

## **Supplementary Material**

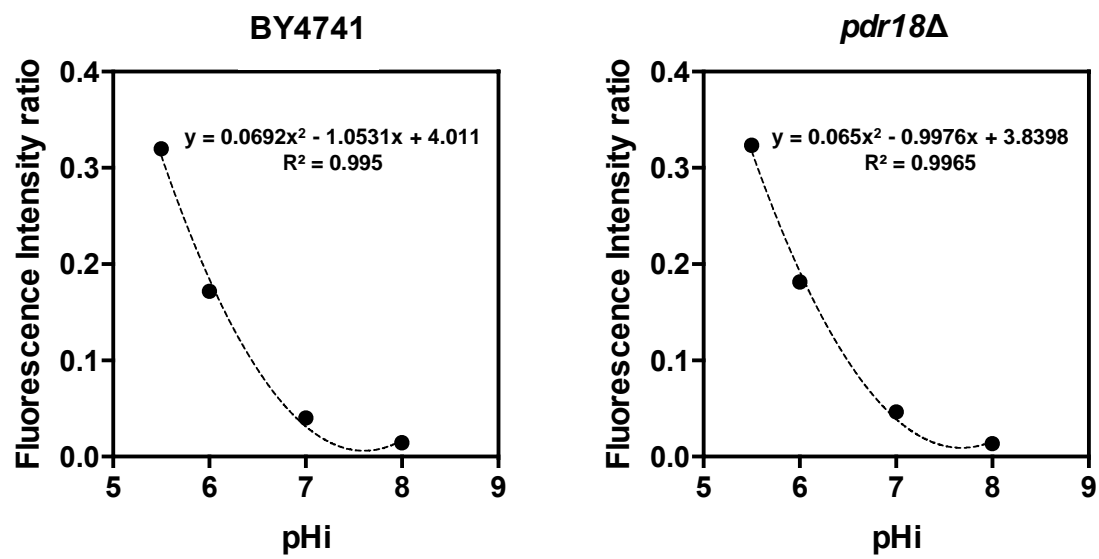
**Crosstalk between yeast cell plasma membrane ergosterol content and cell wall stiffness under acetic acid stress involving**

**Pdr18**

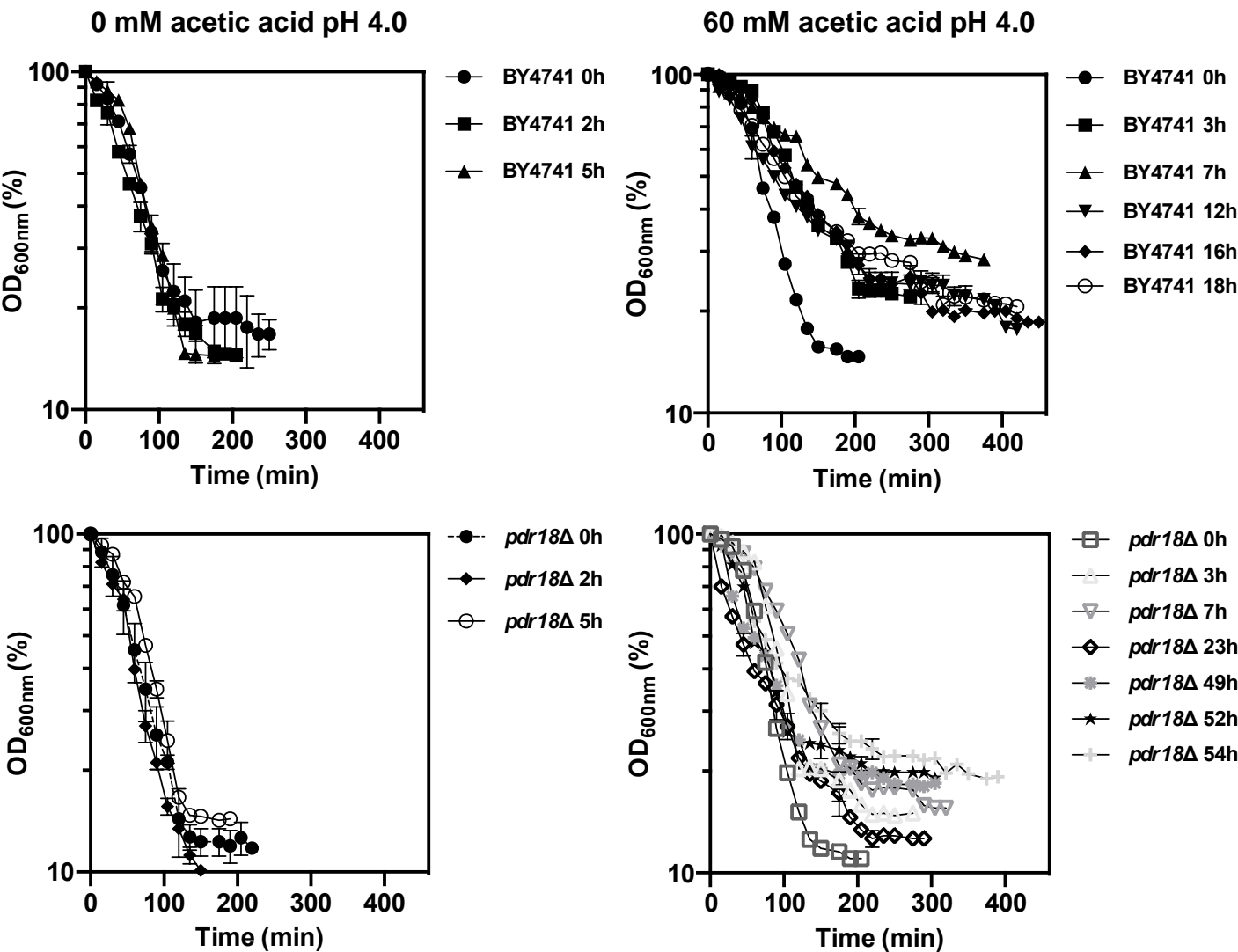
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**Figure S1** – Calibration curves of SNARF-4F-5-(and-6)-carboxylic acid, acetoxymethyl ester, acetate. The calibration curves for parental and *pdr18Δ* populations convert the fluorescence ratio to pHi values. They were obtained by plotting the fluorescence ratio of the different samples as a function of the pH of the buffer, with a defined pHi, in which they were incubated. The equations obtained result from the fitting of a second-order polynomial function.

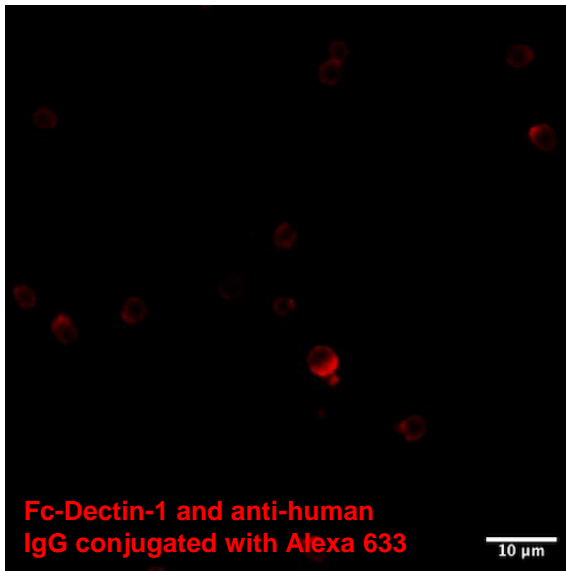


**Figure S2** – Lyticase susceptibility assays using cells of the parental and *pdr18Δ* strains harvest during the growth curve in the absence or presence of acetic acid. The decrease of the OD<sub>600nm</sub> during the incubation time of yeast cell suspensions (in %) following the addition of lyticase is plotted.

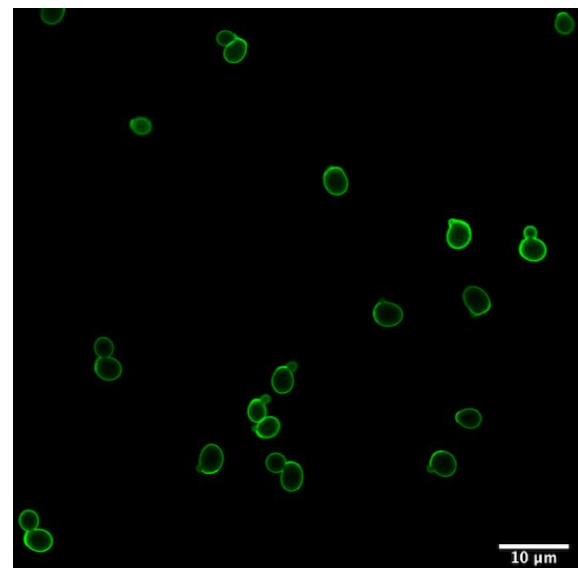
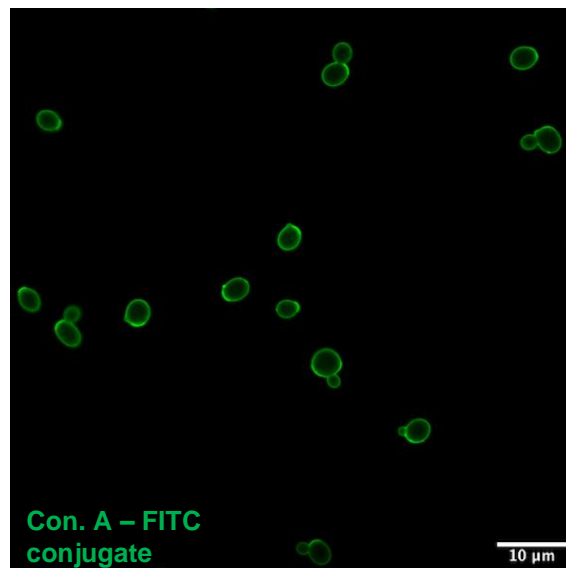
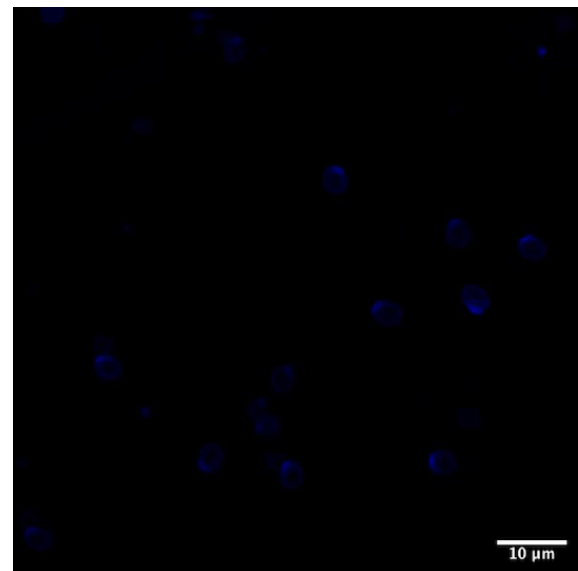
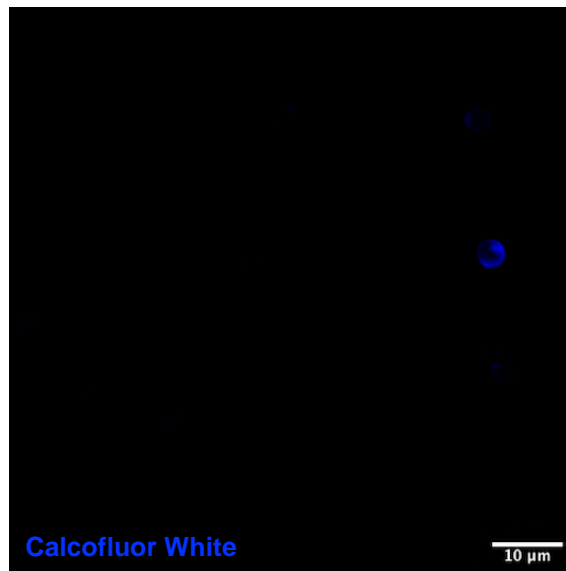
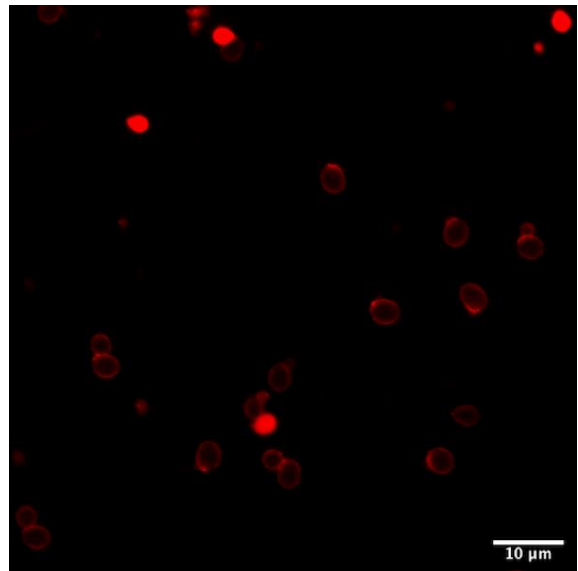


**Figure S3** – Fluorescence microscopy images of fluorescence-stained cell wall  $\beta$ -glucans, chitin and mannans. Illustrative images of stained *pdr18* $\Delta$  cells were taken after 0 hours (A) and 23 hours (B) of exposure to acetic acid, this corresponding to the middle of the acetic acid-induced latency. Confocal microscopy was used to quantify  $\beta$ -glucans and mannans stained with Fc-Dectin 1-Alexa 633 and Concanavalin A-FITC, respectively, and 2-photon excitation microscopy was used to quantify chitin stained with Calcofluor White.

**A**



**B**



**Table S1** - Primers used for qRT-PCR analysis.

Primer	Sequence (5'-3')
<i>ACT1</i>	fw: CTCCACCACTGCTGAAAGAGAA
	rev: CCAAGGCGACGTAACATAGTTTT
<i>CHS3</i>	fw: TCACCTGGATGTTTTACCATCAAG
	rev: CCACTCCGACGAGTTGCAT
<i>FKS1</i>	fw: CATGCTGCTCTGGTCCCTTATT
	rev: CACCGTGGGCAATTCCA
<i>FKS2</i>	fw: GCTCATGTCGTTGGAGCAGTT
	rev: CCAATGGCATTACGGAAAAGA
<i>RLM1</i>	fw: CTTTTTCTGCAACACAGCCATA
	rev: CGCCAGGAATATTCGATGGT
<i>GAS1</i>	fw: AACCGCTGCTGCTTTTTTTTG
	rev: CTTCAATCGCTGGAACATCGT
<i>CRH1</i>	fw: CGCGGCTGCCGAAAG
	rev: GCA GTGCTAGAAGCTGCAGTTG
<i>BGL2</i>	fw: TTTTGTATGGCTAACGCGTTCT
	rev: GAGTAAGAGGCATTTTGCATGGT
<i>PRM5</i>	fw: TTTTCCACACAACATACCCAGTTT
	rev: TCTTTGGCGGGATAATCCATA