

Table S1. Initial sugar concentrations of fermentation broths (immediately after inoculation) for each experiment. Note: Broths contain grape must (carbon source), yeast extract or ammonium sulphate (nitrogen source) and microorganisms.

Experiment	Glucose (g/L)	Fructose (g/L)	Total Sugars (g/L)
Strain comparison (in flasks)	118.95	121.20	240.15
Adjustment of yeast extract dose (in flasks)	119.81	113.99	233.8
Combinations of yeast extract and ammonium sulphate (in flasks)	116.94	126.33	243.27
Free-cell fermentation (bioreactor), $n = 4$	114.36 ± 2.59	122.02 ± 4.83	236.38 ± 7.12
Immobilized fermentation (bioreactor), $n = 4$	114.39 ± 7.27	119.34 ± 10.85	233.73 ± 18.10

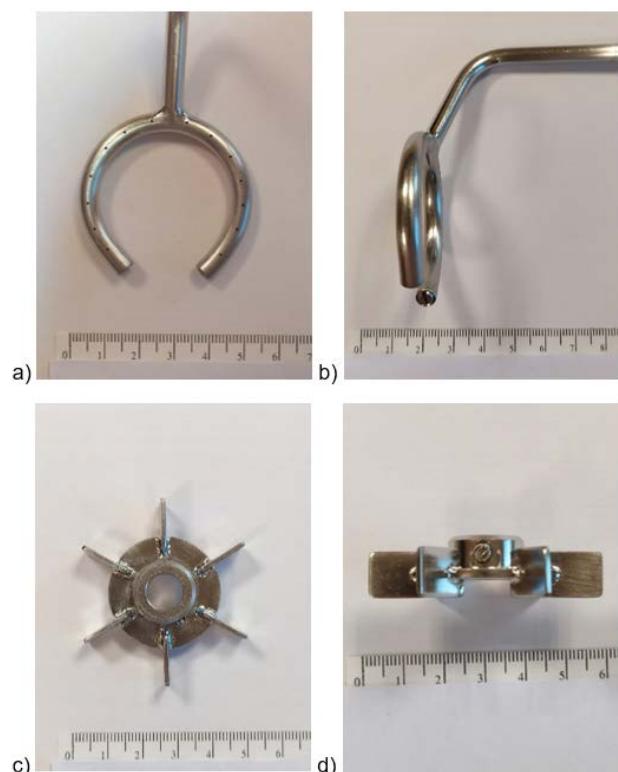


Figure S1. Photographs of (a, b) the sparger and (c, d) the rotor blades of the bioreactor. Note: The ruler shows centimeters.

Table S2. Fermentation parameters of grape must with five different fungal strains in flask experiments after 120 h (average \pm standard deviation; $n = 3$). Nitrogen source: 6.7 g/L yeast extract. Note: For each column, letters between parentheses (a, b, c, d) indicate the existence of statistical differences ($p < 0.05$; Tukey HSD test) among strains; if two strains share the same letter, there are no significant differences between them for that parameter.

Strain	$C_x \times 10^8$ cells/mL	C_{ETH} (g/L)	C_{ERY} (g/L)	C_{GLY} (g/L)	C_{MAN} (g/L)	ΔG (%)	ΔF (%)	ΔS (%)	Y_{ETH} (g/g)	Y_{ERY} (g/g)	Y_{GLY} (g/g)	Y_{MAN} (g/g)
<i>M. acetoabutens</i> DSM 3551	4.38 \pm 0.49 (ab)	2.74 \pm 1.15 (c)	7.85 \pm 0.54 (d)	8.77 \pm 1.55 (c)	0 \pm 0 (a)	100 \pm 0 (a)	32.66 \pm 1.91 (c)	166.02 \pm 0.96 (c)	1.45 \times 10 ⁻² \pm 5.92 \times 10 ⁻³ (c)	4.16 \times 10 ⁻² \pm 2.39 \times 10 ⁻³ (d)	4.65 \times 10 ⁻² \pm 8.47 \times 10 ⁻³ (c)	0 \pm 0 (a)
<i>M. madida</i> CBS 240.79	5.11 \pm 0.59 (b)	12.87 \pm 2.04 (b)	58.05 \pm 0.81 (b)	50.50 \pm 0.20 (b)	1.74 \pm 0.24 (b)	100 \pm 0 (a)	99.05 \pm 0.06 (a)	99.52 \pm 0.03 (a)	4.46 \times 10 ⁻² \pm 6.92 \times 10 ⁻³ (bd)	2.01 \times 10 ⁻¹ \pm 4.12 \times 10 ⁻³ (b)	1.75 \times 10 ⁻¹ \pm 1.99 \times 10 ⁻³ (b)	6.04 \times 10 ⁻³ \pm (b)
<i>M. megachi-liensis</i> CBS 567.85	7.63 \pm 0.96 (c)	8.54 \pm 1.24 (a)	81.10 \pm 1.92 (c)	10.09 \pm 0.91 (c)	0 \pm 0 (a)	100 \pm 0 (a)	90.87 \pm 0.60 (b)	95.39 \pm 0.30 (b)	3.16 \times 10 ⁻² \pm 4.53 \times 10 ⁻³ (ab)	3.00 \times 10 ⁻¹ \pm 5.89 \times 10 ⁻³ (c)	3.73 \times 10 ⁻² \pm 3.35 \times 10 ⁻³ (c)	0 \pm 0 (a)
<i>M. pollinis</i> MUCL 40570	2.80 \pm 0.11 (a)	7.48 \pm 1.20 (a)	100.79 \pm 3.35 (a)	2.44 \pm 0.78 (a)	0 \pm 0 (a)	100 \pm 0 (a)	97.04 \pm 0.62 (a)	98.51 \pm 0.31 (a)	2.66 \times 10 ⁻² \pm 3.96 \times 10 ⁻³ (ac)	3.59 \times 10 ⁻¹ \pm 7.50 \times 10 ⁻³ (a)	8.72 \times 10 ⁻³ \pm 2.88 \times 10 ⁻³ (a)	0 \pm 0 (a)
<i>M. suaveolens</i> var. <i>nigra</i> DSM 2552	3.58 \pm 0.63 (ab)	16.25 \pm 1.45 (b)	49.85 \pm 1.42 (e)	26.55 \pm 2.67 (d)	0.23 \pm 0.20 (a)	100 \pm 0 (a)	98.85 \pm 0.25 (a)	99.42 \pm 0.12 (a)	5.60 \times 10 ⁻² \pm 4.94 \times 10 ⁻³ (d)	1.72 \times 10 ⁻¹ \pm 5.02 \times 10 ⁻³ (e)	9.16 \times 10 ⁻² \pm 9.49 \times 10 ⁻³ (d)	7.83 \times 10 ⁻⁴ \pm (a)

C_x : cell density in the liquid medium; C_{ETH} : ethanol concentration; C_{ERY} : erythritol concentration; C_{GLY} : glycerol concentration; C_{MAN} : mannitol concentration; ΔG : glucose consumption, ΔF : fructose consumption, ΔS : total sugar consumption, Y_{ETH} : ethanol yield, Y_{ERY} : erythritol yield, Y_{GLY} : glycerol yield; Y_{MAN} : mannitol yield.

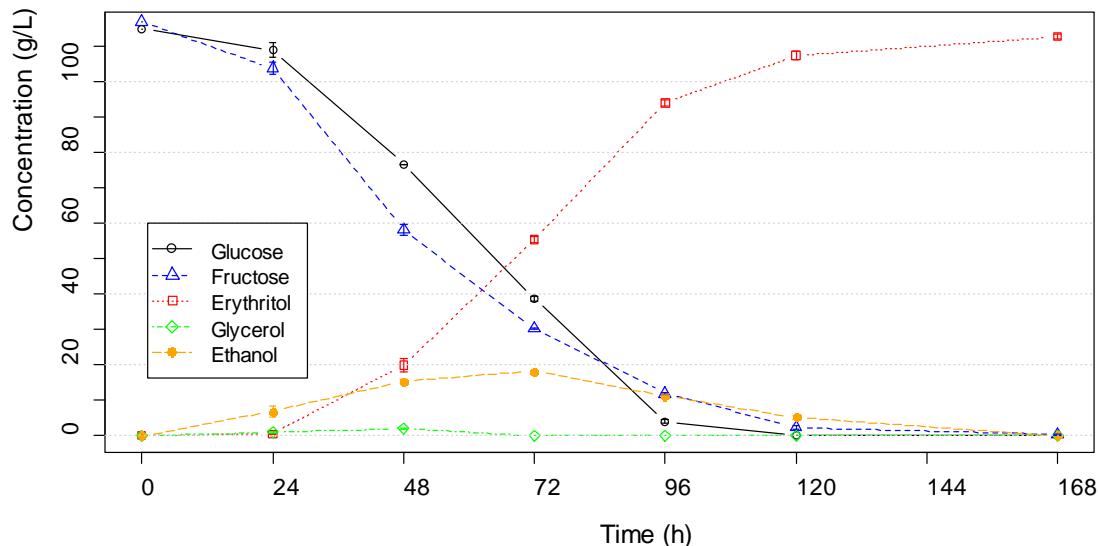


Figure S2. Evolution of grape must fermentation with *M. pollinis* MUCL 40570 in flask experiments under optimized nitrogen dosing (6.88 g/L yeast extract; equivalent to 0.76 g/L TN). Average values \pm standard deviations are shown ($n = 2$).

Table S3. Fermentation parameters (120 h) for grape must in flask experiments with different combinations of yeast extract and ammonium sulphate as nitrogen sources (as described in Table 1 in the manuscript), using *M. pollinis* MUCL 40570 (average \pm standard deviation; $n = 2$). Note: For each column, letters between parentheses (**a**, **b**, **c**, **d**) indicate the existence of statistical differences ($p < 0.05$; Tukey HSD test) among treatments; if two treatments share the same letter, there are no significant differences between them for that parameter.

Cell Den-		C_{ETH} (g/L)	C_{ERY} (g/L)	C_{GLY} (g/L)	ΔG (%)	ΔF (%)	ΔS (%)	Y_{ETH} (g/g)	Y_{ERY} (g/g)	Y_{GLY} (g/g)
Yeast ex-	sist									
tract (%)	(cells/mL) $\times 10^8$	(a)	(a)	(d)	(a)	(a)	(a)	(a)	(a)	(d)
100	2.69 \pm 0.31	5.62 \pm 2.35	89.12 \pm 0.50 (a)	1.29 \pm 0.16 (d)	100 \pm 0 (a)	99.09 \pm 0.39 (a)	99.53 \pm 0.20 (a)	1.95x10 ⁻² \pm 7.96x10 ⁻³ (a)	3.10x10 ⁻¹ \pm 5.29x10 ⁻³ (a)	4.47x10 ⁻³ \pm 6.16x10 ⁻⁴ (d)
80	2.34 \pm 0.22	5.27 \pm 1.96	89.93 \pm 1.19 (a)	0.79 \pm 0.14 (d)	100 \pm 0 (a)	99.49 \pm 0.06 (a)	99.74 \pm 0.03 (a)	1.83x10 ⁻² \pm 6.76x10 ⁻³ (a)	3.10x10 ⁻¹ \pm 5.29x10 ⁻³ (a)	2.74x10 ⁻³ \pm 4.86x10 ⁻⁴ (d)
60	2.14 \pm 0.07	6.91 \pm 3.05	90.17 \pm 0.49 (a)	1.54 \pm 0.19 (d)	100 \pm 0 (a)	98.94 \pm 0.29 (a)	99.45 \pm 0.15 (a)	2.37x10 ⁻² \pm 1.06x10 ⁻² (a)	3.09x10 ⁻¹ \pm 2.40x10 ⁻⁶ (a)	5.25x10 ⁻³ \pm 6.25x10 ⁻⁴ (d)
40	2.13 \pm 0.31	4.86 \pm 1.91	89.73 \pm 0.02 (a)	2.85 \pm 0.49 (c)	100 \pm 0 (a)	97.63 \pm 0.60 (a)	98.77 \pm 0.31 (a)	1.69x10 ⁻² \pm 6.47x10 ⁻³ (a)	3.13x10 ⁻¹ \pm 3.54x10 ⁻³ (a)	9.94x10 ⁻³ \pm 1.83x10 ⁻³ (c)
20	1.80 \pm 0.11	4.52 \pm 0.47	78.67 \pm 2.54 (b)	4.10 \pm 0.05 (b)	99.88 \pm 0.10 (a)	93.36 \pm 1.27 (b)	96.49 \pm 0.71 (b)	1.63x10 ⁻² \pm 1.84x10 ⁻³ (a)	2.83x10 ⁻¹ \pm 6.76x10 ⁻³ (b)	1.47x10 ⁻² \pm 3.02x10 ⁻⁴ (b)
0	2.41 \pm 0.44	0 \pm 0 (a)	62.69 \pm 0.09 (c)	7.84 \pm 0.09 (a)	94.62 \pm 0.28 (b)	83.37 \pm 0.97 (c)	88.78 \pm 0.37 (c)	0 \pm 0 (a)	2.41x10 ⁻¹ \pm 4.64x10 ⁻⁴ (c)	3.01x10 ⁻² \pm 2.51x10 ⁻⁴ (a)

C_{ETH} : ethanol concentration; C_{ERY} : erythritol concentration; C_{GLY} : glycerol concentration; ΔG : glucose consumption, ΔF : fructose consumption, ΔS : total sugar consumption, Y_{ETH} : ethanol yield, Y_{ERY} : erythritol yield, Y_{GLY} : glycerol yield.

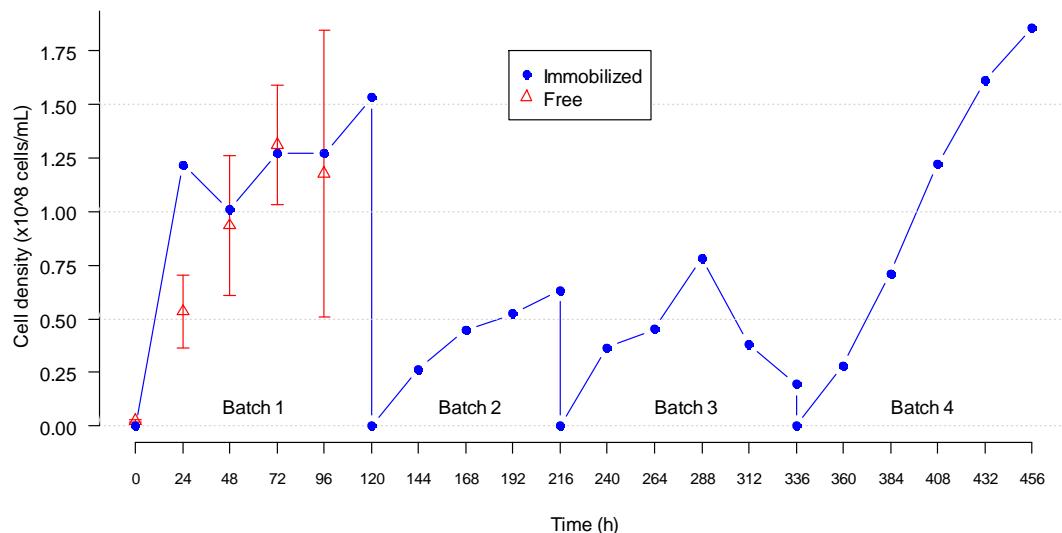


Figure S3. Cell density in the liquid medium of the bioreactor during the four consecutive immobilization batches (filled circles). The values of the free-cell fermentation are shown for comparison (empty triangles).