

# **Uncovering novel plasma membrane carboxylate transporters in the yeast *Cyberlindnera jadinii***

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## Supplementary material

**Table S1.** Identification of *C. jadinii* carboxylate transporter homologs of the distinct transporter families. For ScAto1 and ScJen1 homologs, homology search was performed by BlastP. Protein conserved domains were determined in the Conserved Domain Database (CDD) and the number of transmembrane segments (TMSs) was determined with TMHMM Server v.2.0.

Accession number	Query	Query cover	Identity	E-value	TMSs	ID gene
<b>XP_020070445.1</b>		98 %	56 %	2e <sup>-99</sup>	4	CjAto1
<b>XP_020073179.1</b>	NP_009936.1	96 %	57 %	2e <sup>-96</sup>	6	CjAto2
<b>XP_020073031.1</b>	Ady2p	91 %	55 %	2e <sup>-100</sup>	5	CjAto3
<b>XP_020073178.1</b>	[ <i>S. cerevisiae</i> ]	98 %	57 %	4e <sup>-109</sup>	5	CjAto4
<b>XP_020067765.1</b>		94 %	41 %	6e <sup>-72</sup>	6	CjAto5
<hr/>						
<b>CEP23088.1</b>		82 %	58 %	0	10	CjJen1
<b>CEP21966.1</b>	NP_012705.1	85 %	51 %	1e <sup>-175</sup>	11	CjJen2
<b>CEP22358.1</b>	Jen1p	77 %	40 %	1e <sup>-119</sup>	12	CjJen3
<b>CEP21989.1</b>	[ <i>S. cerevisiae</i> ]	76 %	41 %	5e <sup>-116</sup>	12	CjJen4
<b>CEP21602.1</b>		67 %	43 %	1e <sup>-103</sup>	10	CjJen5
<b>CEP25129.1</b>		82 %	41 %	8e <sup>-127</sup>	12	CjJen6

Accession number	Domain designation	Accession	Description	E-value	TM Ss	ID gene
<b>XP_020067635.1</b>	MFS_MCT_SL C16	cd17352	Monocarboxylate transporter (MCT) family of the Major Facilitator Superfamily of transporters	2.27e <sup>-64</sup>	12	CjSlc16
<b>XP_020068154.1</b>	SLC5-6-like_sbd super family	cl00456	Solute carrier families 5 and 6-like; solute binding domain	3.25e <sup>-14</sup>	11	CjSlc5
<b>XP_020068891.1</b>	TDT super family	cl04176	Tellurite-resistance/Dicarboxylate Transporter (TDT) family	1.19e <sup>-83</sup>	9	CjTDT
<b>XP_020069270.1</b>	SLC13_perme ase	cd01115	Permease SLC13 (solute carrier 13); Sodium/dicarboxylate cotransporter NaDC-1	1.56e <sup>-98</sup>	12	CjSlc13-1
<b>XP_020073044.1</b>			Permease SLC13 (solute carrier 13); Sodium/dicarboxylate cotransporter NaDC-1	7.26e <sup>-103</sup>	12	CjSlc13-2

**Table S2.** Oligonucleotides used for strain construction, cloning and expression.

Name	Sequence
p416-CJAd1_fwd	GCAGGATCCATGTCAGACAAGGAAAGC
p416-CJAd1_rev	GCAGAATTCTAGGAGTGCACTTGAGC
p416-CJAd2_fwd	GCAGGATCCATGTCTAGTATTAATGAG
p416-CJAd2_rev	GCAGAATTCTCACAAATGGCTAGCACC
p416-CJAd3_fwd	GCAGGATCCATGCCGCCAACGTTGAC
p416-CJAd3_rev	GCAGAATTCTCAGGCACGCTTGAC
p416-CJAd4_fwd	GCAGGATCCATGTCGGACAAGGAAAAC
p416-CJAd4_rev	GCAGAATTCTCAAGAACGCTTGAC
p416-CJen1_fwd	GCA GGATCCATGCACAAACTTGAAGAG
p416-CJen1_rev	GCAGAATTCTCACTTCTCTCTGTGG
p416-CJen2-new_Fwd	GCAGGATCCATGACTTCACCATTGCCT
p416-CJen2-new_Rev	GCACTCGAGTCACTCACTTGAAGAGCC
p416-CJen3_fwd	GCAGAATTCATGGCAATGTCTGATGTT
p416-CJen3_rev	GCAGAATTCTCAGGTTTTTCAATTATG
p416-CJen4-new_Fwd	GCAGGATCC ATGACTGCTGGGAGATA
p416-CJen4-new_Rev	GCACTCGAGCTAGTCTCTAGCAGATT
p416-CJen5_fwd	GCAGGATCCATGGACTGGGATGCTTC
p416-CJen5_rev	GCAGAATTCTCACTTGGCTATCTT
Cut-Jen6_Fwd	GCCTCTAGAATGGGATTCAAGACGTAC
Cut-Jen6_Rev	GCCGAATTCTATTAAACCTCAGAAC
ct1.635_Fwd	GCCACTAGTATGACTGAAATCATCA
ct1.635_Rev	GCCGTCGACTCAGAATTACAAATTCT
ct2.154_Fwd	GCCACTAGTATGTTGCAGAGACCGAG
ct2.154_Rev	GCCGTCGACTCAGGAGTTGTAGCACA
ct3.891_Fwd	GCCACTAGTATGACATCTGATGAGAAT
ct3.891_Rev	CCGTCGACTCAATTCTTCACTGTT
ct4.270_Fwd	GCCACTAGTATGAAATTCTCCCTCT
ct4.270_Rev	GCCGTCGACTAACCGTGTAAAGTTGC
ct8.044_fwd	GCCACTAGTATGAAGTTCTCCCATTG
ct8.044_rev	GCCCTCGAGTCACATCCCTGTTAATCT
ct9.7765_fwd	GCCACTAGTATGTCGACCTTCTCTC
ct9.7765_rev	GCCGAATTCTAAACTCTGGTGCATG

**Table S3.** Parameters obtained with HHpred for 3D-model construction.

Protein templates	PDB Hit	Organism	E-value	Similarity	Identity	Score	Probability (%)
<b>ScAto1</b>	5YS3	<i>Citrobacter koseri</i>	3.7e <sup>-32</sup>	0.566	35%	236.5	100.0
<b>EcSatP</b>	5YS3	<i>Citrobacter koseri</i>	1.5e <sup>-36</sup>	1.498	92%	235.4	100.0
<b>CjAto2</b>	5YS3	<i>Citrobacter koseri</i>	6.7e <sup>-33</sup>	0.538	32%	237.2	100.0
<b>CjAto5</b>	5YS3	<i>Citrobacter koseri</i>	7e <sup>-35</sup>	0.525	26%	252.0	100.0

**Table S4.** Residues of Ato homologs presenting strong intramolecular interactions with lactate, succinate and citrate identified by molecular docking studies.

3D-Protein templates	Lactate (-1)	Succinate (-1)	Succinate (-2)	Citrate (-3)
<b>ScATO1</b>	R111; T238; N179; T222; T102; S106; E140; T209; N255	T238; N179; R111; T222; T102; Q133; S208; N145; N255; T209; E140	R111; N179; T238; T222; T209; S208; N145	N255; T209; S208; N89; N145; Q133; T238; H230; R111; N179
<b>CjATO2</b>	K244; S189; N127; N91; T203; I97; T99; Y158	N236; K244; S189; C185; T203; N91; T84; T99; D150; R214; Q219; S211; W144; Y158	S189; K244; N127; T203; W144; T99; Q219; R214; Y158	ND
<b>CjATO5</b>	N101; G220; Q103; T229; K230	E125; K221	K221	N137; S199; K198; N81; K221; Q103; T229; K230; G220

Note: ND - not determined.

**Table S5.** Average of the binding affinity values [kcal/mol] calculated with PyRx software for the docking of Ato proteins with the distinct charged substrates tested.

3D-Protein templates	Average of binding affinities (kcal/mol) at different binding sites						
	Lactate (-1)						
	S4			S3	S2	S1	
	a	b	c			a	b
<b>ScAto1</b>	-	-3.8	-	-4.1	-3.9	-3.3	-3.1
<b>EcSatP</b>	-3	-	-	-3.6	-3.8	-3	-
<b>CjAto2</b>	-3.4	-	-3.4	-4.2	-2.6	-3.4	-3.1
<b>CjAto5</b>	-3.2	-3.1	-	-3.3	-3.8	-3.4	-
Succinate (-1)							
	S4			S3	S2	S1	
	a	b	c			a	b
<b>ScAto1</b>	-	-4.0	-	-5	-4.5	-3.9	-3.8

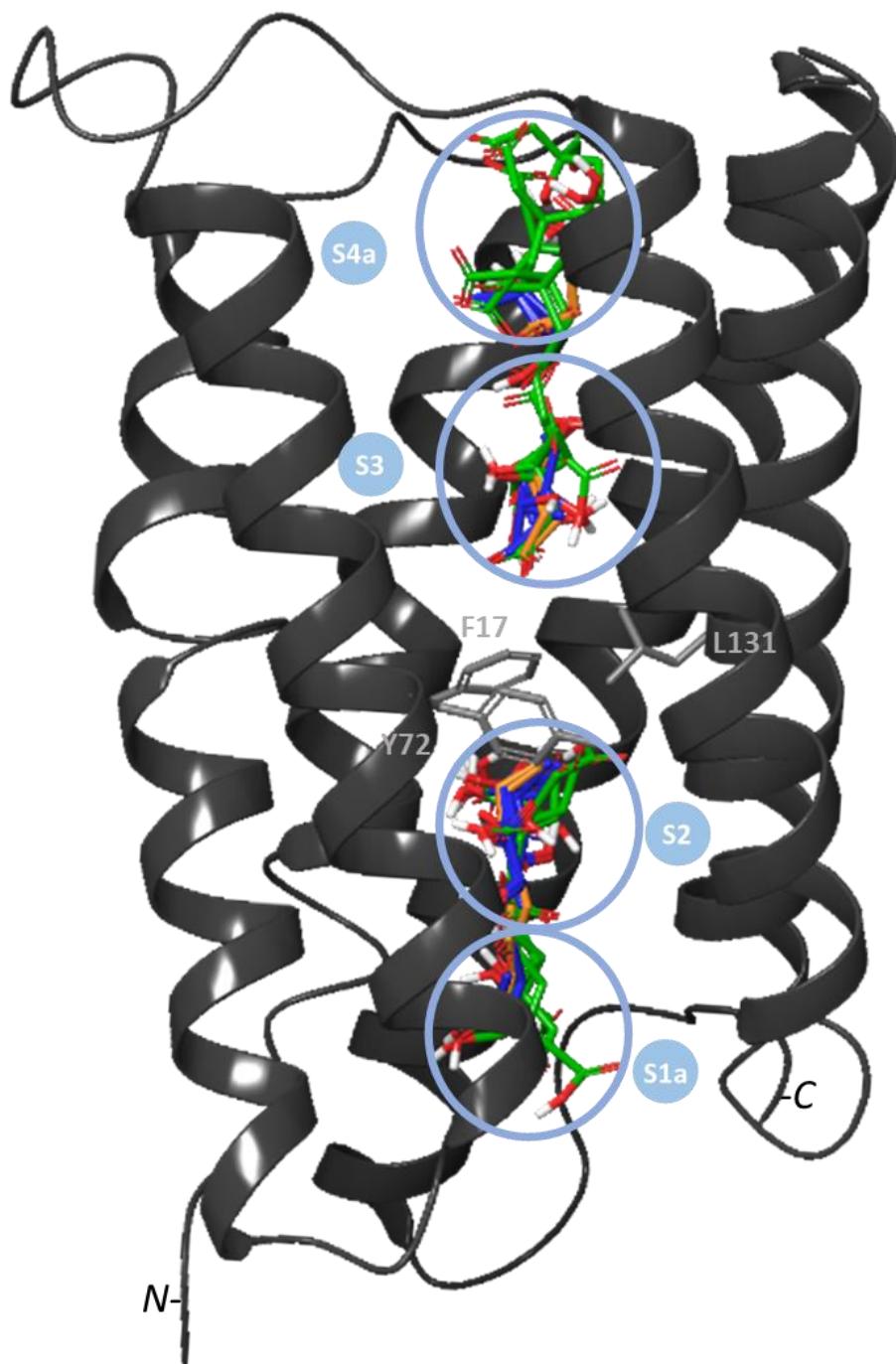
<b>EcSatP</b>	-3.6	-	-	-4.5	-4.6	-4.2	-
<b>CjAto2</b>	-4.2	-	-4.2	-4.7	-3.2	-3.9	-4
<b>CjAto5</b>	-3.8	-	-	-3.9	<b>-4.7</b>	-	-
<b>Succinate (-2)</b>							
<b>S4</b>							
<b>a</b>			<b>S3</b>		<b>S2</b>		<b>S1</b>
<b>b</b>							<b>a</b>
<b>c</b>							<b>b</b>
<b>ScAto1</b>	-	-4.0	-	-5	-4.4	-3.7	-
<b>EcSatP</b>	-3.4	-	-	-4.4	<b>-4.7</b>	-4.3	-
<b>CjAto2</b>	-4	-	-4.1	-4.8	-3.1	<b>-4.1</b>	-4
<b>CjAto5</b>	-3.8	-	-	-3.9	<b>-4.6</b>	-	-
<b>Citrate (-3)</b>							
<b>S4</b>							
<b>a</b>			<b>S3</b>		<b>S2</b>		<b>S1</b>
<b>b</b>							<b>a</b>
<b>c</b>							<b>b</b>
<b>ScAto1</b>	-	-5.0	-	-6.1	-3.7	-4.6	-
<b>EcSatP</b>	-4.2	-	-	-5	<b>-4.3</b>	-4.2	-
<b>CjAto5</b>	-4	-4	-	-3.5	<b>-5.5</b>	-4.5	-4.2

**Figure S1.** Multiple sequence alignment of Ato homologs from *Saccharomyces cerevisiae* (Ato1, Ato2, Ato3), *C. jadinii* (CjAto1-5) and the *Escherichia coli* SatP. The sequence alignment was built with ClustalOmega (<https://www.ebi.ac.uk/Tools/msa/clustalo/>). Localization of transmembrane segments (TMSs) was predicted by the PSI/TM-Coffee software (<http://tcoffee.crg.cat/apps/tcoffee/do:tmcoffee>). Grey background highlights the previously identified and characterized signature motifs of the AceTr family (Ribas et al., 2019). Blue rectangles indicate residues from the narrowest hydrophobic constriction site F98-Y155-L219 (refer to ScAto1p) (Qiu et al., 2018).

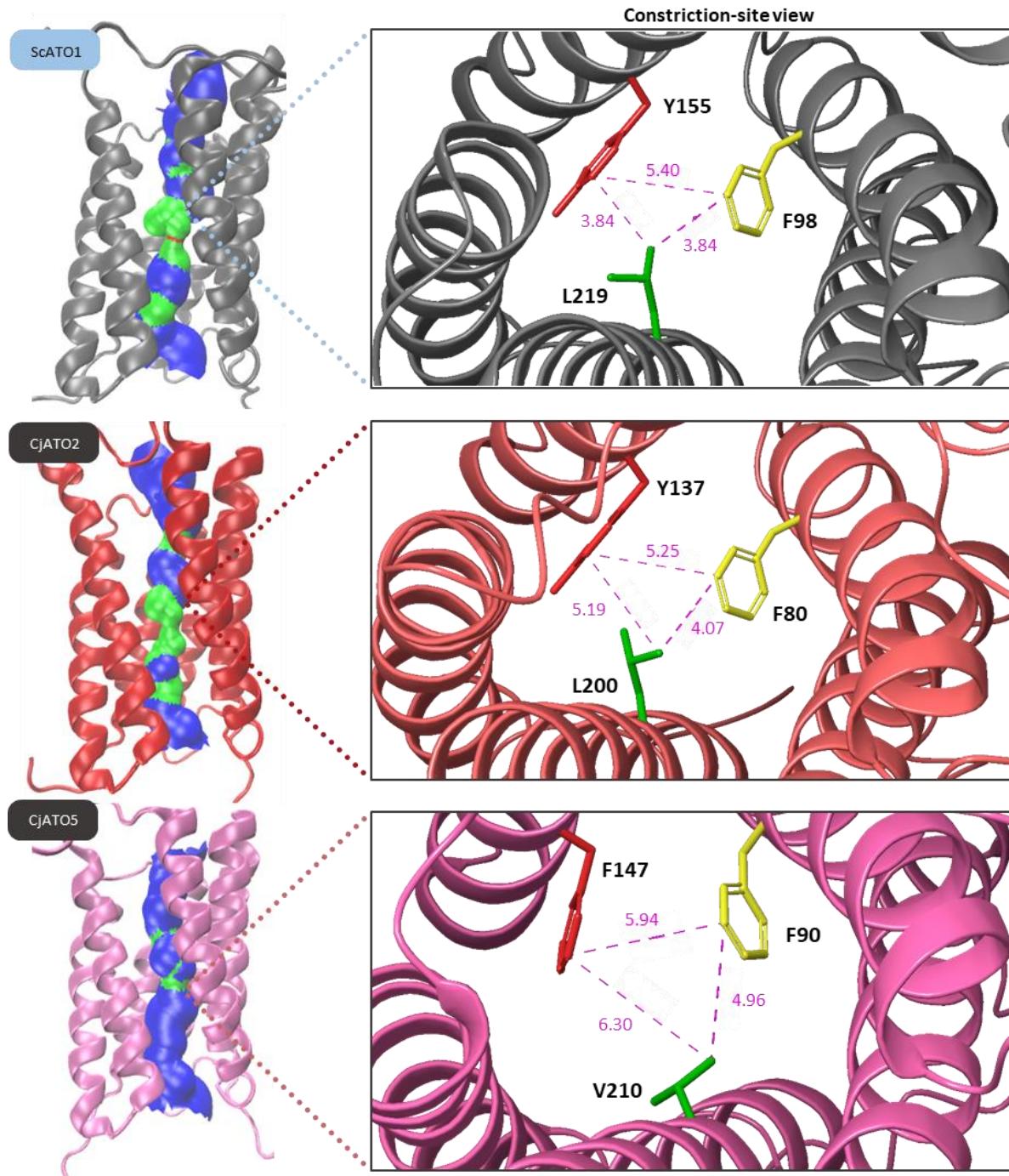
	1 <sup>st</sup> TMS	
ScAto1	GRQKFLKSDLYQAFG-GTINPGLAP-APVHKFANPAPLGLSAFALTTEVLSMFNARAQGI	115
ScAto2	GRQKFLRDDLFEAFG-GTINPGLAP-APVHKFANPAPLGLSGFALTTEVLSMFNARAQGI	114
ScAto3	GSSTYRRRDLLNALDRGDGEEGNCAYTPHQFANPVPLGLASFSLSCLVLSLINANVRGV	112
CjAto1	GRTKVLRSELWNAFG-GDLQPGIHA-PPPRRFANPAPLGLCGFALTTEVLSMSNARAMGI	110
CjAto2	GRMKVRKSELWSAFG-GDLQPGVHA-QPQRKFANPVPILGLCGFALTTLVLSMANARAMGI	97
CjAto3	GRTKVLRSELWNAFG-GDLQPGIHA-TPPRRFANPAPLGLCAFALTTEVLSMVNARAMGI	104
CjAto4	GHQKVLRSELWTAFG-GDLQPGVHA-PPPQRLANPAPLGLCGFALTTEVLSMANARAMGV	106
CjAto5	GNQAFSKKDLFNAFA-GDLQPGLHA-TPHRPMPGNPVPMLTSFCICCEVVSLVNAQARGV	107
EcSatp	-----MGNTKLANPAPLGLMGFGMTTILLNLHNVGYFA-	33
	2 <sup>nd</sup> TMS	
ScAto1	TVPNVVVGCAMFYGGLVQLIAGIWEIALENTFGGTALCSYGFWLSFAAIYI-PWFGILE	174
ScAto2	TIPNVVVGCAMFYGGLVQLIAGIWEIALENTFGGTALCSFGFWLSFGAIYI-PWFGILD	173
ScAto3	TDGKWALSLFMFFGGAIELFAGLLCFVIGDTYAMTVFSSFGFWICYGYGLT-DTDDNLVS	171
CjAto1	TVANAVGPFFYGGIIQLLSGMWEISLDNTFGGTVLSSYGGFWLSWAQI-DWFGIQR	169
CjAto2	RTPNVAVAPAFFYGGFAQILAGMWIEIALENTFGSVLTSYGFWLSSWAQI-DWFGIKA	156
CjAto3	TTPNIVVGLALFYGGFVQLLAGMWIEIALDNTFGGTALSSYGGFWMSYAAQI-DWFGIKS	163
CjAto4	TIPNAAVGAACFYGGLVQLLAGMWIEISLDNTFGGTALSSYGGFWMSWAQI-DWFGIKK	165
CjAto5	TNAKVIASCALFFAGVVETISGLWCLVIENTFAATALGSGGGFWMGYAGLLI-DAFGITS	166
EcSatp	-LDGIILAMGIFYGGIAQIFAGLLEYKKGNTFLTAFTSYGSFWITLVAILLMPKLGTD	92
	3 <sup>rd</sup> TMS	
ScAto1	AYEDNESDLNNALGFYLLGWAIFTFGLTVCTMKSTVMFFLFFFIALTFLLLSIGHFANR	234
ScAto2	AYKDKESDLGNALGFYLLGWALFTFGLSVCTMKSTIMFFALFFFIALTFLLLSIANFTGE	233
ScAto3	GYTDP-TMLNNVIGFFLAGWTVFTFLMLMCTLKSTWGLFLLLTELDTFLLLCIGTFIDN	230
CjAto1	AYDDP-IMLNNGLGFLLGWWIFTLMVLICTVKSTVAFFSLFFELEMFTLLTIGEFTRS	228
CjAto2	AYDDP-IELENAIGFLLGWWIFTFLILLCTMKSTVAFFSMFFILEITFILLTVASFTRH	215
CjAto3	AYTDP-IELANAVSFLLGWTIFTFMILLCTVKSTVSFFSLFFFLEITFLLLTIGDFTRR	222
CjAto4	AYDDP-IMLANAVGFLLGWTIFTFMVLCTVKSTVAFFSLFFFEDITFLLTIGEFTRK	224
CjAto5	SYSTT-EELGNALGFYLTAWTIFAFIMWLCTFKSTWPFFILFLIVVVFLMCIAIGKYNDN	225
EcSatp	A-----PNAQFLGVYLGLWGVFTLFMFFGTLKGARVLQFVFFSLTVLFALLAIGNIAGN	146
	4 <sup>th</sup> TMS	
ScAto1	LGVTTRAGGVVLGVVVAFIAWYNAYAGVATQNSYVLARPFPPLPSTERVIF---	283
ScAto2	VGVTRAGGVVLGVIVAFIAWYNAYAGIATRQNSYIMVHPFALPSNDKVFF---	282
ScAto3	NNLKMAGGYFGILSSCCGWYSLYCSVSPNSYLAFRHTMPNAP-----	275
CjAto1	VGVTRAAGVFGVITSFLGWYNALAGFATRENSYFVATAVPLPGAKRAQVHS-	279
CjAto2	VGCQRAGGVFGVITGFLAWYNAYAGIATKEISYFVPKPWPLPGASHL-----	262
CjAto3	VGVTRAGGVFGVITAFIAWYNNAFAGIATKENSYITIKAWPLPGAKRA-----	269
CjAto4	TGVSRAAGVFGVITSFIAWYNNAFAGLATKENSYVVAIPLPLPGAKRS-----	271
CjAto5	TTATKAGGVVLGVATFVFFFIVYAGVADSSNSYLTIPASPMPHAPRV-----	272
EcSatp	AAIIHFAGWIGLICGASAIYLAMGEVLNEQFGRTVL---PIGESH-----	188
	5 <sup>th</sup> TMS	
	6 <sup>th</sup> TMS	
ScAto1	LGVTRAGGVVLGVVVAFIAWYNAYAGVATQNSYVLARPFPPLPSTERVIF---	283
ScAto2	VGVTRAGGVVLGVIVAFIAWYNAYAGIATRQNSYIMVHPFALPSNDKVFF---	282
ScAto3	NNLKMAGGYFGILSSCCGWYSLYCSVSPNSYLAFRHTMPNAP-----	275
CjAto1	VGVTRAAGVFGVITSFLGWYNALAGFATRENSYFVATAVPLPGAKRAQVHS-	279
CjAto2	VGCQRAGGVFGVITGFLAWYNAYAGIATKEISYFVPKPWPLPGASHL-----	262
CjAto3	VGVTRAGGVFGVITAFIAWYNNAFAGIATKENSYITIKAWPLPGAKRA-----	269
CjAto4	TGVSRAAGVFGVITSFIAWYNNAFAGLATKENSYVVAIPLPLPGAKRS-----	271
CjAto5	TTATKAGGVVLGVATFVFFFIVYAGVADSSNSYLTIPASPMPHAPRV-----	272
EcSatp	AAIIHFAGWIGLICGASAIYLAMGEVLNEQFGRTVL---PIGESH-----	188

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**Figure S2.** Molecular docking of *Escherichia coli* Satp 3D-model, based on SatP\_Ck structure, with the substrates lactate (blue ligand), succinate (orange ligand) and citrate (green ligand). The four binding sites are depicted from S1 to S4 sites, including the localization of N- and C-terminal of the protein. The narrowest hydrophobic constriction of the anion pathway formed by F17, Y72 and L131 is also represented.

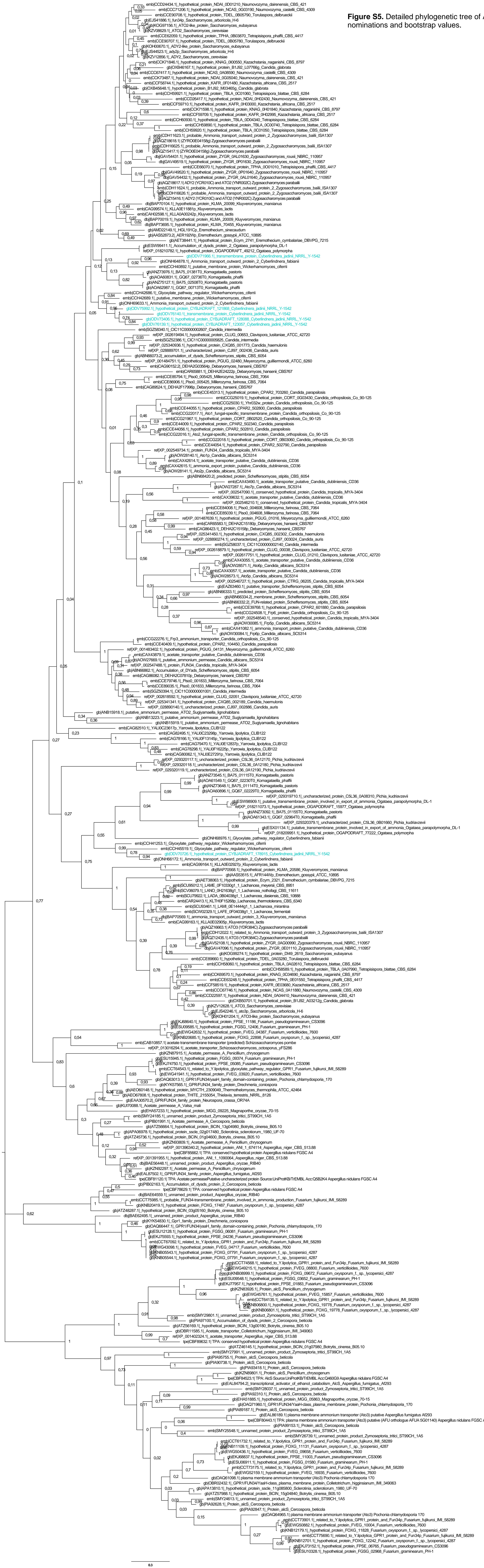


**Figure S3.** Predicted 3D structures of Ato proteins visualized with Maestro and HOLE softwares. On the left are the pore predictions: blue (larger aperture), green (intermediate pore size) and red (constricted pore). The top view of the narrowest hydrophobic constriction site of the anion pathway is also presented, including the predicted distances ( $\text{\AA}$ ) between represented residues.

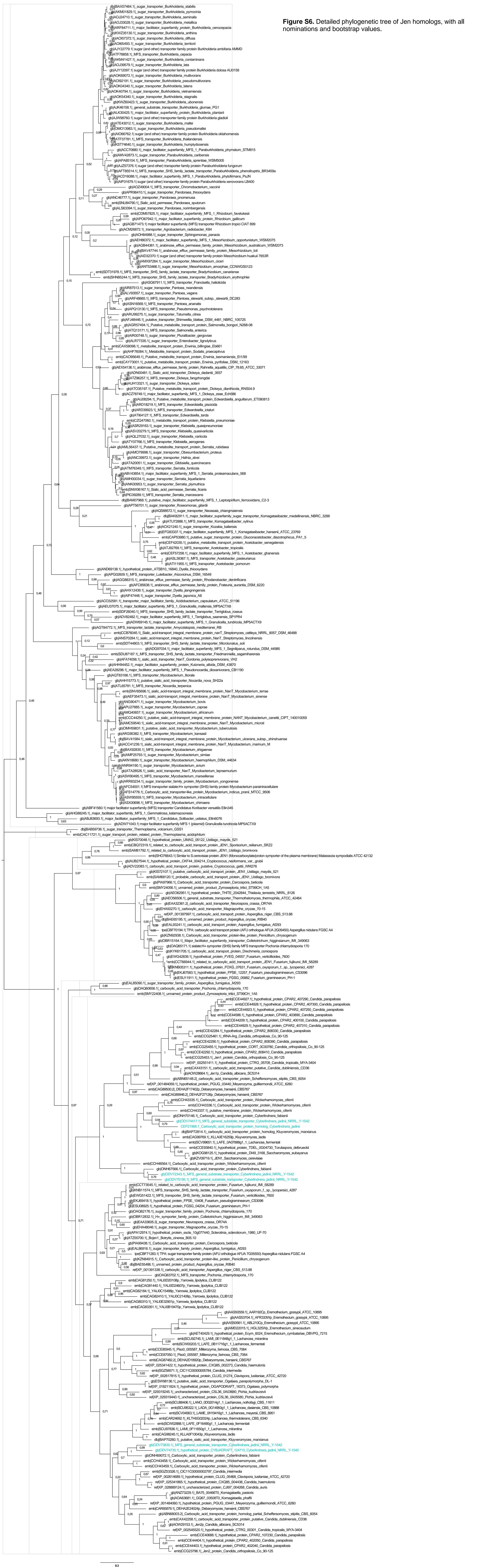




**Figure S5.** Detailed phylogenetic tree of Ato homologs, with all nominations and bootstrap values.



**Figure S6.** Detailed phylogenetic tree of Jen homologs, with all nominations and bootstrap values.



**Figure S7.** Detailed phylogenetic tree of Slc5 homologs, with all nominations and bootstrap values.

