

Supplementary Material

Crosstalk between yeast cell plasma membrane ergosterol content and cell wall stiffness under acetic acid stress involving

Pdr18

Ricardo A Ribeiro, Cláudia P. Godinho, Miguel V. Vitorino,
Tiago T. Robalo, Fábio Fernandes, Mário S. Rodrigues,
Isabel Sá-Correia*

***Corresponding author:** isacorreia@tecnico.ulisboa.pt

Figure S1 – Calibration curves of SNARF-4F-5-(and-6)-carboxylic acid, acetoxymethyl ester, acetate. The calibration curves for parental and *pdr18Δ* populations convert the fluorescence ratio to pHi values. They were obtained by plotting the fluorescence ratio of the different samples as a function of the pH of the buffer, with a defined pH, in which they were incubated. The equations obtained result from the fitting of a second-order polynomial function.

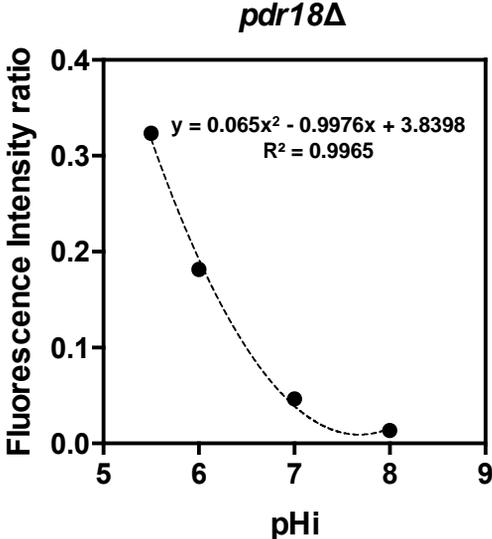
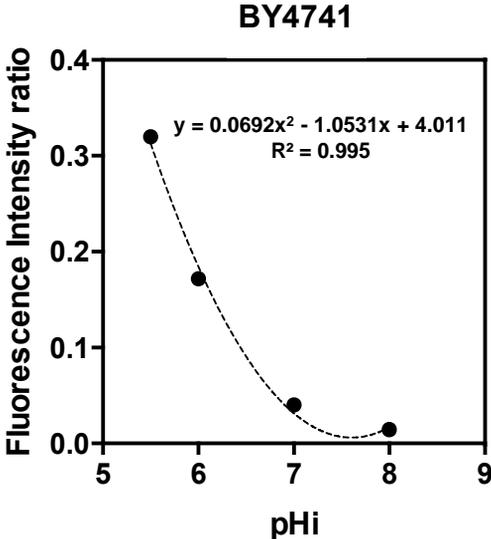


Figure S2 – Lyticase susceptibility assays using cells of the parental and *pdr18*Δ strains harvest during the growth curve in the absence or presence of acetic acid. The decrease of the OD_{600nm} during the incubation time of yeast cell suspensions (in %) following the addition of lyticase is plotted.

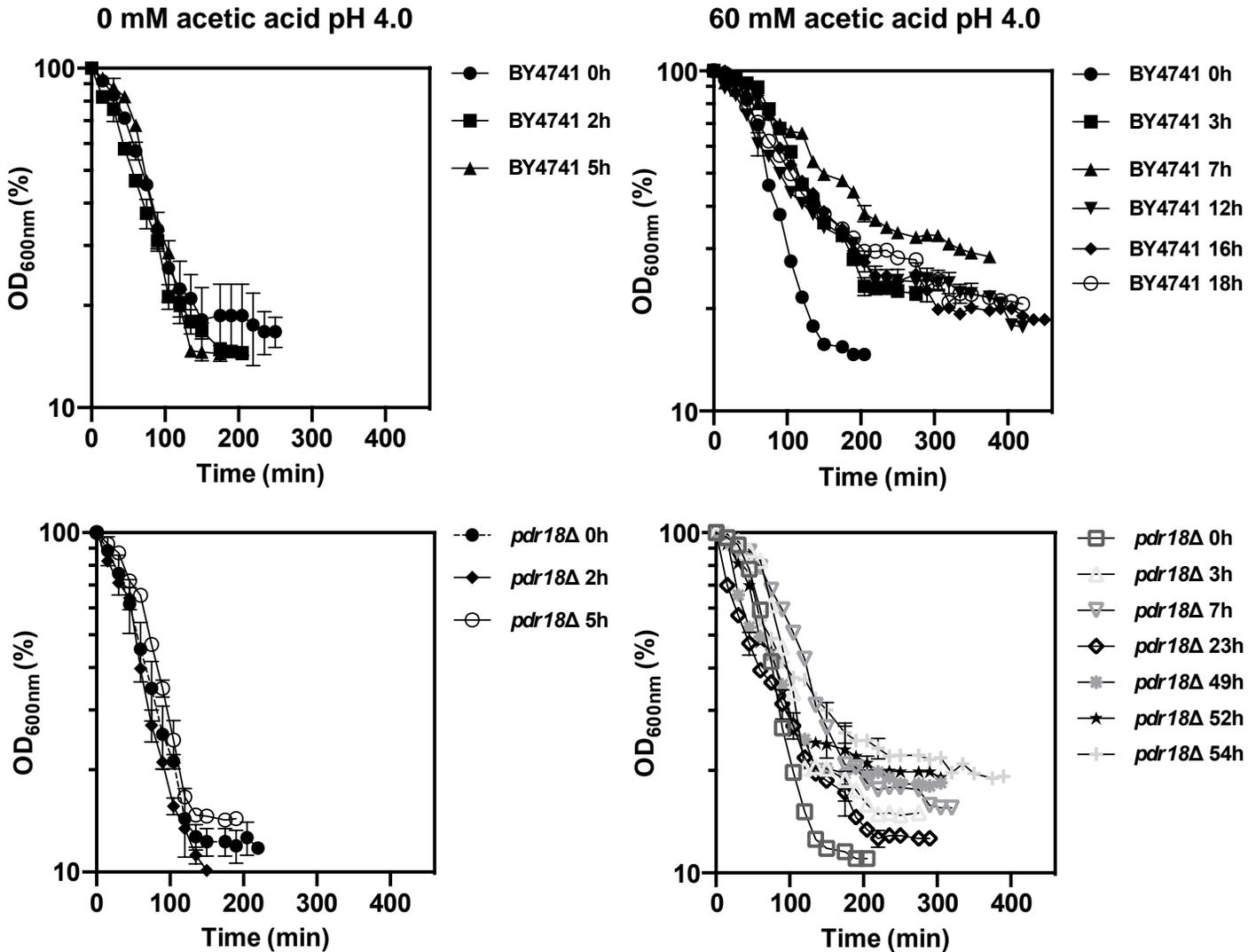
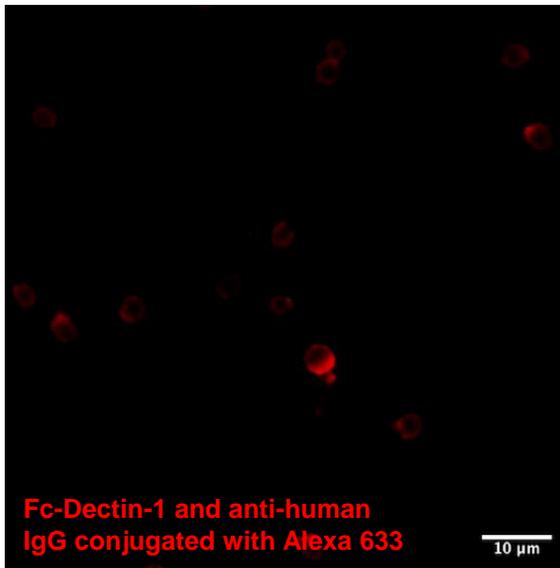


Figure S3 – Fluorescence microscopy images of fluorescence-stained cell wall β -glucans, chitin and mannans. Illustrative images of stained *pdr18* Δ cells were taken after 0 hours (A) and 23 hours (B) of exposure to acetic acid, this corresponding to the middle of the acetic acid-induced latency. Confocal microscopy was used to quantify β -glucans and mannans stained with Fc-Dectin 1-Alexa 633 and Concanavalin A-FITC, respectively, and 2-photon excitation microscopy was used to quantify chitin stained with Calcofluor White.

A



B

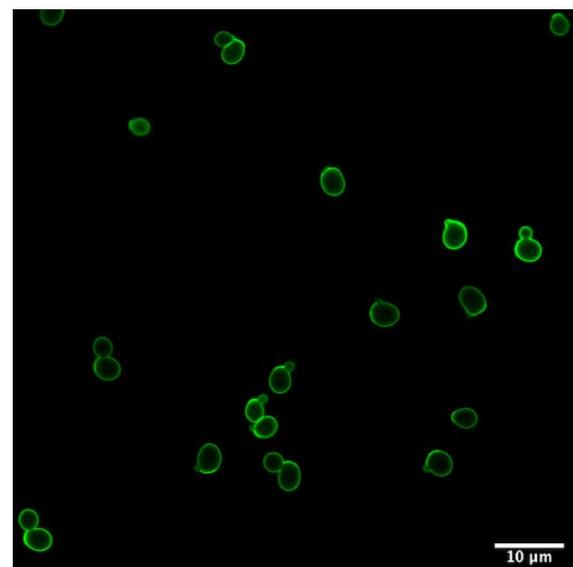
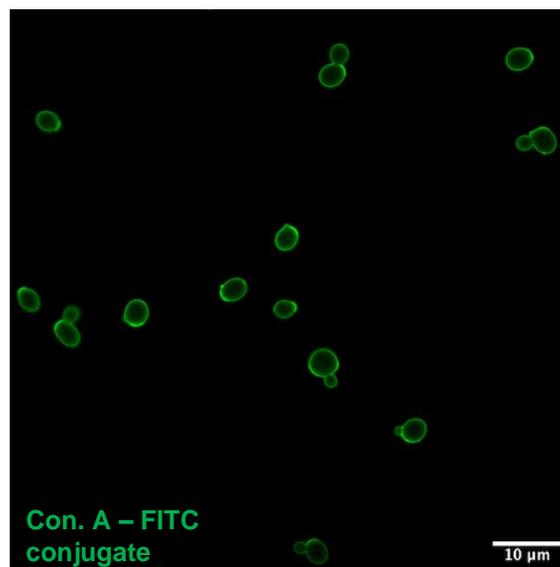
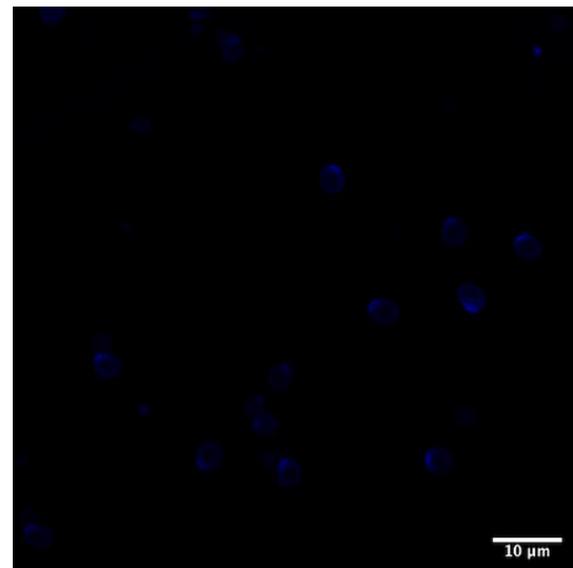
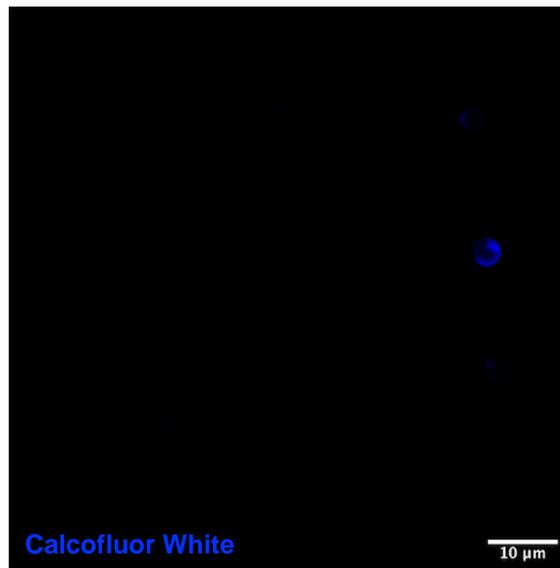
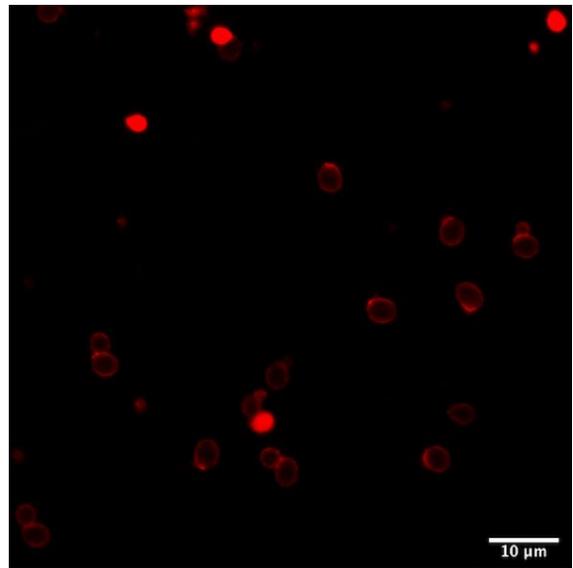


Table S1 - Primers used for qRT-PCR analysis.

Primer	Sequence (5'-3')
<i>ACT1</i>	fw: CTCCACCACTGCTGAAAGAGAA
	rev: CCAAGGCGACGTAACATAGTTTT
<i>CHS3</i>	fw: TCACCTGGATGTTTTACCATCAAG
	rev: CCACTCCGACGAGTTGCAT
<i>FKS1</i>	fw: CATGCTGCTCTGGTCCCTTATT
	rev: CACCGTGGGCAATTCCA
<i>FKS2</i>	fw: GCTCATGTGTTGGAGCAGTT
	rev: CCAATGGCATTACGGAAAAGA
<i>RLM1</i>	fw: CTTTTTCTGCAACACAGCCATA
	rev: CGCCAGGAATATTCGATGGT
<i>GAS1</i>	fw: AACCGCTGCTGCTTTTTTTG
	rev: CTTCAATCGCTGGAACATCGT
<i>CRH1</i>	fw: CGCGGCTGCCGAAAG
	rev: GCA GTGCTAGAAGCTGCAGTTG
<i>BGL2</i>	fw: TTTTGTTATGGCTAACGCGTTCT
	rev: GAGTAAGAGGCATTTTGCATGGT
<i>PRM5</i>	fw: TTTTCCACACAACATACCCAGTTT
	rev: TCTTTGGCGGGATAATCCATA