

*Article Supplementary Information*

# **Contribution of Grape Skins and Yeast Choice on the Aroma Profiles of Wines Produced from Pinot Noir and Synthetic Grape Musts**

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**Table S1.** Formulation of synthetic grape must (SGM) simulating Pinot noir grape juice.

component	chemical/solution	concentration
sugar and major components	D-glucose	115 g L <sup>-1</sup> *
	D-fructose	115 g L <sup>-1</sup> *
	DL-malic acid	3 g L <sup>-1</sup>
	citric acid	0.2 g L <sup>-1</sup>
	K <sub>2</sub> HPO <sub>4</sub>	1.14 g L <sup>-1</sup>
	MgSO <sub>4</sub> ·7H <sub>2</sub> O	1.23 g L <sup>-1</sup>
	CaCl <sub>2</sub> ·2H <sub>2</sub> O	0.44 g L <sup>-1</sup>
	potassium tartrate	5 g L <sup>-1</sup>
	MnCl <sub>2</sub> ·4H <sub>2</sub> O	198.2 µg L <sup>-1</sup>
	ZnSO <sub>4</sub> ·7H <sub>2</sub> O	287.5 µg L <sup>-1</sup>
minerals	CuSO <sub>4</sub> ·5H <sub>2</sub> O	25.3 µg L <sup>-1</sup>
	H <sub>3</sub> BO <sub>3</sub>	5.7 µg L <sup>-1</sup>
	CoCl <sub>2</sub> ·6H <sub>2</sub> O	23.8 µg L <sup>-1</sup>
	NaMoO <sub>4</sub> ·2H <sub>2</sub> O	24.2 µg L <sup>-1</sup>
	KIO <sub>3</sub>	10.8 µg L <sup>-1</sup>
	FeSO <sub>4</sub> ·7H <sub>2</sub> O	70.1 µg L <sup>-1</sup>
	pyridoxine-HCl	2 mg L <sup>-1</sup>
	nicotinic acid	2 mg L <sup>-1</sup>
	calcium pantothenate	1 mg L <sup>-1</sup>
	thiamine-HCl	0.5 mg L <sup>-1</sup>
vitamins	p-aminobenzoic acid	1 mg L <sup>-1</sup>
	riboflavin	1.2 mg L <sup>-1</sup>
	biotin	0.125 mg L <sup>-1</sup>
	myo-inositol	100 mg L <sup>-1</sup>
	folic acid	0.2 mg L <sup>-1</sup>
	L-alanine	146.27 mg L <sup>-1</sup> **
	L-arginine-HCl	513.52 mg L <sup>-1</sup> **
	L-aspartic acid	47.91 mg L <sup>-1</sup> **
	L-asparagine	6.03 mg L <sup>-1</sup> **
	L-cysteine	5.19 mg L <sup>-1</sup> **
amino acids and (NH <sub>4</sub> ) <sub>2</sub> HPO <sub>4</sub>	L-glutamic acid	118.24 mg L <sup>-1</sup> **
	L-glutamine	91.78 mg L <sup>-1</sup> **
	L-glycine	5.93 mg L <sup>-1</sup> **
	L-histidine	19.74 mg L <sup>-1</sup> **
	L-isoleucine	14.42 mg L <sup>-1</sup> **
	L-leucine	21.21 mg L <sup>-1</sup> **
	L-lysine-HCl	5.98 mg L <sup>-1</sup> **
	L-methionine	5.28 mg L <sup>-1</sup> **
	L-phenylalanine	15.79 mg L <sup>-1</sup> **
	L-proline	300 mg L <sup>-1</sup>
L-serine	L-serine	56.49 mg L <sup>-1</sup> **
	L-threonine	106.64 mg L <sup>-1</sup> **
	L-tryptophan	3.5 mg L <sup>-1</sup> **
	L-tyrosine	7.53 mg L <sup>-1</sup> **

	L-valine	20.63 mg L <sup>-1</sup> **
	(NH <sub>4</sub> ) <sub>2</sub> HPO <sub>4</sub>	185.23 mg L <sup>-1</sup> **
ergosterol	ergosterol	15 mg L <sup>-1</sup>
Tween 80	Tween 80	0.5 mL L <sup>-1</sup>
glutathione	glutathione	50 mg L <sup>-1</sup>

\*adjusted so that the total sugars in the SGM was equivalent to the Pinot noir must used in comparable experiments.

\*\*nitrogen components adjusted based on their average concentrations in five Marlborough 2018 Pinot noir juices, with a total YAN of 300 mg L<sup>-1</sup>.

**Table S2.** Ions and retention times (RTs) used to identify aroma compounds and internal standards.

	ions	RTs (mins)
<b>acetate esters</b>		
isobutyl acetate	43, <b>56</b> , 73	9.8
isoamyl acetate	70, 55, <b>87</b>	16.7
ethyl phenylacetate	91, <b>164</b>	47.9
$\beta$ -phenylethyl acetate	104, 43, 91	49.2
<b>alcohols, aldehydes</b>		
isobutanol	43, <b>41</b> , 74	9.3
1-butanol	<b>56</b> , 31, 41	10.9
isoamyl alcohol	55, 42, <b>70</b>	14.2
phenylethyl alcohol	<b>91</b> , 92, 122	51.0
methionol	<b>106</b> , 61, 58	41.1
benzyl alcohol	79, <b>43</b> , 91	49.2
benzaldehyde	77, <b>106</b> , 105	31.1
<b>fatty acid ethyl esters</b>		
ethyl isobutyrate	43, <b>71</b> , 116, 88	8.8
ethyl butanoate	71, <b>88</b> , 101	11.0
ethyl 2-methyl butanoate	57, <b>102</b> , 85	14.5
ethyl isovalerate	88, 85, 115	13.65
ethyl hexanoate	88, 99, 101	25.1
ethyl octanoate	88, <b>101</b> , 127	38.2
ethyl decanoate	88, 101, 155	51.2
<b>fatty acids</b>		
isobutyric acid	43, 73, 88	39.2
isovaleric acid	<b>60</b> , 87	33.6
hexanoic acid	60, 73, 87	48.5
octanoic acid	<b>60</b> , 73, 101	62.0
decanoic acid	60, 73, 129	69.0
<b>cinnamates</b>		
ethyl (di)hydrocinnamate	<b>104</b> , 91, 107	53.7

ethyl cinnamate ( <i>trans</i> )	<b>131</b> , 103, 176	63.88
<b>C<sub>13</sub>-norisoprenoids</b>		
β-damascenone	<b>69</b> , <b>121</b> , 190	53.4
α-ionone	<b>121</b> , 136, 192	55.8
β-ionone	<b>177</b> , 178, 192	59.6
<b>terpenes</b>		
<i>cis/trans</i> -rose-oxide	<b>139</b> , 69, 83	31.93/33.07
linalool	<b>71</b> , <b>93</b> , 121	36.278
α-terpineol	<b>59</b> , 93, 121	43.823
β-citronellol	<b>69</b> , 41, 82, 123	47.395
nerol ( <i>cis</i> -geraniol)	<b>69</b> , 41, 93, 121	48.505
geraniol ( <i>trans</i> -geraniol)	<b>69</b> , 41, 93, 123	51.493
<b>C<sub>6</sub>-compounds</b>		
hexanal	<b>44</b> , 56, 57, 72	11.5
<i>trans</i> -2-hexenal	<b>41</b> , 83, 69	16.5
hexanol	<b>56</b> , 43, 69	22.6
<i>trans</i> -3-hexen-1-ol	<b>41</b> , 67, <b>82</b>	23.597
<i>cis</i> -3-hexen-1-ol	<b>67</b> , 41, 82	25.337
<b>Internal standards</b>		
ethyl butyrate-4,4-d <sub>3</sub>	<b>74</b> , 89	10.9
hexanal-d <sub>12</sub>	<b>46</b> , <b>64</b> , 80	12.3
3-methylbutyl acetate-d <sub>3</sub>	<b>46</b> , <b>90</b>	15.35
ethyl hexanoate-d <sub>11</sub>	<b>91</b> , 110	23.3
<i>n</i> -hexyl-2,2,3,3,4,4,5,5,6,6,6-d <sub>11</sub> alcohol	<b>64</b> , 62, 76	23.4
<i>n</i> -hexyl acetate-d <sub>3</sub>	<b>46</b> , 56, 84	26.4
DL-3-octanol	<b>59</b> , 83, <b>101</b>	27.8
(±)-linalool-d <sub>3</sub>	<b>74</b> , 96, 124	36.187
ethyl octanoate-d <sub>15</sub>	<b>91</b> , <b>105</b>	37.5
4-decanol	<b>73</b> , <b>97</b> , 115	41.0
α-terpineol-d <sub>3</sub>	<b>62</b> , 139, 124	43.7
hexanoic-d <sub>11</sub> acid	<b>63</b> , 77, 93	47.8
2-phenylethyl acetate-d <sub>3</sub>	<b>46</b> , 91	49.0
3,4-dimethylphenol	<b>107</b> , 122	65.07

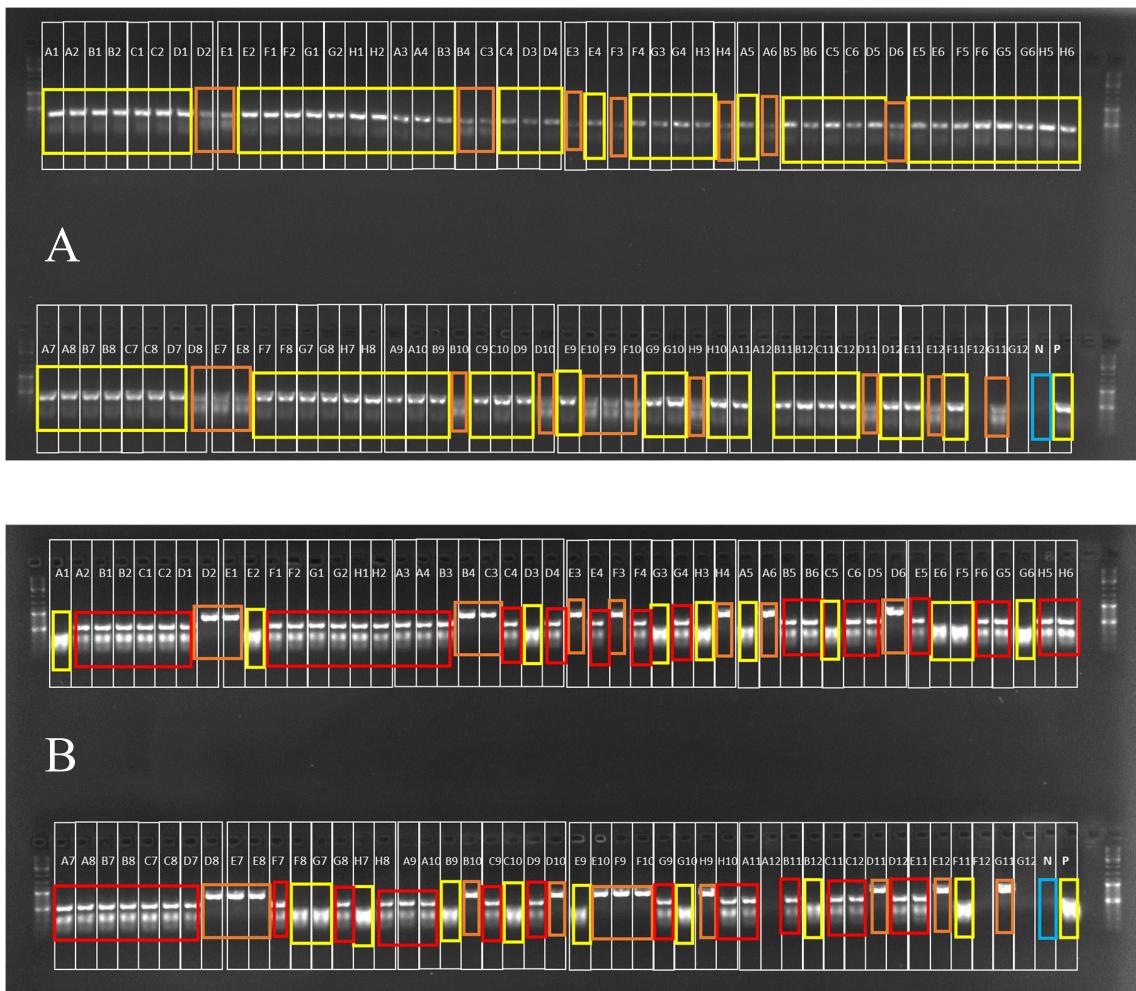
**Table S3.** Species identities of five MSPC isolates (from 96-well positions A3, B3, C3, G2 and H2) selected for Sanger sequencing to represent the three different cohorts identified for MSPC from restriction fragment length polymorphisms (RFLPs) of the ITS1-5.8S rDNA-ITS2 region. BLAST hits of each sequencing were used to identify the closest match for these isolates.

isolate	cohort	name	bit score	description	E value	grade
A3	2	MH041896	981.691	<i>Saccharomyces bayanus</i> strain SF5-310-4I2 26S ribosomal RNA gene, partial sequence	0	100%
A3	2	MH041895	981.691	<i>Saccharomyces uvarum</i> strain SF5-310-4II8 26S ribosomal RNA gene, partial sequence	0	100%
A3	2	LT594198	981.691	<i>Saccharomyces cerevisiae</i> x <i>Saccharomyces eubayanus</i> x <i>Saccharomyces uvarum</i> genomic DNA sequence contains ITS1, 5.8S rRNA gene, ITS2, 28S rRNA gene, strain NCAIM00677	0	100%
B3	1	MK722474	887.512	<i>Saccharomyces cerevisiae</i> strain FBKL2.8022 large subunit ribosomal RNA gene, partial sequence	0	100%
B3	1	MK722467	887.512	<i>Saccharomyces cerevisiae</i> strain FBKL2.8012 large subunit ribosomal RNA gene, partial sequence	0	100%
B3	1	MK722464	887.512	<i>Saccharomyces cerevisiae</i> strain FBKL2.8009 large subunit ribosomal RNA gene, partial sequence	0	100%
C3	2	MH041896	972.458	<i>Saccharomyces bayanus</i> strain SF5-310-4I2 26S ribosomal RNA gene, partial sequence	0	100%
C3	2	MH041895	972.458	<i>Saccharomyces uvarum</i> strain SF5-310-4II8 26S ribosomal RNA gene, partial sequence	0	100%
C3	2	LT009475	972.458	<i>Saccharomyces uvarum</i> partial 26S rRNA gene, strain VA12	0	100%
G2	3	MK581178	922.598	<i>Hanseniaspora uvarum</i> isolate FBKL2.9h702 large subunit ribosomal RNA gene, partial sequence	0	100%
G2	3	MK581177	922.598	<i>Hanseniaspora uvarum</i> isolate FBKL2.9696 large subunit ribosomal RNA gene, partial sequence	0	100%
G2	3	MK581174	922.598	<i>Hanseniaspora uvarum</i> isolate FBKL2.9633 large subunit ribosomal RNA gene, partial sequence	0	100%
H2	2	MH595098	900.439	<i>Saccharomyces uvarum</i> strain UCDFST:48-40 26S ribosomal RNA gene, partial sequence	0	100%
H2	2	MH041896	900.439	<i>Saccharomyces bayanus</i> strain SF5-310-4I2 26S ribosomal RNA gene, partial sequence	0	100%
H2	2	MH041895	900.439	<i>Saccharomyces uvarum</i> strain SF5-310-4II8 26S ribosomal RNA gene, partial sequence	0	100%

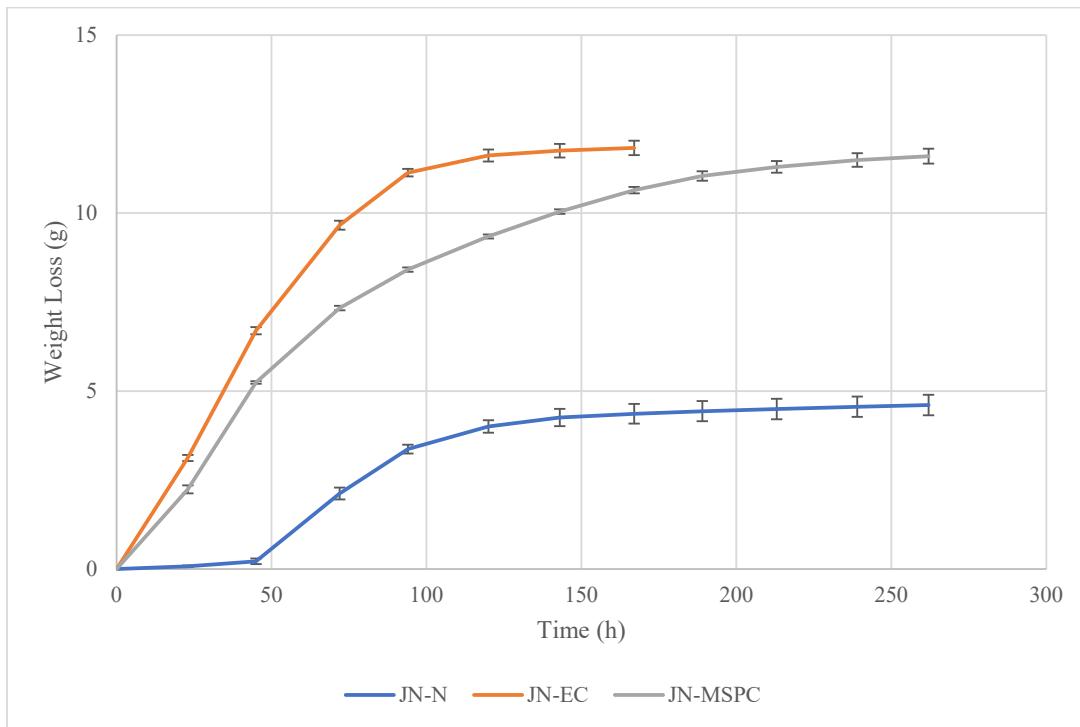
**Table S4.** Microsatellite genotypes of the 19 *S. cerevisiae* isolates from the MSPC community. NP: no peak. EC1118: *S. cerevisiae* EC1118.

sample name/index	A1	A5	B9	B12	C5	C10	D3	E2	E6	E9	F5	F8	F11	G3	G6	G7	G10*	H3	H7	H <sub>2</sub> O	EC1118	
allele size (bp)	YGL139W - 1	122	122	122	122	122	122	122	122	122	122	122	122	122	122	122	125	122	122	NP	114	
	YGL139W - 2	122	122	122	122	122	122	122	122	122	122	122	122	122	122	122	122	122	122	NP	122	
	YLL049W - 1	278	278	278	278	278	278	278	278	278	278	278	278	278	278	278	278	278	278	NP	278	
	YLL049W - 2	278	278	278	278	278	278	278	278	278	278	278	278	278	278	278	278	278	278	NP	291	
	YBR240C - 1	358	358	358	358	358	358	358	358	358	358	358	358	358	358	358	358	358	358	NP	358	
	YBR240C - 2	358	358	358	358	358	358	358	358	358	358	358	358	358	358	358	358	358	358	NP	358	
	YPL009C - 1	446	446	446	446	446	446	446	446	446	446	446	446	446	446	446	446	450	446	446	NP	423
	YPL009C - 2	446	446	446	446	446	446	446	446	446	446	446	446	446	446	446	446	450	446	446	NP	434
	MAT $\alpha$ - 1	457	457	457	457	457	457	457	457	457	457	457	457	457	457	457	457	461	457	457	NP	457
	YFR028C - 1	156	156	156	156	156	156	156	156	156	156	156	156	156	156	156	156	156	156	NP	122	
	YFR028C - 2	156	156	156	156	156	156	156	156	156	156	156	156	156	156	156	156	156	156	NP	132	
	YOL109W - 1	260	260	260	260	260	260	260	260	260	260	260	260	260	260	260	260	260	260	NP	311	
	YOL109W - 2	260	260	260	260	260	260	260	260	260	260	260	260	260	260	260	260	260	260	NP	311	
	MAT $\alpha$ - 1	481	481	481	481	481	481	480	481	481	481	481	480	480	481	481	480	481	481	NP	481	
	YGL014W - 1	146	146	146	146	146	146	146	146	146	146	146	146	146	146	146	146	146	146	NP	139	
	YGL014W - 2	146	146	146	146	146	146	146	146	146	146	146	146	146	146	146	146	146	146	NP	146	
	YML091C - 1	290	290	290	290	290	290	290	290	290	290	290	290	290	290	290	290	290	290	NP	270	
	YML091C - 2	290	290	290	290	290	290	290	290	290	290	290	290	290	290	290	290	290	290	NP	318	
	YDR160W - 1	393	393	393	393	393	393	393	393	393	393	393	393	393	393	393	393	393	393	NP	389	
	YDR160W - 2	393	393	393	393	393	393	393	393	393	393	393	393	393	393	393	393	393	393	NP	484	

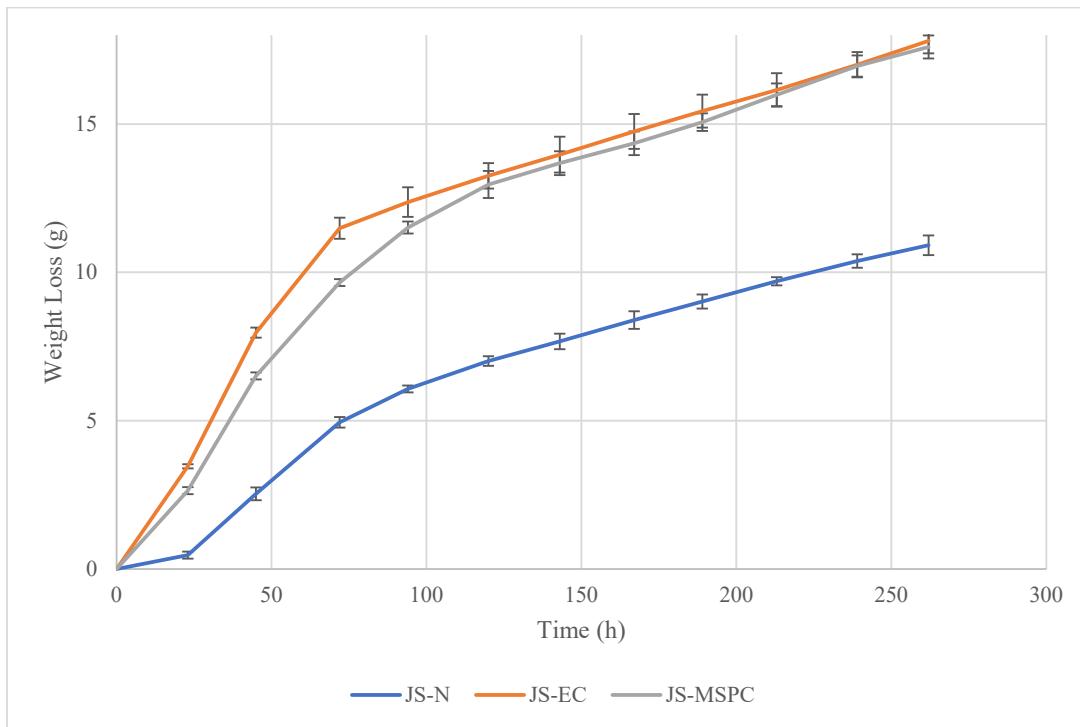
\*data acquired from a second run under the same protocol.



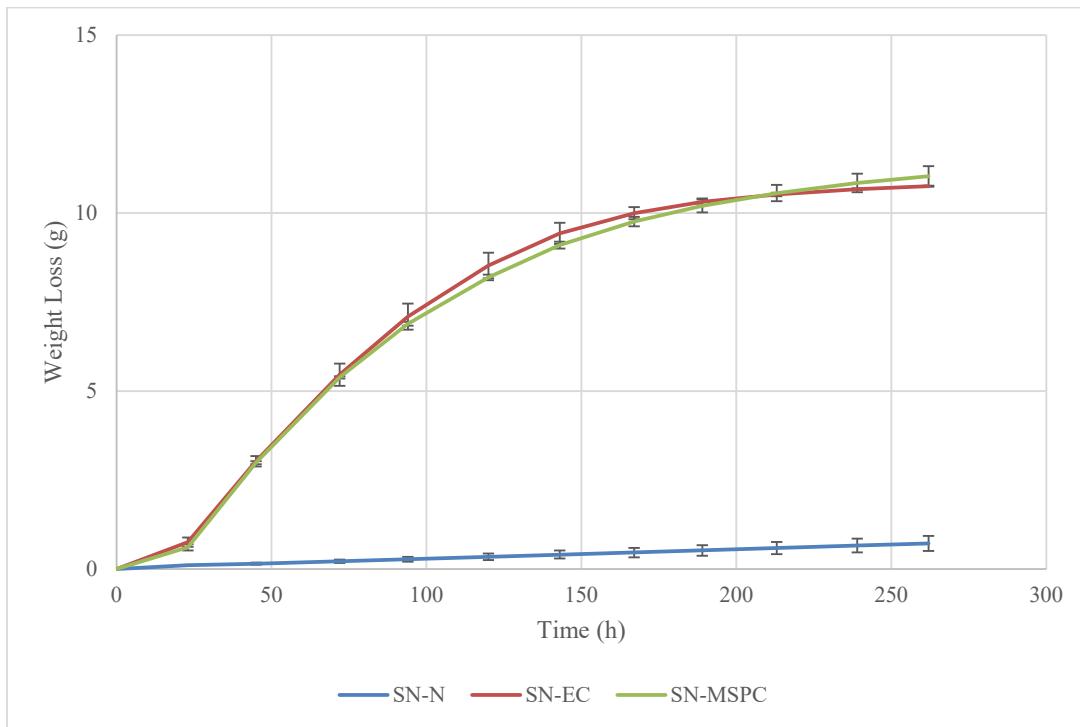
**Figure S1.** Cohort determination of the MSPC single vineyard community using the amplification of the ITS1-5.8S rDNA-ITS2 region, followed by RFLP analysis using *HinfI* (A) and *HaeIII* (B). The 1 kb+ ladder is shown to the left and right of all lanes. N = negative control (no DNA) surrounded by a blue box. P = *S. cerevisiae* positive control. Yellow boxes around *HinfI* and *HaeIII* digests represent cohort 1, *Saccharomyces cerevisiae*. Yellow *HinfI* and red *HaeIII* boxes represent cohort 2, *Saccharomyces uvarum*. Orange *HinfI* and *HaeIII* boxes represent cohort 3, *Hanseniaspora uvarum*.



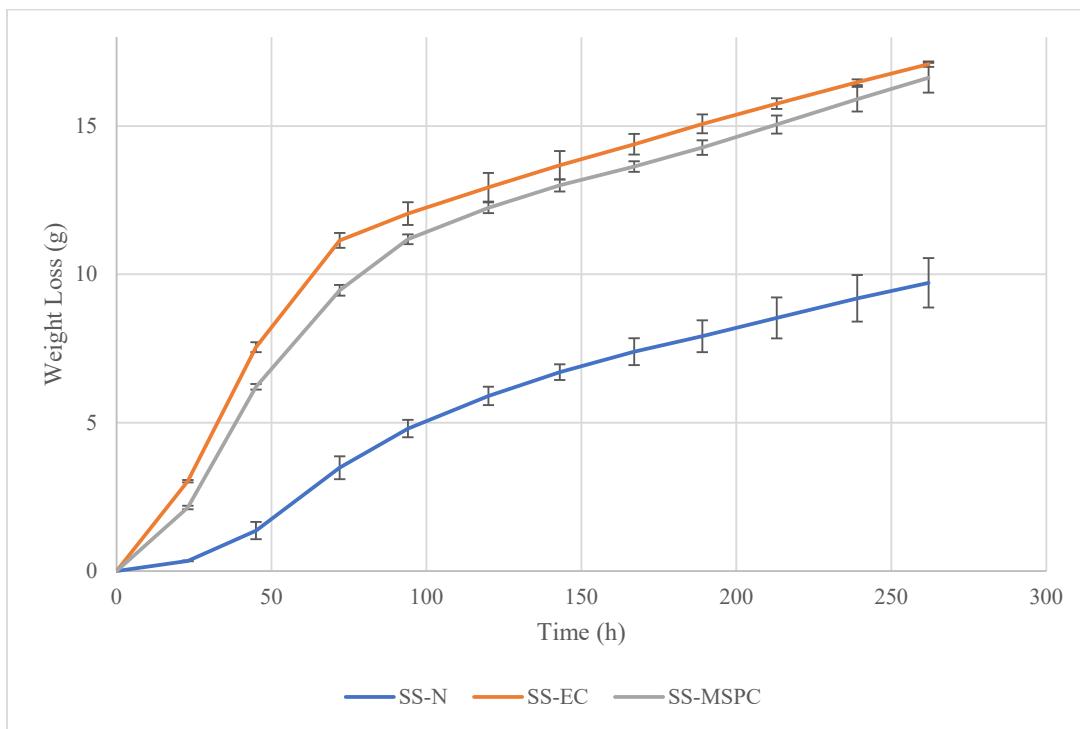
**Figure S2.** Cumulative weight loss (g) over time (h) of fermentations in Pinot noir juice without grape skins (JN). N = no inoculation, EC = inoculation of *S. cerevisiae* EC1118, MSPC = inoculation of the MSPC yeast community. Error bars represent the 95% confidence intervals, ( $n = 3$ ).



**Figure S3.** Cumulative weight loss (g) over time (h) of fermentations in Pinot noir juice with grape skins (JS). N = no inoculation, EC = inoculation of *S. cerevisiae* EC1118, MSPC = inoculation of the MSPC yeast community. Error bars represent the 95% confidence intervals, ( $n = 3$ ).



**Figure S4.** Cumulative weight loss (g) over time (h) of fermentations in synthetic grape medium (SGM) without grape skins (SN). N = no inoculation, EC = inoculation of *S. cerevisiae* EC1118, MSPC = inoculation of the MSPC yeast community. Error bars represent the 95% confidence intervals, ( $n = 3$ ).



**Figure S5.** Cumulative weight loss (g) over time (h) of fermentations in synthetic grape medium (SGM) with grape skins (SS). N = no inoculation, EC = inoculation of *S. cerevisiae* EC1118, MSPC = inoculation of the MSPC yeast community. Error bars represent the 95% confidence intervals, (n = 3).