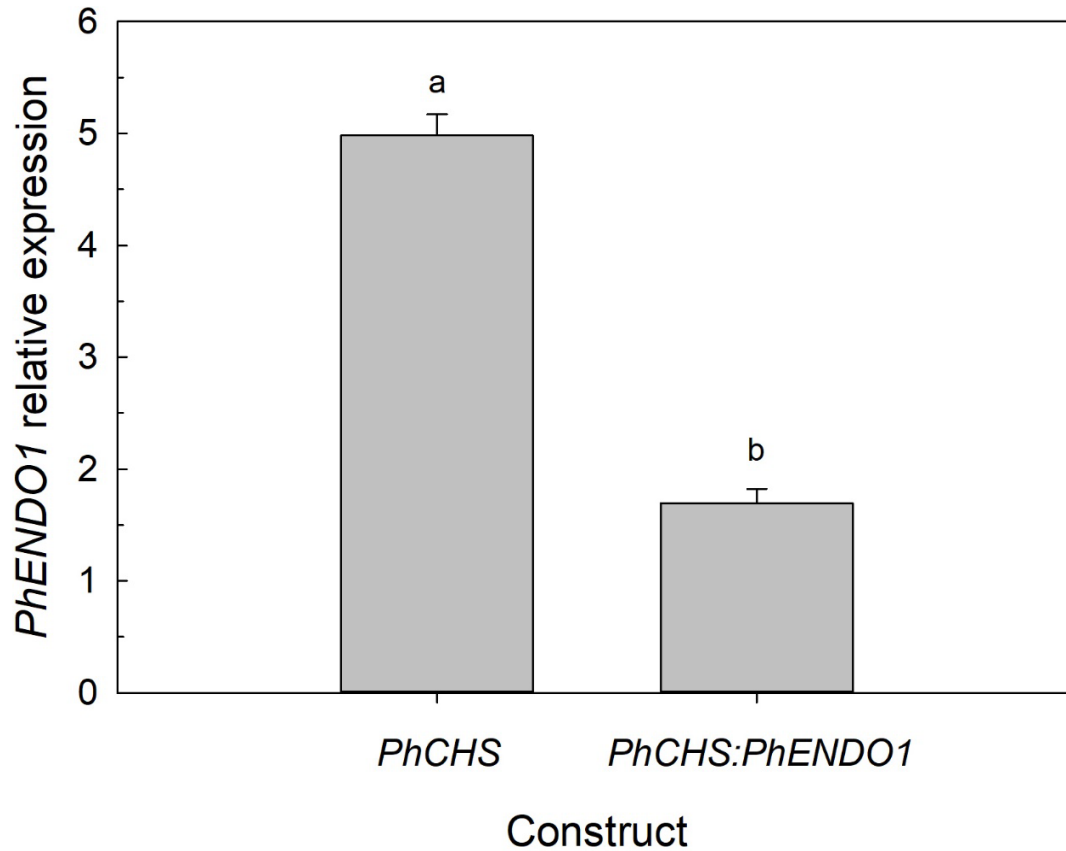


Figure S3. *PhEND1* transcript abundance was reduced in senescing flowers from pTRV2:*PhCHS:PhEND1* plants. Relative abundance of *PhEND1* in senescing corollas (48 h after pollination) was determined by qPCR in pTRV:*PhCHS* and pTRV2:*PhCHS:PhEND1* petunias. Bars represent mean $\pm$ SE (n=3) and least squared means were evaluated at $P < 0.05$. Different letters indicate significant difference between the means.