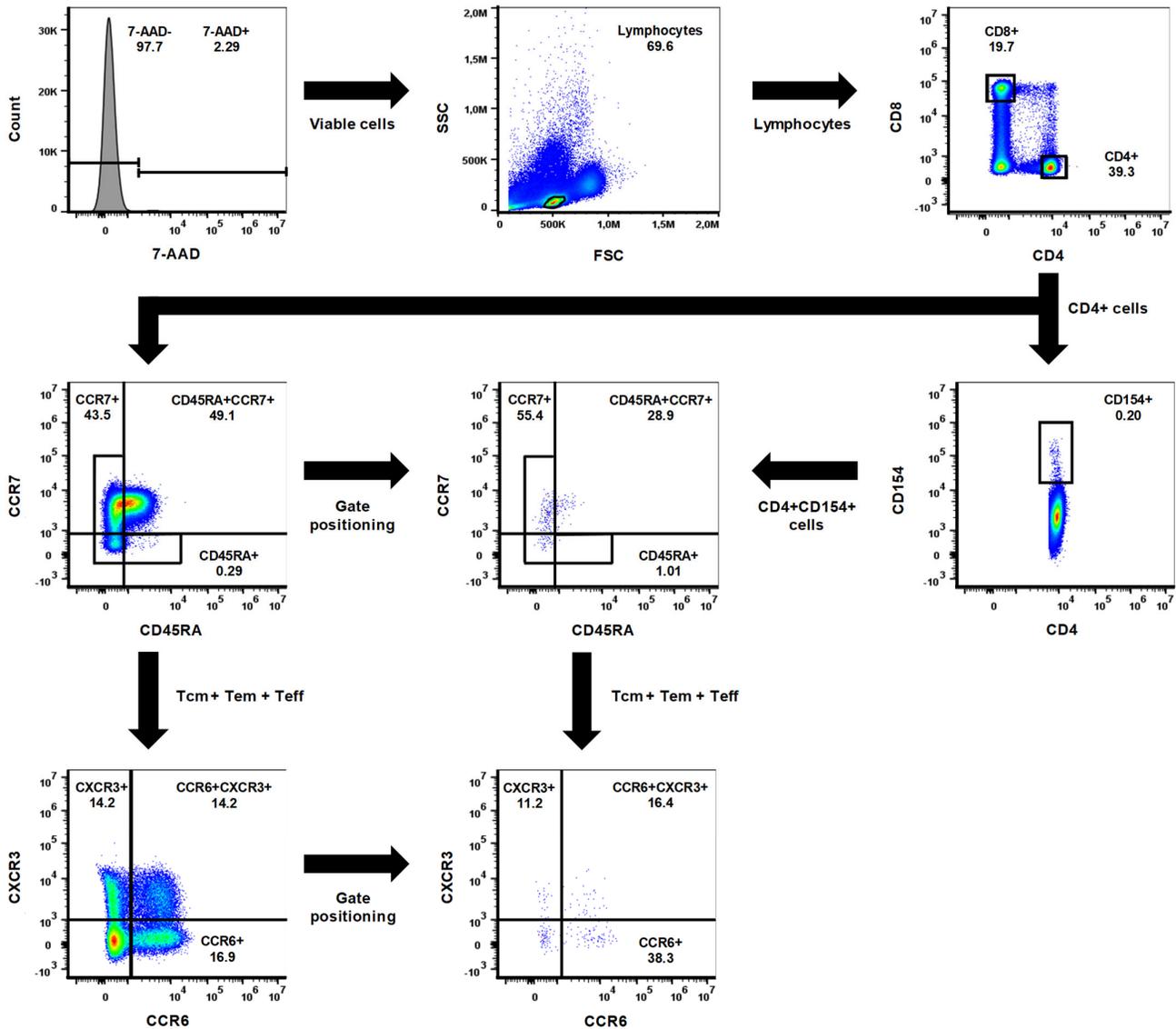


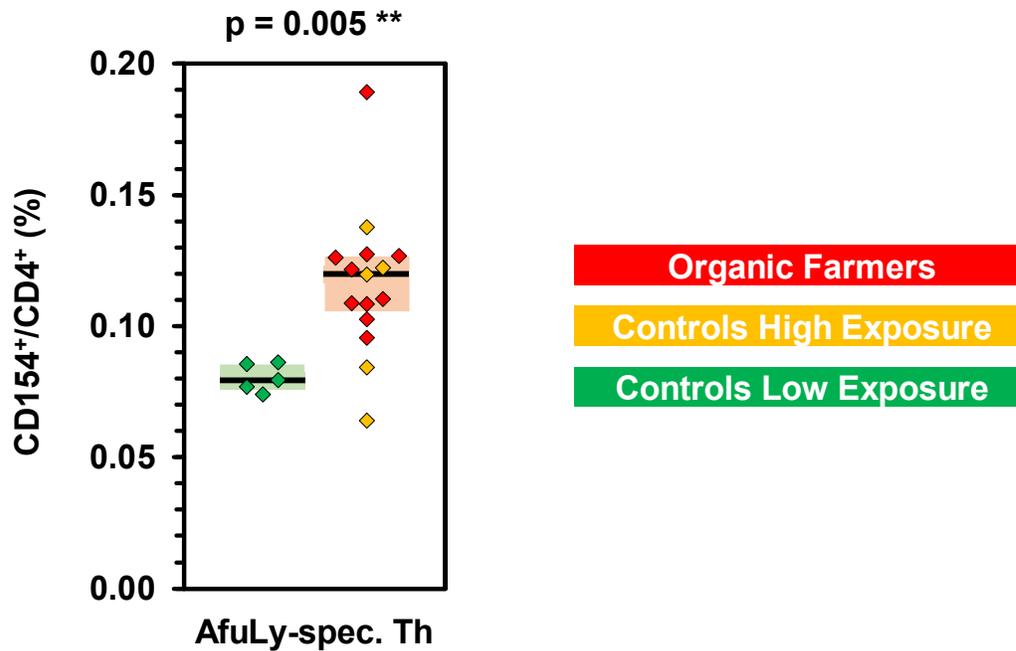
# Chronic occupational mold exposure drives expansion of *Aspergillus*-reactive type 1 and type 2 T-helper cell responses

## Supplementary Figures and Tables

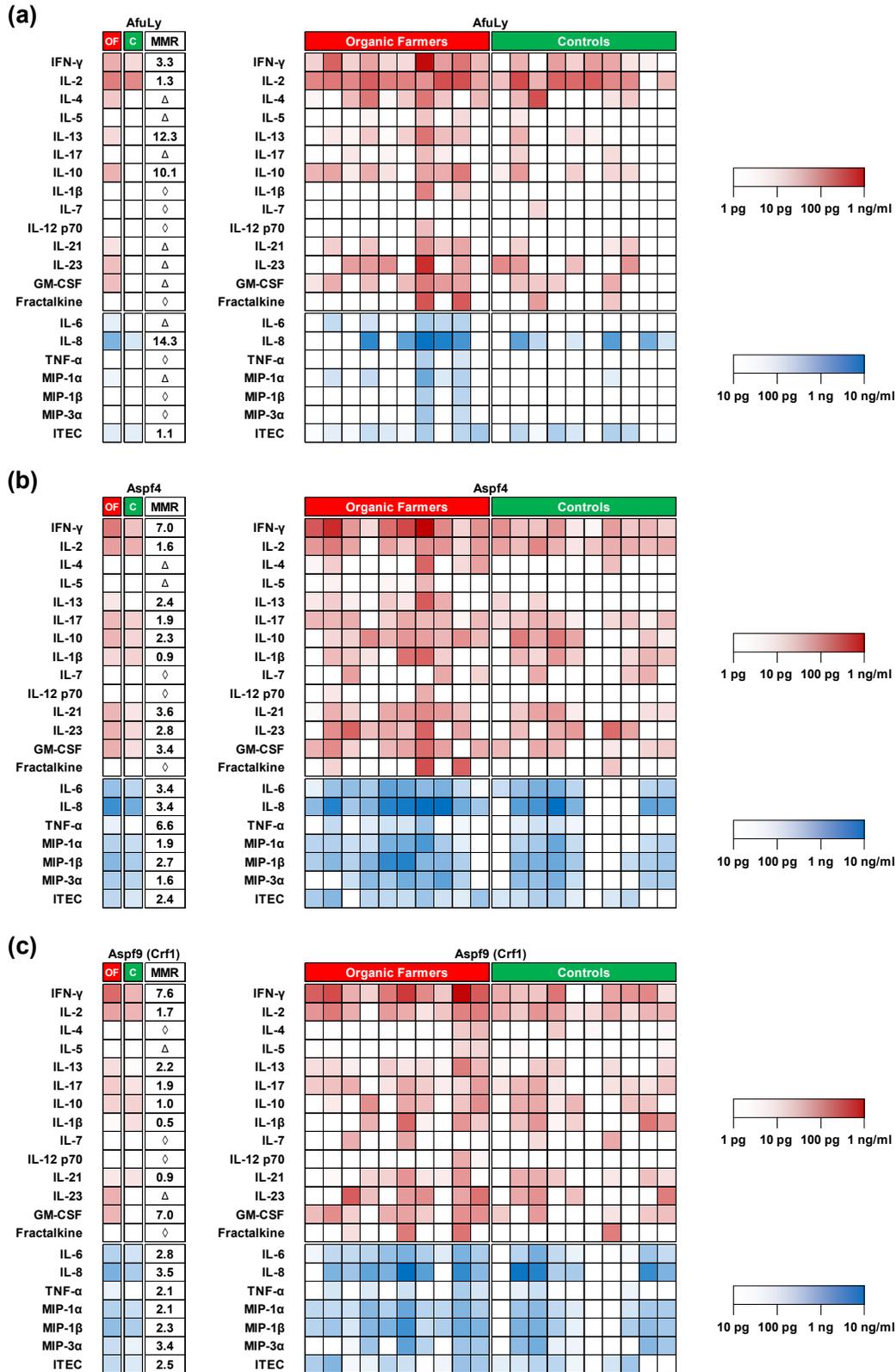


**Figure S1.** Gating strategy and representative data set for flow cytometric analyses. 7-Aminoactinomycin D (7-AAD)-positive (dead) cells were excluded, lymphocytes were identified by light scatter properties, and CD4<sup>+</sup> T-helper (Th) cells were gated. Effector Th cells (Teff; CD4<sup>+</sup>CD45RA<sup>+</sup>CCR7<sup>+</sup>), effector memory Th cells (Tem; CD4<sup>+</sup>CD45RA<sup>+</sup>CCR7<sup>-</sup>), central memory Th cells (Tcm; CD4<sup>+</sup>CD45RA<sup>-</sup>CCR7<sup>+</sup>), and naive Th cells (CD4<sup>+</sup>CD45RA<sup>-</sup>CCR7<sup>-</sup>) were differentiated among CD4<sup>+</sup> cells. An additional gate including the Tcm, Tem, and Teff subsets was positioned in order to exclude naive Th cells from subsequent Th1/Th2/Th17 cell differentiation. Thereafter, the non-naive CD4<sup>+</sup> cells were subdivided into Th1 cells (CD4<sup>+</sup>CCR6<sup>-</sup>CXCR3<sup>+</sup>), Th2 cells (CD4<sup>+</sup>CCR6<sup>+</sup>CXCR3<sup>-</sup>), Th17 cells (CD4<sup>+</sup>CCR6<sup>+</sup>CXCR3<sup>+</sup>), and others including Th1/Th17-type cells (CD4<sup>+</sup>CCR6<sup>-</sup>CXCR3<sup>-</sup>). Antigen-reactive cells among the total CD4<sup>+</sup> Th population were identified by CD154 expression. The CD45RA/CCR7 and CCR6/CXCR3 gates were then transferred to the CD4<sup>+</sup>CD154<sup>+</sup> population in order to define

memory/effector phenotypes and polarization of the antigen-reactive Th cells. Abbreviations: CD = cluster of differentiation, CCR = C-C motive chemokine receptor, CXCR = C-X-C motive chemokine receptor.



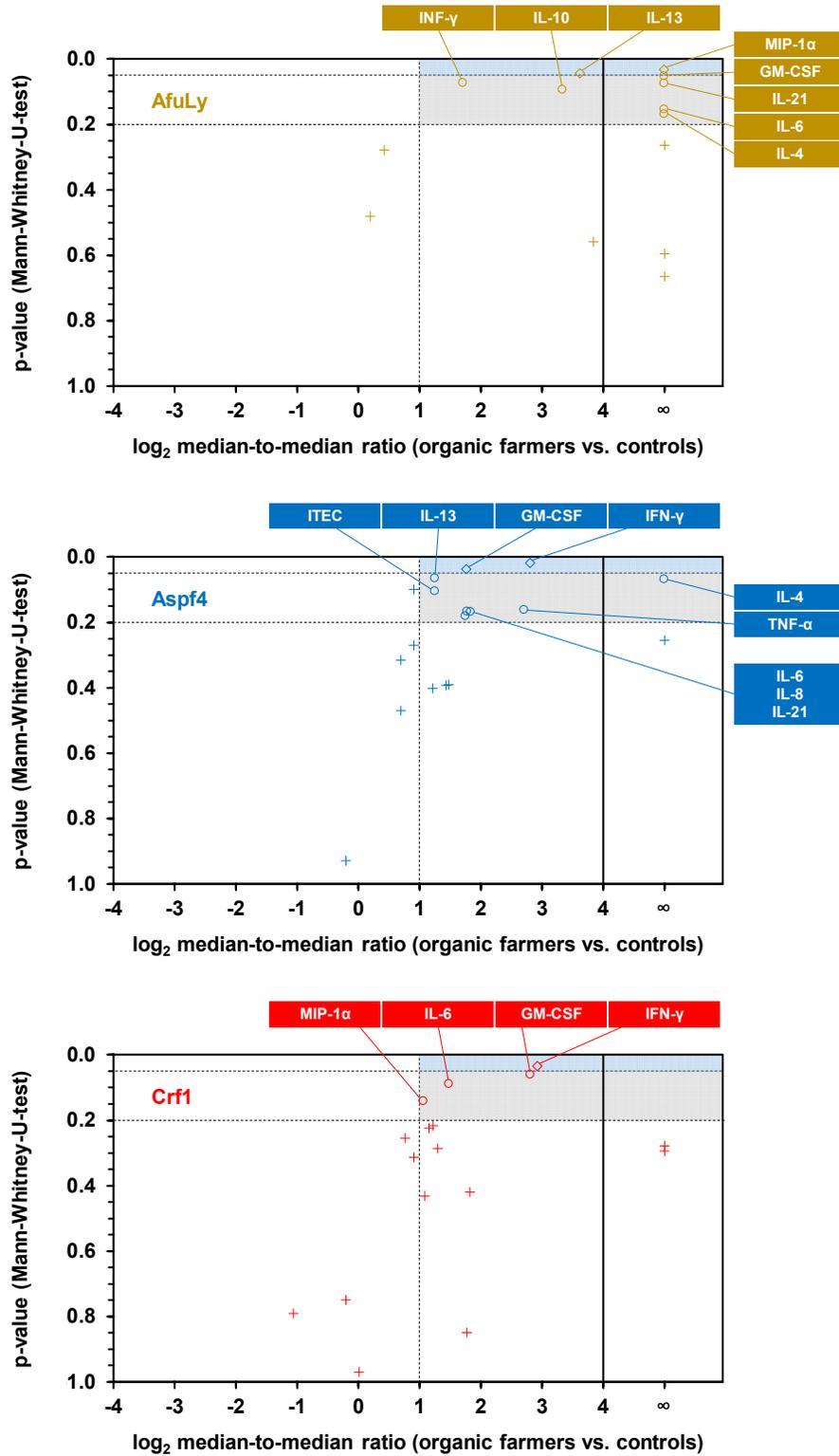
**Figure S2.** Subjects with risk factors for mold exposure in their occupational or residential environment display significantly increased *A. fumigatus*-specific T-helper cell frequencies. PBMCs from 10 organic farmers (red), 5 control subjects with high residential mold exposure (yellow), and 5 control subjects reporting no significant risk factors for occupational or residential exposure (green) were stimulated with *A. fumigatus* mycelial lysate (AfuLy). Frequencies of CD154<sup>+</sup> cells among (CD4<sup>+</sup>) T-helper cells were determined by flow cytometry. Unlike in Fig. 1a, subjects are grouped according to our previously reported classification of mold exposure profiles (Wurster et al., 2017, Mycoses; Page et al., 2018, Intern J Med Microbiol). Individual background-adjusted results, medians (black bars), and inter-quartile ranges (colored boxes) are shown. Two-sided Mann-Whitney-U-test.



**Figure S3.** Heatmaps of median and individual cytokine responses (whole blood-based Luminex assay) to *A. fumigatus* antigens in samples from organic farmers and controls. The heatmaps summarize median and individual background-

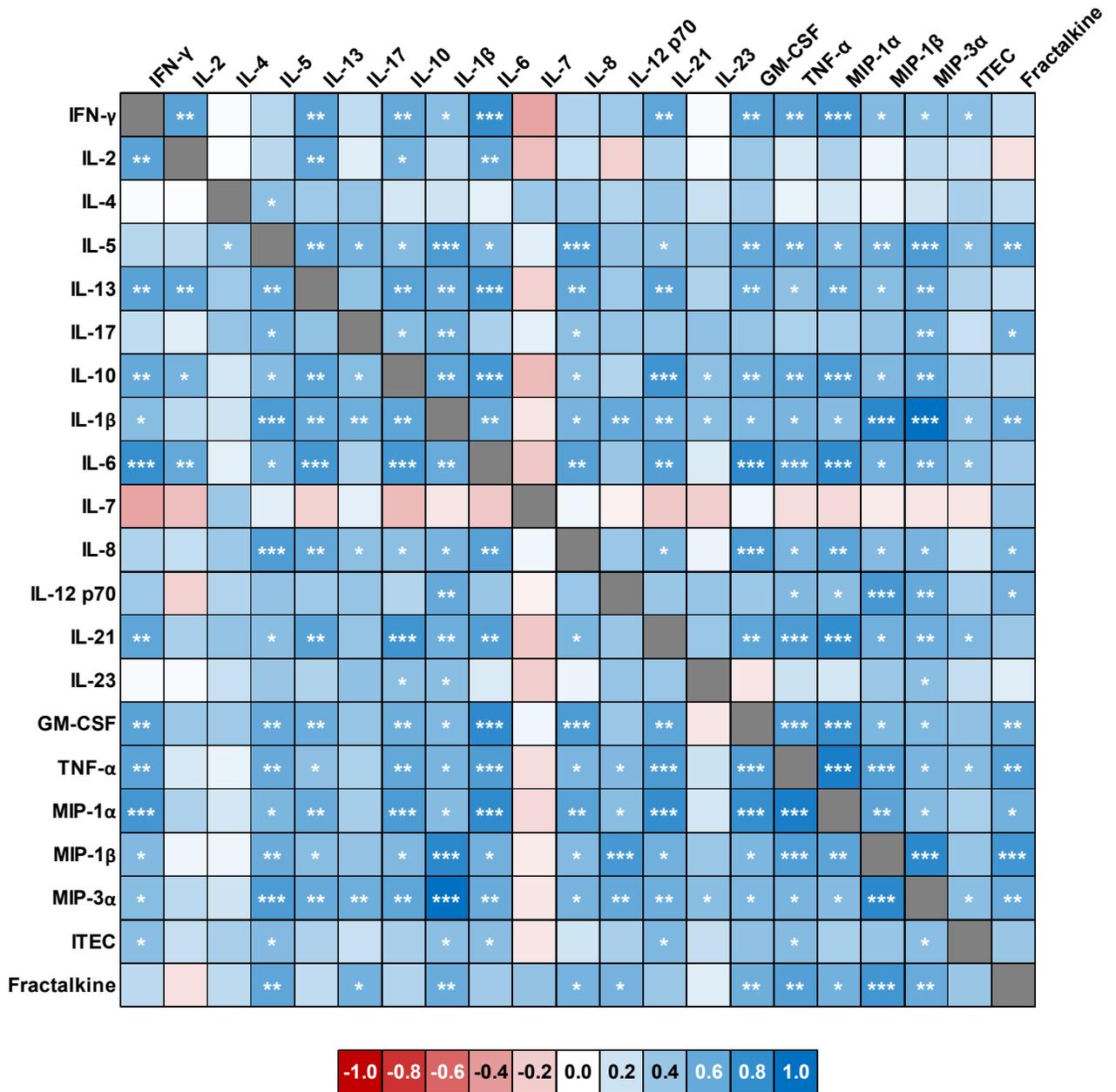
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adjusted cytokine concentrations in organic farmers (“OF”) and control subjects (“C”) depending on the *A. fumigatus* antigen used for stimulation. The numeric values in the MMR column represent median-to-median ratios between the two cohorts, with values >1.0 indicating greater median cytokine concentrations in the organic farmer cohort.  $\Delta$  denotes infinite median-to-median ratios (median = 0 pg/mL in the control cohort).  $\diamond$  denotes undefined median-to-median ratios (median = 0 pg/mL in both cohorts). AfuLy = *A. fumigatus* mycelial lysate.

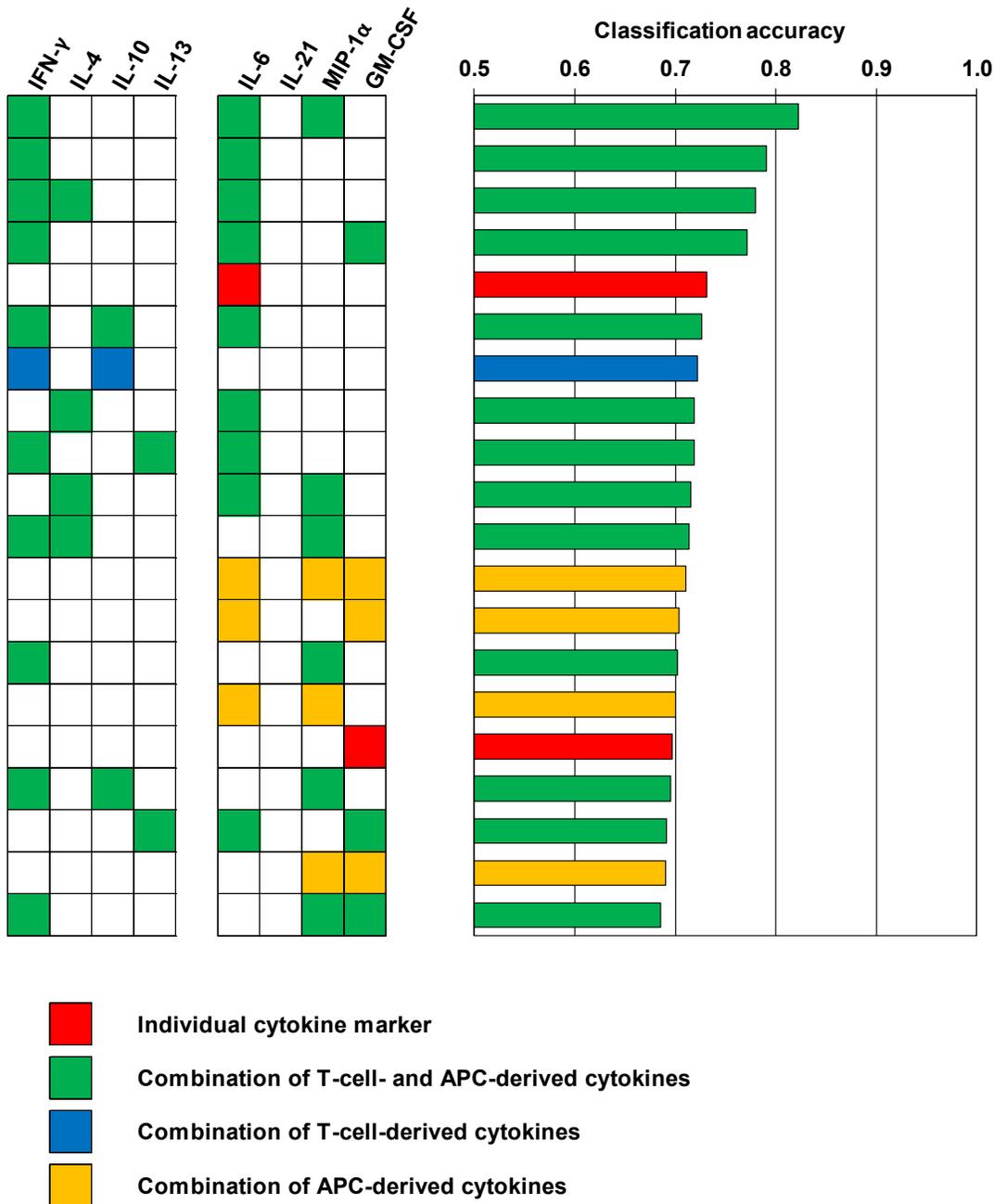


**Figure S4.** Volcano analysis of cytokine responses to *A. fumigatus* antigens identifies a selection of cytokine markers with significant or modest discriminatory power between organic farmers and controls. The volcano plots compare log<sub>2</sub>-transformed ratios of median concentrations for each antigen/cytokine combination in organic farmers and controls (x-values)

with the corresponding p-values (Mann-Whitney-U-test, y-values). Antigen/cytokine pairs are classified as insignificant (“+” symbols, median-to-median ratio < 2 and/or p > 0.2), potentially significant (circles, median-to-median ratio > 2 and 0.05 < p < 0.2), or significant (diamonds, median-to-median ratio > 2 and p < 0.05). Cytokine responses with medians of 0 pg/mL in both cohorts are not displayed.



**Figure S5.** T-cellular and innate immune cell-derived cytokine responses to *A. fumigatus* antigens are strongly correlated in our cohort of organic farmers and controls. Heatmap of Spearman’s rank correlation coefficients of individual cytokine concentrations elicited by *A. fumigatus* mycelial lysate (whole blood-based Luminex assay). FDR-corrected significance of Spearman coefficients is indicated by asterisks. \* p < 0.05, \*\* p < 0.01, \*\*\* p < 0.001.



**Figure S6.** Cytokine concentrations detectable by whole blood-based Luminex analysis modestly differentiate immune responses of organic farmers and controls. The classification accuracy of combinations of up to 3 AfuLy-induced cytokines responses was determined by a random forest algorithm. The top 20 combinations with the highest classification accuracy between organic farmers and controls are shown. Only cytokine markers passing the pre-filtering step (Fig. S4) based on their relative induction in the two cohorts (median-to-median ratio > 2.0) and p-values ( $p < 0.2$ ) were considered. Abbreviations: AfuLy = *A. fumigatus* mycelial lysate, APC = antigen presenting cells.

**Table S1.** Oligonucleotides used for the generation of recombinant *A. fumigatus* proteins.

Designation	Systematic name	Localization	Forward primer Restriction sites are indicated by bold fonts.	Reverse primer Restriction sites are indicated by bold fonts.
Aspf1 (mitogillin)	Afu5g02330	secreted	Aspf1-BamHI-FOR GCAGGATCCACCTGGACATGCAT CAAC	Aspf1-HindIII-REV GCAAAGCTTCTAATGAGAACACAG TCT
Aspf3	Afu6g02280	cytosolic	Aspf3-BamHI-FOR CTGGATCCATGTCTGGACTCAAG GCCGGT	Aspf3-HindIII-REV GCAAAGCTTTTACAGGTGCTTGAG GACGGT
Aspf4	Afu2g03830	secreted	Aspf4-BamHI-FOR CAGGATCCCACGAGCGCCGCCAC CTCCAC	Aspf4-HindIII-REV GCAAAGCTTCTACTCCTTGTAGTCG AGGTT
Aspf6 (MnSOD)	Afu1g14550	mitochondrial	Aspf6-BamHI-FOR AGGGTACCATGTCACAGCAATAC ACG	Aspf6-HindIII-REV GCAAAGCTTCTACAGCTTCATGAA TGG
Aspf8	Afu2g10100	cytosolic	Aspf8-BamHI-FOR GCGGATCCTCCTCTGAGGATGTCA AGGCC	Aspf8-HindIII-REV AGCAAGCTTTTAGTCGAAAAGACC GAAGCC
Aspf9	Afu1g16190	secreted / cell wall	Aspf9-BamHI-FOR CTGGATCCAAGACCTGCCCGCCC AACAAG	Aspf9-HindIII-REV ACTAAGCTTTTAGAATGCCAACAC GGCAG
Aspf22 (enolase)	Afu6g06770	cytoplasm	Eno-BamHI-FOR CGGGATCCATGCCTATCTCCAAG ATC	Eno-PstI-REV GCGCTGCAGTTACAGGTTGACGGC AGT
CatB (catalase)	Afu3g02270	secreted / cell wall	Cat1-BamHI-FOR CAGGATCCGTATGTCCTATATGA CC	Cat1-KpnI-REV AGGGTACCCTCGATTGCATCGTGC AA
CipC	Afu5g09330	cytoplasm	CipC-BamHI-FOR CAGGATCCATGGCTTGGGGCTGG GCAAAGCTTTTACCAACGGTTCGAC	CipC-HindIII-REV
CsnB (chitosanase B)	Afu4g01290	secreted	CsnB-BamHI-FOR TAGGATCCTATAATTTGCCCAACA AC	CsnB-HindIII-REV GTAAAGCTTCTATGCTTTCAAACCA GC
Hly (hemolysin)	Afu3g00590	secreted	Hly-BamHI-FOR CAGGATCCATGGCATCGGTCCAA GCT	Hly-KpnI-REV AGGGTACCTCAGCGCTTCCTTCCA AC

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Pst2	Afu1g02820	cytoplasm	Pst2-BamHI-FOR	Pst2-HindIII-REV
			CAGGATCCTACTCCATGTACGGCC	GCAAAGCTTTCACGCGAAGTTGAC
			AC	CTT

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Abbreviations: Aspf = *Aspergillus fumigatus* allergen, BamHI = *Bacillus amyloliquefaciens* restriction endonuclease HI, CipC = concanamycin-induced protein C, Eno = enolase, FOR = forward, HindIII = *Haemophilus influenzae* restriction endonuclease DIII, KpnI = *Klebsiella pneumoniae* restriction endonuclease I, MnSOD = manganese superoxide dismutase, PstI = *Providencia stuartii* restriction endonuclease I, Pst2 = oxidative stress response protein 2, REV = reverse.

**Table S2.** Preparation of stimulation tubes for whole blood stimulation. .

	$\alpha$ -CD28	$\alpha$ -CD49d	AfuLy	Protein antigens	PHA
<b>Concentration in the ready-to-use stimulation tube</b>	2 $\mu$ g/mL	2 $\mu$ g/mL	100 $\mu$ g/mL	60 $\mu$ g/mL	20 $\mu$ g/mL
<b>Final concentration after injection of 500 <math>\mu</math>L whole blood</b>	1 $\mu$ g/mL	1 $\mu$ g/mL	50 $\mu$ g/mL	30 $\mu$ g/mL	10 $\mu$ g/mL
<b>Unstimulated background control</b>	X	X			
<b>AfuLy stimulation</b>	X	X	X		
<b><i>Aspergillus</i> protein stimulation</b>	X	X		X	
<b>Positive control</b>					X

X indicates that the reagent was used for the respective stimulation mix. The volume of the stimulation mix in the ready-to-use tubes was adjusted to 500  $\mu$ L with RPMI 1640. Abbreviations:  $\alpha$  = anti, AfuLy = *Aspergillus fumigatus* mycelial lysate, CD = cluster of differentiation, PHA = phytohemagglutinin, RPMI = Roswell Park Memorial Institute.