

Review

Advances of Hydrogel Therapy in Periodontal Regeneration—A Materials Perspective Review

Maoxue Li [†], Jiayi Lv [†], Yi Yang, Guoping Cheng, Shujuan Guo, Chengcheng Liu and Yi Ding ^{*}

State Key Laboratory of Oral Diseases, Department of Periodontics, West China School & Hospital of Stomatology, Sichuan University, Chengdu 610041, China

^{*} Correspondence: yidingscu@163.com

[†] These authors contributed equally to this work.

Abstract: Hydrogel, a functional polymer material, has emerged as a promising technology for therapies for periodontal diseases. It has the potential to mimic the extracellular matrix and provide suitable attachment sites and growth environments for periodontal cells, with high biocompatibility, water retention, and slow release. In this paper, we have summarized the main components of hydrogel in periodontal tissue regeneration and have discussed the primary construction strategies of hydrogels as a reference for future work. Hydrogels provide an ideal microenvironment for cells and play a significant role in periodontal tissue engineering. The development of intelligent and multifunctional hydrogels for periodontal tissue regeneration is essential for future research.

Keywords: hydrogel; periodontal tissue regeneration; tissue engineering; periodontitis; delivery system; scaffold material



Citation: Li, M.; Lv, J.; Yang, Y.; Cheng, G.; Guo, S.; Liu, C.; Ding, Y. Advances of Hydrogel Therapy in Periodontal Regeneration—A Materials Perspective Review. *Gels* **2022**, *8*, 624. <https://doi.org/10.3390/gels8100624>

Academic Editor: Mohsen Akbari

Received: 26 August 2022

Accepted: 27 September 2022

Published: 30 September 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Periodontitis is a chronic, destructive inflammation characterized by microbial infection and accelerated loss of alveolar bone, ultimately resulting in the loss of teeth [1]. As the world ages, periodontitis has become one of the major oral diseases, affecting a significant number of people around the world. Epidemiological evidence shows that approximately 20–50% of the global population suffers from periodontal-related diseases, and approximately 10% of the global population is affected by severe periodontitis [2]. There is a pressing need to address the challenges of periodontitis and bone loss in older populations [3]. Furthermore, it is now well accepted that periodontal disease is strongly associated with systemic diseases such as diabetes, cardiovascular disease, Alzheimer's disease, and other inflammatory comorbidities [4]. Whether in developed or developing countries, periodontitis imposes severe medical burdens on the population [5]. Managing periodontal diseases and promoting periodontal tissue regeneration are essential for oral and general health.

Periodontal tissue is the functional system surrounding teeth and has a complex hierarchical structure comprising hard and soft tissue together as a whole. Periodontal tissue regeneration involves the reconstitution of periodontal ligament (PDL) and alveolar bone around the teeth and cementum [6]. The ideal goal of periodontal treatment is to achieve good regeneration of the damaged periodontal tissue. Conventional periodontal therapies such as mechanical debridement and flap surgery are mainly aimed at removing plaque and pathological granulation tissue to prevent the progression of inflammation and the further destruction of periodontal tissue [7]. Reconstruction of the morphology and function of the damaged periodontal tissue is the ideal objective of periodontal regeneration treatment but remains a major challenge [8]. Guided tissue regeneration and bone grafts are currently the clinical approaches to periodontal tissue regeneration, but these techniques have shown limitations in indications such as intraosseous defects and class II fissure defects. The high technical sensitivity also limits their regenerative effect [9]. Periodontal tissue engineering

has emerged as a promising technology to address periodontal diseases and is a technique that uses a combination of stem cells, biological scaffold material implanted in the body, and growth factors to promote periodontal tissue regeneration [10]. In recent years, a large range of scaffold materials has been designed to promote alveolar bone formation and are currently one of the main ways to restore periodontal tissue that has been damaged by inflammation. Ideal biomaterials can effectively recruit regeneration-related functional cells, promote their proliferation and differentiation, and lead to the formation of new periodontal tissues [11]. However, the main challenge in periodontal regeneration therapy currently arises from the simultaneous or sequential repair of the morphology and function of PDL, alveolar bone, and cementum [12]. Traditional two-dimensional biomaterials, such as GTR barriers, provide adhesion sites and prevent soft tissue from growing into bone defects, but biological stimulation for functional cells is limited.

Hydrogels are three-dimensional water-swollen polymeric materials with superior biocompatibility, mechanical strength, and accessibility that have been widely used in biomedical applications such as cell culture [13], drug delivery [14], and tissue engineering [15]. In tissue engineering, biomaterials provide a three-dimensional scaffold for cell adhesion, proliferation, and differentiation. The scaffold should be a porous, three-dimensional, network-like structure providing cells with the necessary space to deposit their extracellular matrix while exchanging cellular substances with the surrounding environment. Due to their distinctive three-dimensional mesh structure, high porosity, superior hydrophilicity and viscoelasticity, and controllable compositions, hydrogels can mimic the microenvironment of the extracellular matrix, which is favorable for cell attachment, proliferation, and differentiation. Through combination with drugs [16], stem cells [17], or growth factors [18], hydrogels show significant potential in periodontal regeneration and have gained a considerable amount of attention in recent years. Periodontal tissue regeneration is a complex and sophisticated process. Hydrogels have been widely applied as scaffolds for regenerative medicine and as a sustained-release system in periodontal tissue engineering. Current research has noted that the composition and structure of hydrogels have a significant impact on periodontal tissue regeneration. However, there is no paper summarizing these impacts which may pave the way for researchers to develop appropriate hydrogel designs in periodontal tissue engineering. Based on the above, this paper reviews the applications of hydrogels in periodontal tissue regeneration research and provides discussions and prospects about their future designs, with the objective of making a valuable contribution to successful periodontal tissue regeneration.

2. Components of Hydrogel in Periodontal Tissue Regeneration

The fundamental components of hydrogels determine the properties and function of the material. Additionally, hydrogels can encapsulate bioactive substances to provide the hydrogels with antibacterial, anti-inflammatory, osteogenetic, and osteoimmunology capabilities and improve the regenerative effect of periodontal tissue as needed. This section focuses on the main components of hydrogels and the substances carried on them, as summarized in Figure 1.

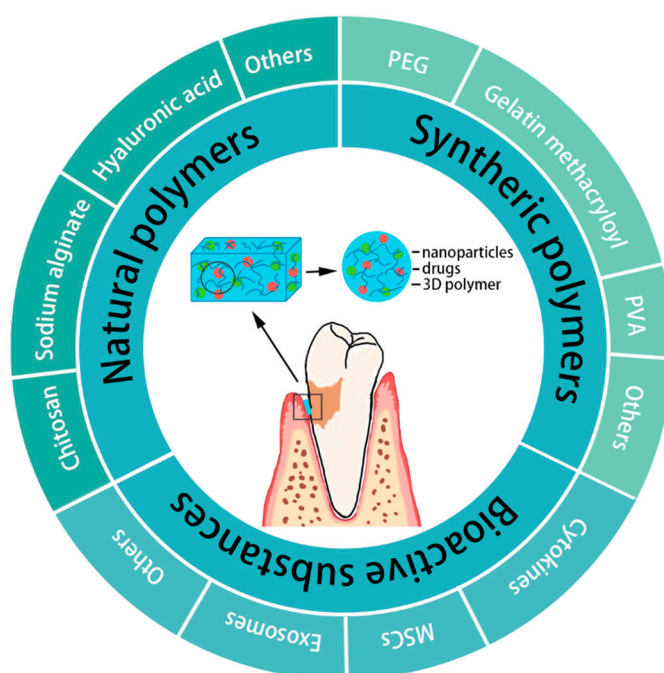


Figure 1. Schematic diagram of the classification of hydrogels in periodontal regeneration.

2.1. The Fundamental Components of Hydrogels in Periodontal Tissue Regeneration

2.1.1. Natural Polymers

The fundamental components of hydrogels are mainly categorized into natural and synthetic polymers. Natural polymer materials are directly derived from natural resources. Since the ECM consists primarily of polysaccharides, glycosaminoglycans, and a variety of proteins, natural polymers generally have excellent biocompatibility and biodegradability, and most of these polymers are water-soluble. The hydrophilic surface facilitates cell adhesion, proliferation, and differentiation. However, the mechanical strength and stability of natural polymers are not as high as those of synthetic hydrogels, which also limits their application to some extent [19].

- Chitosan

Chitosan (CS) is a natural polysaccharide with a chemical structure and biological properties similar to those of glycosaminoglycan, a major component of the extracellular matrix [20]. With good biocompatibility, hemostatic properties [21], much higher adhesiveness [22] and antimicrobial activity [23], CS can potentially be used for the synthesis of various gels in periodontal therapy. Yan, X. Z. et al. [24] reported that enzymatically solidified chitosan hydrogels with or without cell loading showed great potential in periodontal regeneration in terms of functional ligament length. Histological analysis demonstrated that after four weeks of implantation, chitosan hydrogels were largely degraded, without causing any adverse reactions in the surrounding tissue. Furthermore, crosslinking modification of chitosan can form hydrogels with certain swelling properties. Ji, Q. X. et al. [25] developed CS-based thermosensitive hydrogels composed of CS, quaternized CS, and beta-glycerophosphate which showed stronger antibacterial activity toward periodontal pathogens. In addition, CS is the only natural polycationic alkaline polysaccharide possessing a large number of free amino groups, which could enable electrostatic interactions and hydrogen bonding to form hydrogels without using any additional agents [26] and could be modified by introducing other groups (such as carboxyl) to control drug release behaviors [27]. However, CS-only hydrogels have brittle properties and poor mechanical strength and show low solubility at a physiological pH of 7.4, which limits their application [28]. Researchers have reported that drug-loaded chitosan gels and CS gels with the addition of synthetic materials have better mechanical and biomedical properties than

chitosan-only gels [26,29]. Zhang, Y. et al. [30] constructed a poly(ethylene glycol) (PEG) and CS composite gel by utilizing PEG to enhance the high mechanical strength of the hydrogel and encapsulate acetylsalicylic acid (ASA) via electrostatic interactions. The composite gel's sustained release of ASA for over two weeks promoted the proliferation and osteogenic differentiation of periodontal ligament stem cells (PDLSCs) and enhanced bone regeneration in a mouse calvarial bone defect model.

- Sodium alginate

Sodium alginate is a natural, high-productivity polyanionic polysaccharide derived primarily from brown seaweed and bacteria (nitrogen-fixing bacteria and *Pseudomonas*) [31]. Sodium alginate-based hydrogels have been extensively applied in wound healing [32], drug delivery [33], and cell transplantation [34] because of their high water content and favorable biocompatibility. Nevertheless, sodium alginate-based hydrogels have several deficiencies as they are difficult to degrade under physiological conditions in mammals because of the lack of enzymes to degrade, which is undesirable for periodontal regeneration. Recent studies reported that oxidation of alginate by periodate could increase its biodegradation rate and improve its biological safety in the long term [35]. Furthermore, cells prefer to adhere to neutral or cationic interfaces. Sodium alginate-based hydrogels have poor cell adhesion in mammals as a result of the formation of a hydrated surface layer that lacks substances useful for cell growth and adhesion [36]. To address these problems, researchers modified sodium alginate-based hydrogels by incorporating other components or improving crosslinking methods in recent studies. Xiong, X. et al. [37] developed negatively charged alginate and chondroitin sulfate microsphere hydrogels (nCACSMH) for cell delivery. The nCACSMH enhanced the attachment and proliferation of human umbilical vein endothelial cells (HUVECs) and upregulated angiogenesis-related gene expression in endothelial cells. Calcium ions, zinc ions, and other divalent metallic ions are capable of reacting with alginate to form hydrogels [33,38]. However, alginate-based hydrogels exhibit poor stability in the gel structure in the traditional physical crosslinking approach [39].

- Hyaluronic acid

Hyaluronic acid (HA) is a natural linear glycosaminoglycan abundantly found in the human body that consists of repeating units of N-acetyl-d-glucosamine and d-glucuronide; the highest concentrations of this compound are in the eyes and joints [40]. Hyaluronic acid (typically existing in the form of polymers weighing over 106 daltons) can be cleaved by hyaluronidase, and its biological function is related to its molecular weight [41]. Studies have reported that high-molecular-weight HA has anti-inflammatory and immunosuppressive effects, as well as promoting migration in gingival fibroblast cells, which may have beneficial effects on periodontal inflammation [42]. Furthermore, as with most natural polymer hydrogels, hyaluronic acid hydrogels have low cell recognition and adhesion rates due to a lack of cell adhesion sites and anionic properties [43]. To advance their application in periodontal tissue engineering, hyaluronic acid hydrogels must be compounded with other materials to enhance their mechanical properties and adhesion [31,44]. Ansari, S. et al. [31] designed a 3D alginate/HA hydrogel encapsulating PDLSCs that upregulated gene expression related to chondrogenesis (col II, aggrecan, and sox-9) in vitro and showed a greater positive expression of chondrogenic specific protein markers in vivo. To restore the anatomy and functionality of lost periodontal tissue, Babo, P. S. et al. [45] prepared an injectable hydrogel consisting of methacrylate hyaluronic acid (me-HA) and platelet lysate (PL) to release growth factor proteins in situ. Adding PL to me-HA hydrogels improved their viscoelastic properties ($p < 0.02$) and resilience to degradation by hyaluronidase while inhibiting bacterial growth. The versatile hydrogels provided adequate space and stability for cell adhesion and proliferation, showing great potential in periodontal therapy.

- Collagen

Collagen (Col) is an essential component of the extracellular matrix, with a variety of cell signaling binding sites and excellent biocompatibility, and is widely used in the development of periodontal tissue engineering scaffolds [18,46,47]. Collagen can support the adhesion, growth, proliferation, and directed differentiation of functional cells associated with periodontal regeneration [48]. However, collagen-based hydrogels have poor mechanical properties and stability, and terrestrial animal-derived collagen may cause immune reactions. Therefore, some researchers are exploring new collagen sources for biomaterials. Zhou, T. et al. [49] found that the comprehensive properties of blended hydrogels composed of polyvinyl alcohol (PVA) and fish Col can be regulated by controlling the content of PVA and Col to be suitable for guided tissue regeneration. As the proportion of PVA in the blended hydrogel increased, the mechanical properties of the hydrogel increased but were detrimental to human gingival fibroblast (HGF) growth. In contrast, an increase in collagen content enhanced the surface porosity of the hydrogels and their biocompatibility with PDLCs. The PVA/Col (50:50) blended hydrogel exhibited the highest cell proliferation rate for HPDLCs with spread cell morphology.

- Others

Other natural polymers, such as gelatin [50,51], chondroitin sulfate [37], and silk proteins [52], are also used in tissue engineering. These have been shown to have excellent biocompatibility, degradability, and cytocompatibility.

2.1.2. Synthetic Polymers

Synthetic polymers are prepared through chemical reactions. Common synthetic compounds, such as polyethylene glycol (PEG) [53], PVA [49], and poly(lactic-co-glycolic acid) (PLGA) [54], are generally accessible and can be tailored to achieve excellent mechanical properties and hydrogel stability but lack inherent bioactivity. However, the biocompatibility and degradability of synthetic polymer-based hydrogels are not as good as those of natural polymer-based hydrogels [30]. The following section discusses several types of synthetic polymer-based hydrogels.

- PEG

PEG is a hydrophilic and biocompatible synthetic polyether that is a promising hydrophilic biomaterial for periodontal regeneration and is well known for its flexibility, biocompatibility, and hydrophilicity [13]. Since being approved by the FDA, PEG has been widely applied in biomedical research [55]. The well-defined chemistries of PEG enable the precise insertion of cell-responsive and bioactive components into a hydrogel [56,57]. To investigate the specific response of PDLCs to ECM biophysical and biochemical cues, Fraser, D. et al. [58] used PEG hydrogels with peptides to enable MMP-mediated matrix degradation and/or PDLC integrin-matrix binding to mimic the ECM in periodontitis. Additionally, Zhang, Y. et al. [59] fabricated a tetra-PEG hydrogel for the in situ encapsulation of aspirin, ensuring sustained release, anti-inflammatory, and osteoinductive properties to composite hydrogels. In vitro experiments indicated that aspirin-loaded tetra-PEG hydrogels facilitate the proliferation and osteogenic differentiation of human periodontal cells, and in vivo results demonstrated that hydrogels could remarkably promote bone regeneration.

- Gelatin methacryloyl (GelMA)

GelMA is a photosensitive hydrogel material with excellent biocompatibility and the capacity to enable cell encapsulation that is manufactured from methacrylic anhydride (MA) and gelatin (gelatin) [60]. Generally, GelMA hydrogels are cured by UV or visible light and have recently been developed to mimic the 3D cell microenvironment [61]. A study [17] demonstrated that GelMA hydrogels could provide a physical microenvironment for PDLCs to adhere and grow, ensuring, as closely as possible, no loss of PDLCs during the transplantation process and stable performance of their biological functions. Ma, Y. et al. [62] designed a composite gel composed of GelMA and poly (ethylene gly-

col) dimethacrylate (PEGDA) encapsulating PDLs using 3D printing technology. The physical and biological properties of the composite hydrogels differed by adjusting the ratio of GelMA and PEGDA. In this study, cell proliferation, spreading, and osteogenic differentiation increased as the GelMA volume ratio increased. An optimized composition (the 4/1 GelMA/PEGDA hydrogel) was selected to treat periodontal defects, and it showed a significant impact on promoting the regeneration of functional tissue.

- Others

In addition, other synthetic polymers also show significant potential in periodontal therapy. PVA is a water-soluble polymer hydrolyzed from polyvinyl acetate with good biocompatibility, a high modulus of elasticity, and easily adjustable physical properties. The incorporation of PVA and natural polymers significantly compensated for the poor mechanical properties of natural polymer-based hydrogels and remained favorable for cytocompatibility and bioactivity [63,64]. PLGA is a non-toxic, degradable polymeric organic compound made by the polymerization of lactic acid and hydroxyacetic acid which has high biocompatibility and capsule- and film-forming properties as a carrier for cell implantation [65].

2.2. The Multiple Components of Hydrogels in Periodontal Tissue Regeneration

Periodontal tissue has a complex structural and compositional composition, including soft and hard tissue. Moreover, periodontitis is a chronic inflammatory disease caused by bacterial infection with an intricate pathology: pathogenic microorganisms cause inflammation of the gingiva, which overactivated the immune response and results in tissue destruction [66]. As the disease progresses, the host and microorganisms release a variety of proteases and proinflammatory cytokines that stimulate bone resorption [67]. Multiple systems, signaling pathways, and molecules are involved in the regeneration of periodontal tissue, including the skeletal, immune, and circulatory systems [68]. Materials with single components and structured structures are generally difficult to regenerate effectively. To simultaneously fulfill the mechanical and cell-compatible requirements of biomedical applications, multiple components must be arranged chronologically when designing material systems. As needed, hydrogel systems can carry chemicals, growth factors, nanoparticles, exosomes, and stem cells in their polymeric structure, prevent their dissolution, and provide a slow and controlled release to achieve regenerative therapy.

- Antibacterial agents

Hydrogels with antibiotic loading may exhibit enhanced antibacterial properties. Perioline (Perio) is a generally recommended adjunct to scaling and root planning for adult periodontitis in clinical practice and is essentially a 2.1% minocycline gel. However, there are some adverse effects of commercial Perio, including photosensitivity and permanent discoloration of developing teeth [69]. Metronidazole (MTZ) can effectively kill anaerobic bacteria and inhibit bacterial growth to control inflammation. Since MTZ is water-soluble, topical application is easily diluted by saliva and gingival sulcus, resulting in inadequate release. Dong, Z et al. [70] prepared metronidazole microcapsules (CS@MTZ) using the polysaccharide coprecipitation method, while PVA@CS@MTZ hydrogels were prepared by substituting CS@MTZ microcapsules with ions. The safety, biocompatibility, and antibacterial effect of the PVA@CS@MTZ gel were demonstrated through in vitro and in vivo experiments, suggesting that it is a promising therapeutic agent for periodontitis.

- Cytokines

Cytokines are involved in regulating cell proliferation, differentiation, immune response, and intercellular interactions in periodontal hard tissue regeneration and mineralization [71]. Growth factors such as bone morphogenetic proteins (BMPs) [56] and vascular endothelial growth factor (VEGF) [72] have been demonstrated to induce matrix mineralization and participate in bone formation and bone reconstruction. The secretion of proinflammatory cytokines and macrophages polarized into the proinflammatory phenotype would have negative effects on the osteogenesis of MSCs for periodontal regeneration [68]. Some

cytokines could regulate the progression of inflammation and macrophage-polarization. For example, IL-4 and FGF-2 play significant roles in anti-inflammation by polarizing macrophages toward the anti-inflammation phenotype, mitigating the foreign body reaction of hydrogels, and inhibiting osteoclast function, resulting in acceleration of tissue healing and successful periodontal regeneration [50,73]. Researchers [74,75] found that increased gel stiffness supported mesenchymal stem cell proliferation and osteogenic differentiation, whereas stiff gel is more likely to polarize macrophages toward the pro-inflammatory M1 phenotype and increase inflammatory factor release, which is detrimental to bone formation. Based on this, He, X. T et al. [50] introduced interleukin (IL)-4 and stromal cell-derived factor (SDF)-1 α into transglutaminase crosslinked gelatins (TG-gels) to regulate macrophage polarization and promote endogenous stem cell recruitment. The results demonstrated that TG-gels containing both IL-4 and SDF-1 α could induce stem cell homing, modulate cell differentiation, and indeed induce the regrowth of periodontal tissue. SDF-1 α plays a pivotal role in BMSC transplantation therapy and tissue regeneration by recruiting circulating or residing stem cells to the injury site, regulating macrophage polarization, and facilitating osteogenesis in vivo [76]. Tan, J et al. [77] prepared a supramolecular NapFFY hydrogel that encapsulates both SDF-1 and BMP-2, and their study demonstrated that an SDF-1/BMP-2 hydrogel could promote periodontal bone regeneration.

- Mesenchymal stem cells and exosomes

MSC-based tissue engineering combined with injectable hydrogels has been extensively investigated for periodontal regeneration [62,78–80]. The common MSCs applied in periodontal engineering include BMSCs [76], stem cells from human exfoliated deciduous teeth (SHEDs) [78], dental follicle cells [79] and PDLCs [62]. PDLCs were demonstrated to possess multiple differentiation potentials and high proliferative ability and could be differentiated into progenitor cells making up cement, PDL, and alveolar bone, being considered the most suitable MSCs for periodontal regeneration [62].

More recently, cell-free tissue engineering has been significantly developed. Extracellular vesicles (EVs) are indispensable paracrine mediators and have a significant therapeutic effect on periodontal regeneration without any other toxic effect [32,81]. Studies suggest that exosomes play a significant role in tissue regeneration by regulating the immune microenvironment, promoting angiogenesis, balancing bone metabolism, and participating in mineralization [79]. However, it is difficult to achieve therapeutic concentrations of EVs alone for topical or systematic application. Hydrogels are suitable carriers for loading exosomes [32,81]. Huang, C. C et al. [82] developed a 3D encapsulating and tethering photo-crosslinked alginate hydrogel system to prolong EV delivery in vivo and maintain the structural and functional integrity of EVs simultaneously, which had a superior performance in bone regeneration.

- Inorganic nanoparticles

Inorganic nanoparticles have attracted considerable attention in recent years as they could be used both as carriers for delivering drugs and as medicine, showing great potential and safety in the field of medical application [83]. The structural chains of hydrogels contain a significant number of reactive groups that can bond with inorganic nanoparticles [84]. Furthermore, inorganic nanoparticles such as mesoporous silica could serve as carriers for loading various drugs and bioactive substances [78] and have multiple effects such as antibacterial and osteogenic abilities [85]. In light of these findings, combining inorganic nanoparticles with hydrogels could optimize the mechanical properties and biological function of hydrogel materials to achieve the objective of promoting tissue regeneration [86]. Zeolitic imidazolate framework-8 is a porous crystalline material self-assembled by zinc ions and 2-methylimidazole ligands, with a large specific surface area, high porosity, easy synthesis, and controllable dimensions, and it has been applied in the treatment of periodontitis and bone regeneration [87,88]. Liu, Y et al. [89] developed a nano-injectable photosensitive GelMA composite hydrogel loaded with ZIF-8 for the treatment of periodontitis. The ZIF-8/GelMA hydrogel could release Zn²⁺ continuously and had good

cytocompatibility. GelMA-Z effectively upregulated the expression of osteogenic genes and proteins, increased alkaline phosphatase activity, promoted extracellular matrix mineralization in rat bone mesenchymal stem cells, and showed significant antibacterial effects against *Porphyromonas gingivalis*. In vivo, GelMA-Z reduced bacterial load, decreased inflammation, and promoted alveolar bone regeneration in a rat model, thus comprising a promising therapy for periodontitis.

- Natural compounds

Currently, natural compounds derived from herbs have also received substantial attention in medicine. Various natural substances from herbs possess a range of properties, such as anti-inflammatory, antibacterial, antioxidant, and growth factor-promoting properties [26,84]. Puerarin (PUE), a natural flavonoid, exhibits anti-inflammatory, antibacterial, and antioxidant properties [90]. Ferulic acid (FA) is a phenolic compound with excellent antioxidant activity. Ou, Q et al. [91] incorporated PUE and FA into polydopamine (PDA) nanoparticles (NPs) to prepare polyethylene glycol diacrylate (PEG-DA) composite hydrogels, exhibiting excellent mechanical and antioxidant properties. Ginsenoside Rg1, a component derived from the natural extract of ginseng, enhances the proliferation and osteogenic differentiation of hPDLSCs, with a favorable anti-inflammatory ability [92]. However, ginsenoside Rg1 can be hydrolyzed in a short time by matrix metalloproteinase. Guo, H et al. [93] developed an injectable self-healing hydrogel that achieved more than 6 days of sustainable release of ginsenoside Rg1, which might better facilitate periodontal regeneration in periodontitis.

3. Strategies of Hydrogels in Periodontal Tissue Regeneration

Hydrogels provide a survival space for cells to exchange nutrients and gases, regulating cell morphology and function. While hydrogels have many advantages, due to their poor mechanical properties, adjustments to the hydrogel components, network structure, gelation process, and crosslinking are often needed to achieve hydrogels with appropriate mechanical strength to improve tissue regeneration [86,94]. There are two main methods of hydrogel preparation: chemical crosslinking and physical cross-linking. Physical crosslinking refers to connections through ionic interactions, electrostatic interactions, hydrophobic interactions, crystallization, and hydrogen bonding [70,95], while chemical crosslinking reactions include Michael's addition reaction, Schiff's base reaction, the Diels-Alder cycloaddition reaction, and free radical polymerization [35,96,97].

The effect of biomaterials on tissue regeneration is mainly enforced through the interaction of cells with the biomaterial surface. Integrins are heterodimeric receptors on cell membranes that are involved in the regulation of biological behaviors such as cell morphology, migration, proliferation, and differentiation by binding to adhesion proteins on the surface of biomaterials [98]. The chemical composition, mechanical properties, hydrophilicity, and morphology of biomaterials are key factors regulating the control of cellular behaviors by the corresponding materials [99]. As a result, designing and processing the material by selecting the appropriate build-up is crucial to promote periodontal tissue regeneration. On the other hand, conventional three-dimensional hydrogels maintain a fixed shape without actively adapting to the changes occurring within the healing tissue. This leads to the development of four-dimensional hydrogels whose geometry changes with time or external stimulations [100]. Later, we will review the main current strategies for constructing hydrogels in periodontal tissue engineering.

3.1. Biomimetic Hydrogel

Biomimetic materials are biological materials that mimic the composition or structure of natural tissues. The extracellular matrix (ECM) is a 3D meshwork consisting of macromolecular substances (e.g., polysaccharides and proteins) secreted by cells. In the physiological environment, cells exist in a complex microenvironment characterized by both intercellular interactions and heterogeneous ECM. Cues from the ECM could regulate cellular functions, such as proliferation, apoptosis, migration, and differentiation, and affect biosynthesis [101]. As shown in Figure 2A, researchers found that cells cultured on 2D surfaces are flat and have a forced apical-basal polarity, which is unnatural for most mesenchymal cells. However, when embedded in a 3D ECM, cells can regain their physiological form and function [102]. Additionally, mesenchymal cells tend to adhere, proliferate, and differentiate into specific phenotypes when exposed to a matrix of similar tissue-level elasticity [103]. With advances in periodontal tissue engineering and materials biology, scientists have gradually realized that the material does not simply provide a scaffold for cells and growth factors to attach to *in vivo*, but rather, the physicochemical properties of the material could affect the host response and thus tissue regeneration [8,99]. Mimicking the composition and structure of the ECM and providing appropriate biological stimulation to cells are the most attractive advantages of biomimetic hydrogels and show a favorable advantage in periodontal regeneration [104,105]. In light of these findings, utilizing ECM cues to control the activity of cells *in vitro/in vivo* and ultimately design biomaterials has excellent potential to enhance periodontal tissue regeneration [62].

On the other hand, periodontal tissue has a sophisticated composition and structure, in which periodontium, alveolar bone, and cementum form a functionally and structurally multi-layered integration known as the periodontal complex [12]. Once periodontitis progresses, the structural and functional integrity of the periodontal complex is disrupted. According to the characteristics of each hierarchical layer of the periodontal complex (cementum-periodontium-alveolar bone), researchers have designed hierarchical hydrogel scaffolds simulating the “sandwich” structure of the periodontal complex and combined hydrogels with specific drugs/bioactive factors to guide the directional differentiation of PDLSCs, leading to the ideal effect of periodontal tissue regeneration, especially in remodeling the periodontal ligament (PDL) [106,107]. Sowmya, S. et al. [108] developed a porous trilayered nanocomposite hydrogel scaffold similar to the structure of the cementum-periodontium-alveolar bone complex. The hierarchical hydrogel wrapped bioactive molecules in different layers of the scaffold illustrated in Figure 2B to induce tissue regeneration, and the sustained release of growth factors lasted up to 14 days. The cementum layer was composed of chitin-poly (lactic-co-glycolic acid) (PLGA), nanobioactive glass ceramic (nBGC) and cementum protein 1; the PDL layer was composed of chitin, PLGA and fibroblast growth factor 2; and the alveolar bone layer was composed of chitin, PLGA, nBGC and platelet-rich plasma derived growth factors. Both *in vivo* and *ex vivo* experiments also showed complete defect healing, favorable formation of new cementum, fibrous PDL, and alveolar bone with well-defined bony trabeculae, thus demonstrating great potential for periodontal complex regeneration.

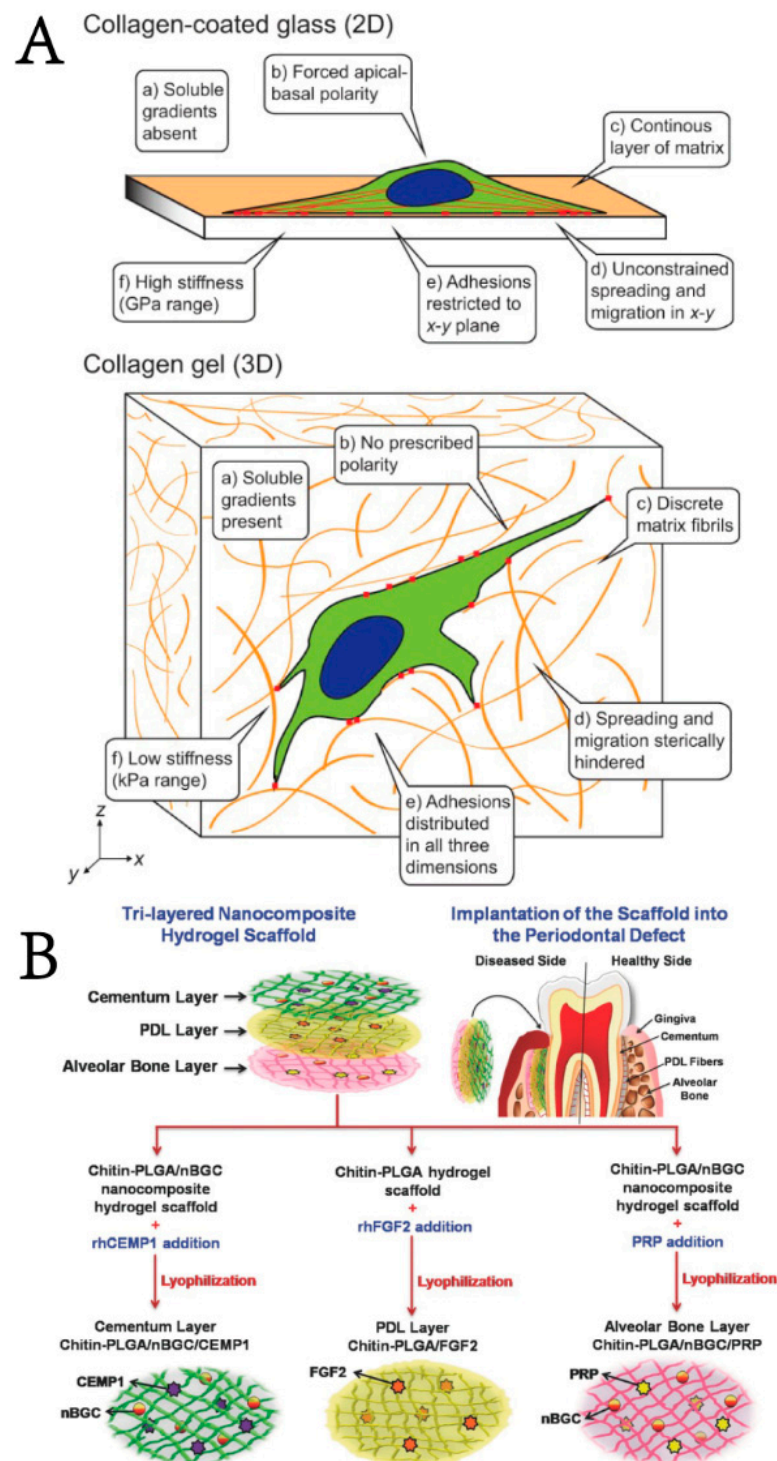


Figure 2. Schematic representation of (A) the different bioactive cues encountered by a cell between 2D substrates or in 3D microenvironments (Used with permission of Ref. [102], permission conveyed through Copyright Clearance Center, Inc, Danvers, MA, USA) and (B) a tri-layered nanocomposite hydrogel scaffold with a similar structure of cementum-periodontium-alveolar bone for simultaneous and complete periodontal regeneration (ab: alveolar bones; pl: periodontal ligaments; d: dentin; CEMP1: cementum protein-1; FGF-2: fibroblast growth factor-2; PRP: platelet-rich plasma; nBGC: nanobioactive glass ceramic; rhCEMP1: recombinant human cementum protein-1; rhFGF: re-combinant human fibroblast growth factor) (Adapted with permission from [108], Copyright 2017 WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim, Germany).

3.2. Intelligent Hydrogels

Intelligent hydrogels, also called stimulation-responsive hydrogels, can be responsive to slight alterations to specific external stimuli. Depending on the specific stimulus perceived, smart hydrogels can be further divided into thermosensitive hydrogels, pH-sensitive hydrogels, photosensitive hydrogels, and other stimulation-responsive hydrogels.

Thermosensitive hydrogels are one of the hydrogels that have received the utmost attention and can react accordingly to external temperature alterations, performing crucial roles in drug delivery [109], cell encapsulation [24], and tissue engineering [110]. The hydrogel exists in a liquid sol at room temperature or lower while transforming the state in situ to gelation when the temperature is higher than the critical dissolution temperature, such as the normal body temperature (37 °C) (Figure 3A) [80,111]. Xu, X et al. [112] prepared an injectable and thermosensitive CS, β -sodium glycerophosphate (β -GP), and gelatin hydrogel to achieve continuous release of aspirin and erythropoietin (EPO) to exert anti-inflammatory and tissue regeneration effects, respectively. In their study, CS/ β -GP/gelatin hydrogels loaded with aspirin/EPO showed efficacy in anti-inflammation and periodontium regeneration, remodeling the height of the alveolar bone, and providing a great alternative to periodontitis treatments.

As for pH-sensitive hydrogels, they typically have ionizable groups, such as carboxylic acid groups and basic primary amines [113]. The acid–base groups are subjected to variable degrees of ionization, resulting in sensitivity to pH and drug delivery and, ensuring the benefit of the natural control of the inflammatory processes when pH is decreased [114]. Bako, J et al. [115] developed a nanocomposite hydrogel as a pH-sensitive drug delivery system to release MTA and chlorhexidine. While MTA was released from the hydrogel within 12 h, chlorhexidine showed a much longer elution time with strong pH dependence, lasting over 7 days as demonstrated by the bactericidal effect, and it could reduce systemic side effects.

Photosensitive hydrogels induce solvation–gelation changes by exposure to long-range photo light. An important type of photosensitive, engineered gelatin-based material is GelMA, which is well suited for encapsulating PDLs and has excellent biocompatibility and tunable physical properties [62]. By ultraviolet (UV) irradiation, solutions form irreversibly covalently crosslinked hydrogels in the presence of photoinitiators. However, photo-crosslinking may have disadvantages due to limited UV light penetration and toxic initiators. UV exposure may cause cell/tissue damage, accelerated tissue aging, and even carcinogenesis, and human osteoblasts are less resistant to UV irradiation. In light of these harmful effects, visible light crosslinking has become a popular method for crosslinking hydrogels in recent years [115,116]. Goto, R. et al. [116] evaluated the in vitro feasibility of a visible-wavelength (VW)-light-crosslinked riboflavin (RF)-gelatin-based hydrogel in bone regeneration. The GelMA–RF hydrogel exhibited suitable stiffness for osteoblast differentiation and displayed significantly higher cell viability and gene expression related to osteoblast differentiation than hydrogels photopolymerized with UV light, which means visible-light-crosslinked hydrogels could also be used as a scaffold in bone tissue regeneration.

In addition to the above typical environment-sensitive hydrogels, researchers have also constructed other stimulation-responsive hydrogels or multi-sensitive hydrogels to combine the advantages of multiple single-stimulus-sensitive hydrogels simultaneously. Antimicrobial photodynamic therapy (PDT) is currently used as a novel treatment for periodontitis which generates reactive oxygen species (ROS) for a bactericidal effect [117]. Leung, B. et al. [118] demonstrated that the use of thermosensitive hydrogels containing methylene blue as a topical antimicrobial photodynamic therapy was a promising alternative to treat infectious wounds. Various studies have identified *Porphyromonas gingivalis* as the most pathogenically critical pathogen in developing periodontitis, and gingipain is a key virulence factor [119,120]. Liu, S. et al. [80] designed a gingipain-responsive thermosensitive hydrogel with sustainable release of SDF-1, which effectively controlled the inflammation caused by *P. gingivalis* and enhanced in situ periodontal bone regeneration in vivo (Figure 3B,C). Thus, multi-sensitive hydrogels combine the advantages of various stimulation-responsive hydrogels simultaneously and exhibit significant potential applications in periodontal tissue engineering.

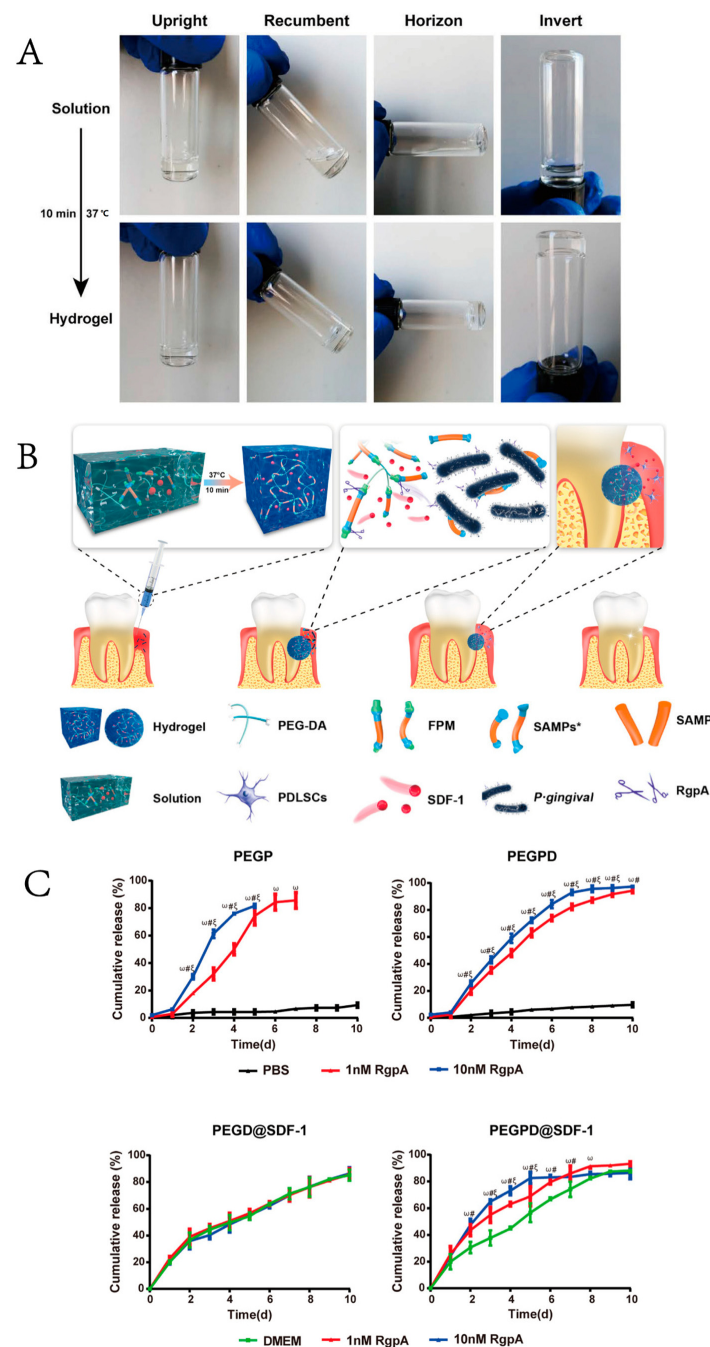


Figure 3. Schematic diagram of the gingipain-responsive thermosensitive hydrogel. (A) Photographs of the solution to hydrogel transition by placing the solution in a 37 °C incubator for 10 min. (B) Preparation and application of the gingipain-responsive thermosensitive hydrogel. (C) Release curves of SAMP (upper) and SDF-1 in different hydrogels. Data are presented as the mean \pm SD, $n = 3$. ω , $p < 0.05$ for the PBS/DMEM group vs. the 1 nM RgpA group. #, $p < 0.05$ for PBS/DMEM group vs. 10 nM RgpA group. ξ , $p < 0.05$ for the 1 nM RgpA group vs. the 10 nM RgpA group. (PEG: polyethylene glycol; DA: diacrylate; FPM: functional peptide module; SDF-1: stromal cell derived factor-1; SAMP: short antimicrobial peptide; RgpA: gingipain R1 protein. Adapted with permission from [80], Copyright 2021 American Chemical Society.

3.3. Self-Healing Hydrogels

Self-healing hydrogels refer to a group of hydrogels with the ability to spontaneously repair their structure and function after damage, inspired by the mechanism of self-healing

ability in biology [121]. The gelation mechanisms of self-healing hydrogels include dynamic covalent bonds [122], supramolecular bonds [123], and multi-mechanism cross-links.

Self-healing hydrogels have been widely used in wound healing and tissue engineering due to their good self-healing properties [124,125]. Lin, T. K et al. [113] synthesized self-healing hydrogels using Schiff base (also known as imine) linkages between difunctional polyurethane (DFPU) and CS. Depending on the properties of the Schiff bases, these hydrogels are sensitive to low pH and amine-containing molecules and have higher degradation rates in acidic microenvironments and internal porosities, which facilitate the release of drugs or substances (Figure 4). Guo, H et al. [93] reported a double-dynamic network polysaccharide-based hydrogel with rapid gelation, injectability, and excellent self-healing properties as a novel therapy for periodontitis. This hydrogel was synthesized by a dynamic Schiff base formation between $-CHO$ in aldehyde-modified HA and $-NH_2$ in glycol CS and a dynamic coordination bond between COO^- in aldehyde-modified HA and Fe^{3+} , which could transform the sol-gel without external stimuli. The CCK-8 assay showed that this self-healing hydrogel has no cytotoxicity. They further investigated the ability of this hydrogel-loaded with ginsenoside Rg1 and amelogenin to promote periodontal regeneration in periodontitis in vivo. Micro-CT, H&E staining, and immunohistochemical stainings analyses of IL-1, TNF- α , and TGF- β and TRAP indicated that the composite hydrogel could promote alveolar bone regeneration in periodontitis. These injectable self-healing hydrogels appear to have a desirable drug delivery ability and could recover hard tissue destruction in periodontitis.

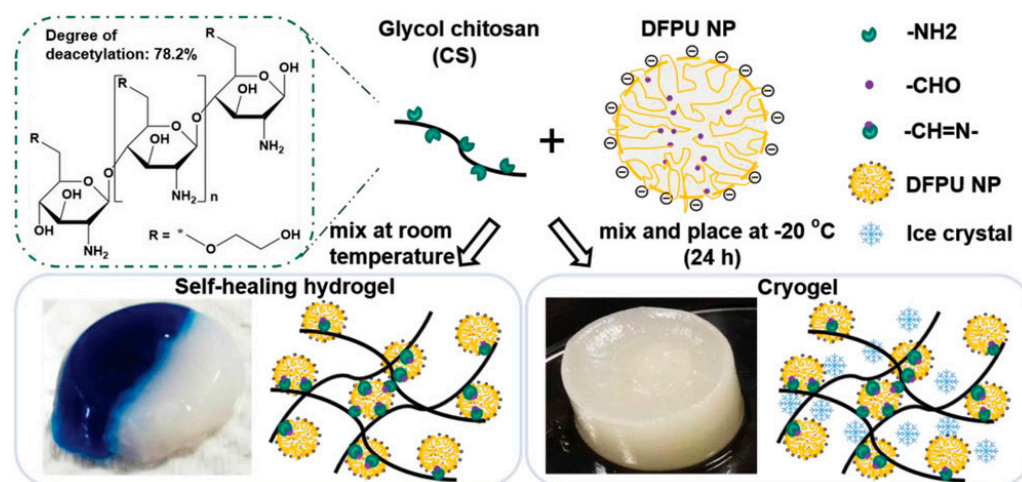


Figure 4. Schematic representation of the simple process to form a self-healing hydrogel or cryogel. (Adapted with permission from Reference [113]).

4. Summary and Challenges

This paper reviews the hydrogels applied for periodontal regeneration therapy in recent years, introducing the components and construction strategies for the preparation of relevant hydrogels. Hydrogels have excellent biocompatibility, water retention, and slow release and provide support for cellular interaction and biological function during periodontal regeneration in terms of promoting mesenchymal cell adhesion, migration, proliferation, and differentiation, reducing the inflammatory response, and regulating the immune environment to remodel the structure and function of periodontal tissues. Although remarkable progress has been made in the development of hydrogel therapy in periodontal regeneration, it remains a challenge to provide sufficient mechanical strength and more biological properties to the hydrogels to achieve ideal regenerative effects, something that needs to be taken into account in the future. The periodontal bone ligament-cementum combination is still the major concern of periodontal tissue engineering, especially the recovery of the periodontal ligament. Meanwhile, less research has been conducted on the regeneration of cementum. At present, there are few clinical studies and long-term

follow-up reports about the effectiveness of hydrogels for periodontal therapy. Further research is needed to address the relevant issues involved.

Author Contributions: Conceptualization, M.L.; methodology, M.L.; validation, Y.Y. and G.C.; writing—original draft preparation, M.L. and J.L.; writing—review and editing, C.L. and Y.D.; visualization, M.L. and J.L.; supervision, S.G., C.L. and Y.D.; project administration, Y.D.; funding acquisition, Y.D. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the National Natural Science Foundation, grant number 82071121, and the Technological Innovation Project from Chengdu Municipal Bureau of Science and Technology, grant number 2020YJ0242.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Slots, J. Periodontitis: Facts, fallacies and the future. *Periodontology 2000* **2017**, *75*, 7–23. [\[CrossRef\]](#) [\[PubMed\]](#)
2. Frencken, J.E.; Sharma, P.; Stenhouse, L.; Green, D.; Laverty, D.; Dietrich, T. Global epidemiology of dental caries and severe periodontitis—A comprehensive review. *J. Clin. Periodontol.* **2017**, *44* (Suppl. S18), S94–S105. [\[CrossRef\]](#) [\[PubMed\]](#)
3. Clark, D.; Kotronia, E.; Ramsay, S.E. Frailty, aging, and periodontal disease: Basic biologic considerations. *Periodontol 2000* **2021**, *87*, 143–156. [\[CrossRef\]](#)
4. Hajishengallis, G.; Chavakis, T. Local and systemic mechanisms linking periodontal disease and inflammatory comorbidities. *Nat. Rev. Immunol.* **2021**, *21*, 426–440. [\[CrossRef\]](#) [\[PubMed\]](#)
5. Chen, M.X.; Zhong, Y.J.; Dong, Q.Q.; Wong, H.M.; Wen, Y.F. Global, regional, and national burden of severe periodontitis, 1990–2019: An analysis of the Global Burden of Disease Study 2019. *J. Clin. Periodontol.* **2021**, *48*, 1165–1188. [\[CrossRef\]](#) [\[PubMed\]](#)
6. Batool, F.; Strub, M.; Petit, C.; Bugueno, I.M.; Bornert, F.; Clauss, F.; Huck, O.; Kuchler-Bopp, S.; Benkirane-Jessel, N. Periodontal Tissues, Maxillary Jaw Bone, and Tooth Regeneration Approaches: From Animal Models Analyses to Clinical Applications. *Nanomaterials* **2018**, *8*, 337. [\[CrossRef\]](#)
7. Aljateeli, M.; Koticha, T.; Bashutski, J.; Sugai, J.V.; Braun, T.M.; Giannobile, W.V.; Wang, H.L. Surgical periodontal therapy with and without initial scaling and root planing in the management of chronic periodontitis: A randomized clinical trial. *J. Clin. Periodontol.* **2014**, *41*, 693–700. [\[CrossRef\]](#)
8. Zhuang, Y.; Lin, K.; Yu, H. Advance of Nano-Composite Electrospun Fibers in Periodontal Regeneration. *Front. Chem.* **2019**, *7*, 495. [\[CrossRef\]](#)
9. Jepsen, S.; Gennai, S.; Hirschfeld, J.; Kalemaj, Z.; Buti, J.; Graziani, F. Regenerative surgical treatment of furcation defects: A systematic review and Bayesian network meta-analysis of randomized clinical trials. *J. Clin. Periodontol.* **2020**, *47*, 352–374. [\[CrossRef\]](#)
10. Chen, F.M.; Jin, Y. Periodontal tissue engineering and regeneration: Current approaches and expanding opportunities. *Tissue Eng. Part B Rev.* **2010**, *16*, 219–255. [\[CrossRef\]](#)
11. Chen, X.; Wu, G.; Feng, Z.; Dong, Y.; Zhou, W.; Li, B.; Bai, S.; Zhao, Y. Advanced biomaterials and their potential applications in the treatment of periodontal disease. *Crit. Rev. Biotechnol.* **2016**, *36*, 760–775. [\[CrossRef\]](#) [\[PubMed\]](#)
12. Liu, J.; Ruan, J.; Weir, M.D.; Ren, K.; Schneider, A.; Wang, P.; Oates, T.W.; Chang, X.; Xu, H.H.K. Periodontal Bone-Ligament-Cementum Regeneration via Scaffolds and Stem Cells. *Cells* **2019**, *8*, 537. [\[CrossRef\]](#) [\[PubMed\]](#)
13. Zhan, H.; Löwik, D.W. A hybrid peptide amphiphile fiber PEG hydrogel matrix for 3D cell culture. *Adv. Funct. Mater.* **2019**, *29*, 1808505. [\[CrossRef\]](#)
14. Abboud, A.R.; Ali, A.M.; Youssef, T. Preparation and characterization of insulin-loaded injectable hydrogels as potential adjunctive periodontal treatment. *Dent. Med. Probl.* **2020**, *57*, 377–384. [\[CrossRef\]](#)
15. Yuan, W.; Wang, H.; Fang, C.; Yang, Y.; Xia, X.; Yang, B.; Lin, Y.; Li, G.; Bian, L. Microscopic local stiffening in a supramolecular hydrogel network expedites stem cell mechanosensing in 3D and bone regeneration. *Mater. Horiz.* **2021**, *8*, 1722–1734. [\[CrossRef\]](#)
16. Zang, S.; Mu, R.; Chen, F.; Wei, X.; Zhu, L.; Han, B.; Yu, H.; Bi, B.; Chen, B.; Wang, Q.; et al. Injectable chitosan/ β -glycerophosphate hydrogels with sustained release of BMP-7 and ornidazole in periodontal wound healing of class III furcation defects. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2019**, *99*, 919–928. [\[CrossRef\]](#)
17. Pan, J.; Deng, J.; Yu, L.; Wang, Y.; Zhang, W.; Han, X.; Camargo, P.H.C.; Wang, J.; Liu, Y. Investigating the repair of alveolar bone defects by gelatin methacrylate hydrogels-encapsulated human periodontal ligament stem cells. *J. Mater. Sci. Mater. Med.* **2019**, *31*, 3. [\[CrossRef\]](#)

18. Momose, T.; Miyaji, H.; Kato, A.; Ogawa, K.; Yoshida, T.; Nishida, E.; Murakami, S.; Kosen, Y.; Sugaya, T.; Kawanami, M. Collagen Hydrogel Scaffold and Fibroblast Growth Factor-2 Accelerate Periodontal Healing of Class II Furcation Defects in Dog. *Open Dent. J.* **2016**, *10*, 347–359. [\[CrossRef\]](#)
19. Kreller, T.; Distler, T.; Heid, S.; Gerth, S.; Detsch, R.; Boccaccini, A.R. Physico-chemical modification of gelatine for the improvement of 3D printability of oxidized alginate-gelatine hydrogels towards cartilage tissue engineering. *Mater. Des.* **2021**, *208*, 109877. [\[CrossRef\]](#)
20. Chen, H.; Wang, H.; Li, B.; Feng, B.; He, X.; Fu, W.; Yuan, H.; Xu, Z. Enhanced chondrogenic differentiation of human mesenchymal stem cells on citric acid-modified chitosan hydrogel for tracheal cartilage regeneration applications. *RSC Adv.* **2018**, *8*, 16910–16917. [\[CrossRef\]](#)
21. Meena, L.K.; Raval, P.; Kedaria, D.; Vasita, R. Study of locust bean gum reinforced cyst-chitosan and oxidized dextran based semi-IPN cryogel dressing for hemostatic application. *Bioact. Mater.* **2018**, *3*, 370–384. [\[CrossRef\]](#)
22. Lehr, C.-M.; Bouwstra, J.A.; Schacht, E.H.; Junginger, H.E. In vitro evaluation of mucoadhesive properties of chitosan and some other natural polymers. *Int. J. Pharm.* **1992**, *78*, 43–48. [\[CrossRef\]](#)
23. Sudarshan, N.R.; Hoover, D.G.; Knorr, D. Antibacterial action of chitosan. *Food Biotechnol.* **1992**, *6*, 257–272. [\[CrossRef\]](#)
24. Yan, X.Z.; van den Beucken, J.J.; Cai, X.; Yu, N.; Jansen, J.A.; Yang, F. Periodontal tissue regeneration using enzymatically solidified chitosan hydrogels with or without cell loading. *Tissue Eng. Part A* **2015**, *21*, 1066–1076. [\[CrossRef\]](#) [\[PubMed\]](#)
25. Ji, Q.X.; Chen, X.G.; Zhao, Q.S.; Liu, C.S.; Cheng, X.J.; Wang, L.C. Injectable thermosensitive hydrogel based on chitosan and quaternized chitosan and the biomedical properties. *J. Mater. Sci. Mater. Med.* **2009**, *20*, 1603–1610. [\[CrossRef\]](#)
26. Hoang Thi, T.T.; Trinh, B.D.T.; Le Thi, P.; Tran, D.L.; Park, K.D.; Nguyen, D.H. Self-antibacterial chitosan/Aloe barbadensis Miller hydrogels releasing nitrite for biomedical applications. *J. Ind. Eng. Chem.* **2021**, *103*, 175–186. [\[CrossRef\]](#)
27. Arpornmaeklong, P.; Sareethammanuwat, M.; Apinyaputham, K.; Boonyuen, S. Characteristics and biologic effects of thermosensitive quercetin-chitosan/collagen hydrogel on human periodontal ligament stem cells. *J. Biomed. Mater. Research. Part B Appl. Biomater.* **2021**, *109*, 1656–1670. [\[CrossRef\]](#)
28. Fakhri, E.; Eslami, H.; Maroufi, P.; Pakdel, F.; Taghizadeh, S.; Ganbarov, K.; Yousefi, M.; Tanomand, A.; Yousefi, B.; Mahmoudi, S.; et al. Chitosan biomaterials application in dentistry. *Int. J. Biol. Macromol.* **2020**, *162*, 956–974. [\[CrossRef\]](#)
29. Işılai Özdoğan, A.; Akca, G.; Şenel, S. Development and in vitro evaluation of chitosan based system for local delivery of atorvastatin for treatment of periodontitis. *Eur. J. Pharm. Sci.* **2018**, *124*, 208–216. [\[CrossRef\]](#)
30. Zhang, Y.; Dou, X.; Zhang, L.; Wang, H.; Zhang, T.; Bai, R.; Sun, Q.; Wang, X.; Yu, T.; Wu, D.; et al. Facile fabrication of a biocompatible composite gel with sustained release of aspirin for bone regeneration. *Bioact. Mater.* **2022**, *11*, 130–139. [\[CrossRef\]](#)
31. Ansari, S.; Diniz, I.M.; Chen, C.; Aghaloo, T.; Wu, B.M.; Shi, S.; Moshaverinia, A. Alginate/hyaluronic acid hydrogel delivery system characteristics regulate the differentiation of periodontal ligament stem cells toward chondrogenic lineage. *J. Mater. Sci. Mater. Med.* **2017**, *28*, 162. [\[CrossRef\]](#) [\[PubMed\]](#)
32. Shafei, S.; Khanmohammadi, M.; Heidari, R.; Ghanbari, H.; Taghdiri Nooshabadi, V.; Farzamfar, S.; Akbariqomi, M.; Sanikhani, N.S.; Absalan, M.; Tavoosidana, G. Exosome loaded alginate hydrogel promotes tissue regeneration in full-thickness skin wounds: An in vivo study. *J. Biomed. Mater. Res. Part A* **2020**, *108*, 545–556. [\[CrossRef\]](#) [\[PubMed\]](#)
33. Chen, L.; Shen, R.; Komasa, S.; Xue, Y.; Jin, B.; Hou, Y.; Okazaki, J.; Gao, J. Drug-Loadable Calcium Alginate Hydrogel System for Use in Oral Bone Tissue Repair. *Int. J. Mol. Sci.* **2017**, *18*, 989. [\[CrossRef\]](#) [\[PubMed\]](#)
34. Diniz, I.M.; Chen, C.; Ansari, S.; Zadeh, H.H.; Moshaverinia, M.; Chee, D.; Marques, M.M.; Shi, S.; Moshaverinia, A. Gingival Mesenchymal Stem Cell (GMSC) Delivery System Based on RGD-Coupled Alginate Hydrogel with Antimicrobial Properties: A Novel Treatment Modality for Peri-Implantitis. *J. Prosthodont. Off. J. Am. Coll. Prosthodont.* **2016**, *25*, 105–115. [\[CrossRef\]](#) [\[PubMed\]](#)
35. Liang, Y.; Liu, W.; Han, B.; Yang, C.; Ma, Q.; Song, F.; Bi, Q. An in situ formed biodegradable hydrogel for reconstruction of the corneal endothelium. *Colloids Surfaces. B Biointerfaces* **2011**, *82*, 1–7. [\[CrossRef\]](#)
36. Tan, F.; Xu, X.; Deng, T.; Yin, M.; Zhang, X.; Wang, J. Fabrication of positively charged poly(ethylene glycol)-diacrylate hydrogel as a bone tissue engineering scaffold. *Biomed. Mater.* **2012**, *7*, 055009. [\[CrossRef\]](#)
37. Xiong, X.; Xiao, W.; Zhou, S.; Cui, R.; Xu, H.H.K.; Qu, S. Enhanced proliferation and angiogenic phenotype of endothelial cells via negatively-charged alginate and chondroitin sulfate microsphere hydrogels. *Biomed. Mater.* **2021**, *16*, 025012. [\[CrossRef\]](#)
38. Iskandar, L.; Rojo, L.; Di Silvio, L.; Deb, S. The effect of chelation of sodium alginate with osteogenic ions, calcium, zinc, and strontium. *J. Biomater. Appl.* **2019**, *34*, 573–584. [\[CrossRef\]](#)
39. Lueckgen, A.; Garske, D.S.; Ellinghaus, A.; Desai, R.M.; Stafford, A.G.; Mooney, D.J.; Duda, G.N.; Cipitria, A. Hydrolytically-degradable click-crosslinked alginate hydrogels. *Biomaterials* **2018**, *181*, 189–198. [\[CrossRef\]](#)
40. Laurent, T.C.; Fraser, J.R. Hyaluronan. *FASEB J. Off. Publ. Fed. Am. Soc. Exp. Biol.* **1992**, *6*, 2397–2404.
41. Lam, J.; Truong, N.F.; Segura, T. Design of cell-matrix interactions in hyaluronic acid hydrogel scaffolds. *Acta Biomater.* **2014**, *10*, 1571–1580. [\[CrossRef\]](#) [\[PubMed\]](#)
42. Chen, M.; Li, L.; Wang, Z.; Li, P.; Feng, F.; Zheng, X. High molecular weight hyaluronic acid regulates P. gingivalis-induced inflammation and migration in human gingival fibroblasts via MAPK and NF- κ B signaling pathway. *Arch. Oral Biol.* **2019**, *98*, 75–80. [\[CrossRef\]](#) [\[PubMed\]](#)
43. Vasi, A.M.; Popa, M.I.; Butnaru, M.; Dodi, G.; Verestiuc, L. Chemical functionalization of hyaluronic acid for drug delivery applications. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2014**, *38*, 177–185. [\[CrossRef\]](#) [\[PubMed\]](#)

44. Miranda, D.G.; Malmonge, S.M.; Campos, D.M.; Attik, N.G.; Grosogeat, B.; Gritsch, K. A chitosan-hyaluronic acid hydrogel scaffold for periodontal tissue engineering. *J. Biomed. Mater. Res. Part B Appl. Biomater.* **2016**, *104*, 1691–1702. [\[CrossRef\]](#)
45. Babo, P.S.; Pires, R.L.; Santos, L.; Franco, A.; Rodrigues, F.; Leonor, I.; Reis, R.L.; Gomes, M.E. Platelet Lysate-Loaded Photocrosslinkable Hyaluronic Acid Hydrogels for Periodontal Endogenous Regenerative Technology. *ACS Biomater. Sci. Eng.* **2017**, *3*, 1359–1369. [\[CrossRef\]](#)
46. Kosen, Y.; Miyaji, H.; Kato, A.; Sugaya, T.; Kawanami, M. Application of collagen hydrogel/sponge scaffold facilitates periodontal wound healing in class II furcation defects in beagle dogs. *J. Periodontol. Res.* **2012**, *47*, 626–634. [\[CrossRef\]](#)
47. Guo, S.; He, L.; Yang, R.; Chen, B.; Xie, X.; Jiang, B.; Weidong, T.; Ding, Y. Enhanced effects of electrospun collagen-chitosan nanofiber membranes on guided bone regeneration. *J. Biomater. Sci. Polym. Ed.* **2020**, *31*, 155–168. [\[CrossRef\]](#)
48. Janjić, K.; Agis, H.; Moritz, A.; Rausch-Fan, X.; Andrukhov, O. Effects of collagen membranes and bone substitute differ in periodontal ligament cell microtissues and monolayers. *J. Periodontol.* **2022**, *93*, 697–708. [\[CrossRef\]](#)
49. Zhou, T.; Zheng, K.; Sui, B.; Boccaccini, A.R.; Sun, J. In vitro evaluation of poly (vinyl alcohol)/collagen blended hydrogels for regulating human periodontal ligament fibroblasts and gingival fibroblasts. *Int. J. Biol. Macromol.* **2020**, *163*, 1938–1946. [\[CrossRef\]](#)
50. He, X.T.; Li, X.; Xia, Y.; Yin, Y.; Wu, R.X.; Sun, H.H.; Chen, F.M. Building capacity for macrophage modulation and stem cell recruitment in high-stiffness hydrogels for complex periodontal regeneration: Experimental studies in vitro and in rats. *Acta Biomater.* **2019**, *88*, 162–180. [\[CrossRef\]](#)
51. Pańczyszyn, E.; Jaśko, M.; Miłek, O.; Niedziela, M.; Męcik-Kronenberg, T.; Hoang-Bujnowicz, A.; Zięba, M.; Adamus, G.; Kowalczyk, M.; Osyczka, A.M.; et al. Gellan gum hydrogels cross-linked with carbodiimide stimulates vacuolation of human tooth-derived stem cells in vitro. *Toxicol. Vitro. Int. J. Publ. Assoc. BIBRA* **2021**, *73*, 105111. [\[CrossRef\]](#)
52. Oliveira, I.M.; Goncalves, C.; Shin, M.E.; Lee, S.; Reis, R.L.; Khang, G.; Oliveira, J.M. Anti-Inflammatory Properties of Injectable Betamethasone-Loaded Tyramine-Modified Gellan Gum/Silk Fibroin Hydrogels. *Biomolecules* **2020**, *10*, 1456. [\[CrossRef\]](#)
53. Valderrama, P.; Jung, R.E.; Thoma, D.S.; Jones, A.A.; Cochran, D.L. Evaluation of parathyroid hormone bound to a synthetic matrix for guided bone regeneration around dental implants: A histomorphometric study in dogs. *J. Periodontol.* **2010**, *81*, 737–747. [\[CrossRef\]](#) [\[PubMed\]](#)
54. Makadia, H.K.; Siegel, S.J. Poly Lactic-co-Glycolic Acid (PLGA) as Biodegradable Controlled Drug Delivery Carrier. *Polymers* **2011**, *3*, 1377–1397. [\[CrossRef\]](#)
55. Chin, S.Y.; Poh, Y.C.; Kohler, A.C.; Sia, S.K. An Additive Manufacturing Technique for the Facile and Rapid Fabrication of Hydrogel-based Micromachines with Magnetically Responsive Components. *J. Vis. Exp. JoVE* **2018**, *137*, e56727. [\[CrossRef\]](#) [\[PubMed\]](#)
56. Cha, J.K.; Jung, U.W.; Thoma, D.S.; Hammerle, C.H.F.; Jung, R.E. Osteogenic efficacy of BMP-2 mixed with hydrogel and bone substitute in peri-implant dehiscence defects in dogs: 16 weeks of healing. *Clin. Oral Implant. Res.* **2018**, *29*, 300–308. [\[CrossRef\]](#)
57. Isaac, A.; Jivan, F.; Xin, S.; Hardin, J.; Luan, X.; Pandya, M.; Diekwisch, T.G.H.; Alge, D.L. Microporous Bio-orthogonally Annealed Particle Hydrogels for Tissue Engineering and Regenerative Medicine. *ACS Biomater. Sci. Eng.* **2019**, *5*, 6395–6404. [\[CrossRef\]](#) [\[PubMed\]](#)
58. Fraser, D.; Nguyen, T.; Benoit, D.S.W. Matrix Control of Periodontal Ligament Cell Activity Via Synthetic Hydrogel Scaffolds. *Tissue Eng. Part A* **2021**, *27*, 733–747. [\[CrossRef\]](#)
59. Zhang, Y.; Ding, N.; Zhang, T.; Sun, Q.; Han, B.; Yu, T. A Tetra-PEG Hydrogel Based Aspirin Sustained Release System Exerts Beneficial Effects on Periodontal Ligament Stem Cells Mediated Bone Regeneration. *Front. Chem.* **2019**, *7*, 682. [\[CrossRef\]](#)
60. Zhu, M.; Wang, Y.; Ferracci, G.; Zheng, J.; Cho, N.J.; Lee, B.H. Gelatin methacryloyl and its hydrogels with an exceptional degree of controllability and batch-to-batch consistency. *Sci. Rep.* **2019**, *9*, 6863. [\[CrossRef\]](#)
61. Chen, X.; Bai, S.; Li, B.; Liu, H.; Wu, G.; Liu, S.; Zhao, Y. Fabrication of gelatin methacrylate/nanohydroxyapatite microgel arrays for periodontal tissue regeneration. *Int. J. Nanomed.* **2016**, *11*, 4707–4718. [\[CrossRef\]](#)
62. Ma, Y.; Ji, Y.; Zhong, T.; Wan, W.; Yang, Q.; Li, A.; Zhang, X.; Lin, M. Bioprinting-Based PDLSC-ECM Screening for in Vivo Repair of Alveolar Bone Defect Using Cell-Laden, Injectable and Photocrosslinkable Hydrogels. *ACS Biomater. Sci. Eng.* **2017**, *3*, 3534–3545. [\[CrossRef\]](#) [\[PubMed\]](#)
63. Berger, J.; Reist, M.; Mayer, J.M.; Felt, O.; Gurny, R. Structure and interactions in chitosan hydrogels formed by complexation or aggregation for biomedical applications. *Eur. J. Pharm. Biopharm.* **2004**, *57*, 35–52. [\[CrossRef\]](#)
64. Mansur, H.S.; Costa, E.D.S.; Mansur, A.A.P.; Barbosa-Stancioli, E.F. Cytocompatibility evaluation in cell-culture systems of chemically crosslinked chitosan/PVA hydrogels. *Mater. Sci. Eng. C* **2009**, *29*, 1574–1583. [\[CrossRef\]](#)
65. Shen, S.; Zhang, Y.; Zhang, S.; Wang, B.; Shang, L.; Shao, J.; Lin, M.; Cui, Y.; Sun, S.; Ge, S. 6-Bromoindirubin-3'-oxime Promotes Osteogenic Differentiation of Periodontal Ligament Stem Cells and Facilitates Bone Regeneration in a Mouse Periodontitis Model. *ACS Biomater. Sci. Eng.* **2021**, *7*, 232–241. [\[CrossRef\]](#)
66. Kato, H.; Taguchi, Y.; Tominaga, K.; Umeda, M.; Tanaka, A. Porphyromonas gingivalis LPS inhibits osteoblastic differentiation and promotes pro-inflammatory cytokine production in human periodontal ligament stem cells. *Arch. Oral Biol.* **2014**, *59*, 167–175. [\[CrossRef\]](#)
67. Hienz, S.A.; Paliwal, S.; Ivanovski, S. Mechanisms of Bone Resorption in Periodontitis. *J. Immunol. Res.* **2015**, *2015*, 615486. [\[CrossRef\]](#)
68. Gruber, R. Osteoimmunology: Inflammatory osteolysis and regeneration of the alveolar bone. *J. Clin. Periodontol.* **2019**, *46* (Suppl. S21), 52–69. [\[CrossRef\]](#)

69. Mou, J.; Liu, Z.; Liu, J.; Lu, J.; Zhu, W.; Pei, D. Hydrogel containing minocycline and zinc oxide-loaded serum albumin nanoparticle for periodontitis application: Preparation, characterization and evaluation. *Drug Deliv.* **2019**, *26*, 179–187. [\[CrossRef\]](#)
70. Dong, Z.; Sun, Y.; Chen, Y.; Liu, Y.; Tang, C.; Qu, X. Injectable Adhesive Hydrogel through a Microcapsule Cross-Link for Periodontitis Treatment. *ACS Appl. Bio Mater.* **2019**, *2*, 5985–5994. [\[CrossRef\]](#)
71. Giannobile, W.V.; Berglundh, T.; Al-Nawas, B.; Araujo, M.; Bartold, P.M.; Bouchard, P.; Chapple, I.; Gruber, R.; Lundberg, P.; Sculean, A.; et al. Biological factors involved in alveolar bone regeneration: Consensus report of Working Group 1 of the 15(th) European Workshop on Periodontology on Bone Regeneration. *J. Clin. Periodontol.* **2019**, *46* (Suppl. S21), 6–11. [\[CrossRef\]](#) [\[PubMed\]](#)
72. Divband, B.; Aghazadeh, M.; Al-Qaim, Z.H.; Samiei, M.; Hussein, F.H.; Shaabani, A.; Shahi, S.; Sedghi, R. Bioactive chitosan biguanidine-based injectable hydrogels as a novel BMP-2 and VEGF carrier for osteogenesis of dental pulp stem cells. *Carbohydr. Polym.* **2021**, *273*, 118589. [\[CrossRef\]](#)
73. Fujihara, C.; Kanai, Y.; Masumoto, R.; Kitagaki, J.; Matsumoto, M.; Yamada, S.; Kajikawa, T.; Murakami, S. Fibroblast growth factor-2 inhibits CD40-mediated periodontal inflammation. *J. Cell. Physiol.* **2019**, *234*, 7149–7160. [\[CrossRef\]](#) [\[PubMed\]](#)
74. He, X.T.; Wu, R.X.; Xu, X.Y.; Wang, J.; Yin, Y.; Chen, F.M. Macrophage involvement affects matrix stiffness-related influences on cell osteogenesis under three-dimensional culture conditions. *Acta Biomater.* **2018**, *71*, 132–147. [\[CrossRef\]](#)
75. Hegedűs, O.; Juriga, D.; Sipos, E.; Voniatis, C.; Juhász, Á.; Idrissi, A.; Zrínyi, M.; Varga, G.; Jedlovsky-Hajdú, A.; Nagy, K.S. Free thiol groups on poly(aspartamide) based hydrogels facilitate tooth-derived progenitor cell proliferation and differentiation. *PLoS ONE* **2019**, *14*, e0226363. [\[CrossRef\]](#)
76. Liu, Q.; Wen, Y.; Qiu, J.; Zhang, Z.; Jin, Z.; Cao, M.; Jiao, Y.; Yang, H. Local SDF-1 α application enhances the therapeutic efficacy of BMSCs transplantation in osteoporotic bone healing. *Heliyon* **2020**, *6*, e04347. [\[CrossRef\]](#) [\[PubMed\]](#)
77. Tan, J.; Zhang, M.; Hai, Z.; Wu, C.; Lin, J.; Kuang, W.; Tang, H.; Huang, Y.; Chen, X.; Liang, G. Sustained Release of Two Bioactive Factors from Supramolecular Hydrogel Promotes Periodontal Bone Regeneration. *ACS Nano* **2019**, *13*, 5616–5622. [\[CrossRef\]](#)
78. Qu, L.; Dubey, N.; Ribeiro, J.S.; Bordini, E.A.F.; Ferreira, J.A.; Xu, J.; Castilho, R.M.; Bottino, M.C. Metformin-loaded nanospheres-laden photocrosslinkable gelatin hydrogel for bone tissue engineering. *J. Mech. Behav. Biomed. Mater.* **2021**, *116*, 104293. [\[CrossRef\]](#)
79. Shi, W.; Guo, S.; Liu, L.; Liu, Q.; Huo, F.; Ding, Y.; Tian, W. Small Extracellular Vesicles from Lipopolysaccharide-Preconditioned Dental Follicle Cells Promote Periodontal Regeneration in an Inflammatory Microenvironment. *ACS Biomater. Sci. Eng.* **2020**, *6*, 5797–5810. [\[CrossRef\]](#)
80. Liu, S.; Wang, Y.N.; Ma, B.; Shao, J.; Liu, H.; Ge, S. Gingipain-Responsive Thermosensitive Hydrogel Loaded with SDF-1 Facilitates In Situ Periodontal Tissue Regeneration. *ACS Appl. Mater. Interfaces* **2021**, *13*, 36880–36893. [\[CrossRef\]](#)
81. Liu, L.; Guo, S.; Shi, W.; Liu, Q.; Huo, F.; Wu, Y.; Tian, W. Bone Marrow Mesenchymal Stem Cell-Derived Small Extracellular Vesicles Promote Periodontal Regeneration. *Tissue Eng. Part A* **2021**, *27*, 962–976. [\[CrossRef\]](#) [\[PubMed\]](#)
82. Huang, C.C.; Kang, M.; Shirazi, S.; Lu, Y.; Cooper, L.F.; Gajendraredy, P.; Ravindran, S. 3D Encapsulation and tethering of functionally engineered extracellular vesicles to hydrogels. *Acta Biomater.* **2021**, *126*, 199–210. [\[CrossRef\]](#) [\[PubMed\]](#)
83. Razavi, M.; Khandan, A. 14—Safety, regulatory issues, long-term biotoxicity, and the processing environment. In *Nanobiomaterials Science, Development and Evaluation*; Razavi, M., Thakor, A., Eds.; Woodhead Publishing: Sawston, UK, 2017; pp. 261–279.
84. Cui, P.; Pan, P.; Qin, L.; Wang, X.; Chen, X.; Deng, Y.; Zhang, X. Nanoengineered hydrogels as 3D biomimetic extracellular matrix with injectable and sustained delivery capability for cartilage regeneration. *Bioact. Mater.* **2023**, *19*, 487–498. [\[CrossRef\]](#)
85. Farazin, A.; Aghadavoudi, F.; Motifard, M.; Saber-Samandari, S.; Khandan, A. Nanostructure, Molecular Dynamics Simulation and Mechanical Performance of PCL Membranes Reinforced with Antibacterial Nanoparticles. *J. Appl. Comput. Mech.* **2021**, *7*, 1907–1915. [\[CrossRef\]](#)
86. Xu, Y.; Zhao, S.; Weng, Z.; Zhang, W.; Wan, X.; Cui, T.; Ye, J.; Liao, L.; Wang, X. Jelly-Inspired Injectable Guided Tissue Regeneration Strategy with Shape Auto-Matched and Dual-Light-Defined Antibacterial/Osteogenic Pattern Switch Properties. *ACS Appl. Mater. Interfaces* **2020**, *12*, 54497–54506. [\[CrossRef\]](#) [\[PubMed\]](#)
87. Liu, Y.; Zhu, Z.; Pei, X.; Zhang, X.; Cheng, X.; Hu, S.; Gao, X.; Wang, J.; Chen, J.; Wan, Q. ZIF-8-Modified Multifunctional Bone-Adhesive Hydrogels Promoting Angiogenesis and Osteogenesis for Bone Regeneration. *ACS Appl. Mater. Interfaces* **2020**, *12*, 36978–36995. [\[CrossRef\]](#) [\[PubMed\]](#)
88. Mustfa, S.A.; Maurizi, E.; McGrath, J.; Chiappini, C. Nanomedicine Approaches to Negotiate Local Biobarriers for Topical Drug Delivery. *Adv. Ther.* **2020**, *4*, 2000160. [\[CrossRef\]](#)
89. Liu, Y.; Li, T.; Sun, M.; Cheng, Z.; Jia, W.; Jiao, K.; Wang, S.; Jiang, K.; Yang, Y.; Dai, Z.; et al. ZIF-8 modified multifunctional injectable photopolymerizable GelMA hydrogel for the treatment of periodontitis. *Acta Biomater.* **2022**, *146*, 37–48. [\[CrossRef\]](#)
90. Zhang, S.; Ou, Q.; Xin, P.; Yuan, Q.; Wang, Y.; Wu, J. Polydopamine/puerarin nanoparticle-incorporated hybrid hydrogels for enhanced wound healing. *Biomater. Sci.* **2019**, *7*, 4230–4236. [\[CrossRef\]](#)
91. Ou, Q.; Zhang, S.; Fu, C.; Yu, L.; Xin, P.; Gu, Z.; Cao, Z.; Wu, J.; Wang, Y. More natural more better: Triple natural anti-oxidant puerarin/ferulic acid/polydopamine incorporated hydrogel for wound healing. *J. Nanobiotechnol.* **2021**, *19*, 237. [\[CrossRef\]](#)
92. Yin, L.H.; Cheng, W.X.; Qin, Z.S.; Sun, K.M.; Zhong, M.; Wang, J.K.; Gao, W.Y.; Yu, Z.H. Effects of ginsenoside Rg-1 on the proliferation and osteogenic differentiation of human periodontal ligament stem cells. *Chin. J. Integr. Med.* **2015**, *21*, 676–681. [\[CrossRef\]](#) [\[PubMed\]](#)

93. Guo, H.; Huang, S.; Yang, X.; Wu, J.; Kirk, T.B.; Xu, J.; Xu, A.; Xue, W. Injectable and Self-Healing Hydrogels with Double-Dynamic Bond Tunable Mechanical, Gel-Sol Transition and Drug Delivery Properties for Promoting Periodontium Regeneration in Periodontitis. *ACS Appl. Mater. Interfaces* **2021**, *13*, 61638–61652. [[CrossRef](#)] [[PubMed](#)]
94. Alipour, M.; Ashrafihelan, J.; Salehi, R.; Aghazadeh, Z.; Rezabakhsh, A.; Hassanzadeh, A.; Firouzmandi, M.; Heidarzadeh, M.; Rahbarghazi, R.; Aghazadeh, M.; et al. In vivo evaluation of biocompatibility and immune modulation potential of poly(caprolactone)-poly(ethylene glycol)-poly(caprolactone)-gelatin hydrogels enriched with nano-hydroxyapatite in the model of mouse. *J. Biomater. Appl.* **2021**, *35*, 1253–1263. [[CrossRef](#)] [[PubMed](#)]
95. Xing, Q.; Yates, K.; Vogt, C.; Qian, Z.; Frost, M.C.; Zhao, F. Increasing mechanical strength of gelatin hydrogels by divalent metal ion removal. *Sci. Rep.* **2014**, *4*, 4706. [[CrossRef](#)]
96. Choi, Y.; Kim, H.J.; Min, K.S. Effects of proanthocyanidin, a crosslinking agent, on physical and biological properties of collagen hydrogel scaffold. *Restor. Dent. Endod.* **2016**, *41*, 296–303. [[CrossRef](#)]
97. Moshaverinia, A.; Xu, X.; Chen, C.; Akiyama, K.; Snead, M.L.; Shi, S. Dental mesenchymal stem cells encapsulated in an alginate hydrogel co-delivery microencapsulation system for cartilage regeneration. *Acta Biomater.* **2013**, *9*, 9343–9350. [[CrossRef](#)]
98. Madl, C.M.; Heilshorn, S.C. Engineering Hydrogel Microenvironments to Recapitulate the Stem Cell Niche. *Annu. Rev. Biomed. Eng.* **2018**, *20*, 21–47. [[CrossRef](#)]
99. Li, Y.; Xiao, Y.; Liu, C. The Horizon of Materiobiology: A Perspective on Material-Guided Cell Behaviors and Tissue Engineering. *Chem. Rev.* **2017**, *117*, 4376–4421. [[CrossRef](#)]
100. Champeau, M.; Heinze, D.A.; Viana, T.N.; de Souza, E.R.; Chinellato, A.C.; Titotto, S. 4D Printing of Hydrogels: A Review. *Adv. Funct. Mater.* **2020**, *30*, 1910606. [[CrossRef](#)]
101. Dangaria, S.J.; Ito, Y.; Walker, C.; Druzinsky, R.; Luan, X.; Diekwisch, T.G. Extracellular matrix-mediated differentiation of periodontal progenitor cells. *Differ. Res. Biol. Divers.* **2009**, *78*, 79–90. [[CrossRef](#)]
102. Baker, B.M.; Chen, C.S. Deconstructing the third dimension: How 3D culture microenvironments alter cellular cues. *J. Cell Sci.* **2012**, *125*, 3015–3024. [[CrossRef](#)]
103. Engler, A.J.; Sen, S.; Sweeney, H.L.; Discher, D.E. Matrix Elasticity Directs Stem Cell Lineage Specification. *Cell* **2006**, *126*, 677–689. [[CrossRef](#)] [[PubMed](#)]
104. Moshaverinia, A.; Chen, C.; Xu, X.; Akiyama, K.; Ansari, S.; Zadeh, H.H.; Shi, S. Bone regeneration potential of stem cells derived from periodontal ligament or gingival tissue sources encapsulated in RGD-modified alginate scaffold. *Tissue Eng. Part A* **2014**, *20*, 611–621. [[CrossRef](#)]
105. Fawzy El-Sayed, K.M.; Mekhemar, M.K.; Beck-Broichsitter, B.E.; Bähr, T.; Hegab, M.; Receveur, J.; Heneweer, C.; Becker, S.T.; Wiltfang, J.; Dörfer, C.E. Periodontal regeneration employing gingival margin-derived stem/progenitor cells in conjunction with IL-1ra-hydrogel synthetic extracellular matrix. *J. Clin. Periodontol.* **2015**, *42*, 448–457. [[CrossRef](#)] [[PubMed](#)]
106. Varoni, E.M.; Vijayakumar, S.; Canciani, E.; Cochis, A.; De Nardo, L.; Lodi, G.; Rimondini, L.; Cerruti, M. Chitosan-Based Trilayer Scaffold for Multitissue Periodontal Regeneration. *J. Dent. Res.* **2018**, *97*, 303–311. [[CrossRef](#)] [[PubMed](#)]
107. Shah, A.T.; Zahid, S.; Ikram, F.; Maqbool, M.; Chaudhry, A.A.; Rahim, M.I.; Schmidt, F.; Goerke, O.; Khan, A.S.; Rehman, I.U. Tri-layered functionally graded membrane for potential application in periodontal regeneration. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2019**, *103*, 109812. [[CrossRef](#)]
108. Sowmya, S.; Mony, U.; Jayachandran, P.; Reshma, S.; Kumar, R.A.; Arzate, H.; Nair, S.V.; Jayakumar, R. Tri-Layered Nanocomposite Hydrogel Scaffold for the Concurrent Regeneration of Cementum, Periodontal Ligament, and Alveolar Bone. *Adv. Healthc. Mater.* **2017**, *6*, 1601251. [[CrossRef](#)]
109. Petit, C.; Batool, F.; Stutz, C.; Anton, N.; Klymchenko, A.; Vandamme, T.; Benkirane-Jessel, N.; Huck, O. Development of a thermosensitive statin loaded chitosan-based hydrogel promoting bone healing. *Int. J. Pharm.* **2020**, *586*, 119534. [[CrossRef](#)]
110. Chien, K.H.; Chang, Y.L.; Wang, M.L.; Chuang, J.H.; Yang, Y.C.; Tai, M.C.; Wang, C.Y.; Liu, Y.Y.; Li, H.Y.; Chen, J.T.; et al. Promoting Induced Pluripotent Stem Cell-driven Biomineralization and Periodontal Regeneration in Rats with Maxillary-Molar Defects using Injectable BMP-6 Hydrogel. *Sci. Rep.* **2018**, *8*, 114. [[CrossRef](#)]
111. Li, D.D.; Pan, J.F.; Ji, Q.X.; Yu, X.B.; Liu, L.S.; Li, H.; Jiao, X.J.; Wang, L. Characterization and cytocompatibility of thermosensitive hydrogel embedded with chitosan nanoparticles for delivery of bone morphogenetic protein-2 plasmid DNA. *J. Mater. Sci. Mater. Med.* **2016**, *27*, 134. [[CrossRef](#)]
112. Xu, X.; Gu, Z.; Chen, X.; Shi, C.; Liu, C.; Liu, M.; Wang, L.; Sun, M.; Zhang, K.; Liu, Q.; et al. An injectable and thermosensitive hydrogel: Promoting periodontal regeneration by controlled-release of aspirin and erythropoietin. *Acta Biomater.* **2019**, *86*, 235–246. [[CrossRef](#)] [[PubMed](#)]
113. Lin, T.W.; Hsu, S.H. Self-Healing Hydrogels and Cryogels from Biodegradable Polyurethane Nanoparticle Crosslinked Chitosan. *Adv. Sci.* **2020**, *7*, 1901388. [[CrossRef](#)] [[PubMed](#)]
114. Farjadian, F.; Rezaeifard, S.; Naeimi, M.; Ghasemi, S.; Mohammadi-Samani, S.; Welland, M.E.; Tayebi, L. Temperature and pH-responsive nano-hydrogel drug delivery system based on lysine-modified poly (vinylcaprolactam). *Int. J. Nanomed.* **2019**, *14*, 6901–6915. [[CrossRef](#)]
115. Bako, J.; Toth, F.; Gall, J.; Kovacs, R.; Csík, A.; Varga, I.; Sculean, A.; Zelko, R.; Hegedus, C. Combined Release of Antiseptic and Antibiotic Drugs from Visible Light Polymerized Biodegradable Nanocomposite Hydrogels for Periodontitis Treatment. *Pharmaceutics* **2022**, *14*, 957. [[CrossRef](#)]

116. Goto, R.; Nishida, E.; Kobayashi, S.; Aino, M.; Ohno, T.; Iwamura, Y.; Kikuchi, T.; Hayashi, J.I.; Yamamoto, G.; Asakura, M.; et al. Gelatin Methacryloyl-Riboflavin (GelMA-RF) Hydrogels for Bone Regeneration. *Int. J. Mol. Sci.* **2021**, *22*, 1635. [[CrossRef](#)] [[PubMed](#)]
117. Chambrone, L.; Wang, H.L.; Romanos, G.E. Antimicrobial photodynamic therapy for the treatment of periodontitis and peri-implantitis: An American Academy of Periodontology best evidence review. *J. Periodontol.* **2018**, *89*, 783–803. [[CrossRef](#)]
118. Leung, B.; Dharmaratne, P.; Yan, W.; Chan, B.C.L.; Lau, C.B.S.; Fung, K.P.; Ip, M.; Leung, S.S.Y. Development of thermosensitive hydrogel containing methylene blue for topical antimicrobial photodynamic therapy. *J. Photochem. Photobiol. B Biol.* **2020**, *203*, 111776. [[CrossRef](#)]
119. Kariu, T.; Nakao, R.; Ikeda, T.; Nakashima, K.; Potempa, J.; Imamura, T. Inhibition of gingipains and Porphyromonas gingivalis growth and biofilm formation by prenyl flavonoids. *J. Periodontol. Res.* **2017**, *52*, 89–96. [[CrossRef](#)]
120. Xu, W.; Zhou, W.; Wang, H.; Liang, S. Roles of Porphyromonas gingivalis and its virulence factors in periodontitis. *Adv. Protein Chem. Struct. Biol.* **2020**, *120*, 45–84. [[CrossRef](#)]
121. Wei, Z.; Yang, J.H.; Zhou, J.; Xu, F.; Zrínyi, M.; Dussault, P.H.; Osada, Y.; Chen, Y.M. Self-healing gels based on constitutional dynamic chemistry and their potential applications. *Chem. Soc. Rev.* **2014**, *43*, 8114–8131. [[CrossRef](#)]
122. Shen, J.; Zhou, Z.; Chen, D.; Wang, Y.; He, Y.; Wang, D.; Qin, J. Poly(aspartic acid) based self-healing hydrogels with antibacterial and light-emitting properties for wound repair. *Colloids Surf. B Biointerfaces* **2021**, *200*, 111568. [[CrossRef](#)] [[PubMed](#)]
123. Yu, G.; Chen, X. Host-Guest Chemistry in Supramolecular Theranostics. *Theranostics* **2019**, *9*, 3041–3074. [[CrossRef](#)] [[PubMed](#)]
124. Geng, X.; Qi, Y.; Liu, X.; Shi, Y.; Li, H.; Zhao, L. A multifunctional antibacterial and self-healing hydrogel laden with bone marrow mesenchymal stem cell-derived exosomes for accelerating diabetic wound healing. *Biomater. Adv.* **2022**, *133*, 112613. [[CrossRef](#)]
125. Bai, S.; Zhang, M.; Huang, X.; Zhang, X.; Lu, C.; Song, J.; Yang, H. A bioinspired mineral-organic composite hydrogel as a self-healable and mechanically robust bone graft for promoting bone regeneration. *Chem. Eng. J.* **2021**, *413*, 127512. [[CrossRef](#)]