

Article

An Attempt to Relate Oleogel Properties to Wax Ester Chemical Structures

Henriette Brykczynski ^{1,*} , Birgit Hetzer ²  and Eckhard Flöter ¹

¹ Chair of Food Process Engineering, Technische Universität Berlin, 10623 Berlin, Germany

² Department of Food Technology and Bioprocess Engineering, Max Rubner-Institut, 76131 Karlsruhe, Germany

* Correspondence: h.brykczynski@tu-berlin.de

Abstract: Wax esters are considered to have a dominant contribution in the gelling properties of wax-based oleogels. To understand their gelling behavior, oleogels of seven different wax esters (total carbon number from 30 to 46; $c = 10\%$ [m/m]) in medium-chain triglycerides oil were characterized. Scanning electron microscopy revealed that wax esters crystallize in rhombic platelets with a thickness of 80 to 115 monomolecular layers. Bright field microscopy showed that the regularity and face length of the crystals increased with the total carbon number and molecular symmetry of the respective wax ester. Oscillatory rheology was used to characterize the gel rigidity (G_{max}^*). Here, wax ester oleogels with smaller total carbon numbers yielded higher G_{max}^* values than those of wax esters with higher total carbon numbers. The gel rigidity (G_{max}^*) inversely correlated with the crystal face length. Smaller and optically less well-defined platelets promoted higher gel rigidities. In the case of the microstructure of a specific oleogel composition being manipulated by a variation in the cooling rates (0.8; 5; 10 K/min), this relationship persisted. The information compiled in this manuscript further elucidates the crystallization behavior of wax esters in oleogels. This contributes to the understanding of the composition–structure–functionality relationship of wax-based oleogels supporting future food applications.

Keywords: oleogels; natural waxes; wax esters; microstructure; viscoelastic behavior; cooling rate



Citation: Brykczynski, H.; Hetzer, B.; Flöter, E. An Attempt to Relate Oleogel Properties to Wax Ester Chemical Structures. *Gels* **2022**, *8*, 579. <https://doi.org/10.3390/gels8090579>

Academic Editors: Julia Keppler and Anja Maria Wagemans

Received: 17 August 2022

Accepted: 7 September 2022

Published: 12 September 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The current research on alternatives for triglyceride-based structuring of semi-solid fat phases covers numerous different structuring agents [1]. Natural waxes are low-molecular weight oleogelators with broad availability at reasonable procurement costs that form oleogels at low concentrations [2]. Consequently, they are promising candidates for future industrial application. Hitherto, natural waxes such as beeswax (BWX), candelilla wax (CLX) and carnauba wax (CRX) are, for example, already applied as coating agents in confectionery. This implies that they are generally recognized as safe (GRAS-status) [3,4] and are widely accepted by consumers. In addition, other natural waxes such as sunflower wax (SFX) and rice bran wax (RBX) show substantial potential as oleogelators [2,5]. Nonetheless, their application in oleogels remains difficult.

Natural waxes are composed of hydrocarbons (HC), fatty acids (FA), fatty alcohols (FaOH), wax esters (WE) and some minor components. The proportions of HC, FA, FaOH and WE present in a wax depend not only on the originating plant but also on the extraction and purification processes. Undoubtedly, the oil structuring abilities of waxes result from the overall wax composition. However, long-chained WE are considered dominant for gelling behavior. Figure 1 illustrates characteristic properties of the aforementioned natural waxes. The literature data on the melting temperature [2,6], WE mass fraction [2,7] and characteristics concerning WE fraction are displayed. The data presented on the degree of homogeneity (DoH) [8] and the average total carbon numbers (CN) of FA, WE and FaOH fraction [7] are normalized to ranges typical for natural waxes. The DoH is a parameter

defining the homogeneity of a wax, considering the WE concentration as well as the chain length distribution within the WE fraction, as described, in detail, elsewhere [8]. For natural waxes, the DoH ranges between 0.23 (CLX) and 14.27 (SFX). Accordingly, this range is depicted in Figure 1 with CLX located in the center (0%) and SFX at the outer of the scale (100%). The values given for the averaged mean CN of the WE fraction as well as for the FA and FaOH constituting the WE fraction were based on data given by Doan et al. [7]. The axes for these properties were normalized such that the single FA occurring between a CN of 14 and 24 and a single FaOH with a CN between 16 and 32 could be depicted. The mean CN of the WE fraction was calculated assuming a statistical recombination of the respective FA and FaOH moieties, as previously described [9]. In line with the other parameters, the axis for the mean CN of the WE fraction was from the shortest CN present (CN = 30) to the longest CN present (CN = 56). The fat gray shaded areas indicate, respectively, which range is covered by the pure WE studied in this contribution.

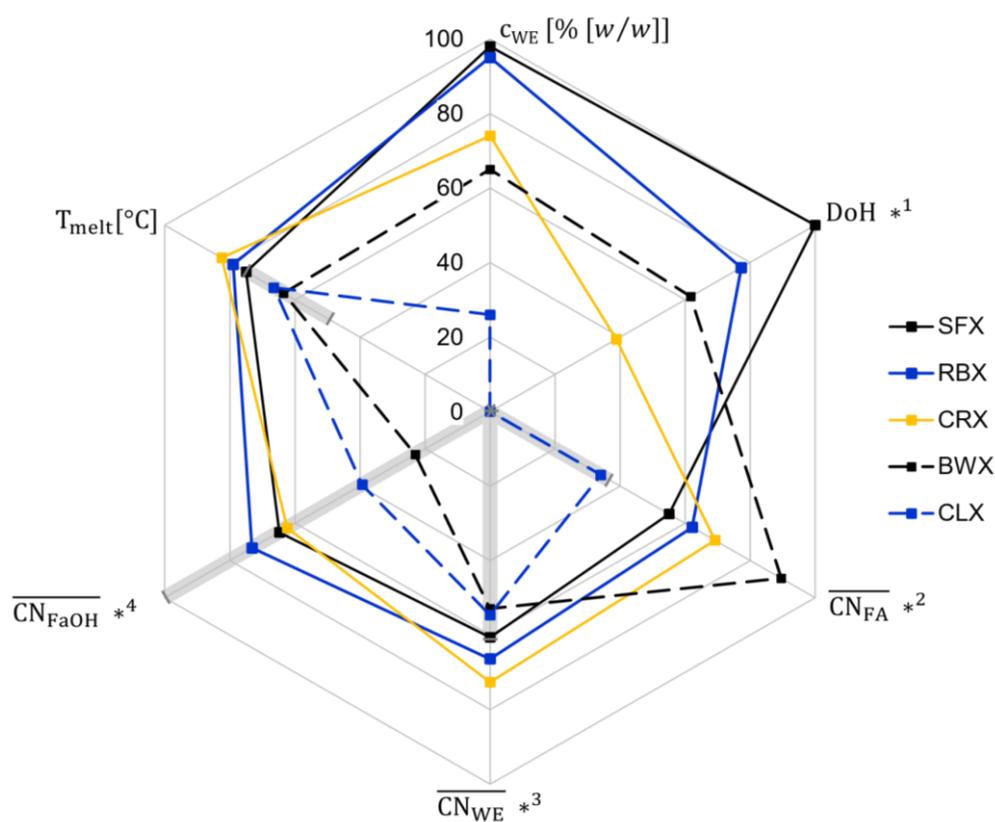


Figure 1. Characteristic parameters of different natural waxes: melting temperature (T_{melt}), total wax ester (WE) content (% [w/w]), degree of homogeneity (DoH), mean total carbon number (CN) of the WE fraction, mean CN of the fatty acid (FA) and fatty alcohol (FaOH) moieties in the WE fraction. Span of axis from center to outer: *1: DoH from 0.23 to 14.27; *2: CN_{FA} from 14 to 24; *3: CN_{WE} from 30 to 56; *4: CN_{FaOH} from 16 to 32. Gray areas: range covered by the WE studied. SFX = sunflower wax, RBX = rice bran wax, CRX = carnauba wax, BWX = beeswax, CLX = candelilla wax.

The apparent compositional variation of natural waxes propagates into their respective performance as oleogelators. It is well established that wax-based oleogels are particle gels based on the aggregation of wax crystals forming a three-dimensional network [1,10]. The structure and functionality of wax-based oleogels has extensively been studied by several authors, e.g., [5,11,12] but cannot be reviewed within this study.

Irrespective of the type of molecules—WE, FA, FaOH or HC—the crystal structure results from the parallel assembly of alkyl chains [13,14]. The effectiveness in oleogelation is related to the platelet-like crystal habit. The crystal habit is driven by the growth rates in different directions. Dassanayake et al. previously emphasized this during the discussion

of synchrotron radiation X-ray spectra and micrographs of oleogels prepared with RBX and salad oil [15].

During gel formation, nano-platelets aggregate to form a three-dimensional network [16]. It has been reported that the particle network can be manipulated by variation of, e.g., the oil to be entrapped [8,17] and the process conditions applied [18,19]. The description of the three-dimensional scaffolding is not univocal. Cardhouse-like and spherulitic structures are mainly considered [2,20–22], but also dendritic networks are described [11]. The platelet-based cardhouse networks are often interpreted to be needle-based. This misinterpretation of light-microscopic images was previously pointed out by Hwang et al. in 2015 [23] and repeatedly confirmed, e.g., [9,18,24].

The differences in gelling behavior and microstructure displayed by different natural waxes is related to variations in wax composition [20,25]. Nonetheless, until now, no relationship between the molecular composition, the process parameters and the resulting structure and functionality for wax-based oleogels has been identified. Many recent publications indicate WE to be decisive for the network formation of natural waxes [9,23,26]. In a previous contribution of ours [9], the relevance of WE crystallization for wax-based oleogels was illustrated by comparing the microstructures of either WE-based or SFX-based oleogels. Similar crystal habits could be generated by the adaption of the cooling rate. This illustrates the relevance of the crystallization of pure WE for oleogels based on natural waxes [9].

A WE crystal is composed of n stacked lamellae, each composed of m individual WE molecules, arranged in monomolecular layers (lamellae). This is schematically illustrated by Figure 2. The crystal surface per mass of solid material essentially results from the planar extension, here simplified as length and width (a and b axis) of the crystal and the platelet thickness (c axis). This depends on the number of stacked lamellae and the lamellar thickness. These theoretical considerations lead to the conclusion that the CN has a profound effect on the crystal size and hence, the crystallization behavior of WE: the higher the CN, the larger the size of a single WE molecule and thus, the lamella thickness d . Simultaneously, the lateral interactions between the alkyl chains, predominantly van der Waals (vdW) interactions (indicated by black arrows), also increase. Assuming an equal mass of solid material and a constant number of lamellae in a crystal, the crystal surface is reduced with the increasing molecular weight of the WE. This is because the number of ester bonds in the system is reduced.

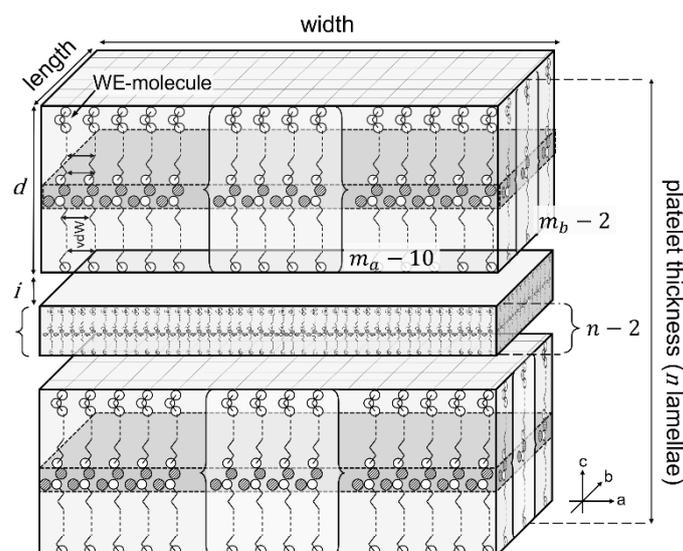


Figure 2. Schematic representation of WE crystal platelets built up of n stacked single lamellae each being composed of $m_a * m_b$ individual WE molecules in monomolecular layers. The platelet thickness results from the height of the WE molecules (d), the number of lamellae (n) and the interlamellar distance i .

The C design of wax-based oil structuring, particularly the solid phase mixing behavior, has yet to be understood. So far, this has not been studied much for oleogels based on pure WE. In this context, the influence of the chain length distribution and the symmetry of the respective WE molecules needs to be unraveled.

Recently, a first study by Avendaño-Vasquez et al. [27] reported the thermal, viscoelastic and microstructural properties of oleogels based on symmetric (CN 28; 32; 36; 40; 44) and asymmetric (FaOH_FA: 18_14; 18_16; 18_20; 18_22) WE. Previously, it was shown that pure WE form oleogels in high oleic safflower oil at low inclusion levels of 3% [w/w] [27]. Even though the publication is fairly comprehensive in its approach, revealing a systematic behavior depending on the WE structure, CN and related X-ray data, many questions remain open. The thermal properties of oleogels based on pure WE given herein and the pure WE given in another contribution [9] correspond sufficiently well. Both contributions also find that the maximum elastic moduli (G') of WE-based oleogels decrease with increasing CN.

However, based on earlier X-ray diffraction data on pure WE [28,29], it was summarized that WE crystallize in a monomolecular chain packing, either in an orthogonal or a tilted orientation of the alkyl chains within the layer [9]. The existing data indicate that the preferred polymorph is a function of the position of the ester bond within the WE molecule): the orthogonal chain arrangement seems to occur only for WE with a FaOH moiety two carbon atoms longer ($\Delta CN = +2$) or four carbon atoms shorter ($\Delta CN = -4$) than the FA moiety [9]. This is actually not in agreement with the data given in the contribution by Avendaño-Vasquez et al. [27]. Anyhow, variations in the (micro-) structure of WE-based oleogels can result not only from asymmetry but also from different polymorphs and crystal habits. This lack of certainty requires more investigations of the viscoelastic behavior and the microstructure of WE-based oleogels.

Consequently, this manuscript focuses on the viscoelastic behavior and the microstructure of WE-based oleogels. The WE investigated cover a range from 30 to 46 carbon atoms. According to pre-existing knowledge [9], all but one WE crystallize in an orthogonal chain arrangement. Hence, the difference between the CN of the FaOH and the FA residue, further designated as ΔCN , is either +2 or -4. This paper intends investigating the effect of CN independently of the respective polymorph. To eliminate the effect of differences in the number of ester bonds present in the system on the viscoelastic behavior, the dosage is on a molar basis. The microstructure of oleogels based on different WE is examined with respect to the size and shape of the crystal structures. Hence, this research aims to find a relationship between the microstructure and the resulting viscoelastic behavior.

2. Results and Discussion

2.1. Viscoelastic Behavior

In earlier publications, oleogels based on WE of different CNs were prepared on a weight basis [9,27]. Accordingly, oleogels based on WE and MCT oil were investigated at a WE inclusion level of 10% [w/w] in our previous contribution [9]. For weight fraction-based equal dosage, it remains unclear whether the reduced number of WE molecules with increasing molecular weight of the WE is responsible for the decrease in the maximum complex shear modulus (G_{max}^*). To eliminate this possible contribution, dosage was chosen to be equimolar in this study. Consequently, the number of ester bonds, potentially defining the total planar crystal area remained constant throughout the experimental plan. In Table 1, the WE weight fractions to the respective 10% molar fraction are given.

All oleogel systems were studied within their linear viscoelastic range (LVR). This was ensured using a prior amplitude strain test. (Data not shown.) The maximum complex shear modulus (G_{max}^*) was determined after 30 min crystallization time at 5 °C. The values which were determined are shown as a function of CN in Figure 3. In addition, with the molar dosage, G_{max}^* in general, decreased with increasing CN. This confirms the earlier findings which were based on dosage on an equal weight basis [9,27]. Within the homologous series, $\Delta CN = +2$, the relationship between G_{max}^* and CN appeared to be linear in the range from

CN 30 to CN 42. G_{max}^* decreased from 4.82×10^6 Pas (CN 30 (16_14)) to 8.39×10^5 Pas (CN 42 (22_20)).

Table 1. Total weight fraction for wax esters of different total carbon numbers (CN) for an inclusion level of 10% on a molar basis (% [m/m]) and, respectively, on a weight basis (% [w/w]).

Total Carbon Number (CN)	% [m/m]	% [w/w]
30	10	8.93
34	10	9.93
36	10	10.42
38	10	10.91
42	10	11.86
46	10	12.80

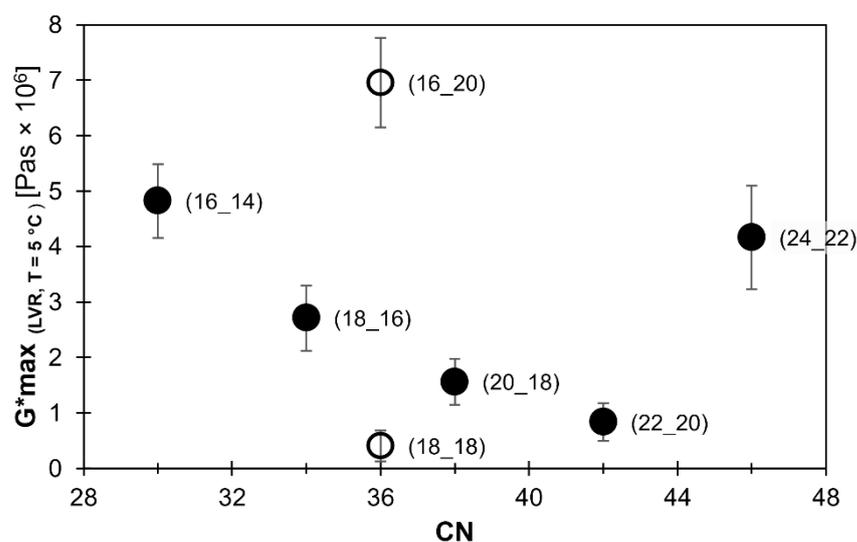


Figure 3. Maximum complex shear modulus (G_{max}^*) (in linear viscoelastic range (LVR); $T = 5$ °C) as a function of CN for mono-ester gels consisting of a pure WE (10% [m/m]) and medium chain triglycerides (MCT) oil. Filled circles: homologous series of $\Delta CN = +2$, hollow circles: not part of homologous series. All WE are denoted as (FaOH_FA).

DSC measurements of WE-based oleogels confirmed that the WE solubility in MCT oil decreases with increasing CN [9]. Taken that all gels are crystallized at the same temperature, this results in higher supersaturations for higher CNs and hence, relates to higher nucleation rates. It is reasoned elsewhere that the incorporation time into a crystal lattice is longer for larger molecules than for smaller molecules, further promoting smaller crystals for larger WE [30]. Consequently, one would expect smaller crystals for WE with higher CN and hence, G_{max}^* to increase with increasing CN. The observations made actually demonstrated the opposite trend. However, it is remarkable that the longest WE studied (CN 46 (24_22)) yielded a considerably higher value, G_{max}^* (CN 46 (24_22)) = 4.16×10^6 Pas, than the approximately linear correlation covering the other WE of the homologous series suggests. Hence, there were dramatic changes observed on the increase in CN: first, a linear decrease and then, a step change from CN 42 to 46. Thus, the observations made cannot be straightforwardly related to considerations of supersaturation, nucleation and growth rates, as the crystallization theory implies [31,32].

Figure 3 further illustrates the effect of ΔCN . At CN 36, two WE with different ΔCN , thus different lengths of the respective FA or FaOH chain, were investigated. WE CN 36 (18_18), $\Delta CN = 0$, yielded the lowest value obtained (G_{max}^* (CN 36 (18_18)(18_18)) = 4.08×10^5 Pas). In contrast, the WE CN 36 (16_20), $\Delta CN = -4$, exhibited the highest G_{max}^* value (G_{max}^* (CN 36 (16_20)) = 6.95×10^6 Pas). Comparison of the data revealed that the G_{max}^* values of WE with equal CN but different ΔCN varied within a broader range than the G_{max}^* values

of the WE of the homologous series in the CN range considered in this study. Accordingly, these data emphasize the importance of molecular symmetry (ΔCN) on the structuring performance of WE.

Overall, these observations are in line with earlier oleogel systems studies. Further, this shows that the effect of dosage on either weight or molar basis is insignificant. It was previously reported that molecular symmetry matters. This was indicated by symmetric WE ($\Delta CN = 0$) showing lower values of viscoelastic parameters than respective asymmetric WE at equal CN [9,27]. For instance, it was found that oleogels with WE at CN 36 are the softest oleogels at $\Delta CN = 0$ and most rigid at $\Delta CN = 8$ [9].

It is noteworthy to realize that the G_{max}^* values found for WE-based oleogels are comparable to those based on natural waxes [8,33] and shortenings based on fully hydrogenated oils [34]. Several parameters contribute to the viscoelastic behavior of oleogels. For TAG-based semi-solid phases, it is widely accepted that van der Waals forces are the predominant attractive interaction between nanocrystalline particles dispersed in the oil phase [35,36]. It is fair to assume that WE crystals gel oils according to the same principles. The solid mass fraction is obviously determined by the WE inclusion level and the solubility of WE [37–40]. However, the latter relates to the CN, symmetry and polymorph of WE considered. The structure efficiency additionally depends on the crystal habit and crystal dimension-driven contributions.

In summary, the hypothesis that the G_{max}^* levels of WE-based oleogels would increase with increasing CN at equimolar dosage (% [m/m]) was shown to be wrong. The dosage changes, compared to previous studies, were between minus 1% and plus 3% [w/w] (Table 1).

This only marginally affected the gel rigidity, emphasizing once more the importance of the WE molecular symmetry for WE-based oil structuring. Hence, a more in-depth understanding of the crystals and the crystal–crystal interaction is required to understand and control wax-based oleogels.

2.2. Microstructure

2.2.1. Cryo-Scanning Electron Microscopy (C-SEM)

Cryo-SEM images of a de-oiled sample of WE CN 42 (22_20) are shown in Figure 4. The micrographs illustrate that the WE crystals formed in MCT oil are well-defined platelets with straight faces. It appears that the crystal habit is rudimentarily rhombic, but the distinction of single crystals is quite obscured so that a separation of primary crystals is difficult. Individual, very thin plate-like crystals lie on top of each other. The apparent absence of a three-dimensional network structure is possibly an artifact induced by the preparation procedure. Figure 4a (2500 \times magnification) reveals that the planar area predominantly includes angles between the crystal faces of 70 $^\circ$ or 110 $^\circ$. The striking perfection of the crystal faces is illustrated by Figure 4b, showing a well-defined angle of 70 $^\circ$. Beside a rough appearance, caused by the platinum coating, the crystal surface shows no defects, and the faces of the crystal are straight.

This observation on the crystal habit is in line with the description of palmityl palmitate (CN 36 (18_18)) single crystals by Kohlhaas [41]. These were characterized to be thin rhombic platelets with rhombic angles of 74.1 $^\circ$ and 106.5 $^\circ$, which is different from the angles observable in Figure 4. The minute differences in the angles found might be due to the crystallization process, effects of the solvent or even the angle of observation.

Figure 5 shows the cryo-SEM micrograph of an oleogel sample containing MCT oil and 10% [w/w] of WE CN 42 (22_20) at 2500 \times magnification. The crystal habit observed above (Figure 4) can also be identified despite the remaining oil droplet. In this partially de-oiled sample, the spatial orientation of the crystals appears fairly random. Neither solid bridges between individual crystals nor a distinct three-dimensional network structure is identifiable.

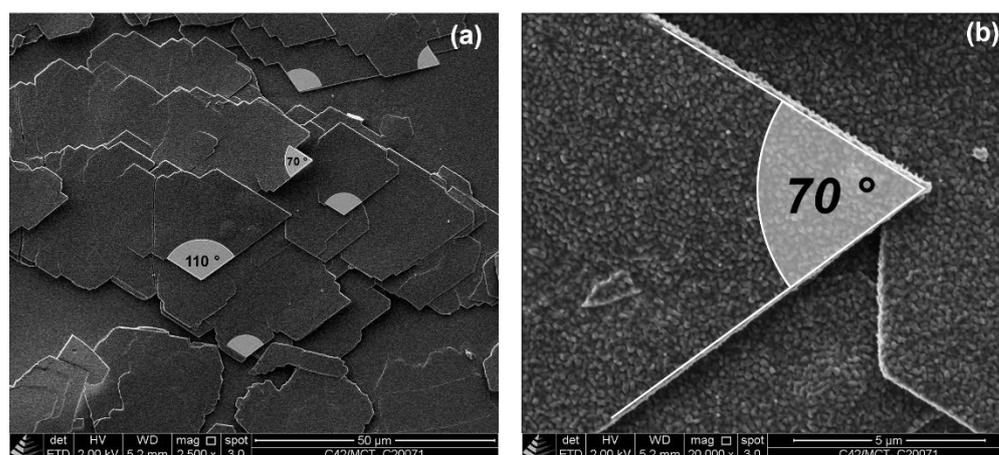


Figure 4. Scanning electron microscopy (SEM) micrographs of WE CN 42 (22_20) for a de-oiled 10% [w/w] oleogel sample: 2500× magnification (a); 20,000× magnification (b).

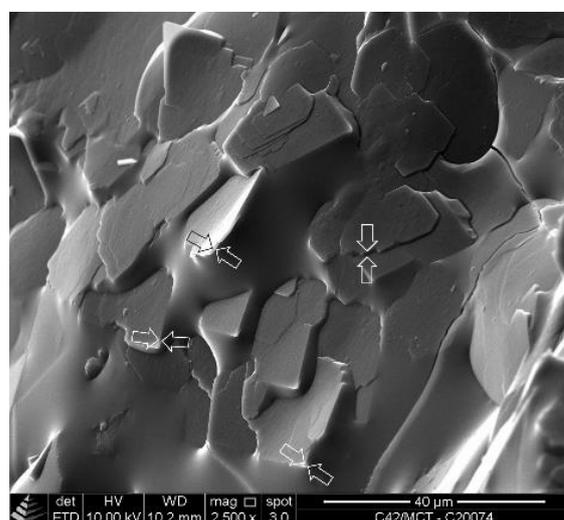


Figure 5. SEM micrograph of oleogel containing MCT oil and 10% [w/w] of WE CN 42 (22_20) (2500× magnification). Arrows: measuring positions for the determination of the crystal thickness.

The thickness of single platelets was determined based on different crystals oriented orthogonally to the projection area (indicated by arrows Figure 5). The values derived are between 460 and 570 nm.

These thickness values and layer thickness values calculated based on earlier publications [9,42] were used to derive the number of molecular layers. This resulted in 80 to 100 molecular WE layers for the orthogonal chain arrangement (Figure 6a) and 90 to 112 molecular WE layers assuming a tilted chain arrangement (Figure 6b).

Both the platelet thickness and number of stacked unimolecular layers are significantly higher than assessed for triglyceride (TAG) crystals [35]. For crystals of fully hydrogenated canola oil, Acevedo et al. [35] stated a stacking of eight lamellae which relates to crystal thicknesses much smaller (factor 15) for the TAGs than derived here for the WE.

Hence, the presented data suggest different lateral and vertical growth rates for the different aliphatic molecules. In a first approximation, possibly ignoring the effects of different growth regimes, it appears unlikely that the methyl end planes of TAGs and WE differ too much [43]. This could indicate similar growth rates orthogonal to the methyl end planes. It is worth noting that TAGs are tri-esters and thus have one ester bond per alkyl chain, whereas WE are mono-esters and have only one ester bond per two alkyl chains.

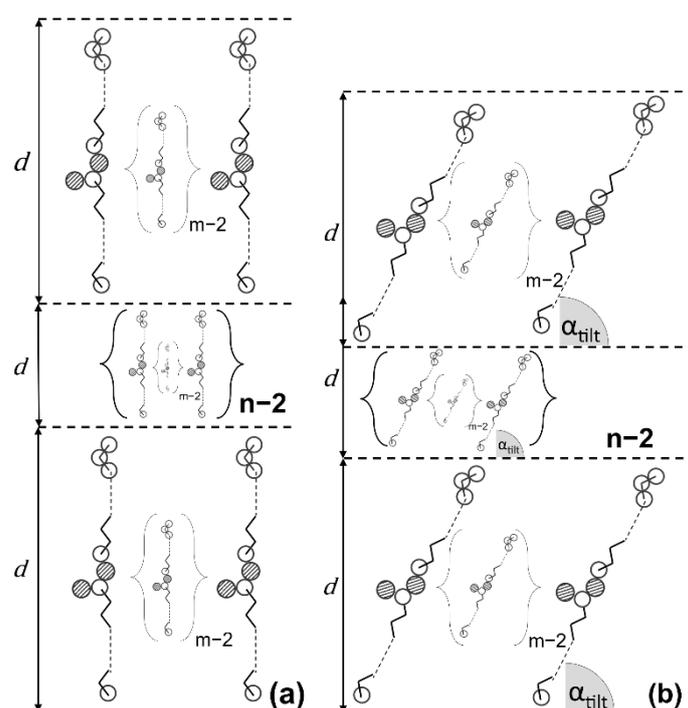


Figure 6. Schematic representation of WE molecules in unimolecular stacking: orthogonal (a) or tilted (b) chain arrangement related to the methyl end plane. White empty circles: carbon atoms, hatched circles: oxygen atoms, d = layer thickness, m = number of molecules in lateral direction, n = number of molecular layers.

This gives a first indication that the alkyl chain to ester bond ratio affects growth rates in the planar direction. However, the lamellar stacking of WE crystals certainly requires more detailed consideration.

To discuss this initial finding (i.e., the ratio of alkyl chain to ester bond in aliphatic chain crystals seem to influence the lamellar stacking in the crystalline structure) is certainly beyond the scope of this contribution but warrants further investigations.

2.2.2. Bright Field Microscopy (BFM)

The BFM images of oleogel samples containing MCT oil and 10% [m/m] of a single WE of different CNs are given in Figure 7. The WE with the lowest CN (CN 30 (16_14)) is on top, and the WE with the highest CN (CN 46 (24_22)) is at the bottom. Herein, the platelet-like morphology visible in the cryo-SEM images is confirmed. Closer examination reveals that crystal size and pronunciation of the well-defined habit increase with CN. Accordingly, the microstructure of WE CN 30 (16_14) has a low regularity, not revealing defined angles or long straight edges. The usually rhombic morphology cannot be identified. Only for this WE, needle-like crystal structures, as described for SFX-oleogels, are discernible. Conversely, all larger WE develop crystals with longer crystal faces and appear as fused rhombic structures. Consistent with the cryo-SEM images, the angles of WE CN 42 (22_20) are found to be 110° and 70° . Most other WE show essentially the same angles. Within the WE with CN 36, remarkably different microstructures are observed. The symmetric WE CN 36 (18_18) exhibits large platelets. Here, in addition to 110° and 70° , other obtuse angles such as 120° and 105° were also found. In contrast, the WE CN 36 (16_20) exhibits shorter crystal faces and a much less regular morphology. The angles identified are again 110° and 70° .

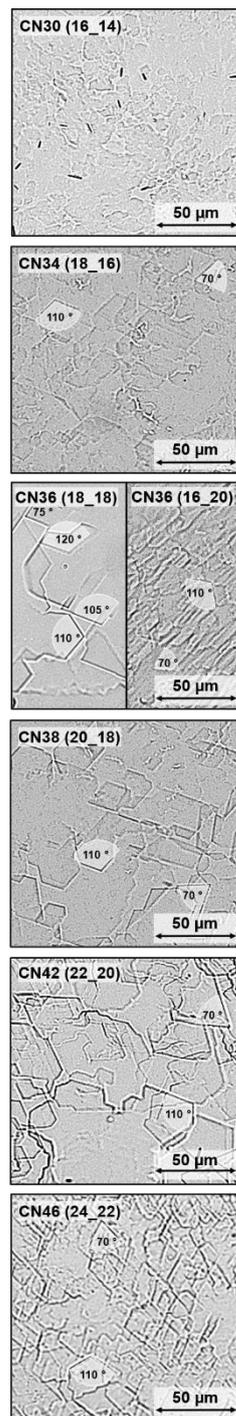


Figure 7. Bright field microscopy (BFM) images of oleogels containing MCT oil and 10% [m/m] of WE of different CN, indicated in the respective BFM micrograph. Cooling rate: 5 K/min. All WE are denoted as CN (FaOH_FA).

Due to the thickness of the specimen and single platelets being grown together, it is impossible to characterize single crystals. To further quantify the morphology, the length of the crystal faces (L_{face}) was determined using ImageJ. For each WE, the respective visible maximum face length was determined as the average of the ten longest faces identifiable in Figure 7. These characteristic values representing the crystal size are listed in Table 2.

Table 2. Face length (L_{face}) WE crystals in mono-ester gels (10% [m/m] of WE in MCT oil). L_{face} : average of ten longest straight faces, respectively. Cooling rate: 5 K/min.

WE (FaOH_FA)	L_{face} [μm]	Standard Deviation
CN 30 (16_14)	5.9	± 1.5
CN 34 (18_16)	17.6	± 3.5
CN 36 (18_18)	30.0	± 11.5
CN 36 (16_20)	11.2	± 3.1
CN 38 (18_20)	21.9	± 5.8
CN 42 (22_20)	30.8	± 6.9
CN 46 (24_22)	14.9	± 2.4

The relationship of the determined face length and the respective CN is illustrated by Figure 8. The pattern found is qualitatively inverted to that shown and described in Figure 3. Again, the homologous series from WE CN 30 (16_14) to WE CN 42 (22_20) suggests a linear relationship. Along this line, the face length increases by a factor of five. Analogous to Figure 3, the longest WE CN 46 [22_24] deviates from this pattern, showing significantly shorter face lengths than the correlation would suggest. Likewise, the data on the face length emphasize that the symmetry (ΔCN) of a WE is of utmost importance. As visible in the micrographs (Figure 7), WE CN 36 (16_20) has much shorter face lengths than CN 36 (18_18).

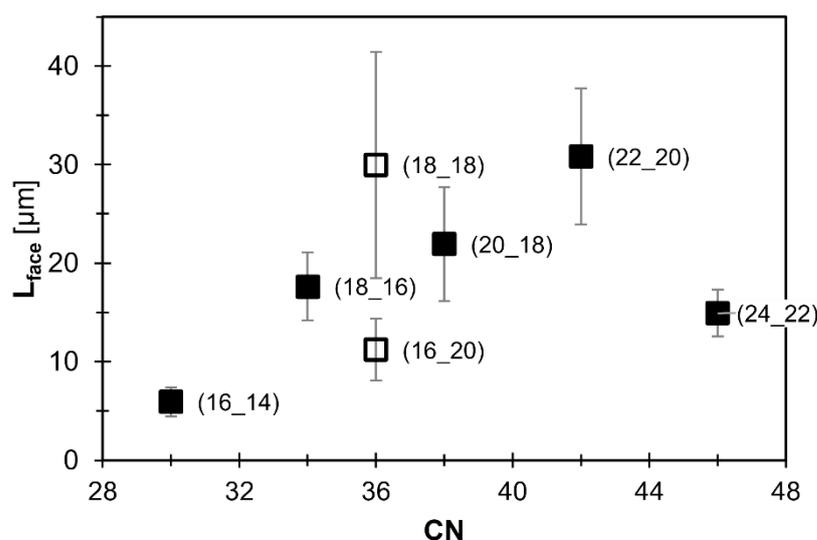


Figure 8. L_{face} of WE crystals in oleogels containing MCT oil and 10% [m/m] of WE (different CN) as a function of CN. L_{face} was determined using measurement of the ten longest straight faces. The oleogel samples were crystallized at a cooling rate of 5 K/min. Filled squares: homologous series of $\Delta\text{CN} = +2$, hollow squares: not part of the homologous series. All WE are denoted as (FaOH_FA).

2.3. Relationship between Microstructure and Viscoelastic Behavior

The simplest attempt to correlate characteristic parameters on the microstructure and the viscoelastic behavior is presented in Figure 9. The figure reveals a surprisingly strong relationship. Except for the oleogel based on WE C36 (16_20), the data suggest a linear dependency of the G_{max}^* on the face length. Obviously, this simple relationship is rather an artifact from scaling the face length to the internal surface and more precisely, to rheological properties. Anyhow, this finding indicates that the interaction of the crystals in the three-dimensional network structure does not change with the CN of WE. Accordingly, it is reasonable to assume that the variation in structure efficiency for different WE is primarily in their crystal size. The two WE with equal CN but different symmetry, CN 36 (18_18) and CN 36 (16_20), are already distinguished by a different microstructural appearance (Figure 7). Anyhow, the data for these respective oleogels adhere to the

relationship displayed in Figure 9. This documents the importance of the planar extension of WE platelets for the gelling ability of WE.

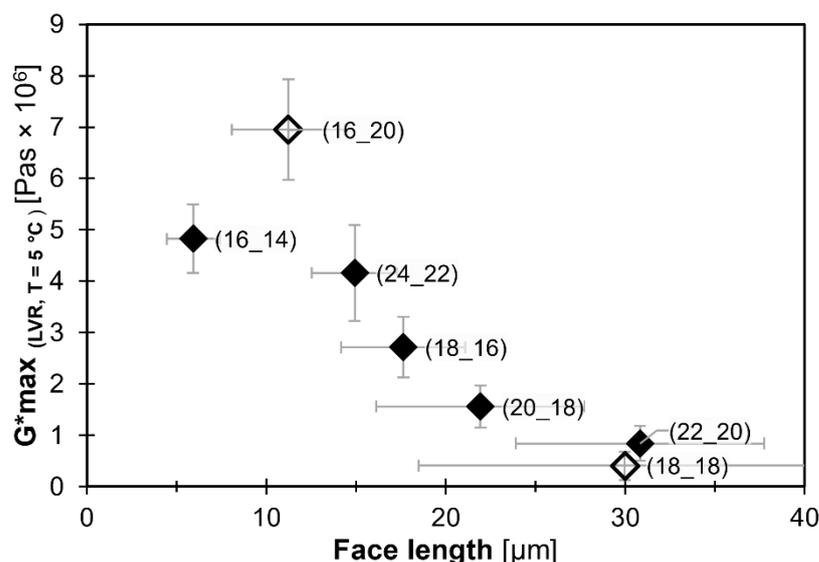


Figure 9. Respective level of G_{max}^* (within LVR; $T = 5\text{ }^{\circ}\text{C}$) as function of L_{face} for mono-ester gels (10% [m/m] WE in MCT oil). Cooling rate: 5 K/min. Filled diamonds: homologous series of $\Delta CN = +2$, hollow diamonds: not part of homologous series. All WE are denoted as (FaOH_FA).

2.4. Effect of Cooling Rate

For two oleogel systems, containing either 10% [w/w] WE CN 30 (16_14) or CN 42 (22_20), the effect of crystallizing at different cooling rates was assessed. The viscoelastic behavior (G_{max}^*) and microstructure (including the maximum face length) of the oleogels were determined.

The microstructure of the respective oleogels is illustrated by the BFM images shown in Figure 10. From top to bottom, the cooling rate increases from 0.8 K/min (slow) via 5 K/min (standard rate, Figure 7) to 10 K/min (fast). At any cooling rate, for CN 30 (16_14), neither clear straight edges nor the usual rhombic morphology could be clearly identified. However, for both WE, higher cooling rates (bottom) yield less regular and smaller platelets with shorter faces. This effect is clearly visible for WE CN 42 (22_20) due to better distinguishable structures. These micrographs prove that for WE also, the microstructure does not only change as a function of the molecular make-up but can also be manipulated by variation of the crystallization process. As generally established, increasing cooling rates result in higher nucleation rates and reduced induction times [24]. Furthermore, the results confirm that low cooling rates appear to induce more regular crystal habits as shown elsewhere [9]. The data presented here and in an earlier contribution [9] point out how arbitrary choices of cooling rates and applied supersaturations actually are.

Figure 11a illustrates that the G_{max}^* values of the oleogels increase with an increasing cooling rate. The maximum crystal face length in the respective oleogels also systematically changes, as illustrated by Figure 11b. These data are in line with expectations based on generally acknowledged crystallization kinetics, e.g., for wax-based oleogels [24,25]. They confirm that smaller crystals in compositionally equal systems typically yield more rigid gels. Figure 11c is an extension of Figure 9 and illustrates that WE oleogels produced at different cooling rates adhere to the relationship displayed in Figure 9: smaller crystals yield more rigid gels. It emphasizes the validity of the relationship found between face length and G_{max}^* , irrespective of WE-type and cooling rate.

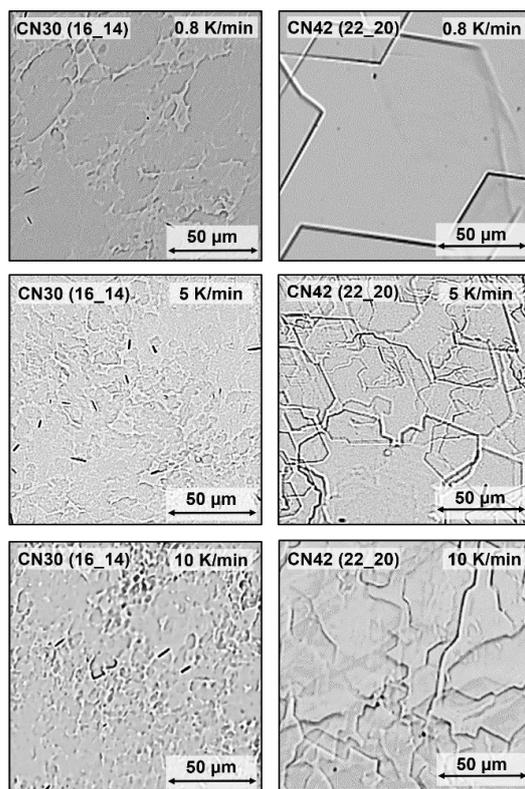


Figure 10. BFM images of WE oleogels (10% [w/w] of WE CN 30 (16_14) or CN 42 (22_20) and MCT oil) crystallized at three different cooling rates. The WE are denoted as CN (FaOH_FA).

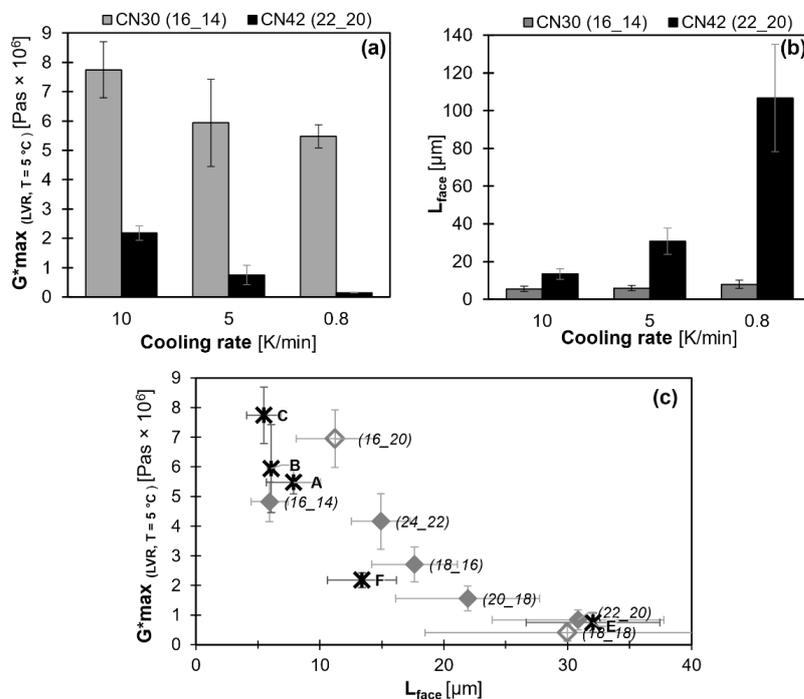


Figure 11. Effect of the cooling rate on mono-ester gels (10% [w/w] of the respective WE and MCT oil) for (a) G^*_{max} (within LVR; $T = 5^\circ\text{C}$) and (b) L_{face} ; (c) relationship between microstructure (L_{face}) and viscoelastic behavior (G^*_{max}). A: CN 30; 0.8 K/min, B: CN 30; 5 K/min, C: CN 30; 10 K/min, D: CN 42; 0.8 K/min exceeds the range of face length displayed and is not shown, E: CN 42; 5 K/min, F: CN 42; 10 K/min. Filled diamonds: homologous series of $\Delta\text{CN} = +2$, hollow diamonds: not part of homologous series. All WE are denoted as (FaOH_FA).

The data reveal that, in the range studied, the effect of cooling rate is not able to compensate the inferior structuring potential of CN 42 (22_20): even the G_{max}^* value of WE CN 30 (16_14) ($G_{max}^* (CN\ 30\ (16_14),\ 0.8\ K/min) = 5.48 \times 10^6\ Pas$) at the lowest cooling rate is higher than the G_{max}^* value of WE CN 42 (22_20) ($G_{max}^* (CN\ 42\ (22_20),\ 10\ K/min) = 2.18 \times 10^6\ Pas$) at the highest cooling rate. This illustrates that even though the effect of cooling rate is strong, it is not sufficient to compensate the material specific crystallization behavior in the range studied.

This study reveals again how difficult it is to perform correct like-for-like comparisons. However, one must acknowledge that, for practical reasons, experiments at realistic cooling rates derived from anticipated applications seem most justified. For stored products, this could mean low cooling rates. In conclusion, the responses of the WE-based oleogels studied at different cooling rates are in line with published data on wax-based oleogels [24,25,44].

3. Conclusions

In this contribution, the viscoelastic behavior and microstructure of different WE-based oleogels are discussed. The oleogels were composed of MCT oil and 10% of a pure WE, respectively. The respective chain length of the WE ranged from 30 to 46 carbon atoms. The WE were mainly selected as a homologous series with a carbon number difference (ΔCN) of +2. Irrespective of CN, WE with such a molecular symmetry assume an orthogonal chain arrangement in crystals. Additionally, two WE with CN 36, one symmetric ($\Delta CN = 0$), one asymmetric ($\Delta CN = -4$), were investigated. Earlier publications showed that asymmetry entails the least effects on the caloric properties of WE-based oleogels.

To normalize the WE inclusion regarding the number of ester bonds present, mostly samples with an inclusion level of 10% on a molar basis ($[m/m]$) were studied. However, changing from mass to molar dosage only marginally affected the viscoelastic oleogel properties. This proved the hypothesis that structuring functionality is driven by the cumulative planar crystal area wrong.

Bright field microscopy and scanning electron microscopy revealed that WE in oleogels crystallize in very regular, almost two-dimensional platelets with a stacking between 80 and 115 monomolecular layers. Among the primary platelet-like crystals, no sintering was observed. Anyhow, the molecular make-up of the WE regarding CN and ΔCN was found to have a strong effect on crystal habit and size. For increasing CN, more distinct and regular crystals were observed. The projection of planar crystals revealed angles between straight faces of consistently 70° and 110° for most WE. It was found that WE-based oleogels in which no regular, rhombic crystals could be identified by BFM exhibit significantly higher G_{max}^* values. The length of the identifiable straight faces in BFM images was defined as a characteristic value for crystal size. It was found that within the homologous series of WE, this length decreased linearly with increasing CN.

To study the effect of the cooling rate on gel structure, oleogels of WE CN 30 (16_14) and CN 42 (22_20), respectively, were investigated. In line with expectations, higher cooling rates yielded smaller crystals and higher G_{max}^* values. For the rigidity of the oleogels, it was found that G_{max}^* strongly correlated with the characteristic size parameter of the crystals, the maximum face length (L_{face}). More rigid gels related to smaller crystals. However, all oleogels studied here, irrespective of the WE employed or the cooling rate applied, adhered to the apparently linear relationship of G_{max}^* and the characteristic face length. This indicates that in WE-based oleogels, the crystal size is next to dosage as the dominant factor defining gel strength. Furthermore, these findings suggest that the crystal–crystal interaction defining the three-dimensional network structure remains unchanged throughout the experimental set of gels investigated here.

Admittedly, studies on oleogels based on MCT oil and pure WE appear to be of a rather academic nature compared to product applications of wax-based oleogels. However, the observations of these rather pure systems resemble those made for wax-based oleogels. Even though the fundamental insights are already relevant for future wax-based oleogel

applications in foods, further extension into mixtures of wax esters is recommended. This way, a knowledge base can be generated, enabling an a priori design of wax-based structuring systems.

4. Materials and Methods

4.1. Material

The WE used in this study were purchased from Larodan AB (Solna, Sweden): specifically, palmityl myristate (CN 30 (16_14)), stearyl palmitate (CN 34 (18_16)), myristyl behenate (CN 36 (14_22)), stearyl stearate (CN 36 (18_18)), behenyl myristate (CN 36 (22_14)), arachidyl palmitate (CN 36 (20_16)), arachidyl stearate (CN 38 (20_18)), behenyl arachidate (CN 42 (22_20)) and lignoceryl behenate (CN 46 (24_22)), all with a purity >99%. In this manuscript, the carbon number of the FaOH residue is indicated by the first number, and the carbon number of the FA residue is indicated by the second number.

Medium-chain triglycerides (MCT) oil was obtained from Caesar & Lorentz GmbH (Hilden, Germany). By GC analysis of FA methyl esters according to DGF method C-VI 10a (00), the FA composition of the MCT oil was determined to be 56% caprylic acid (10:0) and 44% capric acid (8:0).

All materials were used without further purification or modification.

4.2. Methods

4.2.1. Oleogel Preparation

To form oleogels, 10% [*m/m*] of a single pure WE were dissolved in MCT oil. The specific WE were selected to investigate the effects of the CN for WE that accept an orthogonal chain arrangement ($\Delta CN = +2$ or -4). Further, at CN 36, a symmetric WE ($\Delta CN = 0$) was studied. In 1.5 mL screw neck vials, 300 mg (weighing precision: ± 0.03 mg) of oleogel were prepared as stock solution. The samples were heated ($T = 95$ °C) under agitation (250 rpm) to ensure complete dissolution. For the different analytical methods applied, the samples were directly transferred to the respective device configuration while still liquid at high temperature. This was done to avoid changes in the gel composition and network structure.

4.2.2. Viscoelastic Behavior (Rheology)

The viscoelastic behavior of the oleogel systems was studied on a MCR 302 rheometer (Anton Paar GmbH, Graz, Austria). To avoid slippage, a steel plate–plate geometry with sandblasted surface (PP25-S; $d = 25$ mm) was used. Accurate temperature control was ensured by a top and base Peltier system of the measurement geometry (measuring gap: 0.2 mm).

For sample preparation, the plate was preheated ($T = 90$ °C) and around 100 mg of the hot stock solution were poured on the center. The samples were sheared ($\dot{\gamma} = 10$ s⁻¹; $t = 5$ min) for standardization and then stabilized under quiescent conditions (1 min). Gel-sol transitions were observed during the cooling from 90 °C to 5 °C. Afterwards, an isothermal stabilization step was implemented ($T = 5$ °C, $t = 30$ min). After that, the isothermal amplitude strain test was performed from 0.005% to 100% at an angular viscosity of 10 rad/s. Prior to repeating the same procedure, samples were heated to 90 °C and standardized. For all measurements, the angular frequency was 10 rad/s, and for temperature and time tests, the applied strain was 0.005%. For the characterization of the viscoelastic behavior, the heating or cooling rates were 5 K/min. For the study on the effect of cooling rate variation, the stated cooling rates were applied. In order to compensate changes in the sample density, the gap width was automatically regulated during the test ($F_N = 0$ N) and reset to 0.2 mm after each heating step. The crystallization was performed in quadruple, and the amplitude sweep was performed in duplicate.

4.2.3. Microstructure

Scanning Electron Microscopy (SEM)

Since the available amount of wax ester oleogel was very low, about 50 mg of the samples were placed on a silicon wafer and partially de-oiled with isobutanol which was poured over the sample. Additionally, individual crystalline platelets from the oleogels were isolated using deposition of a few drops of the retained de-oiling solution (isobutanol with traces of washed out MCT oil) on another silicon wafer. Following freezing by liquid nitrogen, the samples were transferred to the cryo chamber (PP2000 T, Quorum Technologies Ltd., Laughton, UK) and pre-cooled at $-135\text{ }^{\circ}\text{C}$. The samples were sublimated ($T = -90\text{ }^{\circ}\text{C}$, $t = 15\text{ min}$) inside the cryo chamber to eliminate eventual ice contamination on the sample's surface. The samples were coated with platinum in Argon atmosphere ($t = 30\text{ s}$ for coating, current: ca. 5–10 mA). This, in order to minimize charging problems. Lastly, samples were loaded to the cryo stage in the SEM chamber ($T = -135\text{ }^{\circ}\text{C}$). For imaging, Quanta 250 FEG field emission scanning electron microscope (FEI, Brno, Czech Republic) under high vacuum ($3 \times 10^{-7}\text{ mbar}$) was used with an Everhart–Thornley detector (working distance = 5 mm; accelerating voltage = 10 kV). This procedure was kindly performed by the Department of Food Technology and Bioprocess Engineering, Max Rubner-Institut, Karlsruhe, 76131 Germany.

Bright Field Microscopy (BFM)

The objective slides for the sample preparation were pre-heated ($T = 90\text{ }^{\circ}\text{C}$), then a 10 μL -droplet of the hot oleogel stock solution was poured on it, and a cover slip was placed. A thermoelectric stage was used to apply different cooling rates in a temperature range from $90\text{ }^{\circ}\text{C}$ to $5\text{ }^{\circ}\text{C}$. Stabilization of the samples was performed prior to the image recording ($T = 5\text{ }^{\circ}\text{C}$; $t = 1\text{ h}$). All BFM images were captured at ambient temperature.

Author Contributions: Conceptualization, H.B. and E.F.; methodology, H.B. and B.H.; validation, H.B. and B.H.; formal analysis, H.B. and B.H.; investigation, H.B.; writing—original draft preparation, H.B.; writing—review and editing, E.F.; visualization, H.B.; supervision, E.F.; project administration, E.F. All authors have read and agreed to the published version of the manuscript.

Funding: Part of the study was funded by the Elsa Neumann Scholarship (Henriette Brykczynski).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Flöter, E.; Wettlaufer, T.; Conty, V.; Scharfe, M. Oleogels-Their Applicability and Methods of Characterization. *Molecules* **2021**, *26*, 1673. [[CrossRef](#)] [[PubMed](#)]
2. Blake, A.I.; Co, E.D. Marangoni AG Structure and Physical Properties of Plant Wax Crystal Networks and Their Relationship to Oil Binding Capacity. *J. Am. Oil. Chem. Soc.* **2014**, *91*, 885–903. [[CrossRef](#)]
3. No 231/2012, Commission Regulation (EU). Publications Office of the European Union: Brussels, Belgium, 2012.
4. Title 21: Foods and Drugs CFR: § 172.890. In *Direct Food Substances Affirmed as Generally Recognized as Safe*; 2012; Volume 01.04.2021. Available online: <https://www.govinfo.gov/content/pkg/CFR-2021-title21-vol3/pdf/CFR-2021-title21-vol3.pdf> (accessed on 17 August 2022).
5. Doan, C.D. Structure-function of wax-based oleogels prepared in rice bran oil. Ph.D. Thesis, Ghent University, Ghent, Belgium, 2017.
6. Holloway, P.J. The composition of beeswax alkyl esters. *J. Am. Oil. Chem. Soc.* **1969**, *46*, 189–190. [[CrossRef](#)]
7. Doan, C.D.; To, C.M.; De Vrieze, M.; Lynen, F.; Danthine, S.; Brown, A.; Dewettinck, K.; Patel, A.R. Chemical profiling of the major components in natural waxes to elucidate their role in liquid oil structuring. *Food Chem.* **2017**, *214*, 717–725. [[CrossRef](#)] [[PubMed](#)]
8. Wettlaufer, T.; Brykczynski, H.; Flöter, E. Wax-Based Oleogels—Properties in Medium Chain Triglycerides and Canola Oil. *Eur. J. Lipid. Sci. Technol.* **2021**, *124*, 2100114. [[CrossRef](#)]
9. Brykczynski, H.; Wettlaufer, T.; Flöter, E. Revisiting pure component wax esters as basis of wax-based oleogels. *J. Am. Oil. Chem. Soc.* **2022**. [[CrossRef](#)]
10. Marangoni, A.G.; Garti, N. *Edible Oleogels: Structure and Health Implications*; AOCS Press: Urbana, IL, USA, 2011.

11. Blake, A.I.; Toro-Vazquez, J.F.; Hwang, H.-S. *Wax Oleogels*; Academic Press: London, UK; AOCS Press: London, UK, 2018; pp. 133–171.
12. Patel, A.R. Natural Waxes as Oil Structurants. In *Alternative Routes to Oil Structuring*; Patel, A.R., Ed.; Springer International Publishing: Cham, Germany, 2015; pp. 15–27.
13. Ensikat, H.J.; Boese, M.; Mader, W.; Barthlott, W.; Koch, K. Crystallinity of plant epicuticular waxes: Electron and X-ray diffraction studies. *Chem. Phys. Lipids* **2006**, *144*, 45–59. [[CrossRef](#)] [[PubMed](#)]
14. Larsson, K. Lipids: Molecular organization, physical functions and technical applications. In *Oily Press lipid library*; Oily Press: Dundee, UK, 1994; Volume 5.
15. Dassanayake, L.S.K.; Kodali, D.R.; Ueno, S.; Sato, K. Crystallization kinetics of organogels prepared by rice bran wax and vegetable oils. *J. Oleo Sci.* **2012**, *61*, 1–9. [[CrossRef](#)]
16. Marangoni, A.G.; Westorp, L.H. (Eds.) *Structure and Properties of Fat Crystal Networks*, 2nd ed.; CRC Press: Boca Raton, FL, USA, 2013.
17. Martins, A.J.; Cerqueira, M.A.; Fasolin, L.H.; Cunha, R.L.; Vicente, A.A. Beeswax organogels: Influence of gelator concentration and oil type in the gelation process. *Food Res. Int.* **2016**, *84*, 170–179. [[CrossRef](#)]
18. Blake, A.I.; Marangoni, A.G. The Use of Cooling Rate to Engineer the Microstructure and Oil Binding Capacity of Wax Crystal Networks. *Food Biophys.* **2015**, *10*, 456–465. [[CrossRef](#)]
19. Blake, A.I.; Marangoni, A.G. The Effect of Shear on the Microstructure and Oil Binding Capacity of Wax Crystal Networks. *Food Biophys.* **2015**, *10*, 403–415. [[CrossRef](#)]
20. Wettlaufer, T.; Hetzer, B.; Flöter, E. Characterization of Oleogels Based on Waxes and Their Hydrolyzates. *Eur. J. Lipid. Sci. Technol.* **2021**, *123*, 2000345. [[CrossRef](#)]
21. Ögütçü, M.; Arifoğlu, N.; Yılmaz, E. Preparation and Characterization of Virgin Olive Oil-Beeswax Oleogel Emulsion Products. *J. Am. Oil. Chem. Soc.* **2015**, *92*, 459–471. [[CrossRef](#)]
22. Rocha JC, B.; Lopes, J.D.; Mascarenhas MC, N.; Arellano, D.B.; Guerreiro LM, R.; da Cunha, R.L. Thermal and rheological properties of organogels formed by sugarcane or candelilla wax in soybean oil. *Food Res. Int.* **2013**, *50*, 318–323. [[CrossRef](#)]
23. Hwang, H.S.; Kim, S.; Evans, K.O.; Koga, C.; Lee, Y. Morphology and networks of sunflower wax crystals in soybean oil organogel. *Food Struct.* **2015**, *5*, 10–20. [[CrossRef](#)]
24. Wettlaufer, T.; Flöter, E. Effect of Cooling Rate on the Oleogel Properties of Wax–Wax-Hydrolyzate Mixtures. *Food Biophys.* **2022**. [[CrossRef](#)]
25. Doan, C.D.; Tavernier, I.; Okuro, P.K.; Dewettinck, K. Internal and external factors affecting the crystallization, gelation and applicability of wax-based oleogels in food industry. *Innov. Food Sci. Emerg. Technol.* **2018**, *45*, 42–52. [[CrossRef](#)]
26. Blake, A.I. Marangoni AG Plant wax crystals display platelet-like morphology. *Food Struct.* **2015**, *3*, 30–34. [[CrossRef](#)]
27. Avendaño-Vásquez, G.; De la Peña-Gil, A.; Charó-Alvarado, M.E.; Charó-Alonso, M.A.; Toro-Vazquez, J.F. Self-Assembly of Symmetrical and Asymmetrical Alkyl Esters in the Neat State and in Oleogels. *Front. Sustain. Food Syst.* **2020**, *4*, 132. [[CrossRef](#)]
28. Iyengar, B.T.; Schlenk, H. Melting points of synthetic wax esters. *Lipids* **1969**, *4*, 28–30. [[CrossRef](#)]
29. Aleby, S.; Fischmeister, I.; Iyengar, B.T.R. The infrared spectra and polymorphism of long chain esters: IV. Some esters from tetradecanol, hexadecanol, octadecanol, eicosanol, docosanol and dodecanoic, tetradecanoic, hexadecanoic, octadecanoic and eicosanoic acid. *Lipids* **1971**, *6*, 421–425. [[CrossRef](#)]
30. Himawan, C.; Starov, V.M.; Stapley, A.G.F. Thermodynamic and kinetic aspects of fat crystallization. *Adv. Colloid Interface Sci.* **2006**, *122*, 3–33. [[CrossRef](#)] [[PubMed](#)]
31. Callau, M.; Sow-Kébé, K.; Nicolas-Morgantini, L.; Fameau, A.L. Effect of the ratio between behenyl alcohol and behenic acid on the oleogel properties. *J. Colloid. Interface Sci.* **2020**, *560*, 874–884. [[CrossRef](#)] [[PubMed](#)]
32. Gandolfo, F.G.; Bot, A.; Flöter, E. Structuring of edible oils by long-chain FA, fatty alcohols, and their mixtures. *J. Am. Oil. Chem. Soc.* **2004**, *81*, 1–6. [[CrossRef](#)]
33. Choi, K.O.; Hwang, H.S.; Jeong, S.; Kim, S.; Lee, S. The thermal, rheological, and structural characterization of grapeseed oil oleogels structured with binary blends of oleogelator. *J. Food Sci.* **2020**, *85*, 3432–3441. [[CrossRef](#)]
34. Ahmadi, L.; Wright, A.J.; Marangoni, A.G. Structural and Mechanical Behavior of Tristearin/Triolein-rich Mixtures and the Modification Achieved by Interesterification. *Food Biophys.* **2009**, *4*, 64–76. [[CrossRef](#)]
35. Acevedo, N.C. Characterization of the Nanostructure of Triacylglycerol Crystal Networks. In *Structure-Function Analysis of Edible Fats*; Marangoni, A.G., Ed.; AOCS Press: Champaign, IL, USA, 2018; pp. 1–19.
36. Nederveen, C. Dynamic mechanical behavior of suspensions of fat particles in oil. *J. Colloid Sci.* **1963**, *18*, 276–291. [[CrossRef](#)]
37. Aydın, A.A. High-chain fatty acid esters of 1-octadecanol as novel organic phase change materials and mathematical correlations for estimating the thermal properties of higher fatty acid esters' homologous series. *Sol. Energy Mater. Sol. Cells* **2013**, *113*, 44–51. [[CrossRef](#)]
38. Aydın, A.A. High-chain fatty acid esters of 1-hexadecanol for low temperature thermal energy storage with phase change materials. *Sol. Energy Mater. Sol. Cells* **2012**, *96*, 93–100. [[CrossRef](#)]
39. Aydın, A.A.; Okutan, H. High-chain fatty acid esters of myristyl alcohol with even carbon number: Novel organic phase change materials for thermal energy storage—1. *Sol. Energy Mater. Sol. Cells* **2011**, *95*, 2752–2762. [[CrossRef](#)]
40. Floeter, E.; Haeupler, M.; Sato, K. Molecular Interactions and Mixing Phase Behavior of Lipid Crystals. In *Crystallization of Lipids: Fundamentals and Applications in Food, Cosmetics, and Pharmaceuticals*, 1st ed.; Sato, K., Ed.; Wiley: Hoboken, NJ, USA, 2018; Volume 149, pp. 61–104.

41. Kohlhaas, R. Röntgenographische Untersuchung von definierten Einkristallen des Palmitinsäure-Cetylesters. *Z. Für Krist. Cryst. Mater.* **1938**, *98*, 418–438. [[CrossRef](#)]
42. Kreger, D.R.; Schamhart, C. On the long crystal-spacings in wax esters and their value in micro-analysis of plant cuticle waxes. *Biochim. Et Biophys. Acta* **1956**, *19*, 22–44. [[CrossRef](#)]
43. Sato, K. (Ed.) *Crystallization of Lipids: Fundamentals and Applications in Food, Cosmetics, and Pharmaceuticals*, 1st ed.; Wiley: Hoboken, NJ, USA, 2018.
44. Yao, Y.; Zhou, H.; Liu, W.; Li, C.; Wang, S. The Effect of Cooling Rate on the Microstructure and Macroscopic Properties of Rice Bran Wax Oleogels. *J. Oleo Sci.* **2021**, *70*, 135–143. [[CrossRef](#)] [[PubMed](#)]