

## Article

# Functional Significance of miR-4693-5p in Targeting HIF1 $\alpha$ and Its Link to Rheumatoid Arthritis Pathogenesis

Mohd Saquib <sup>1,2</sup>, Prachi Agnihotri <sup>1,2</sup>, Ashish Sarkar <sup>1,2</sup>, Swati Malik <sup>1,2</sup>, Sonia Mann <sup>1</sup>, Debolina Chakraborty <sup>1,2</sup>, Lovely Joshi <sup>1,2</sup>, Rajesh Malhotra <sup>3</sup> and Sagarika Biswas <sup>1,2,\*</sup> 

- <sup>1</sup> Council of Scientific and Industrial Research (CSIR), Institute of Genomics and Integrative Biology, Delhi University Campus, Mall Road, Delhi 110007, India; saquibmkn@gmail.com (M.S.); prachiagnihotri21@gmail.com (P.A.); ashish2sarkar@gmail.com (A.S.); swatimalik5577@gmail.com (S.M.); mannsonia23@gmail.com (S.M.); debochak55@gmail.com (D.C.); lovelyabhilasha98@gmail.com (L.J.)
- <sup>2</sup> Academy of Scientific and Innovative Research (AcSIR), Ghaziabad 201002, India
- <sup>3</sup> All India Institute of Medical Science (AIIMS), Ansari Nagar, New Delhi 110029, India; rmalhotra62@gmail.com
- \* Correspondence: sagarika.biswas@igib.res.in; Tel.: +91-11-27667602; Fax: +91-11-27667471 or +91-98-18-004740

**Abstract:** Rheumatoid arthritis (RA) is a chronic inflammatory autoimmune disease that causes joint inflammation and destruction with an unknown origin. Our study aims to elucidate the molecular mechanism behind HIF1 $\alpha$  overexpression in RA. Dysregulated miRNA expressions are known to influence gene behavior, thereby enhancing cell proliferation, inflammation, and resistance to apoptosis, contributing to RA development. Our earlier finding indicated that exogenous miRNA similar to miR-4693-5p may modulate RA-related targets. However, the specific role of miR-4693-5p and its targets in RA remain unexplored. In this study, we found that miR-4693-5p was significantly reduced in PBMCs of RA patients, with evidence suggesting it targets the 3' UTR of HIF1 $\alpha$ , thereby potentially contributing to its overexpression in RA. In vitro overexpression of miR-4693-5p leads to the knockdown of HIF1 $\alpha$ , resulting in inhibited expression of Survivin to disrupt apoptosis resistance, inflammation suppression, and a reduction in the total cellular ROS response in SW982 and RAFLS cells. The results were validated using the CIA Rat model. In conclusion, this study provides a crucial foundation for understanding the functional role of miR-4693-5p. These findings improve our understanding and provide novel insights into the molecular mechanisms underlying RA pathogenesis.

**Keywords:** miRNA; rheumatoid arthritis; inflammation; apoptosis; synovial fibroblast; SW982



**Citation:** Saquib, M.; Agnihotri, P.; Sarkar, A.; Malik, S.; Mann, S.; Chakraborty, D.; Joshi, L.; Malhotra, R.; Biswas, S. Functional Significance of miR-4693-5p in Targeting HIF1 $\alpha$  and Its Link to Rheumatoid Arthritis Pathogenesis. *Non-Coding RNA* **2024**, *10*, 22. <https://doi.org/10.3390/ncrna10020022>

Academic Editors: Y-h. Taguchi and Hsiuying Wang

Received: 6 March 2024

Revised: 26 March 2024

Accepted: 29 March 2024

Published: 10 April 2024



**Copyright:** © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Rheumatoid arthritis (RA) is a chronic inflammatory autoimmune disease that causes joint destruction with unknown origin. Approximately 1% worldwide and 0.92% of the Indian population are affected by RA [1]. Currently, anti-citrullinated protein antibodies (ACCPA) and rheumatoid factor (RF) are the most widely available non-specific markers, and treatments include DMARDs, NSAIDs, etc., which have severe side effects [2].

Previous reports showed that the high metabolic requirement of proliferative cells and intraarticular pressure generates an oxygen demand, causing hypoxia of the synovial joint. As a result, the prominent protein HIF1 $\alpha$  (Hypoxia Inducible Factor 1 Subunit Alpha) gets overexpressed, contributing to RA pathogenesis [3]. However, the molecular mechanism of HIF1 $\alpha$  overexpression in RA is unknown. As RA is affected by various factors like cytokines, immune cell activities, joint inflammation, and immune complex accumulation [4], it is necessary to study the possible regulatory molecule that targets HIF1 $\alpha$ . Therefore, we focused on knockdown studies of HIF1 $\alpha$  to understand the above underlying factors. Recently, the involvement of non-coding RNA (ncRNAs), particularly

microRNA (miRNA), regulating the gene expression associated with the disease was reported [5]. It was also reported that HIF1 $\alpha$  can be regulated by miRNAs and that miR-210 regulates HIF1 $\alpha$  negatively, affecting the dynamic equilibrium of Th17/Treg cells in RA [6]. Another report claimed that pharmacological inhibition of HIF1 $\alpha$  by a short hairpin RNA (shRNA) lentiviral expression vector improved the clinical manifestations of RA [7].

As miRNA is a potent gene expression regulator, we speculated that a specific miRNA might be involved in HIF1 $\alpha$  regulation. We previously reported that exogenous miRNA similar to miR-4693-5p may modulate RA-related targets [8,9]. The altered miRNA expressions are key controls on gene behavior, contributing to the development of inflammatory conditions [10]. For example, Legg-Calvé-Perthes is an inflammatory disease with high expression of miR-4693-5p that promotes endothelial cell dysfunction and osteoclastogenesis [11]. In another report, miR-4693-5p targeted thyroid hormone receptor-interacting protein 13 (TRIP) 13 to promote apoptosis and suppress cell proliferation [12]. In this study, we found that miR-4693-5p was significantly reduced in the peripheral blood mononuclear cells (PBMCs) of RA patients. This result led us to believe that miR-4693-5p may play a role in the pathogenesis of RA as an endogenous molecule, which has not been reported to date. Therefore, we focused our research on validating the functional role of miR-4693-5p, speculating that miR-4693-5p may be involved in RA pathogenesis. We hypothesized that employing endogenous molecules miR-4693-5p to modulate HIF1 $\alpha$  expression may play an important role in preventing RA development. Therefore, *in vitro* validation of miR-4693-5p using SW982 and rheumatoid arthritis fibroblast-like synoviocytes (RAFLS) cells, followed by *in vivo* validation using a collagen-induced arthritis (CIA) rat model, was attempted. Overexpression of miR-4693-5p confirms that it regulates HIF1 $\alpha$  directly, playing a role in maintaining homeostasis of RA joints by inducing apoptosis, proinflammatory cytokines, and total cellular reactive oxygen species (ROS) suppression. These findings improved our understanding and provided novel insights into the molecular mechanisms driving the pathogenesis of RA.

## 2. Results

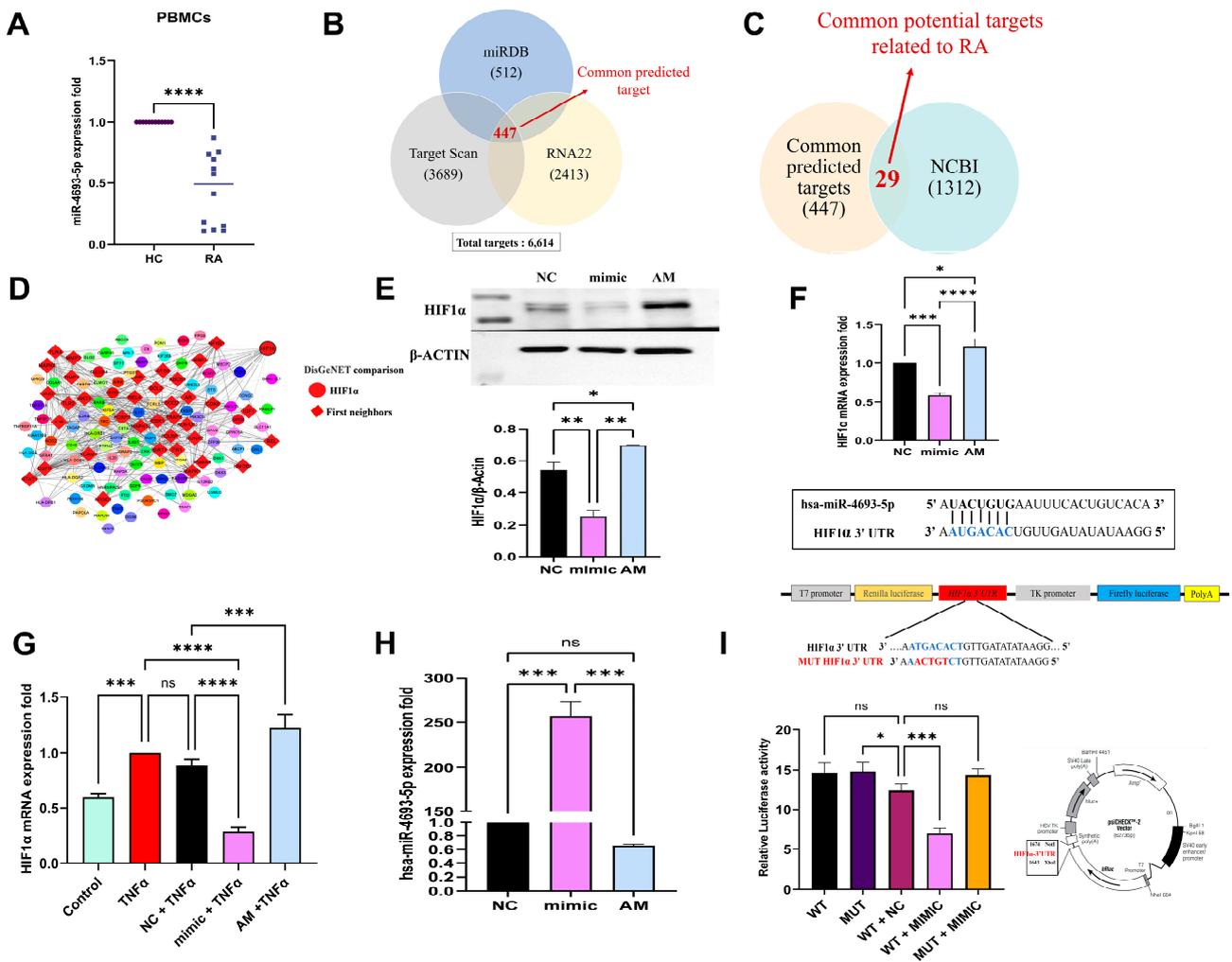
### 2.1. miR-4693-5p Expression Decreased in RA PBMCs

PBMCs can imitate conditions in synovial tissue, and their dysregulated miRNAs play important roles in RA pathogenesis and are employed as disease activity indicators [13]. Hence, we checked the expression of miR-4693-5p in PBMCs. We observed that the expression of miR-4693-5p was significantly downregulated in the PBMCs of RA ( $n = 12$ ) patients compared with Healthy Controls (HC,  $n = 12$ ) (Figure 1A), confirming the link between miR-4693-5p and RA.

Next, we explored the regulatory mechanism behind the reasons for the HIF1 $\alpha$  overexpression in RA. We considered miR-4693-5p as a potential regulator of HIF1 $\alpha$  expression because miRNA is known for its powerful influence on gene expression.

### 2.2. *In Silico* Target Prediction of miR-4693-5p

Target prediction analysis revealed 447 common targets from the three miRNA prediction databases: miRDB (512), TargetScan (3689), and RNA22 (2413) (Figure 1B) [14]. Targets were matched with RA-related genes acquired from NCBI (1312), yielding 29 RA-specific targets (Figure 1C). HIF1 $\alpha$  was selected because of its high total context score, 8mer binding site, and increased expression in RA. The 447 common targets were compared with DisGeNET (RA disease-related targets) [15] for better potential target selection. Therefore, 159 proteins having gene-disease association (GDA) scores of  $\geq 0.1$  were identified, and  $\sim 1/3$ rd of the proteins (first neighbors, red color) (Figure 1D) were found to be directly regulated by HIF1 $\alpha$ , showing an important regulator in the development of RA.



**Figure 1.** Target identification of miR-4693-5p: (A) Downregulated expression level of miR-4693-5p in RA PBMCs compared with Healthy Controls ( $n = 12$  each) by stem-loop PCR (B) Venn diagram of common 447 predicted targets identified using miRDB (512), TargetScan (3689), and RNA22 (2413). (C) Identified 29 common potential targets related to the RA database of NCBI represented by Venn diagram. (D) Cytoscape interpretation of first neighbor or directly regulated (in Red) targets of HIF1 $\alpha$ . (E) Western blot analysis showing significantly decreased HIF1 $\alpha$  levels (~2.2 fold) in miR-4693-5p transfected SW982 cells at 25 nM. (F) Downregulated HIF1 $\alpha$  mRNA expression by qRT-PCR in uninduced and (G) TNF $\alpha$ -induced miR-4693-5p-transfected SW982 cells at 25 nM. (H) Post-transfection overexpression levels of miR-4693-5p in transfected (25 nM) SW982 cells. (I) Dual luciferase assay of human HIF1 $\alpha$  3' UTR binding sequence of miR-4693-5p significantly downregulated (~2 fold) the luciferase activity in Luc-HIF1 $\alpha$ -WT 3' UTR. (NC: Negative Control, mimic: synthetic miR-4693-5p, AM: AntimiR, GAPDH: Glyceraldehyde 3-phosphate dehydrogenase, TNF- $\alpha$ : Tumor Necrosis Factor-Alpha, HC: Healthy Control, RA: Rheumatoid arthritis, WT: Wild type, MUT: Mutant, level of statistical significance: \* =  $p \leq 0.05$ , \*\* =  $p \leq 0.01$ , \*\*\* =  $p \leq 0.001$ , \*\*\*\* =  $p \leq 0.0001$ ).

### 2.3. miR-4693-5p Directly Targets HIF1 $\alpha$

HIF1 $\alpha$  is a transcription factor strongly expressed in the intimal lining of synovial tissue and plays a role in cell proliferation, metabolism, angiogenesis, and cell survival in RA [16]. In silico results revealed that miR-4693-5p may target HIF1 $\alpha$ . To validate the regulation of HIF1 $\alpha$  by miR-4693-5p, dose standardization experiments were performed. WB analysis of HIF1 $\alpha$  expression at various 5–50 nM concentrations was conducted (Supplementary Figure S1). Significant downregulation of HIF1 $\alpha$  expression at 25 nM was observed; therefore, this concentration was selected for further experiments.

Further, after transfection of miR-4693-5p in SW982 cells, significant downregulation of HIF1 $\alpha$  was revealed at 25 nM by WB (~2.2 fold) (Figure 1E) and qRT-PCR (Figure 1F) compared with the Negative Control (NC) and recovered in the presence of miRNA inhibitor (AM). A similar downregulated profile of HIF1 $\alpha$  (mimic + TNF $\alpha$ ) at mRNA level was also revealed in TNF $\alpha$ -induced SW982 cells compared with TNF $\alpha$  alone and Negative Control (NC) + TNF $\alpha$  (Figure 1G). An overexpressed level of miR-4693-5p was seen at 25 nM in transfected SW982 cells compared with the Negative Control (NC) (Figure 1H). These results indicate that miR-4693-5p significantly downregulates HIF1 $\alpha$  expression.

#### 2.4. miR-4693-5p Binds to 3' UTR of HIF1 $\alpha$

To validate that HIF1 $\alpha$  is a direct target of miR-4693-5p, a reporter vector containing a luciferase coding gene with predicted specific binding sites of miR-4693-5p was constructed. Wild type and mutated sequence 3' UTR of HIF1 $\alpha$  was constructed (Supplementary Figure S2) to check the regulation of miR-4693-5p. We found that miR-4693-5p significantly downregulates (~2 fold) the luciferase activity in Luc-HIF1 $\alpha$ -WT 3' UTR but not in Luc-HIF1 $\alpha$ -MUT 3' UTR (Figure 1I). Therefore, the results indicate that miR-4693-5p was directly bound to the seed region of HIF1 $\alpha$  3' UTR, ultimately reducing luciferase activity in the WT + mimic set.

#### 2.5. TNF $\alpha$ Induces HIF1 $\alpha$ Expression and Establishment of RA

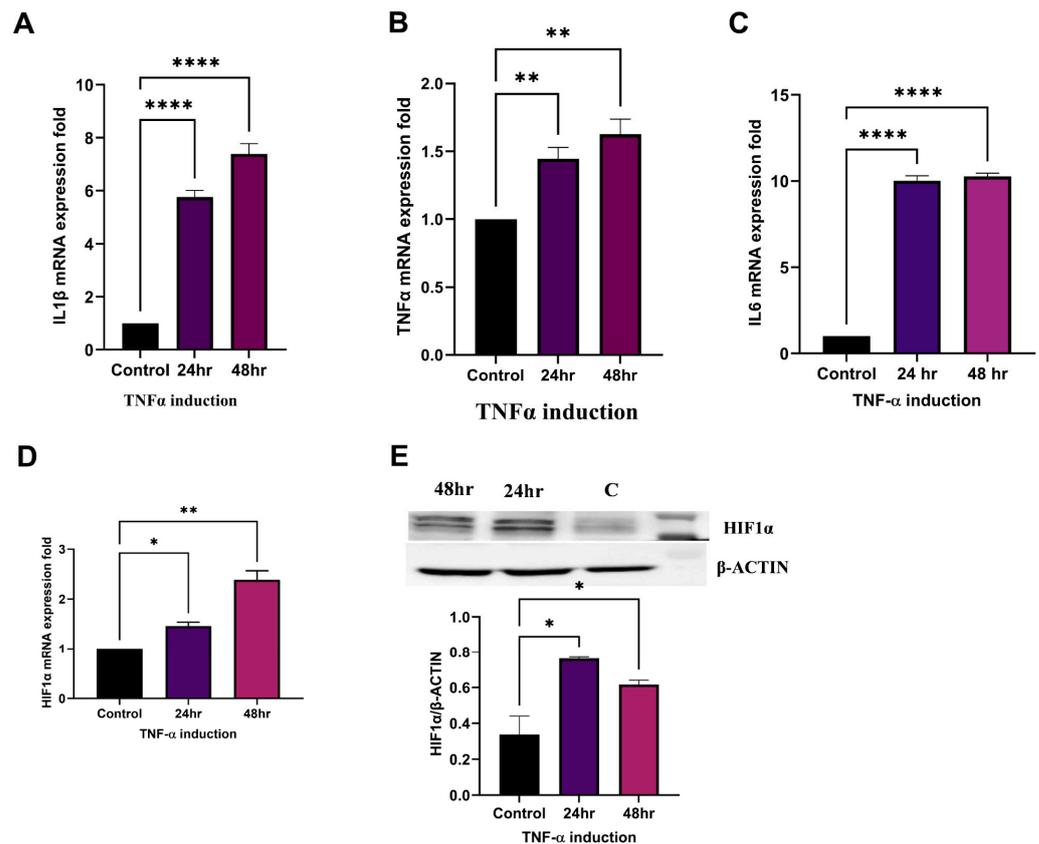
In order to mimic the conditions of RA, we assessed the levels of proinflammatory cytokines in TNF $\alpha$ -induced SW982 cells at concentrations of 10 ng/mL [17] measured at both 24 h and 48 h. The results showed increased mRNA levels of IL1 $\beta$  (Figure 2A), TNF $\alpha$  (Figure 2B), and IL6 (Figure 2C), confirming the establishment of an inflammatory state. These findings support the pivotal role of TNF- $\alpha$  in the pathogenesis of RA. HIF1 $\alpha$  helps cells to survive in low-oxygen conditions, potentially speeding up RA development. Surprisingly, even in normal oxygen levels, HIF1 $\alpha$  can be triggered by various parameters like thrombin, vasoactive peptides, growth factors, insulin, inflammatory agents (LPS), proinflammatory cytokines (TNF- $\alpha$ , IL-1 $\beta$ ), ROS, etc., thereby stabilizing HIF1 $\alpha$  protein (32). Our results show that upregulated levels of HIF1 $\alpha$  in TNF $\alpha$  induced SW982 cells at 24 and 48 h at both mRNA (Figure 2D) and protein levels compared with the controls (Figure 2E) (~2 fold). The results thus indicate that TNF $\alpha$  induces the expression of HIF1 $\alpha$ , contributing to the establishment of an inflammatory condition.

#### 2.6. In Silico Target Interaction of HIF1 $\alpha$

The literature suggests that HIF1 $\alpha$  is a critical transcriptional regulator of Survivin, facilitating binding to the Survivin promoter directly and stimulating its transcription [18]. Our protein–protein (PPI) interaction (STRING and STITCH databases) result demonstrated a significant role of HIF1 $\alpha$  in the pathogenesis of RA by directly interacting with proinflammatory cytokines (TNF $\alpha$ , IL6, IL1 $\beta$ ) and Survivin (BIRC5) (Figure 3A). The highest interaction score was observed between HIF1 $\alpha$  and Survivin (0.903) compared with proinflammatory cytokines IL1 $\beta$  (0.866), TNF $\alpha$  (0.650), and IL6 (0.504) (Figure 3B), indicating a direct relation of Survivin (downstream target) with HIF1 $\alpha$ . Survivin was, therefore, chosen for downstream analysis of miR-4693-5p to emphasize its potential significance in the regulatory mechanisms underlying RA.

#### 2.7. miR-4693-5p Reduced the Expression of Survivin, a Downstream Target of HIF1 $\alpha$

In normoxic conditions, HIF1 $\alpha$  regulates Survivin expression directly, and in RA, Survivin level is highly upregulated compared with Healthy Controls [18]. Therefore, the expression level of Survivin was checked again in the present study using ELISA of the plasma samples of RA ( $n = 8$ ) and HC ( $n = 8$ ), and an increased expression of Survivin was revealed, indicating its direct relation with disease pathogenesis (Supplementary Figure S3).



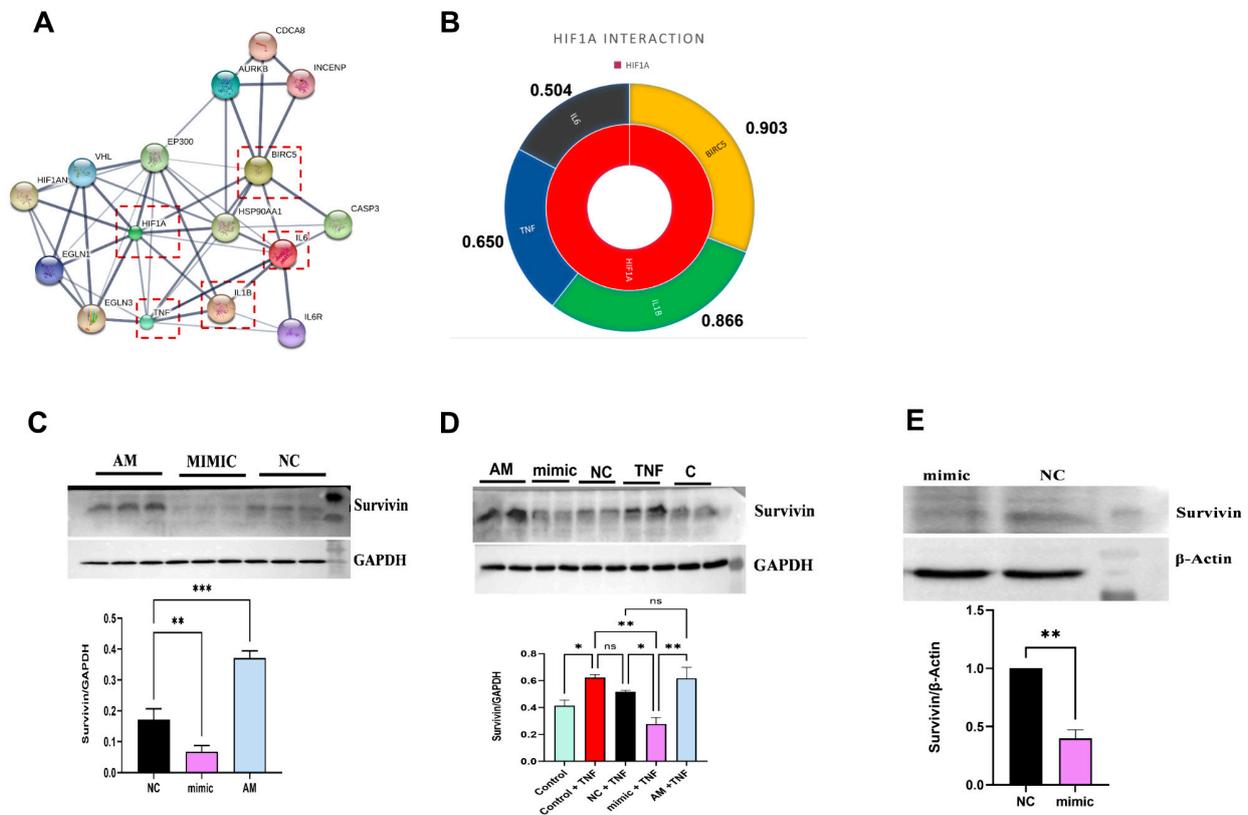
**Figure 2.** HIF1 $\alpha$  expression in TNF $\alpha$ -induced SW982 cells: Increased mRNA level of pro-inflammatory cytokines showing the establishment of inflammatory conditions. (A) IL1 $\beta$ , (B) TNF $\alpha$ , (C) IL6, and (D) HIF1 $\alpha$  mRNA expressions after induction with TNF $\alpha$  at 10 ng/mL for 24 h and 48 h in SW982 cells by qRT-PCR. (E) An increased level of HIF1 $\alpha$  (~2 fold) was examined after induction with TNF $\alpha$  at 10 ng/mL for 24 h and 48 h in SW982 cells using Western blot analysis, using  $\beta$ -actin as a loading control. (NC: Negative Control, mimic: synthetic miR-4693-5p, AM: AntimiR, GAPDH: Glyceraldehyde 3-phosphate dehydrogenase, TNF- $\alpha$ : Tumor Necrosis Factor-Alpha, level of statistical significance: \* =  $p \leq 0.05$ , \*\* =  $p \leq 0.01$ , \*\*\*\* =  $p \leq 0.0001$ ).

Further, significant downregulation of Survivin was revealed after transfection of miR-4693-5p in SW982 cells without (Figure 3C) (~2.6 fold) and with (Figure 3D) (~1.7 fold) TNF $\alpha$ -induction and also in RAFLS compared with the Negative Control (NC) (Figure 3E) (~2 fold). These findings strongly suggest that the knockdown of HIF1 $\alpha$  leads to significant inhibition of Survivin expression levels, both in the absence and presence of TNF $\alpha$  induction, in SW982 and RAFLS cells. This reinforces the intricate relationship between HIF1 $\alpha$ , Survivin, and the regulatory mechanisms contributing to the pathogenesis of RA.

## 2.8. miR-4693-5p Promotes Apoptosis in RA

Survivin is an antiapoptotic protein and is overexpressed in RA, inhibiting Cytochrome C's expression, proapoptotic proteins, and effector caspases [19]. To investigate the impact of miR-4693-5p on apoptosis, Western blot (Cyt C, Bax, and Bcl2) and Caspase-Glo 3/7 assays were attempted. The results showed upregulation of the proapoptotic protein CytC without (Figure 4A) (~1.6 fold) and with (Figure 4B) (~2 fold) TNF $\alpha$ -induced SW982 cells and also upregulation of Bax in TNF $\alpha$ -uninduced (Figure 4C) (~2 fold) and -induced (Figure 4D) (~1.5 fold) SW982 cells, with downregulation of antiapoptotic Bcl2 expression in TNF $\alpha$ -uninduced (Figure 4E) (~1.3 fold) and -induced (Figure 4F) (~1.5 fold) SW982 cells. Similarly, the upregulation of CytC (Figure 4G) (~1.3 fold) and Bax (Figure 4H) (~1.4 fold) and downregulation of Bcl2 (Figure 4I) (~1.3 fold) were also revealed by RAFLS. Further, the upregulation of relative luminescence (~1.2 fold) of caspase 3/7 (Figure 4J) in

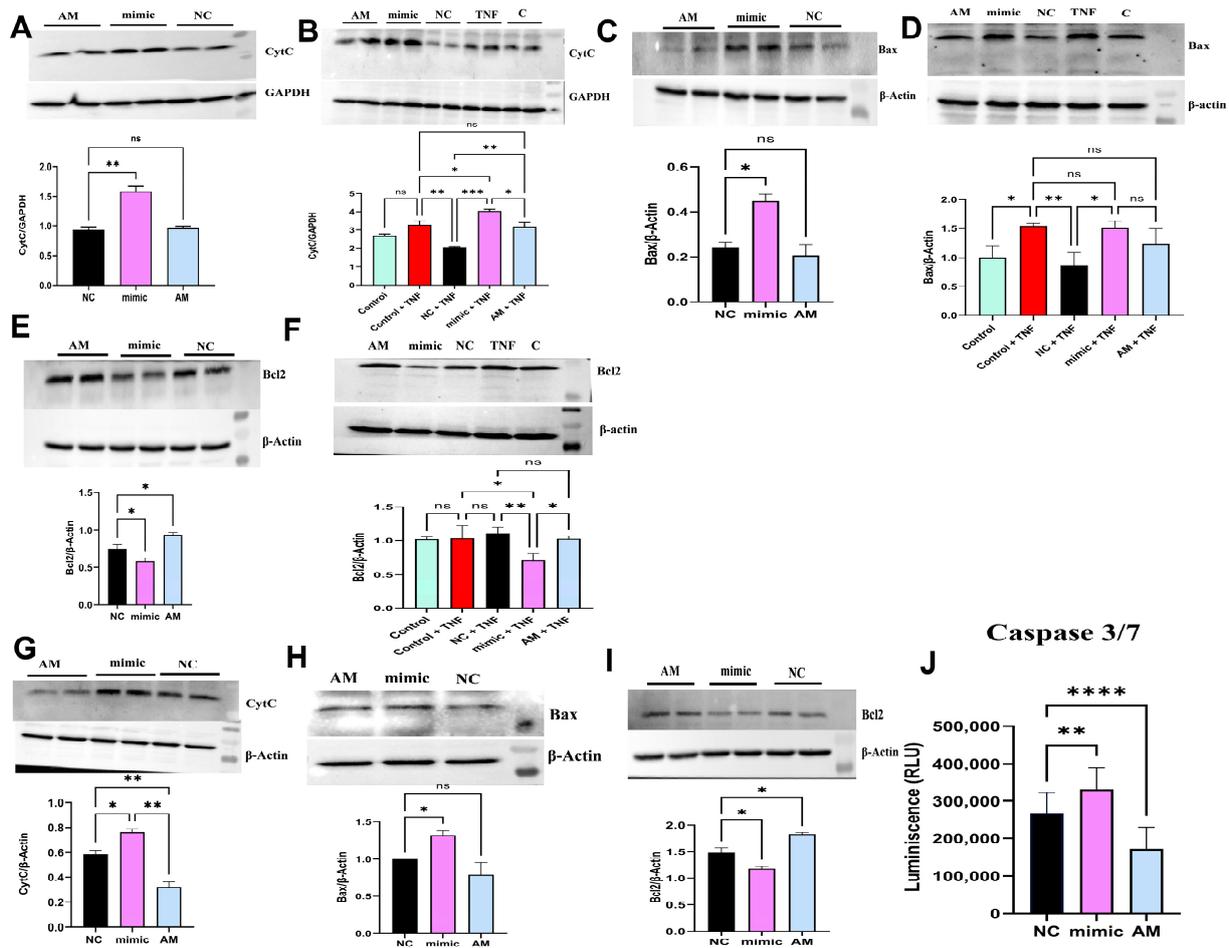
RAFLS was also demonstrated. These results collectively indicate that miR-4693-5p induces apoptosis stimulation in both SW982 cells and RAFLS, suggesting a potential regulatory role in apoptosis-related pathways.



**Figure 3.** Downstream target analysis and connection with HIF1α: (A) STRING network interaction of HIF1α with Survivin (BIRC5) and proinflammatory cytokines IL1β, TNFα, and IL6. (B) The interaction score of HIF1α to Survivin is higher (0.903) compared with proinflammatory cytokines IL1B (0.866), TNFα (0.650), and IL6 (0.504). (C) Western blot analysis showing downregulated Survivin expression in transfected miR-4693-5p (25 nM) uninduced (~2.6 fold) and (D) in TNFα-induced (~1.7 fold) SW982 cells and (E) in RAFLS (~2.0 fold) for 48 h. β-actin and GAPDH were used as loading controls. (NC: Negative Control, mimic: synthetic miR-4693-5p, AM: AntimiR, GAPDH: Glyceraldehyde 3-phosphate dehydrogenase, TNF-α: Tumor Necrosis Factor-Alpha, level of statistical significance: \* =  $p \leq 0.05$ , \*\* =  $p \leq 0.01$ , \*\*\* =  $p \leq 0.001$ ).

### 2.9. miR-4693-5p Inhibits Inflammatory Cytokines Production

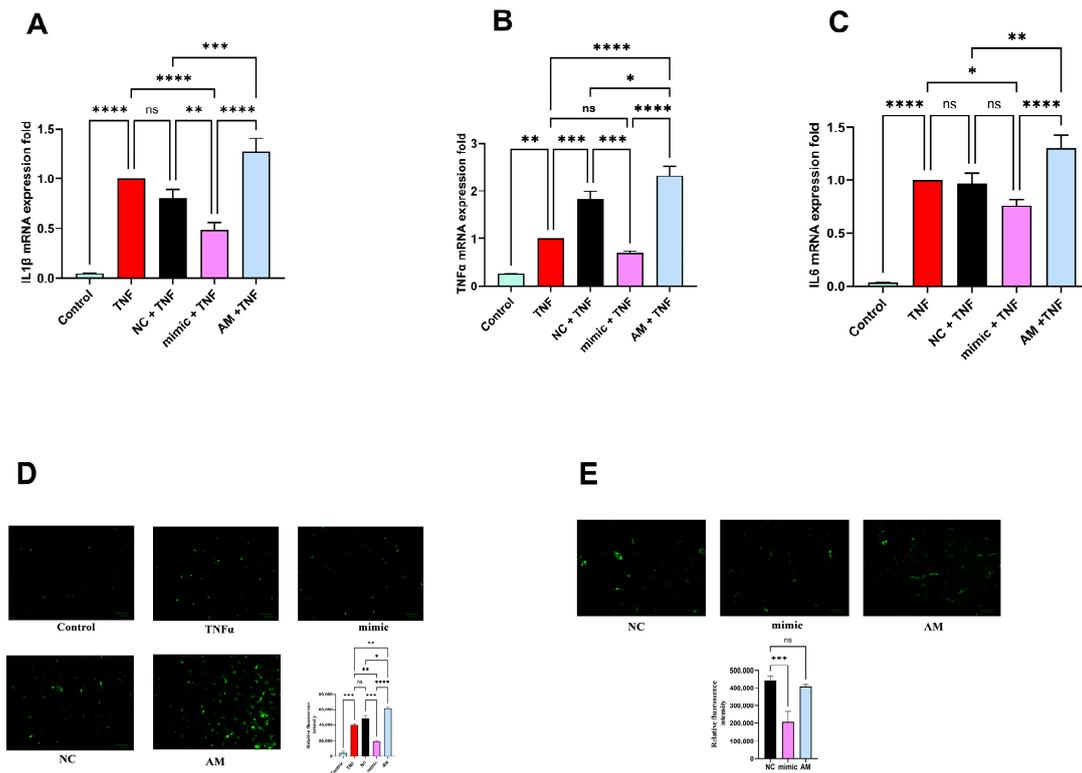
The major characteristics of RA joints are inflammation and hypoxia, which activate HIFs to regulate the expression of pro-inflammatory cytokines [16]. A recent report showed that HIF1α increases the production of proinflammatory cytokines like IL-6, IL-8, TNF-α, and IL-1β [20]. Therefore, we evaluated the effect of HIF1α knockdown on proinflammatory cytokines levels and found downregulation of the proinflammatory cytokines IL1β (Figure 5A), TNFα (Figure 5B), and IL6 (Figure 5C) at mRNA level compared with the Negative Control (NC) in transfected TNFα-induced SW982 cells, suggesting that knockdown of HIF1α significantly reduced the expression levels of proinflammatory cytokines. These findings indicate that miR-4693-5p exhibits anti-inflammatory potential by targeting HIF1α.



**Figure 4.** miR-4693-5p promoted cell apoptosis in SW982 and RAFLS cells: Western blot analysis in transfected miR-4693-5p at 25 nM in SW982 and RAFLS cells for 48 h. (A) Upregulated CytC expression in uninduced cells (~1.6 fold) and (B) TNF $\alpha$ -induced cells (~2 fold). GAPDH was used as the loading control. (C) Increased expression of Bax (~2 fold) in uninduced cells and (D) in TNF $\alpha$ -induced (~1.5 fold) SW982 cells.  $\beta$ -Actin was used as the loading control. (E) Downregulated expression of Bcl2 (~1.3 fold) in uninduced and TNF $\alpha$ -induced (F) (~1.5 fold) SW982 cells.  $\beta$ -Actin was used as a loading control. (G) Western blot analysis in transfected miR-4693-5p at 25 nM in RAFLS cells for 48 h, showing increased expression of CytC (~1.3 fold) and (H) Bax (~1.4 fold).  $\beta$ -Actin was used as the loading control. (I) Downregulated expression of Bcl2 (~1.3 fold).  $\beta$ -Actin was used as the loading control. (J) Increased Relative Luminescence (~1.2 fold) of caspase 3/7 was measured in RAFLS cells using the Caspase-Glo<sup>®</sup> 3/7 assay in miR-4693-5p-transfected RAFLS cells. (NC: Negative Control, mimic: synthetic miR-4693-5p, AM: AntimiR, GAPDH: Glyceraldehyde 3-phosphate dehydrogenase, B-Actin:  $\beta$ -Actin, TNF- $\alpha$ : Tumor Necrosis Factor-Alpha, level of statistical significance: \* =  $p \leq 0.05$ , \*\* =  $p \leq 0.01$ , \*\*\* =  $p \leq 0.001$ , \*\*\*\* =  $p \leq 0.0001$ ).

### 2.10. miR-4693-5p Reduces Cellular ROS Level

RA is also associated with elevated levels of ROS, and maintaining a balance between HIF1 $\alpha$  and ROS production is critical to minimize oxidative damage [21]. In this study, we assessed oxidative stress by measuring total intracellular ROS in miR-4693-5p transfected TNF $\alpha$ -induced SW982 cells and RAFLS. Fluorescence signals from DCFDA dye indicated a substantial increase in intracellular ROS production in TNF- $\alpha$ -induced cells compared with untreated control SW982 cells. However, miR-4693-5p transfection inhibited TNF- $\alpha$ -induced intracellular ROS production in SW982 cells (Figure 5D) and RAFLS (Figure 5E). These results suggest the potential of miR-4693-5p in modulating oxidative stress, providing insights into its therapeutic relevance in managing the oxidative aspects of RA.



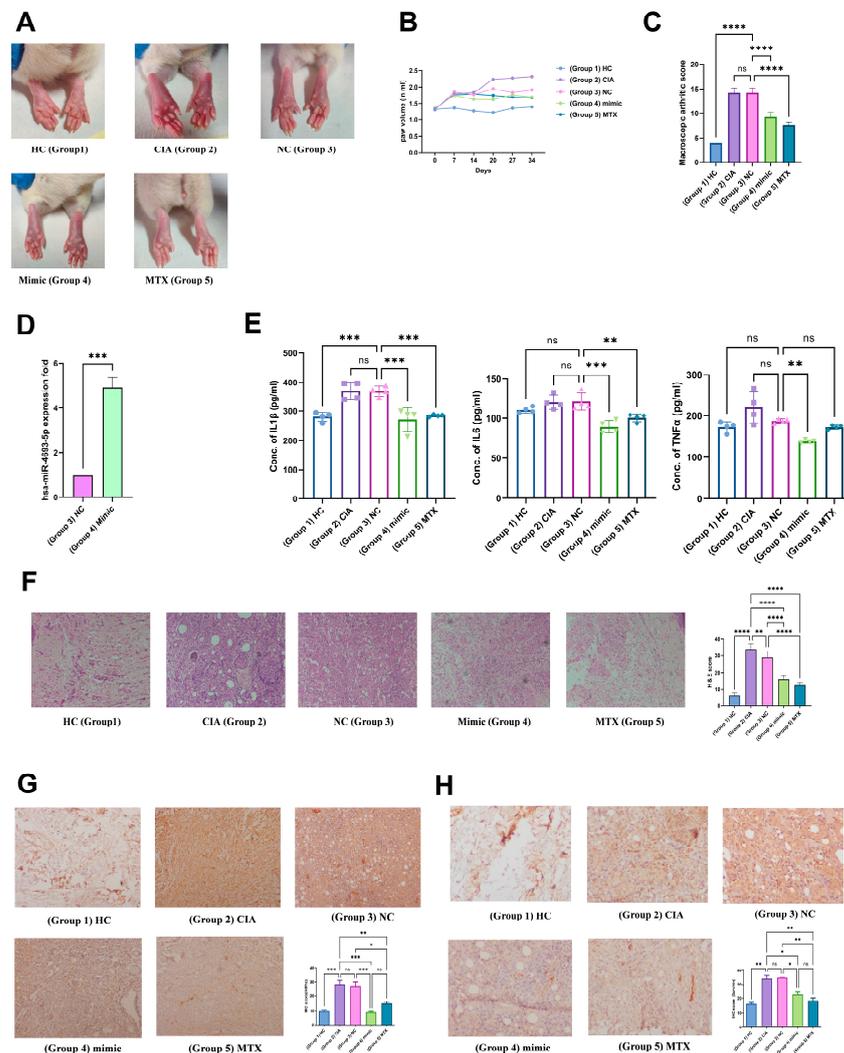
**Figure 5.** Anti-inflammatory and ROS inhibition effect of miR-4693-5p in SW982 cells and RAFLS: Decreased mRNA level of pro-inflammatory cytokines by qRT-PCR. (A) IL1 $\beta$ , (B) TNF $\alpha$ , and (C) IL6 expressions in SW982 cells transfected with miR-4693-5p after TNF $\alpha$  induction. GAPDH was used as an internal control. Intracellular ROS analyzed in transfected miR-4693-5p in (D) TNF $\alpha$  induced SW982 cells and (E) RAFLS. miR-4693-5p-inhibited ROS production, measured by relative fluorescence intensity compared with TNF $\alpha$ -induced and NC. DCFDA green fluorescence positive cell population was measured and normalized with a grayscale image. (TNF- $\alpha$ : Tumor Necrosis Factor-Alpha, IL: interleukin, NC: Negative Control, mimic: synthetic miR-4693-5p, AM: AntimiR, GAPDH: Glyceraldehyde 3-phosphate dehydrogenase, level of statistical significance: \* =  $p \leq 0.05$ , \*\* =  $p \leq 0.01$ , \*\*\* =  $p \leq 0.001$ , \*\*\*\* =  $p \leq 0.0001$ ).

### 2.11. In Vivo Validation

#### 2.11.1. miR-4693-5p Reduces Macroscopic Arthritic Score and Pro-Inflammatory Cytokines

The CIA rat model is widely used to mimic the RA condition [22]. We used this model to investigate the effects of miR-4693-5p. Images of rat paws taken on the 34th day for all groups (Group 1 (HC), Group 2 (CIA), Group 3 (CIA + Negative Control (NC)), Group 4 (CIA + mimic), and Group 5 (CIA + MTX)) before scarification are shown (Figure 6). We observed that Groups 4 (CIA + mimic) and 5 (CIA + MTX) showed less redness and swelling compared with Groups 2 (CIA) and 3 (CIA + Negative Control (NC)) (Figure 6A). On days 0, 7, 14, 21, 28, and 34, the paw volume was measured using a plethysmometer to confirm the onset of the disease. After day 14, average paw volume decreased in Groups 4 and 5, whereas paw volume increased in Groups 2 and 3 (Figure 6B). The macroscopic arthritic score considerably decreased in Group 4 compared with Groups 3 and 2 (Figure 6C). After sacrificing the rats, the injected miRNA mimic was assessed in the rat synovium of Groups 3 and 4 by qRT-PCR to evaluate the overexpression of miR-4693-5p. We observed that Group 4 had higher levels of miR-4693-5p than Group 3, indicating an upregulated mimic level in rat synovium (Figure 6D). Levels of pro-inflammatory cytokines were also measured by ELISA in the rat plasma of all groups. Downregulation of pro-inflammatory cytokines (IL-1 $\beta$ , IL-6, and TNF $\alpha$ ) was also revealed in rat plasma in Groups 4 and 5 compared with Groups 2 and 3 (Figure 6E). Therefore, our results suggest that miR-4693-5p might be responsible for the downregulation of pro-inflammatory cytokines and may have a

protective effect in CIA rats by reducing paw volume, macroscopic arthritic score, and pro-inflammatory cytokine levels.



**Figure 6.** The effect of miR-4693-5p on the Collagen-Induced Arthritis (CIA) Rat model: (A) Visual representation of the hind paws of rats from each group, where edema and redness were reduced in Groups 4 and 5 compared with Groups 2 and 3. (B) Graphical representation of measured paw volumes from day 0 to day 34, depicting the changes in paw volume in Group 4 compared with Groups 2, 3, and 5. (C) The macroscopic arthritic score was measured on the 28th day, and a significant reduction in redness and swelling was observed in Group 4 compared with Groups 2 and 3. (D) Fold change of miR-4693-5p in rat synovium by qRT-PCR found significantly upregulated levels of miR-4693-5p in Group 4 compared with Group 3 after normalization with U6 loading control. (E) The proinflammatory cytokine levels were measured using quantitative ELISA analysis in rat plasma in Groups 1 to 5, showing the downregulation of IL1  $\beta$ , IL-6, and TNF $\alpha$  levels in Group 4 compared with Groups 2 and 3. (F) The H&E staining shows decreased cell inflammation (pink color) in Groups 4 and 5 compared with Groups 2 and 3. The analysis of cell infiltration in the synovium was measured and found to be downregulated in Groups 4 and 5 compared with Groups 2 and 3. (G,H) Immunohistochemistry assay for HIF1 $\alpha$  and Survivin expression in rat synovium of Group 1 to 5 rats. Typical images of (G) HIF1 $\alpha$  and (H) Survivin expression in synovial tissues showing that HIF1 $\alpha$  and Survivin were expressed at low levels in the synovium of Groups 4 and 5 compared with Groups 2 and 3. (Group 1: Healthy or HC, Group 2: CIA, Group 3: CIA + Negative Control (NC), Group 4: CIA + miR-4693-5p mimic, Group 5: CIA + MTX (standard drug), level of statistical significance: \* =  $p \leq 0.05$ , \*\* =  $p \leq 0.01$ , \*\*\* =  $p \leq 0.001$ , \*\*\*\* =  $p \leq 0.0001$ ).

### 2.11.2. Validation of the Anti-Inflammatory Effect of miR-4693-5p

To further validate the anti-inflammatory activity of miR-4693-5p, histological tests were performed on rat synovium by H&E staining (Figure 6F). The pink color represents cytoplasm, which correlates with the synovium's inflammation. The purple color represents the number of nuclei present, and is used to determine the number of cells infiltrated into the given region to quantify inflammation [22]. The H&E scan analysis revealed that the group injected with miR-4693-5p (Group 4) and Group 5 (MTX) exhibited much less cell infiltration compared with Groups 3 and 2. This histological examination supports the idea that Group 4 selectively regulated pro-inflammatory cytokines through the NF- $\kappa$ B crosstalk, significantly reducing inflammation *in vivo*. These findings provide additional evidence of the therapeutic potential of miR-4693-5p in mitigating inflammation in a CIA rat model of RA.

### 2.11.3. Immunohistochemical Analysis of HIF-1 $\alpha$ and Survivin Expression

The expression of HIF1 $\alpha$  in the synovial tissue in all groups was also measured to strengthen our findings further and was found to have less brown deposition in Group 4 compared with Groups 2 and 3, indicating less relative percentage area expression in Group 4 compared with Groups 2 and 3 (Figure 6G). Similarly, Survivin expression was reduced in Groups 4 and 5 compared with Groups 2 and 3 (Figure 6H). These findings confirm that miR-4693-5p suppressed the expression of HIF1 $\alpha$  and Survivin, suggesting a protective role in impeding the progression of the disease. The downregulation of HIF1 $\alpha$  and Survivin aligns with the observed anti-inflammatory effects of miR-4693-5p in the CIA rat model, further supporting its potential therapeutic significance in RA.

## 3. Discussion

RA is an autoimmune disease that causes joint damage and inflammation, but its exact cause remains unknown [1]. Studies show that overexpression of HIF1 $\alpha$  in synovial joints leads to hyperplasia, immune cell infiltration, and resistance to apoptosis, aiding the development of RA pathogenesis [23], but the cause for overexpression of HIF1 $\alpha$  in RA is unclear [6]. Given the potent regulatory role of miRNA in gene expression, several miRNAs, including miR-221, miR-448, miR-124, and miR-551b, have been identified in RA, playing regulatory roles in vital cellular processes like cell proliferation, angiogenesis, and inflammation, while their dysregulation contributes to RA pathogenesis [24].

It was reported that the miRNA expression of PBMCs might mimic conditions found in synovial tissue, making it easier to study in large samples [25]. Certain miRNAs such as miR146, miR145 [13], and miR21 [26] found in PBMCs are involved in RA pathogenesis. Our study is the first to report that miR-4693-5p was significantly downregulated in PBMCs of RA, suggesting miR-4693-5p may be a potential regulator in RA development. Therefore, we were interested in investigating the potential role of miR-4693-5p and attempted *in silico* studies followed by *in vitro* and *in vivo* studies. Next, our objective was to find the favorable target for miR-4693-5p, and our *in silico* target prediction results revealed that miR-4693-5p might target and bind to the 3' UTR of HIF1 $\alpha$ . Then, we validated the target *in vitro*, and the results confirmed that miR-4693-5p downregulates the expression of HIF1 $\alpha$  in SW982 cells (Figure 1), demonstrating that miR-4693-5p is a potential regulator in RA development.

Pro-inflammatory cytokines initiate destructive processes in RA joints, leading to structural damage, reduced mobility, and functional decline, ultimately resulting in disability [27]. The report shows that proinflammatory cytokines can induce or stabilize HIF1 $\alpha$  protein [3]. We explored the specific time relationship with HIF1 $\alpha$  and TNF $\alpha$  (major cytokine involved in RA pathogenesis) and revealed that TNF $\alpha$  induces HIF1 $\alpha$  expression in SW982 cells (24 and 48 h) in normoxic conditions (Figure 2D,E).

It is known that HIF1 $\alpha$  regulates many proteins that are involved in carbohydrate metabolism, invasion, angiogenesis, cell proliferation, and apoptosis pathways [28]. While exploring more, PPI interaction results show that Survivin was an interacting protein

of HIF1 $\alpha$ , having the highest interaction score (Figure 3B) compared with the known proinflammatory cytokines (IL1 $\beta$ , TNF $\alpha$ , and IL6) involved in RA pathogenesis. It was reported that Survivin is transcriptionally regulated by HIF1 $\alpha$  and also gets overexpressed in RA and inhibits intrinsic and extrinsic apoptotic pathways by interacting with caspase 3 and 9 to promote cell proliferation and hyperplasia, which aids disease progression [29]. Therefore, a study of the inhibition of Survivin expression in SW982 and RAFLS cells by HIF1 $\alpha$  knockdown was conducted and validated (Figure 3C–E).

Our findings showed that miR-4693-5p upregulated the expression of proapoptotic proteins (Bax, CytC, and caspase 3/7) and downregulated the expression of antiapoptotic protein (Bcl2) (Figure 4) via HIF1 $\alpha$  knockdown to promote apoptotic stimulation in SW982 and RAFLS cells, indicating that miR-4693-5p inhibits apoptosis resistance in RA to control disease progression [30].

The interaction between HIFs and NF- $\kappa$ B has been observed, and increased levels of both NF- $\kappa$ B and HIFs contribute to inflammatory responses by upregulating the expression of pro-inflammatory cytokines [20]. Moreover, NF- $\kappa$ B and HIF1 $\alpha$  crosstalk in regulating the immune response in various immune cell types has also been observed, resulting in the stabilization of HIF1 $\alpha$  in normoxia [20]. Recently, a report demonstrated that HIF1 $\alpha$  increases the production of proinflammatory cytokines such as IL6, IL-8, TNF $\alpha$ , and IL1 $\beta$  [6]. Therefore, we checked the cytokine levels after HIF1 $\alpha$  knockdown and found them to be downregulated (Figure 5A–C), demonstrating the anti-inflammatory activity of miR-4693-5p in RA (Figure 5) [25].

Hypoxia-inducible factor (HIF) accumulation and ROS formation occur simultaneously during hypoxia [31]. It was reported that downstream transcriptional targets of HIF could either activate or inhibit ROS formation, and an elevated level of ROS leads to HIF1 $\alpha$  stabilization, resulting in increased HIF1 $\alpha$  expression, and this stabilization governs ROS formation [21], which is also supported by our findings. We found that knockdown of HIF1 $\alpha$  minimized the hypoxia conditions by suppressing ROS levels in TNF $\alpha$ -induced SW982 cells and RAFLS compared with the controls (Figure 5D,E), indicating that miR-4693-5p can reduce oxidative stress by reducing total cellular ROS levels.

Our findings were further validated by the CIA rat model. In total, downregulation of HIF1 $\alpha$  and Survivin (Figure 6G,H) in the synovium of Group 4 (CIA + mimic) facilitated the decreased expression of pro-inflammatory cytokines compared with Groups 2 (CIA) and 3 (CIA + Negative Control (NC)) (Figure 6E). The current study provides a crucial foundation for understanding the potential role of miR-4693-5p as a promising therapeutic candidate and provides novel insights into the molecular mechanisms underlying RA pathogenesis.

## 4. Materials and Methods

### 4.1. Sample Collection

Blood and biopsy tissue samples were obtained from RA patients ( $n = 12$ ) at the Orthopedic Department, All India Institute of Medical Sciences (AIIMS), New Delhi, India. Inclusion criteria: Patients were diagnosed with RA and met the revised 2010 American College of Rheumatology (ACR) and European League Against Rheumatism (EULAR) Rheumatism diagnostic criteria [32], and NSAIDs (nonsteroidal anti-inflammatory drugs, such as Ibuprofen, Diclofenac), DMARDs (Methotrexate), and surgery were advised as treatment options. Healthy Control (HC) blood samples ( $n = 12$ ) were collected from individuals with no history of joint disease. Patients who were pregnant, alcoholic, or had other diseases such as diabetes, cardiovascular disease, or any other inflammatory diseases were excluded from the study. Detailed medical histories were recorded for all patients (Supplementary Table S1). Informed consent was obtained from all participants.

### Blood Sample Processing

Plasma was separated from blood samples by centrifugation at  $1300\times g$  for 15 min ( $4\text{ }^{\circ}\text{C}$ ). The resulting plasma was processed, aliquoted, and stored at  $-80\text{ }^{\circ}\text{C}$  for subsequent analysis [33].

#### 4.2. miRNA Quantification in PBMCs

For quantification of miR-4693-5p, PBMCs were isolated using Histopaque (Sigma, St. Louis, MO, USA), and total RNA was extracted using Tri-Xtract Reagent (G-biosciences, St. Louis, MO, USA). cDNA synthesis was performed using a cDNA Synthesis Kit (G-biosciences) with stem-loop PCR. Expression analysis of miR-4693-5p was conducted using real-time PCR (Roche Light Cycler<sup>®</sup> 480 Instrument-II, Mannheim, Germany). Data were quantitatively assessed using the  $2^{-\Delta\Delta CT}$  formula and normalized to U6 as an internal control [34]. Primer sequences are provided in Supplementary Table S2.

#### 4.3. Target Prediction

Three miRNA target prediction databases (miRDB, RNA22, and TargetScan) were utilized to identify miR-4693-5p targets [14]. Common targets across databases were matched with RA-related genes from NCBI and DisGeNET. Selection criteria for favorable targets included a high total context score and 8mer binding site among targets, as well as the maximum number of proteins that interacted with the target [35].

#### 4.4. In Vitro Studies

##### 4.4.1. Cell Culture (SW982)

SW982 cells, commonly used to study synovitis in RA, were purchased from the National Centre for Cell Science (NCCS). Cells were cultured in DMEM medium with antibiotics and 10% FBS at 37 °C with 5% CO<sub>2</sub>. Cells were then stimulated with Tumor Necrosis Factor-Alpha (TNF $\alpha$ ) (10 ng/mL) [27].

##### 4.4.2. Primary Cell (RAFLS) Isolation

Synovium or synovial tissue was minced, digested with collagenase (0.5 mg/mL) (Sigma, St. Louis, MO, USA), plated, and incubated in DMEM containing 10% FBS at 37 °C with 5% CO<sub>2</sub>. Cells were used for the treatment after the second passage [29].

##### 4.4.3. Cell Transfection

For functional validity of miR-4693-5p, cells were transfected in a six-well plate until 60–70% confluency using Lipofectamine RNAiMAX Transfection Reagent (Invitrogen, Waltham, MA, USA) with 25 nM of miRNA Negative Control (NC), miR-4693-5p (mimic), and miRNA inhibitor (AntimiR or AM) (Invitrogen) in Opti-MEM media (Gibco, Waltham, MA, USA). After transfection (5 h), media were replaced with DMEM containing 10% FBS [10].

##### 4.4.4. Dual-Luciferase Reporter Assay

The TargetScan database was used to identify possible binding sites of miR-4693-5p to the 3' UTR of HIF1 $\alpha$ . Luciferase reporter vectors were constructed [36], and SW982 cells were co-transfected with miRNA and vectors. Five combinations were made: (1) Wild type (WT), (2) Mutant (Mut), (3) Wild type + Negative Control (WT + NC), (4) Wild type + mimic (WT + mimic), and (5) Mutant + mimic (MT + mimic). Luciferase activities were measured using a Tecan i-control infinite 200 Pro device (see details in Supplementary File).

##### 4.4.5. Real-Time PCR

After transfection, cells (SW982) were stimulated with TNF $\alpha$  (10 ng/mL). Total RNA was isolated and reverse-transcribed, and mRNA expression was determined by real-time PCR using human-specific primer sequences (Supplementary Table S2). The values were normalized by GAPDH and quantitatively evaluated using the  $2^{-\Delta\Delta CT}$  formula. Stem-loop primer was used to check the overexpression of miR-4693-5p in in vitro experiments using a specific primer and U6 as an internal control [27] (see details in Supplementary File).

#### 4.4.6. Western Blot (WB)

Cells were lysed in RIPA buffer, proteins were estimated using the BCA method [27], and 40 µg protein was separated by running SDS-PAGE. Then, proteins were transferred to the Nitrocellulose Membrane. The membrane was blocked (2 h) with 3% BSA, incubated overnight (4 °C) separately with the primary antibodies (Santacruz, Santa Cruz, CA, USA) Anti-human-HIF1α (1:500), Survivin (1:3000), CytC (1:2000), Bax (1:2000), and Bcl2 (1:2000), washed with 1XTBST, and incubated again (1 h) with HRP-conjugated secondary antibody (1:10,000, Jackson, MI, USA) at room temperature (RT). The blots were then developed with ECL and analyzed using the ChemiDoc system [34] (Bio-Rad Laboratories (Singapore) Pte. Ltd.) (see details in Supplementary File).

#### 4.4.7. Protein–Protein Interaction (PPI)

Interactive partners of HIF1α were identified using STRING and STITCH databases [35]. The downstream targets of HIF1α and their interactions with known pro-inflammatory cytokines were found to be related to RA pathogenesis.

#### 4.4.8. Caspase-Glo 3/7 Assay

The Caspase-Glo 3/7 Assay System (Promega, Madison, WI, USA) was used to evaluate caspase activity or apoptosis. After transfection, as described above, 100 µL of Caspase-Glo 3/7 reagent was added, followed by 3 h of incubation [37]. The luminescence was then measured using a Tecan i-control (see details in Supplementary File).

#### 4.4.9. Total ROS Estimation

Intracellular ROS formation was measured using DCFDA (Dichlorodihydrofluorescein diacetate). RAFLS and SW982 cells were transfected and induced with TNFα followed by 10 µM DCFDA (Invitrogen) addition into each well, incubation (30 min), and analysis of fluorescence intensity [38] (see details in Supplementary File).

### 4.5. *In Vivo* Studies

#### 4.5.1. Development of Collagen-Induced Arthritis (CIA) Rat Model

Female Wistar rats were obtained from the Indian Council of Medical Research–National Institute of Nutrition, Hyderabad, India. After two weeks of acclimatization, five groups ( $n = 4$  rats in each group) were created: Healthy Control (HC) (Group 1), CIA (Group 2), Negative Control (NC)+ CIA (Group 3), CIA+ mimic (Group 4), and CIA+ MTX (Methotrexate, standard drug) (Group 5). CIA (not in the HC group) was caused by inducing 2 mg/mL collagen Type II from chicken (Sigma, USA) dissolved in 0.01 M acetic acid and combined (1:1) with complete adjuvant (Sigma, USA). miRNA (miR-4693-5p) (1 ng/gm of body weight or 200 ng/Rat) was administered to the rats intraperitoneally [39]. The rats were then sacrificed on day 34 [22,40] (see details in Supplementary File).

#### 4.5.2. Macroscopic Arthritis Score Evaluation

Arthritis severity was graded based on swelling, edema, and redness in all four paws of CIA rats using a 1 to 4 scale. A plethysmometer was used to calculate and quantify the swelling of the joints [41] (see details in Supplementary File).

#### 4.5.3. miRNA Isolation from Rat Synovium

Synovium was separated, and miRNA was extracted using a mirVana™ miRNA Isolation Kit (Invitrogen). Expression of miR-4693-5p was determined by qRT-PCR [42] (see details in Supplementary File).

#### 4.5.4. Enzyme-Linked Immunosorbent Assay (ELISA)

Rat plasma was separated and added (100 µL) to the pre-coated ELISA plate, following the manufacturer's guidelines. TNFα, IL(Interleukin)1β, and IL-6 cytokines were quantified using ELISA kits (ELK Biotechnology, Wuhan, China) [33].

#### 4.5.5. Hematoxylin and Eosin Staining (H & E)

For histological analysis, rat synovium slices (5  $\mu\text{m}$  thick) were made using a microtome and stained with H&E, as per standard histological analysis protocol [40] (see details in Supplementary File).

#### 4.5.6. Immunohistochemistry (IHC)

For IHC analysis, rat synovium slices were immersed in 3%  $\text{H}_2\text{O}_2$  solution (25 min) at room temperature (RT), blocked with 3% BSA (2 h), incubated overnight with HIF1 $\alpha$  (1:500) and Survivin (1:500) antibodies (Santacruz) separately at 4  $^\circ\text{C}$ , and washed after incubation. The slices were then incubated (1 h) with the secondary antibody HRP conjugate, followed by incubation (10 min) with DAB (Diaminobenzidine) peroxidase substrate. Images of the slides were taken at a 10 $\times$  magnification and examined using Image-J software (Version 1.54d) [22] (see details in Supplementary File).

#### 4.6. Statistical Analysis

Graph Pad Prism (version 9.0) was used for statistical analysis, employing Student's *t*-test, Mann–Whitney U, Analysis of variance (ANOVA), and Chi-square as appropriate ( $p < 0.05$ ). Each experiment was repeated at least three times (see details in Supplementary File).

### 5. Conclusions

In our study, we observed a downregulation of miR-4693-5p in PBMCs of RA patients, indicating its role as a regulatory element of HIF1 $\alpha$ . Additionally, we revealed that miR-4693-5p modulates apoptosis, inflammatory cytokine production, and cellular ROS response by targeting HIF1 $\alpha$  and its downstream mediator, Survivin. These findings improve our understanding and provide novel insights into the molecular mechanisms driving the pathogenesis of RA. A limitation of the study was the inability to validate the expression of miR-4693-5p in large numbers of samples to consider miR-4693-5p as an effective therapeutic agent to ameliorate RA.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/ncrna10020022/s1>, Figure S1: Western blot analysis of HIF1 $\alpha$  expression at different concentrations of hsa-miR-4693-5p ranging from 5–50 nm.; Figure S2: (A) Cloned confirmation of HIF1 $\alpha$  3'UTR in psiCHECK2 vector, (B) Site directed mutagenesis of seed region of HIF1 $\alpha$  3'UTR (mutated sequence shown in highlighted box); Figure S3: Significant overexpression of survivin confirmed in HC and RA plasma samples ( $n = 8$  each) by ELISA; Table S1: The clinical demography characteristics of healthy control and patients with RA; Table S2: The amplification primers sequences of human-specific genes.

**Author Contributions:** Conceptualization, M.S. and P.A.; Data curation, M.S.; Formal analysis, M.S. and P.A.; Funding acquisition, S.B.; Investigation, M.S.; Methodology, M.S., A.S., S.M. (Swati Malik), S.M. (Sonia Mann), D.C. and L.J.; Project administration, S.B.; Resources, R.M. and S.B.; Visualization, P.A.; Writing—original draft, M.S. and P.A.; Writing—review & editing, M.S., P.A., A.S., S.M. (Swati Malik), S.M. (Sonia Mann), D.C., L.J., R.M. and S.B. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was supported by the Council of Scientific and Industrial Research (CSIR), Project code MLP 2013, Government of India, New Delhi, India.

**Institutional Review Board Statement:** All animals received care in compliance with the Principles of Laboratory Animal Care and National standards. The study protocol was approved by the ethics committee of the institution, the Council of Scientific and Industrial Research (CSIR)–Institute of Genomics and Integrative Biology, Delhi, India IGIB/IAEC/3/3/March 2023. The study protocol received ethical approval from the All India Institute of Medical Sciences (AIIMS), New Delhi, India (Reg No IEC-237/07.05.2021, RP-18/2021). Additionally, signed consents were obtained from all participants involved in the study.

**Informed Consent Statement:** Written informed consent has been obtained from the patient(s) to publish this paper.

**Data Availability Statement:** For all original data and protocols, please contact Sagarika Biswas (sagarika.biswas@igib.res.in).

**Acknowledgments:** We would like to acknowledge the Council of Scientific and Industrial Research (CSIR), New Delhi, India, for financial support, and providing fellowship support to Mohd Saquib, Lovely Joshi, Ashish Sarkar, and Debolina Chakraborty, while Prachi Agnihotri and Swati Malik received a fellowship from DST, and Sonia Mann from ICMR. Special thanks go to Pankaj Yadav for transporting biological samples. We are thankful to Rajesh Malhotra for providing patient samples from AIIMS. Additionally, we acknowledge the CSIR–Institute of Genomics and Integrative Biology, Delhi, India, for research support and the Academy of Scientific and Innovative Research (AcSIR) for academic support.

**Conflicts of Interest:** The authors declare no conflicts of interest.

## References

1. Guo, Q.; Wang, Y.; Xu, D.; Nossent, J.; Pavlos, N.J.; Xu, J. Rheumatoid Arthritis: Pathological Mechanisms and Modern Pharmacologic Therapies. *Bone Res.* **2018**, *6*, 15. [[CrossRef](#)] [[PubMed](#)]
2. Sparks, J.A. In the Clinic<sup>®</sup> Rheumatoid Arthritis. *Ann. Intern. Med.* **2019**, *170*, ITC1–ITC15. [[CrossRef](#)]
3. Quiñonez-Flores, C.M.; González-Chávez, S.A.; Pacheco-Tena, C. Hypoxia and Its Implications in Rheumatoid Arthritis. *J. Biomed. Sci.* **2016**, *23*, 62. [[CrossRef](#)]
4. Yap, H.Y.; Tee, S.Z.Y.; Wong, M.M.T.; Chow, S.K.; Peh, S.C.; Teow, S.Y. Pathogenic Role of Immune Cells in Rheumatoid Arthritis: Implications in Clinical Treatment and Biomarker Development. *Cells* **2018**, *7*, 161. [[CrossRef](#)] [[PubMed](#)]
5. Kaikkonen, M.U.; Lam, M.T.Y.; Glass, C.K. Editor's Choice: Non-Coding RNAs as Regulators of Gene Expression and Epigenetics. *Cardiovasc. Res.* **2011**, *90*, 430. [[CrossRef](#)] [[PubMed](#)]
6. Wang, H.; Flach, H.; Onizawa, M.; Wei, L.; Mcmanus, M.T.; Weiss, A. Negative Regulation of Hif1a Expression and TH17 Differentiation by the Hypoxia-Regulated MicroRNA MiR-210. *Nat. Immunol.* **2014**, *15*, 393–401. [[CrossRef](#)]
7. Storti, P.; Bolzoni, M.; Donofrio, G.; Airoidi, I.; Guasco, D.; Toscani, D.; Martella, E.; Lazzaretti, M.; Mancini, C.; Agnelli, L.; et al. Hypoxia-Inducible Factor (HIF)-1 $\alpha$  Suppression in Myeloma Cells Blocks Tumoral Growth in Vivo Inhibiting Angiogenesis and Bone Destruction. *Leukemia* **2013**, *27*, 1697–1706. [[CrossRef](#)] [[PubMed](#)]
8. Sharma, A.; Sahu, S.; Kumari, P.; Gopi, S.R.; Malhotra, R.; Biswas, S. Genome-Wide Identification and Functional Annotation of MiRNAs in Anti-Inflammatory Plant and Their Cross-Kingdom Regulation in Homo Sapiens. *J. Biomol. Struct. Dyn.* **2017**, *35*, 1389–1400. [[CrossRef](#)] [[PubMed](#)]
9. Saquib, M.; Agnihotri, P.; Monu, X.; Biswas, S. Exogenous MiRNA: A Perspective Role as Therapeutic in Rheumatoid Arthritis. *Curr. Rheumatol. Rep.* **2021**, *23*, 43. [[CrossRef](#)]
10. Hou, C.; Wang, D.; Zhang, L. MicroRNA-34a-3p Inhibits Proliferation of Rheumatoid Arthritis Fibroblast-like Synoviocytes. *Mol. Med. Rep.* **2019**, *20*, 2563. [[CrossRef](#)]
11. Huang, Q.; Li, B.; Lin, C.; Chen, X.; Wang, T.; Liu, J.; Liu, Y.; Lu, R.; Liao, S.; Ding, X. MicroRNA Sequence Analysis of Plasma Exosomes in Early Legg-Calvé-Perthes Disease. *Cell. Signal.* **2022**, *91*, 110184. [[CrossRef](#)]
12. Chen, Y.; Chen, D.; Qin, Y.; Qiu, C.; Zhou, Y.; Dai, M.; Li, L.; Sun, Q.; Jiang, Y. TRIP13, Identified as a Hub Gene of Tumor Progression, Is the Target of MicroRNA-4693-5p and a Potential Therapeutic Target for Colorectal Cancer. *Cell Death Discov.* **2022**, *8*, 35. [[CrossRef](#)] [[PubMed](#)]
13. Pauley, K.M.; Satoh, M.; Chan, A.L.; Bubb, M.R.; Reeves, W.H.; Chan, E.K.L. Upregulated MiR-146a Expression in Peripheral Blood Mononuclear Cells from Rheumatoid Arthritis Patients. *Arthritis Res. Ther.* **2008**, *10*, R101. [[CrossRef](#)] [[PubMed](#)]
14. Riolo, G.; Cantara, S.; Marzocchi, C.; Ricci, C. MiRNA Targets: From Prediction Tools to Experimental Validation. *Methods Protoc.* **2021**, *4*, 1. [[CrossRef](#)] [[PubMed](#)]
15. Kaloni, D.; Chakraborty, D.; Tiwari, A.; Biswas, S. In Silico Studies on the Phytochemical Components of *Murraya Koenigii* Targeting TNF- $\alpha$  in Rheumatoid Arthritis. *J. Herb. Med.* **2020**, *24*, 100396. [[CrossRef](#)]
16. Hu, F.; Mu, R.; Zhu, J.; Shi, L.; Li, Y.; Liu, X.; Shao, W.; Li, G.; Li, M.; Su, Y.; et al. Hypoxia and Hypoxia-Inducible Factor-1 $\alpha$  Provoke Toll-like Receptor Signalling-Induced Inflammation in Rheumatoid Arthritis. *Ann. Rheum. Dis.* **2014**, *73*, 928–936. [[CrossRef](#)]
17. Khansai, M.; Phitak, T.; Klangjorhor, J.; Udomrak, S.; Fanhchaksai, K.; Pothacharoen, P.; Kongtawelert, P. Effects of Sesamin on Primary Human Synovial Fibroblasts and SW982 Cell Line Induced by Tumor Necrosis Factor-Alpha as a Synovitis-like Model. *BMC Complement. Altern. Med.* **2017**, *17*, 532. [[CrossRef](#)]
18. Bai, H.; Ge, S.; Lu, J.; Qian, G.; Xu, R. Hypoxia Inducible Factor-1 $\alpha$ -Mediated Activation of Survivin in Cervical Cancer Cells. *J. Obstet. Gynaecol. Res.* **2013**, *39*, 555–563. [[CrossRef](#)]
19. Mobahat, M.; Narendran, A.; Riabowol, K. Survivin as a Preferential Target for Cancer Therapy. *Int. J. Mol. Sci.* **2014**, *15*, 2494. [[CrossRef](#)]
20. D'Ignazio, L.; Bandarra, D.; Rocha, S. NF- $\kappa$ B and HIF Crosstalk in Immune Responses. *FEBS J.* **2016**, *283*, 413. [[CrossRef](#)]

21. Tafani, M.; Sansone, L.; Limana, F.; Arcangeli, T.; De Santis, E.; Polese, M.; Fini, M.; Russo, M.A. The Interplay of Reactive Oxygen Species, Hypoxia, Inflammation, and Sirtuins in Cancer Initiation and Progression. *Oxid. Med. Cell. Longev.* **2016**, *2016*, 3907147. [[CrossRef](#)]
22. Bai, L.; Bai, Y.; Yang, Y.; Zhang, W.; Huang, L.; Ma, R.; Wang, L.; Duan, H.; Wan, Q. Baicalin Alleviates Collagen-Induced Arthritis and Suppresses TLR2/MYD88/NF-KB P65 Signaling in Rats and HFLS-RAs. *Mol. Med. Rep.* **2020**, *22*, 2833. [[CrossRef](#)]
23. Falconer, J.; Murphy, A.N.; Young, S.P.; Clark, A.R.; Tiziani, S.; Guma, M.; Buckley, C.D. Synovial Cell Metabolism and Chronic Inflammation in Rheumatoid Arthritis. *Arthritis Rheumatol.* **2018**, *70*, 984. [[CrossRef](#)]
24. Fitriana, M.; Hwang, W.L.; Chan, P.Y.; Hsueh, T.Y.; Liao, T.T. Roles of MicroRNAs in Regulating Cancer Stemness in Head and Neck Cancers. *Cancers* **2021**, *13*, 1742. [[CrossRef](#)]
25. Zhu, X.; Wu, L.; Mo, X.; Xia, W.; Guo, Y.; Wang, M.; Zeng, K.; Wu, J.; Qiu, Y.; Lin, X.; et al. Identification of PBMC-Expressed MiRNAs for Rheumatoid Arthritis. *Epigenetics* **2020**, *15*, 386. [[CrossRef](#)]
26. Saquib, M.; Agnihotri, P.; Monu, X.; Biswas, S. Interrelated Grid of Non-Coding RNA: An Important Aspect in Rheumatoid Arthritis Pathogenesis. *Mol. Biol. Rep.* **2023**, *50*, 6217–6232. [[CrossRef](#)]
27. Rakhecha, B.; Agnihotri, P.; Dakal, T.C.; Saquib, M.; Monu, X.; Biswas, S. Anti-Inflammatory Activity of Nicotine Isolated from Brassica Oleracea in Rheumatoid Arthritis. *Biosci. Rep.* **2022**, *42*, 20211392. [[CrossRef](#)]
28. Lee, S.H.; Golinska, M.; Griffiths, J.R. HIF-1-Independent Mechanisms Regulating Metabolic Adaptation in Hypoxic Cancer Cells. *Cells* **2021**, *10*, 2371. [[CrossRef](#)]
29. Mokuda, S.; Miyazaki, T.; Ito, Y.; Yamasaki, S.; Inoue, H.; Guo, Y.; Kong, W.S.; Kanno, M.; Takasugi, K.; Sugiyama, E.; et al. The Proto-Oncogene Survivin Splice Variant 2B Is Induced by PDGF and Leads to Cell Proliferation in Rheumatoid Arthritis Fibroblast-like Synoviocytes. *Sci. Rep.* **2015**, *5*, 9795. [[CrossRef](#)]
30. Mita, A.C.; Mita, M.M.; Nawrocki, S.T.; Giles, F.J. Survivin: Key Regulator of Mitosis and Apoptosis and Novel Target for Cancer Therapeutics. *Clin. Cancer Res.* **2008**, *14*, 5000–5005. [[CrossRef](#)]
31. Bae, T.; Hallis, S.P.; Kwak, M.K. Hypoxia, Oxidative Stress, and the Interplay of HIFs and NRF2 Signaling in Cancer. *Exp. Mol. Med.* **2024**, *1–14*. [[CrossRef](#)]
32. Hua, C.; Daien, C.L.; Combe, B.; Landewe, R. Diagnosis, Prognosis and Classification of Early Arthritis: Results of a Systematic Review Informing the 2016 Update of the EULAR Recommendations for the Management of Early Arthritis. *RMD Open* **2017**, *3*, e000406. [[CrossRef](#)] [[PubMed](#)]
33. Biswas, S.; Sharma, S.; Saroha, A.; Bhakuni, D.S.; Malhotra, R.; Zahur, M.; Oellerich, M.; Das, H.R.; Asif, A.R. Identification of Novel Autoantigen in the Synovial Fluid of Rheumatoid Arthritis Patients Using an Immunoproteomics Approach. *PLoS ONE* **2013**, *8*, e56246. [[CrossRef](#)] [[PubMed](#)]
34. Monu, X.; Agnihotri, P.; Saquib, M.; Sarkar, A.; Chakraborty, D.; Kumar, U.; Biswas, S. Transthyretin and Receptor for Advanced Glycation End Product's Differential Levels Associated with the Pathogenesis of Rheumatoid Arthritis. *J. Inflamm. Res.* **2021**, *14*, 5581–5596. [[CrossRef](#)] [[PubMed](#)]
35. Chakraborty, D.; Sarkar, A.; Mann, S.; Monu, X.; Agnihotri, P.; Saquib, M.; Malik, S.; Kumavat, R.; Mathur, A.; Biswas, S. Estrogen-Mediated Differential Protein Regulation and Signal Transduction in Rheumatoid Arthritis. *J. Mol. Endocrinol.* **2022**, *69*, R25–R43. [[CrossRef](#)]
36. Wangyang, Y.; Yi, L.; Wang, T.; Feng, Y.; Liu, G.; Li, D.; Zheng, X. MiR-199a-3p Inhibits Proliferation and Induces Apoptosis in Rheumatoid Arthritis Fibroblast-like Synoviocytes via Suppressing Retinoblastoma 1. *Biosci. Rep.* **2018**, *38*, BSR20180982. [[CrossRef](#)] [[PubMed](#)]
37. Payne, A.M.; Zorman, J.; Horton, M.; Dubey, S.; Ter Meulen, J.; Vora, K.A. Caspase Activation as a Versatile Assay Platform for Detection of Cytotoxic Bacterial Toxins. *J. Clin. Microbiol.* **2013**, *51*, 2970. [[CrossRef](#)]
38. Kim, H.; Xue, X. Detection of Total Reactive Oxygen Species in Adherent Cells by 2',7'-Dichlorodihydrofluorescein Diacetate Staining. *J. Vis. Exp.* **2020**, *2020*, e60682. [[CrossRef](#)]
39. Kang, X.; Jiao, Y.; Zhou, Y.; Meng, C.; Zhou, X.; Song, L.; Jiao, X.; Pan, Z. MicroRNA-5112 Targets IKK $\gamma$  to Dampen the Inflammatory Response and Improve Clinical Symptoms in Both Bacterial Infection and DSS-Induced Colitis. *Front. Immunol.* **2022**, *13*, 779770. [[CrossRef](#)]
40. Bendele, A.; Mccomb, J.; Gould, T.; Mcabee, T.; Sennello, G.; Chlipala, E.; Guy, M. Animal Models of Arthritis: Relevance to Human Disease. *Toxicol. Pathol.* **1999**, *27*, 134–142. [[CrossRef](#)]
41. af Klint, E.; Catrina, A.I.; Matt, P.; Neregråd, P.; Lampa, J.; Ulfgren, A.K.; Klareskog, L.; Lindblad, S. Evaluation of Arthroscopy and Macroscopic Scoring. *Arthritis Res. Ther.* **2009**, *11*, R81. [[CrossRef](#)] [[PubMed](#)]
42. Iwamoto, N.; Furukawa, K.; Endo, Y.; Shimizu, T.; Sumiyoshi, R.; Umeda, M.; Koga, T.; Kawashiri, S.Y.; Igawa, T.; Ichinose, K.; et al. Methotrexate Alters the Expression of MicroRNA in Fibroblast-like Synovial Cells in Rheumatoid Arthritis. *Int. J. Mol. Sci.* **2021**, *22*, 11561. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.