



Comprehensive Assessment of Ameliorative Effects of AMF in Alleviating Abiotic Stress in Tomato Plants

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Abstract: Population growth and food necessity envisaged the dire need for supplementation to a larger community balance in food production. With the advent of the green revolution, agriculture witnessed the insurrection of horticultural fruit crops and field crops in enormous modes. Never-theless, chemical fertilizer usage foresees soil pollution and fertility loss. Utilization of biocontrol agents and plant growth promotion by microbial colonization enrooted significant restoration benefits. Constant reliability for healthy foods has been emancipated across the globe stressing high nutritive contents among indigenous field crops like tomato (*Solanum lycopersicum*). However, stress tolerance mechanisms and efficient abatement require deeper insights. The applicability of arbuscular mycorrhizal fungi (AMF) poses as an ultimate strategy to minimize the deleterious consequences of abiotic stress such as salt, drought, temperature and heavy metal stress sustainably. The rational modality employing the application of AMF is one of significant efforts to lessen cell damages under abiotic stress. The novelty of the compilation can be redressed to cohesive literature for combating stress. The literature review will provide agricultural scientists worldwide in providing a rational approach that can have possible implications in not only tomato but also other vegetable crops.

Keywords: Arubuscular mycorrhizha; salt stress; drought stress; heavy metal stress; temperature stress

1. Introduction

Tomato (Solanum lycopersicum) remains the most widely cultivated and largely consumed vegetables, next to potato universally. Tomato is regarded as a staple food crop to all sections of the economy irrelevant of developed or underdeveloped nations over a wide array of population. Large area under cultivation of nearly 4.2 million ha yielding 100 million tons per year enunciates the ultimate significance for the vegetable crop. Horticultural applicability of the fleshy vegetable nature of tomato has invigorated for its wide applications in food and feed ingredients. Major incorporations in salads and all varieties of dishes ranging from pasta, pizza, and other bakery products render them as an effective food supplement. Health-enhancing components in tomato comprise vitamins like C and E, beta-carotene, lycopene, flavonoids, and lutein. Macronutrient contents like potassium (K), phosphorous (P), magnesium (Mg), and calcium (Ca) along with trace elements composition of iron (Fe), manganese (Mn), zinc (Zn), and copper (Cu) stress the biocompatibility of staple and sustainable nature of tomato [1]. When foods containing tomato are subjugated for nutritional capabilities in arresting debilitating diseases like cancer and cardiovascular complications provide explicit strategies for nutraceutical benefits addressing food as medicine [2–4].

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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/). Tomato production and yield despite of variability patterns of region-specificity, soil fertility, and productivity are hampered by stress complications like drought, salt stress, temperature and environmental complexities especially heavy metal contamination accounting for nearly 70% loss eventually affecting food security. Subsequent stress emancipated through abiotic stress modalities addresses a large-scale hurdle in tomato plants growth and productivity throughout the world necessitating an effective alternate mode in efficient abatement of plant stress.

Precisely, abiotic stress factors are on an alarming rise emphasizing the requirement for arresting the devastating predicaments of salt stress, drought and soil contamination that accounts for vulnerability in sustainable agriculture of over 10% of arable land ensuing major yield loss of nearly 50% inclusive of key horticultural crops like tomato. Therefore, improvement of tolerance of plants against abiotic stresses remains a major focus of agricultural research. In day-to-day agriculture, the sustainability of crop productivity and yield are largely affected by abiotic stress thus corroborating to greater hazards both for the soils and the plant immunity. Climate change and ever-increasing industrialization are attributed to high range of abiotic stresses that affect yield and productivity. Thus, cost-effective strategies for the application of arbuscular mycorrhizal fungi (AMF) were assessed for biofertilization, commercialization, and commodity selection for ensuring maximal benefits. Many studies have demonstrated an ameliorative effect of AMF in alleviating abiotic stress in tomato plants (Figure 1). Holistically, there have been ample reviews about the AMF application for improvising tomato cultivation; the present review combines multi-disciplinary approaches in employing AMF and the potential benefits. Innovativeness can be confronted on the compilation of updated AMF profiles and their mechanistic molecular implications of AMF in abiotic stress amelioration for enhanced tomato productivity. Tolerating heat is attributed to many factors as enlisted in Figure 1. Heat stress poses serious consequences due to global warming. Reduction in electrolyte leakage, malondialdehyde (MDA), H2O2 and O2- levels along with escalating photosystem II (PS II), chlorophyll concentration, photosynthetic rate, tomato seedling growth rate, antioxidant levels, nutrient uptake, and biomass are discussed appropriately in the review. Similarly, the factors for drought tolerance (increased biomass, nutrient uptake, photosynthetic rate (Pn), stomatal conductance (Gs), water uptake, root hydraulic conductivity, root osmotic potentials, leaf water potentials, water use efficiency (WUE), stress hormone levels, and osmolytes accumulation), heavy metal tolerance (increase in root changes, nutritional status, heavy metal adsorption, secondary metabolites, glomalin, auxin, organic acids, antioxidants and nutrient uptake), and salt tolerance (increase in biomass, nutrient uptake, K⁺/Na⁺ ratio, Ca²⁺/Na⁺, Mg²⁺/Na⁺, Pn, Gs, transcription rate (Tr), water uptake, antioxidant status, osmolyte accumulation and diminished MDA content) that require deeper and intricate mechanism revelation are concisely analyzed.

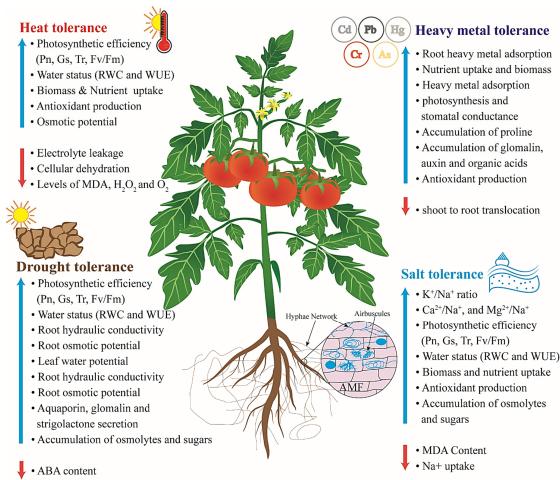


Figure 1. Effects of arbuscular mycorrhizal inoculation on abiotic stress alleviation in tomato.

2. Arbuscular Mycorrhizal Fungi

AMF, belonging to the Glomeromycota, are obligate biotrophs that establish mutualistic symbiotic associations, with most flowering plants revealing plant beneficiary attributes for a wide range of plants [5,6]. AMF principally are segregated into four classes inclusive under Glomerales, Archaeosporales, Paraglomerales, and Diversisporales, which are further sub-divided based on morphological parameters into 25 genera grouped under the sub-phylum [7]. Upon successful colonization, the fungi occupy a vast surface area involving extraradical hyphae networking like cloud embedded in a matrix of the root cortex. These hyphae upon increased accumulation render the soil mass enhancement and aid in augmented quenching of nutrient uptake stressing soil fertility and alternatively for plant growth promotion [8]. Nutrient exchange between the partners reveals the involvement of specialized structures termed arbuscules in the fungi that assist in nutrient exchange and efficient uptake. The mycorrhizal pathway renders the effective uptake of P, N and micronutrients that are not easily available for plant uptake from the soil through thick external mycelial mass surpassing the depletion zone enveloping the plant roots. In return, the plants' aid compatible, mutual benefits to the AMF in an obligate biotrophic association by providing carbohydrates as a food source [9-11], and life cycle management through lipids utilization [12]. AMF not only facilitates nutrient uptake but also plant resistance mechanisms against various stress mechanisms like salinity, drought, and temperature fluctuations together with heavy metal toxicity [8]. AMF also acts as an essential, facultative, obligate endosymbionts offering sustainable agriculture by ensuring ecosystem modulation and phytobiome engineering for successful crop production [13].

3. AMF-Mediated Growth and Yield Enhancement in Tomato

Soil fertility throughout the world depends on several soil parameters focusing plant growth nutrient availability that confronts to an increased usage of chemical fertilizers to capitalize huge benefits based on production and yield [14,15]. Many researchers and agriculturalists worldwide have proved AMF as an efficient growth regulator irrespective of nutrient availability ranging from poor to medium fertility. Further, AMF inoculation and optimistic response on tomato yield had adhered to critical effects in combinatorial benefits with (nitrogen phosphorus potassium) NPK fertilizers or varying P levels effectuating plant growth through enhancement of nutrient availability [16,17]. Even under low P availability, particular AMF for a definite crop provide the biofertilizer appropriateness for novel and innovative eco-sustainable practice for increased crop profitability for growers. These facts represent AMF inoculation under low P availability for enhanced P uptake, enriching biomass and thereby establishing high plant growth [18,19] and tissue level escalated deposition of P content [20].

Funneliformis mosseae altogether resulted in augmented total yield and slender rise in fruit size show pertinent effects of AMF on tomato production [21]. Application of commercial AMF inoculum (F. mosseae and Septoglomus constrictum) caused an increase in the overall height of tomato plants despite chemical fertilization in inoculated plants (48.4 cm plant-1) than non-inoculated plants (39.7 cm plant-1) emphasizing 18% gain. When the effect of mycorrhizal inoculation was studied for overall yield impact, 23% gain was found in inoculated plants (57.1 t ha⁻¹) compared to non-inoculated (43.93 t ha⁻¹) counterparts. Quantitative assessments of fruit numbers yield were escalated to 35 numbers in inoculated plants (30.6 fruit plant⁻¹) rather than non-inoculated ones (19.9 fruit plant⁻¹). The above results affirm that AMF inoculation resulted in fruit numbers regardless of the fertilization criterion in tomato. Hence, AMF that are newly introduced than the indigenous AMF unlike the conventional practices employing *Trichoderma* that reveal the contrary of inoculation benefits compared to AMF. Nevertheless, these results were derived from rigorous field trials indicating the need for further research in authenticating long-term field applications [21]. AMF inoculation had also positive impacts upon flowering that have inducing effects on overall yield consequently [17]. Rhizophagus intraradices inoculation articulated elevated P revival better than the plants which were not inoculated [17]. Rhi*zophagus etunicatum* under enriched P environment widened leaf surface area before flowering and total flower production per plant [16]. The results summarily show an increased flower and fruit production upon AMF inoculation in both the cultivars and escalated seed numbers per fruit in one cultivar. Thus, the impact of mycorrhizal infection upon plant reproduction, and vegetative growth will increase the significance of mycorrhizal research in agriculture, forestry, and land reclamation ensuring food production and security for meeting the issue of global population explosion.

4. Drought Stress

Among the abiotic stresses, drought stress accounts for water unavailability posing as the sole criteria in deciding the growth and development of tomato crop plants eventually causing reduction in nutrient uptake hampering the efficiency of production and yield [22]. The molecular mechanism of the stress-induced deficit can be attributed to obstacles in selective permeability which alter active transport characterized phenomenally to the reduction of transpiration rates [22], which advertently result in the culmination of physiology and metabolic pathways hampering that ultimately leads to lessened plant growth due to plant growth promotion, respiration vs. photosynthesis and hindered translocation of photo-assimilates. The effective way to combat water deficit stress has been focused on AMF symbiosis apart from agricultural methodologies and the ecosystem perspective for sustainable agricultural practices [23,24].

The mechanism of AMF symbiosis in mitigating water deficit corresponds to a combinatorial benefit emancipated by nutritional, physical, and cellular effects resulting in the primary effect [25]. The mechanisms that underlie the above said phenomenon are correlated to, 1) higher nutrients absorption [26], 2) articulation of water uptake by external hyphal mass by escalating hydraulic conductivity rendering high water status for the host plants [26], 3) osmotic adjustment [27], 4) increase in antioxidant activity [28], 5) modification of hormonal balance [29]. Non-availability of water in surplus amounts has key retardations ranging from gene expression, secondary metabolite synthesis which in turn affects yield and growth parameters on the tomato plants [30–33]. There are plentiful reports that establish the AMF symbiosis with plants in resisting drought havoc [34–38]. The variability patterns in the data can be assorted to the ubiquity of the host plants and noncorrelation of the AMF under various drought stresses based on region-specific anomalies [39].

4.1. Plant Growth and Nutrient Uptake

AM symbiosis and colonization has been amply reported to enhance plant growth and productivity despite drought stress and water deficit conditions [25]. Tomato seedlings inoculated with R. etunicatum produced higher dry biomass than non-mycorrhizal plants [40]. R. clarum encouraged higher growth in colonized tomato plants aerial biomass than in root biomass under drought stress [34] because AM colonization causes a proportionally greater allocation of carbohydrates to the shoot than to the root tissues [41]. AMF colonization also provides an increase in leaf surface area of inoculated plants and shoot biomass, wherein wild type tomato revealed a high rate of mycorrhizal population than mutant tomato plants under drought stress [42]. S. constrictum inoculation under water stress overpowered S. deserticola pretreatment indicating mycorrhizal colonization to have profound effects with 14–18% high root and shoot dry weight rather than the non-colonized plants [28]. Moreover, F. mosseae, R. irregulare, and R. etunicatum colonization showed similar results and thereby characterizing growth promotion as a positive regulator with the degree of stress mediated by collective AMF-inoculation [43]. The increase in root biomass eventually can be correlated as crucial since increased soil volume harnessing water intake under scarcity regulated by AMF.

Several studies have illustrated the varied plant responses to drought owing to the specificity of AMF to plant roots [30-43]. Randomized metabolomic analysis in tomato roots colonized by three AM fungi of different genera revealed that some responses to drought and salt stress were commonly mediated by most of the AM fungi, whereas some were specifically associated with single isolates. Single AMF inoculation on comparison with combinatorial assessment of three AMF species in studying tomato tolerance in mitigating water limitation showed variation benefits. Two AM fungal inocula (Myc_Rhizo and MULTISTRAIN) revealed a consistent difference compared to mixed inoculums that showed root inoculation pattern variation [44]. Consequently, mixed inocula (e.g., MUL-TISTRAIN vs. Myc_Rhizo) both in control and drought plants depicted varied significance affirming species-specificity in affecting physiological traits [44]. F. mosseae and R. intraradices were assessed and proved for multitude of benefits concerning species specificity in plant-microbe symbiotic interactions both above and below ground levels [45]. This effect can be attributed to increasing in internodes and its ratio for plant growth promotion in R. intraradices and considerably no positive implications upon F. mosseae inoculation showing discrepancy [29]. The discrepancy with the research by Rivero et al. [43] can be acclimatized to a variety of cultivars impacting varied growth results. So, from these results, we can conclude that indigenous AMF isolated along the individual regions owing to environmental implications will have prominent results rather than introducing a new family of AMF than ad hoc families could yield considerable results [46,47]. AM symbiosis enables mycorrhiza-host plant-mediated drought tolerance based on nutritional availability showing that even under low nutrition, mycorrhizal colonization aids in host plant nutrition and survival [48] (Table 1).

Mycorrhizal Species	Effect	Reference
Septoglomus deserticola and Septoglomus constrictum	Increased root and shoot dry weight. Increased stomatal conductance, relative water content and antioxidant enzyme activities	[28]
<i>Funneliformis mosseae</i> and <i>Rhizophagus intraradices</i>	Increased photosynthetic rate and water use efficiency, reduction in ABA, higher proline accumulation,	[29]
Glomus clarum	Increased aerial biomass, photosynthetic activity and stomatal conductance	[34]
Rhizophagus intraradices	Increased shoot biomass, number of flowers, fruits, N and P uptake.	[35]
Rhizophagus intraradices	Increased shoot dry weight, stomatal conductance, photosystem II efficiency, and abscisic acid	[38]
Rhizophagus intraradices	Lower transpiration rate and increased water use efficiency, increased shoot biomass and P concentration	[42]
Funneliformis mosseae, Rhizoglomus irregulare and Rhizophagus etunicatum	Increased shoot and root dry weight.	[43]
<i>Funneliformis mosseae</i> and <i>Rhizophagus intraradices</i>	<i>Rhizophagus intraradices</i> efficient in water use efficiency, <i>F. mosseae</i> in volatile organic compounds	[45]

Table 1. Application of arbuscular mycorrhizal fungi on drought stress alleviation.

R. intraradices colonization under varied drought stress intensities resulted in increased N and P levels in both onion and tomato plants, wherein the former host plant was able to survive due to less negative water potential. The latter host plant depicted improved P nutrition along with higher N content in both roots and shoots. The mode of action corresponds to an increase in N demand impeding the mobility of NO₃ ions under water deficit altering nutritional status during mycorrhizal colonization [35]. Further, R. *intraradices* inoculation resulted in a 14% increase in shoot P concentration of wild type tomato plants under well-watered conditions and a 23% increase under drought stress [42]. Pi uptake, transfer and delivery have been largely investigated in AM roots, resulting in the characterization of a symbiotic inorganic phosphate (Pi) uptake pathway [49–51]. Eight PHT1 genes regulate tomato roots uptake of Pi in tomato [52] and PT genes are synthesized inductively by mycorrhiza [53,54]. AMF-induced LePT3, LePT4, and LePT5 PT genes were detected in water surplus conditions as evident from previous research indicating the differential regulation of PT genes under drought stress [53–55]. Under water stress, they found a different regulation of the considered PT genes. Particularly, it is worth noting the opposite trend for the two genes involved in the direct Pi uptake from soil (LePT1 and LePT2), independently from the presence of the AM fungus. Among the AM-inducible PT genes, LePT3 seems not involved in response to water deficit [45]. LePT4 and *LePT5* transcript levels both increased under water deficit with highest values in *F*. mosseae, in agreement with the higher P content found in the leaves of these plants in respect to control ones. Overall results suggest a major role played by *LePT2* and *LePT4* in promoting tolerance to water deficit, particularly under a severe condition, although this is dependent on the fungal species.

Multi-omics and culture-omics have been utilized largely nowadays for deriving at better plant varieties in sustainable agriculture. Tomato root transcriptome analysis revealed that cytochrome P450 genes were mainly up-regulated in *R. intraradices* under drought stress [56]. Handa et al., [57] showed the importance of cytochrome P450 in the synthesis of sterols for membrane biogenesis during arbuscule formation, it could be inferred that their over expression reflects a change in fungal development under water deficit. Other most highly up-regulated fungal genes include a "conidiation protein 6" domain and domains involved in signaling transduction were significantly influenced by

the WS treatment, being mainly up-regulated. Fungal conidiation can be induced by nutrient deprivation or mycelium desiccation, under drought stress. Moreover, they found three glutathione S-transferases (GSTs) were overexpressed in *R. intraradices* upon water stress. GSTs are recognized as a candidate in the cell protection from oxidative damage under drought stress both in tomato and fungi [56,58]. Such analysis for stress management will provide deeper insights for symbiotic management for multiple stress modalities.

4.2. Photosynthesis and Water Status

Tolerance elicited by AMF symbiosis in enhancing plants efficiency in fighting water insufficiency comprise increased water intake through Gs [39], efficient water use and prevention of reactive oxygen species production to reduce oxidative stress damages encompassing various antioxidant mechanisms, both enzymatically and non-enzymatically [59,60]. Affirmatively, a better performance of PS II was correlated to AM tomato plant interaction during water deficit and surplus [38]. Under field conditions also the symbiosis confirmed higher leaf relative water content (RWC) irrespective of various drought stress conditions in mycorrhizal inoculated tomato plants [35]. In a similar study, escalated photosynthetic activity, Gs levels in well-watered conditions and an increase in Pn and Gs values under stressed conditions provide clues for the efficiency of mycorrhizal infection in tomato plant growth upsurge. This was proved by the fact that recovery under fully watered conditions yielded neither Pn nor Gs [34]. The enhanced Gs probably caused an increase in intracellular CO₂, which could also enhance the photosynthetic activity of the infected plants. These increases in the photosynthetic rate and leaf conductance were accompanied by the better uptake of water by the roots, as can be seen from the root hydraulic conductivity values. The results confirm that leaf water potential remains higher in mycorrhizal plants under drought conditions because of higher water uptake.

In a more recent study, *R. intraradices* colonization caused high Tr of mutant plants for abscisic acid (ABA) genes under well-watered conditions (p < 0.05) but not for Tr of wild type plants under drought stress. On the contrary, WUE of wild type plants under drought stress (p < 0.05) increased. Nevertheless, WUE under well-watered conditions did not show a considerable rise. Whereas Gs of non-inoculated tomato plants (p < 0.05) show a consequent increase when compared to the negative effect on colonized tomato plants. Drought significantly decreased the photosynthetic rate of non-inoculated plants (p < p0.01), but not for inoculated plants. Further, mycorrhizal inoculation depicted positive outcomes on the photosynthetic efficiency on two genotypes subjected to drought stress (p < 0.05) [42]. AM plants under drought stress substantially enhanced the Gs (increased by 62% with S. deserticola and by 200% with S. constrictum) in comparison to non-AM plants. Besides, leaf water potential and relative water content were higher in the presence of mycorrhiza, suggesting that AM plants could improve the water status under water deficit. Application of S. constrictum triggered a higher Fv/Fm in plants compared with non-AM and S. deserticola colonized plants under drought stress, which supports the results of Ruiz-Lozano et al. [38] that showed similar results under moderate and severe drought conditions [28]. AM plants inoculation illustrated efficient PS II and plant growth right from early inoculation than non-AM plants during drought stress [38]. Chitarra et al. [29] found that R. intraradices inoculation significantly increased the photosynthetic rate and water use efficiency. As a result of higher water status and improved nutritional status, mycorrhizal tomato plants have produced higher biomass under varying intensities of drought stress. The improved nutritional status and relative water content caused by mycorrhizal colonization would have alleviated drought impacts and promoted tomato fruit production under varying intensities of drought stress. Because mycorrhizal treatments consistently increased fruit yields under varying intensities of drought, WUE of AM plants were much higher than the control plants [29].

Also, the fungal external hyphae can penetrate pores that are beyond the root zone or inaccessible for the root systems, thus allowing colonized roots to access additional water reservoir in the soil [39]. Therefore, the differences found between mycorrhizal and non-mycorrhizal tomato plants were not due to osmotic adjustment, and the enhanced development of extraradical hyphae in soil containing mycorrhizal plants could be responsible for improving the root hydraulic conductivity and reducing drought stress in infected tomato plants [61]. Infection plays a pivotal role in increased water transport by hyphae directly than non-infected roots due to increased hydraulic conductivity and transpiration rate. Hydraulic conductivity transmits and sends the hydraulic message from the roots to the host plants as affirmed by high Gs levels [62].

4.3. Modification of Hormonal Balance

Furthermore, AM fungi modulated root hydraulic properties, regulated patterns of expression of aquaporin genes [63,64], induced changes in phytohormone ABA, JA [29,60], and altered proline content in the host plants [29]. AM symbiosis is critically mediated by ABA for successful regulation [65], on the other hand, AMF also regulates the ABA signaling pathway [66]. However, few evidences were available as to the interplay of AM symbiosis with the functional genes in ABA signal transduction under drought stress. In Xu et al.'s [42] study the ABA-deficient tomato mutant *not*, together with its wild-type was adopted to reveal the influence of AM symbiosis on 14-3-3 genes in response to drought stress. Expression variant profiling revealed high expression of 14-3-3 genes *TFT2*, *TFT3* and *TFT5*, *TFT7*, *TFT9* and *TFT10* in wild type plants and *not* plants, respectively. These genes further mediate the stomatal behavior, subsequently improving plant WUE and drought tolerance.

Neither drought nor mycorrhizal inoculation influenced the ABA concentrations in not leaves [42]. Furthermore, AM-induced alterations such as hormonal changes, improvement of gas exchange, regulation of osmotic adjustment, and mediated aquaporin genes in host plant as well as in fungal symbiont can also contribute to the host water status [25,29,64]. The expression of LeNCED1 was not regulated by drought or by ABA in tomato plants under drought stress [36,57,67]. However, the expression pattern of the ABA-responsive gene Le4 perfectly matched with that of ABA levels, indicating an efficient activation of the ABA signaling pathway under drought. In addition to its role as a "stress phytohormone," ABA is also important for symbiosis establishment and functioning [65,68]. Therefore, the increased ABA levels in stressed plants would serve not only to promote tolerance against stresses in non-AM and AM plants but also to enhance and maintain the symbiosis in mycorrhizal plants. Hormonal results, together with those of other physiological parameters, support that AM symbiosis improves plant fitness under water-related stress conditions. Although ABA has not been detected in Rivero et al. [43] study, instead they found higher concentrations of ABA-GE in mycorrhizal roots, a reservoir for the rapid production of active ABA. Same results were found in Chitarra et al. [29], under severe WS conditions (21.3 MPa), tomato AMF inoculated plants showed significantly lower levels of ABA in roots compared with control plants. The expression of the ABA biosynthetic gene LeNCED1 was low and constant in the root of NS plants, without significant changes due to the presence of AM symbionts. Under drought stress, a strong increase in the transcription of this gene was observed in control plants, while in AM-colonized plants under drought stress, LeNCED1 was only up-regulated in F. mosseae plants, whereas it did not undergo significant expression changes in roots from the R. intraradices inoculated plants [29]. At the same time, the gene was also vague in showing less regulation because AM colonization could vary in yielding significant results based on the experimental setup and system variations [38].

ABA biosynthetic gene *SINCED* was downregulated under *S. constrictum* colonization under drought conditions. Nevertheless, the levels were normal and optimized without stress [58]. Conversely, significantly higher expression levels of JA-biosynthetic gene *SILOXD* in roots colonized by *S. constrictum* were observed under non-stress and drought conditions. There are ample reports that confirm ABA in structuring and maintaining AMF symbiosis [65,68] at the same time, JA plays a pivotal role and is involved in organizing and developing AM symbiosis [69,70]. Under drought stress and high salinity conditions, water loss is prevented by closure of stomata by ABA production [71], therefore down-regulated *SINCED* in mycorrhizal plants relates to higher stomatal opening and conductance and elevated water status of the host plants. The above reasons can be enlisted for stress tolerance in AMF inoculations. Increased production of strigolactones has been reported in lettuce and tomato plants under drought in the presence of *R. irregularis,* indicating that AMF symbiosis induces striglolactones biosynthesis [38].

Expression profiling for *SICCD7* and *SICCD8* genes responsible for striglolactones biosynthesis revealed upregulation of *SICCD7* under drought stress without *SICCD8* gene alterations. Strigolactones are increased upon drought stress to promote AM symbiosis in the regulation of host plant development. But further research is needed to confirm the intrinsic correlation between striglolactones biosynthesis and AM symbiosis under drought conditions. Striglolactones – ABA crosstalk in plant retorting to water stress show the mechanistic background for colonization criteria for AM symbiosis. *F. mosseae* inoculated plants show high proline values under inoculation in drought plants rather than control plants. AM symbiosis could not be relied on drought conditions, as varied expressions of proline content in treated plants than in control plants remains clueless [37,72–74]. Our results, showing a higher content of proline in AM inoculated plants during drought concerning control plants, could suggest a better resistance of colonized plants to drought.

Physiological mechanisms in averting drought stress are regulated by the expression of genes encoding aquaporin proteins (AQPs) that controls water movement in plants [75]. Furthermore, AM symbiosis can have potential implications in regulation of the expression profiles of AQP genes to enhance root hydraulic conductivity as well as the plant water status and tolerance under water deficiency. Currently, five major subfamilies of AQPs have been characterized based on sequence similarity [76]. Drought affected the transcriptional pattern of these AQP genes differently: *LeNIP3;1* was over-expressed in AMF-inoculated plants, particularly in those inoculated with *F. mosseae;* conversely, *LePIP1;1* and *LeTIP2;3* were down-regulated in both AMF inoculated and control plants [29]. AM fungal AQPs can also have a role in drought tolerance during AM symbiosis, the expression profiling for two *R. intraradices* AQP genes (*RiAQPF1* and *RiAQPF2*) indicated up-regulation of *RiAQPF2* gene under drought suggesting putative roles in drought stress response [29]. The simultaneous induction of both fungal and plant AQP genes also affirm that the two symbionts strictly cooperate to regulate the mycorrhizal drought stress response resulting in enhanced physiological performance [29].

5. Salt stress

Salinization of soil is a serious agricultural and eco-environmental problem and is increasing steadily in many parts of the world, in arid and semi-arid areas. Salinity inhibits plant growth by lowering soil water potential and increasing the potentially toxic ions (Na⁺ and Cl⁻), which in turn lead to water deficit and nutritional imbalance. Increasing Na⁺ concentrations disturb the nutrient balance, osmotic regulation, and cause ion toxicity [77,78]. Salinity stress significantly reduced the root, stem, and leaf dry matter and leaf area compared with the control treatment due to direct effects of ion toxicity or indirect effects in osmotic imbalance due to increase in saline ions between soil and plant. In the areas with an optimal climate for tomato cultivation, salinity is a serious constraint [79]. Salinity irrevocably affects plant length, dry weight, and growth of tomato plants. Salinity also reduced the fresh and dry shoot and root weight of tomato [80,81]. The reduction of dry weights due to increased salinity may be a result of a combination of osmotic and specific ion effects of Cl and Na [82]. Salinity increased the absorption of Na⁺, whereas it decreased N, P, K, and Mg uptake, and K⁺/Na⁺ ratio. AMF has been extensively identified as a sensible fungus in salinity adversities on host plants under salt stress [83] by negatively regulating salinity impacts by mycorrhizal symbiosis [84]. Nevertheless, many literatures show that AMF can enhance salt tolerance of many plants and some physiological changes occurred in AM symbiosis under salt stress [80,82,85]. In recent years, studies indicated that AMF can increase plant growth and uptake of nutrients, decrease yield losses of tomato under saline conditions and improve the salt tolerance of tomato [80,82,86,87].

5.1. Tomato Growth and Nutrient Uptake

Previously, some studies have indicated that AMF can enhance the plant growth and uptake of nutrients, decrease yield losses of tomato under saline conditions, and improve salt tolerance of tomato [86–88]. Colonization with F. mosseae significantly improved dry matter and leaf area in the salt-stressed tomato plants. This effect of AM on the dry matter was more pronounced in aerial biomass than in root biomass which may be because arbuscular mycorrhizal colonization was proportionally greater in roots than in shoots concerning the distribution of carbohydrates. Under salinity, R. intraradices -mediated growth stimulation was higher in more salt-tolerant tomato cultivar, Piazar than sensitive cultivar, Behta. High salinity reduced the dry matter production of plants in both cultivars [89]. Interestingly, soil salinity accounts for the AMF inoculated tomato plants to show increased dry matter (0.098 g plant-1) content than non-inoculated seedlings (0.082 g plant-1) [90]. Beneficial effects of AM fungi have been appropriated not to root biomass rather on above-ground biomass [83]. P nutrition was partially linked to better growth under salinity due to mycorrhizal inoculation in tomato plants [91]. AMF-enriched plants grown in saline soils (1.5 mS cm⁻¹ to 6.0 mS cm⁻¹) showed amplified fruit yield, quality, mineral composition, and antioxidant status. As for fruit mineral composition [88], the application of AMF-based formulates resulted in higher content of K, Ca, Mg, P, S, and NO₃ compared to the control, whereas Cl was not significantly affected by the beneficial microorganisms.

AMF inoculation showed low Na levels in fruits apart from non-inoculated counterparts due to concomitant accretion of K, Ca, and Mg. N, P, Mg, Ca, Mn, and Fe uptake was subsequently high in tomato seedlings inoculated with AMF under salt stress than non-inoculated ones. In other research [15], plants inoculated with R. intraradices produced fruits with a higher content of K, Ca, P, and Zn compared to control plants. Moreover, mycorrhization increased P, K, and Ca contents more in salt-tolerant Piazar than salt-sensitive Behta [82]. Different mycorrhizal responsiveness of cultivars could be the result of plant- and/or AMF-related mechanisms. The enhancement of plant P and K uptake by AMF have been reported and was considered one of the main reasons for the amelioration of growth in salt-affected plants colonized by AMF [92]. Earlier studies found that the improved growth of mycorrhizal plants in saline conditions is highly related to the mycorrhizal-mediated enhancement of host plant P nutrition [93]. AMF in salt-stress is responsible for increased P uptake due to integral vacuolar membranes and compartmentalization Na⁺ ions regardless of integral P in host plants [94]. Recently, Ebrahim et al. [85] found that the AMF inoculation (R. fasciculatus) increased the accumulation of N, P, K, and Mg, it reduced the Na⁺ concentration. Moreover, AMF can increase the accumulation of osmotic regulators such as soluble sugar [95], proline, betaine, and polyamines [96,97] in plants under salt stress. Kong et al. [98] confirmed that the tomato plants and mixed inoculation of AMF (R. clarum and R. intraradices) formed a symbiotic relationship that significantly improved the growth of plants and increased the concentration of vitamin C, soluble sugar and lycopene in the tomato fruit (Table 2). Compared with the control, the single fruit weight and the yield per plant in the AMF treatment were significantly increased. AMF promoted tomato plants absorption of N and reduced the absorption of Na⁺. Under NaCl salinity, R. etunicatum than other AMF counterparts F. mosseae and R. irregular enhanced plant growth promotion mainly the root growth [41].

Level of Salinity	Mycorrhizal Species	Effect	Reference
NaCl solution (0, 0.5 and 1%) 4.2 and 7.1 dS m ⁻¹	Funneliformis mosseae	Reduced MDA content and increased antioxidant enzymes SOD, POD, APX and CAT	[80]
without salt (EC= 0.63 dS m ⁻¹), with low (EC=5 dS m ⁻¹), or high (EC=10 dS m ⁻¹) salinity.	Rhizophagus intraradices	Increased growth, stomatal conductance, photosynthetic activity, proline and ROS enzymes	[82]
1.4 (control), 4.9 (medium) and 7.1 dS m ⁻¹ (high salt stress)	Funneliformis mosseae	Shoot dry matter, yield and leaf area were higher, the contents of P, K, Zn, Cu, and Fe were higher	[87]
1.5, 3.0, 4.5, 6.0 mS cm ⁻¹ EC	Rhizophagus etunicatum, Funneliformis mosseae, Glomus aggregatum, Rhizophagus intraradices	s Increased yield and size of fruits	[88]
0, 50 and 100 mM NaCl.	Funneliformis mosseae	Increased growth, leaf area, cholorphyll, fruit fresh weight and yield. Increased P and K uptake. Increased SOD, CAT, and POD reduced MDA content	[89]
0, 50, and 100 mM, NaCl	Mixtures of <i>Glomus</i> sp.	Increased root biomass, P, N, Ca uptake	[90]
EC of 4.56 dS m ⁻¹	Glomus clarum and Rhizophagus intraradices	Increased soluble sugar, proline accumulation and vitamin C. Increased the chlorophyll concentration, Pn, Gs and Tr of plants.	[98]

Table 2. Application of arbuscular mycorrhizal fungi on salt stress alleviation.

5.2. K+/Na+ Ratio

Because reduced growth under salinity is partially caused by ion imbalances and/or non-availability of nutrient ions due to their competition with major ions (Na+ and Cl-) in the soil [99], the sustained growth of AMF+ plants under salinity is partially based on improved uptake of nutrients and maintaining favorable ionic ratios [100]. AMF inoculation can maintain the K⁺/Na⁺ balance under salt stress. AMF restricted the transport of Na from roots to shoots in plants, which increased K⁺/Na⁺, Ca²⁺/Na⁺, and Mg²⁺/Na⁺ in leaves and stems, in addition to protecting the photosynthetic organs from damage. Certain ion ratios, such as K/Na, are accepted indicators for the evaluation of salinity tolerance in tomato cultivars. Hajiboland et al. [82] reaffirmed that AMF inoculation depicted high content of K/Na and Ca/Na levels equally in roots and shoots than non-inoculated plants. Mycorrhizal F. mosseae plants had a higher concentration of K at both salinity levels [89]. Na concentration was lower in mycorrhizal than non-mycorrhizal plants regardless of the salinity level. Growth improvement was observed in AMF-inoculated tomato plants under salinity conditions and was mainly associated with ionic factors such as higher K concentration and K⁺/Na⁺ ratio [101]. Indeed, although K concentration and the K⁺/Na⁺ ratio in leaves were positively correlated with the growth parameters, those correlations were especially apparent in the AMF-inoculated plants. The concentration of Na⁺ in roots was reduced in mycorrhizal treatments, while the accumulation of Na⁺ in leaves was significantly lower in plants colonized by R. etunicatum [43]. Notably, R. etunicatum was the only salt-adapted AMF compared to F. mosseae, R. irregulare. Higher K accumulation by mycorrhizal plants in saline soil could be beneficial by maintaining a high K/Na ratio and by influencing the ionic balance of the cytoplasm or Na efflux from plants. The higher K accumulation in AM plants under salt stress conditions resulted in maintaining a high K/Na ratio, preventing the disruption of metabolic processes and the inhibition of protein synthesis [82,102]. AQP gene (GintAQP1) in R. intraradices expressed upon homeostasis relying on two Na⁺/H⁺ antiporters in salinity stressed tomato plants depict insignificant expression levels of LeNHX1 and LeNHX2 genes under AMF symbiosis [103].

5.3. Photosynthesis and Water Status

AMF also increased the chlorophyll concentration, Pn, Gs, and Tr of plants. AMF infection effectively improves plant photosynthetic capacity [104,105], and salt stress-induced stomatal closure [106]. Colonization with AMF enhanced CO₂ assimilation rate of salt-affected tomato plants. R. intraradices-inoculated tomato plants had greater transpiration and stomatal conductance. AMF inoculation consequently lowered WUE than noninoculated counterparts. R. fasciculatus role in enhancing chlorophyll concentration can be ascribed to increase in N and Mg concentrations and reduction in absorption of sodium in tomato leaves. Light-harvesting protein complexes variability patterns can be corroborated to salinity stress tolerance and adaptations based on the rise in Chl a/b ratio [85]. AMF also enable processing tomato plants to accumulate more proline and to reduce membrane peroxidation damage [98]. Oxylipins such as the phytohormone JA and derivatives reduced the negative effect of drought and salinity stress on plant fitness when exogenously applied [107]. The AM colonization resulted in the accumulation of JA; the higher the JA concentration in the roots colonized by *R. etunicatum*, the more efficient the bio-protector against salt stress. ABA glucosyl ester (ABA-GE), b-ionone, and blumenol C glucoside belonging to the carotenoid pathway are other metabolites quantified in the R. *etunicatum*-colonized AM roots apart from oxylipins [43].

5.4. Antioxidant Enzymes

Reactive oxygen species (ROS) such as superoxide radical (O_2^{-}), hydrogen peroxide (H_2O_2) , hydroxyl radical (OH), and singlet oxygen (O_2^-) are generated in plants during the salt stress. The oxidative stress results in oxidative stress-mediated damage to the vital cell components like proteins, nucleic acids, and lipids. This led to the change in selective permeability of bio-membranes and thereby membrane leakage and change in the activity of enzymes bound to the membrane occurred. The induction of ROS-scavenging enzymes, such as superoxide dismutase (SOD), peroxidase (POD), ascorbate peroxidase (APX), and catalase (CAT) is the most common mechanism for detoxifying ROS synthesized during stress responses. AMF can also reduce the level of ROS by increasing the activities of antioxidant enzymes, and other enzymes under stressful conditions. The activity of antioxidant enzymes (SOD, CAT, POD, and APX) increased in F. mosseae-inoculated tomato plants under salinity (except SOD and CAT activity at 100 mM NaCl). However, this higher activity did not provide enough protection against ROS, as judged by simultaneous enhancement of MDA [89]. Under saline conditions, ionic balance involved in photochemical reactions and gaseous exchange is regulated by enhanced proline concentration, lowered lipid peroxidation, and increased SOD activity [82].

Inoculation with AMF increased the activity of antioxidant enzymes and protein contents of salt-affected plants. AMF colonization was accompanied by an enhancement of the activity of APX, CAT, and SOD in both salt-affected and control tomato plants. Besides, inoculation with AMF caused increase in the proline concentrations and reduction of hydrogen peroxide and MDA [82]. The role of ROS-scavenging enzymes in AM tomato under salt stress showed the positive effects of AMF on tomato growth [80], which corroborates to a similar study wherein lower leakage value was found for macromolecules and MDA content in AM plants under salt or saltless condition. The lower leakage values for macromolecules and MDA content illustrates lower cell membrane damage or higher salt tolerance [87]. AM inoculation-enhanced tomato salt tolerance assessment shows that cell membrane damage was lower in AM symbiosis compared to non-AM plants and SOD, APX, and POD activity was contributed to protecting the plant from salinity injury, the induced SOD, APX, and POD activity in AM symbiosis may be an important mechanism to improve salt resistance of AM plants [80].

6. Temperature Stress

The optimal temperature for tomato cultivation ranges from 20 to 30 °C. Stress due to heat will have profound implications on growth parameters and result in major threats in a plethora of plant parameters inducted by adversities in physiological, biochemical, morphological, anatomical, and genetic responses in plants [108], eventually depressing yield and the quality of crops. Tomato usually prefers thermophilic environment stating that the plant is susceptible to various temperature conditions. Physiology and metabolic regulations are the key components in growth and improvement in plants which are impeded by a reduction in temperature. Low-temperature conditions will affect the normalized distribution, water potentials, and photosynthetic ability. The entire scenario can be associated with stomatal conductance irregularities by altercations in hydraulic conductance [109,110]. Also, low temperature decreases the capacity and efficiency of photosynthesis through the change in pigment composition, a decline in chlorophyll fluorescence, and impaired chloroplastic development [105]. When the plant is subjected to low-temperature stress, the cell membrane is first affected with increased membrane permeability. At the same time, a variety of ROS, such as O_2^- , OH, and H_2O_2 , are induced, causing a loss in the balance between production and scavenging in the cell or organism, which causes membrane lipid peroxidation [111].

Most of the cultivated processing tomato genotypes are sensitive to low temperatures (0–12 °C) in all growth stages [112]. The negative effects of chilling are more remarkable during germination and at the seedling stage [113]. Chilling damage could limit processing tomato growth and production in an open field. Inoculation with F. mossae was the most effective treatment in reducing electrolyte leakage while increasing the efficacy of PSII and regrowth capacity of seedlings exposed to severe chilling stress (1 °C) [114]. F. mosseae has been proved responsible for escalations in redox compounds enrooting stability of tomato roots in optimal temperature conditions [115]. The phenomenon underlying is being reinstated of the chilling stress in seedlings to overcome the temperature complications. Similarly, they also proved that AMF-inoculated tomato seedlings exhibited significantly higher fresh weight and dry weight than non-AMF control plants under both control (25/15 °C) and low temperature (8 °C/4 °C) treatments. Under chilling stress, AMF inoculation significantly reduced the level of MDA, H₂O₂, and O₂⁻ than in the non-AMF control because of calcium precipitation in tomato roots apoplast and vacuole. Furthermore, AMF inoculation induced activities of antioxidant enzymes and transcripts of related genes under chilling stress.

AM fungi have largely been studied to tackle low temperature in addition to boosting tolerance to stress mechanisms both biotic and abiotic together with plant growth promotion [110,116]. F. mosseae grow better than non-mycorrhizal plants under low temperature [117]. A low chlorophyll concentration in non-mycorrhizal and low temperature-stressed tomato plant leaves indicated a reduced synthesis rate of chlorophyll and an increase in chlorophyll breakdown. Mycorrhiza and low temperature were reported to modify the protein content in tomato, whereas an increase in soluble proteins was related to lowtemperature tolerance [117]. F. mosseae could enhance chlorophyll concentration of tomato leaves at low temperature, which was in agreement with the results of wheat and maize under cold stress by Paradis et al. [118] and Zhu et al. [110]. Proline agglomeration can cause deleterious effects of temperature stress; mycorrhizal plants evidenced lower accumulation than non-mycorrhizal plants. MDA content in mycorrhizal plants remained lower than that in non-mycorrhizal plants indicating that leaf proline can have indirect consequences in addressing osmotic imbalance and lipid peroxidation mechanism alleviation by AM fungi will have protective effects than lowering proline accumulation in the tomato leaves [117]. They also reported the activities of SOD, POD, and APX in AM tomato plants were higher than the non-AM plants under low-temperature stress. An increase in MDA level and H2O2 accumulation in the leaves of stressed plants, although the levels of MDA and H₂O₂ were lower in AM plants than in non-AM plants according to the different stresses [28]. Similar results were obtained from *R. versiforme* inoculation during drought [119] and *R. intraradices* inoculation under salt stress salinity [82], show similar consistent results for leaves and roots of citrus seedlings. All these results summarily prove AM colonization is responsible for enhancing the survival rates under extreme temperature by lipid peroxidation and amelioration by ROS deleterious effects. Recently, Haddidi et al. [120] showed the beneficial role of AMF symbiosis in the alleviation of ROS accumulation caused by combined drought and heat, and combined drought and heat shock stress. They revealed that the accumulation of H₂O₂ and lipid peroxidation was much higher in leaves than in roots. However, inoculation with different AMF strains, and especially, *F. moseae*, could enhance tomato plants' tolerance by lowering H₂O₂ and MDA content, and changed the activities of antioxidant enzymes (Table 3).

Type of Temperature	Mycorrhizal Species	Effect	Reference
42 °C	Septoglomus deserticola and Septoglomus constrictum	Decreasing the lipid peroxidation, hydrogen peroxide level and improving leaf and root antioxidant enzyme activities	[28]
1 °C	Funneliformis mosseae	Reduced the cell membrane injuries in term of electrolytic leakage and efficiency of photosystem II	[114]
8 °C/4 °C	Funneliformis mosseae	induced activities of antioxidant enzymes, significantly reduced the level of malondialdehyde (MDA), H2O2, and O2- along with increased calcium precipitates in the apoplast and vacuole of root cells	[115]
8 °C	Funneliformis mosseae	Decreased MDA content in leaves. The contents of photosynthetic pigments, sugars and soluble protein in leaves were higher, but leaf proline content was lower and increased the activities of antioxidant enzymes	[117]
45 °C	Rhizophagus irregularis, Funneliformis mosseae, and Funneliformis coronatum	A decrease in hydrogen peroxide and malondialdehyde content and increased antioxidant enzyme activities	[120]

Table 3. Application of arbuscular mycorrhizal fungi on temperature stress alleviation.

7. Heavy Metal Stress

Heavy metals (HMs) such as arsenic (As), lead (Pb), mercury (Hg), cadmium (Cd), chromium (Cr), nickel (Ni) and copper (Cu) are toxic based on their toxic effects on human beings [121]. Not all metals are harmful to possess severe and devastating health effects, rather micronutrients potential pose some of the metals to have immense biological properties for the welfare of human beings and agricultural potentialities. Whereas non-essential metals have irrevocable consequences on the environment and ecosystem thereby soil health and fertility are affected badly. The food chain and food web are massively affected posing severe threats to human health by long-term effects on plants and food products. Biological methods of plants-microbe's interactions can be used for the bioremediation of contaminated areas with heavy metals. AM fungi colonization and benefits do not depend on soil pollution even in the extreme terrestrial environment [122]. AMF can be adapted to various environmental conditions even in metal-contaminated sites [123] (Table 4). AMF-colonizing plant roots will alter the morphological parameters of plant roots and nutritional status of the soil [124,125], mycelial culmination and adsorption of heavy metals [126], and secondary metabolites-mediated mitigation of toxic HMs concentration and metal resistance, finally aiding efficient growth through the accumulation of key molecules like glomalin, organic acids, and auxin [127,128]. Tomato plants subjected to Cd stress showed reduced growth in terms of length and weight [129]. Cd alters growth, triggers leaf necrosis and impedes cell division, elongation, and reduced growth attributes in tomato [130]. Chlorophyll content in AMF-inoculated tomato plants and subsequent recovery of Cd-stressed plants may have effects due to the effect of AMF on magnesium uptake. Proline and sugar accumulation help plants to maintain cellular water potential well below that of the soil solution [129]. Cd stress has positive attributions for proline accumulation in tomato [131] which emancipates a positive correlation between AMF symbiosis and proline accumulation under severe stress. The role of proline together with AM inoculation suggests increased levels of scavenging enzymes like SOD, CAT, POD, GR, and APX under Cd stress through antioxidant quenching of ROS [129]. F. mosseae when inoculated in Cd contaminated tomato plants considerably enhanced biomass of shoots and roots. These effects were substantiated to Cd sequestration in non-vital mycorrhizal roots and Cd immobilization due to microbial adsorption, accumulation, and/or chelation by rhizosphere beneficial effects [132]. Further, F. mosseae drastically reduced toxic effectuations upon cadmium accumulation. The results depict that pre-transplant inoculation incited plant dry matter accumulation than post-transplant inoculation despite cadmium contamination richness in the soil [133]. On the contrary, AM inoculation did not elicit considerable alleviation of Cd contamination due to intra-radical compartmentalization in root cell vacuoles which results in translocation of Cd in aerial roots resulting in normal growth and productivity. Furthermore, tomato grafting with Maxifort rootstock minimized Cd-induced oxidative injury hydrogen peroxide, lipid peroxidation, and electrolyte leakage in tomato leaves and thereby promoting the growth performance of tomato plants. These results are attributed to alterations in plant nutritional status, photosynthetic pigments, the photochemical activity of PSII, increase the capacity of antioxidant enzymes (CAT, APX), proline and metabolites linked to oxidative stress clearly related to Cd tolerance (i.e., phytochelatin, fructans, and inulins) [134]. F. mosseae BEG167 under field conditions were evaluated for pot experiment analysis for tomato plants contaminated with five As levels (0, 25, 50, 75, and 150 mg·kg⁻¹) and found that mycorrhizal colonization increased plant biomass at application rates of 25, 50, and 75 mg kg⁻¹. Mycorrhizal colonization under the rate of 75 and 150 mg kg⁻¹ concentration levels were escalated in shoots, whereas decreased in 50 mg kg⁻¹. Total P was supplemented at the levels of 25, 50, and 75 mg kg⁻¹, and increased supply was administered to mycorrhizal colonized roots which demonstrated higher shoot and root P/As ratio than the non-mycorrhizal counterparts. pH values of the soils were high in uninoculated controls indicating that AM fungi can have enormous applications in phytostabilization of HMs-contaminated soils and phytoextraction of the contaminated metal pollutants for soil enrichment and enhancement for growth optimization [135].

Mycorrhizal colonization was subjugated for evaluation under arsenic supplemented soils for tomato seedlings upsurge and enhanced shoot height and root length was found in inoculated soils than in non-inoculated control soils. Vegetative growth was linked toward the positive outcomes and nutrient availability and uptake. High chlorophyll content with a decrease in arsenic levels in mycorrhizal colonized plants can also be connected to translocation of metal ions from soil to tomato plants. This criterion can also be rationalized for AMF effect on P uptake [136]. Cu pose as yet another useful micronutrient that is indispensable for plant growth and productivity. *R. intraradices* and *R. etunicatum* inoculation were experimented against challenged soils with increased Cu and illustrated that dry weight of roots and shoots increased quantitatively and qualitatively [137] (Table 4).

Level of Heavy Metals	Mycorrhizal Species	Effect	Reference
Cd concentrations (50 and 100 mg kg ⁻¹)	Funneliformis mosseae	Increased dry weights, reduction in transcolation of Cd from root to shoot	[66]
Cd (50 µM CdCl2)	Funneliformis mosseae Rhizophagus intraradices and Rhizophagus etunicatum	Increased antioxidant enzymes, Proline and phenol content	[129]
Cd (30 and 60 mg of CdSO4)	Funneliformis mosseae	Increased Cd absorption and dry weights	[133]
0 and 25 μMCd	Rhizophagus irregularis	Induced photosynthetic pigments, photosystem II efficiency, antioxidant enzyme and proline accumulation and reduced lipid peroxidation products. Increased nutritional status such as P, K, Ca, Fe, Mn, and Zn	[134]
As levels (0, 25, 50, 75 and 150 mg kg ⁻¹).	Funneliformis mosseae	Increased biomass and P uptake, higher shoot and root P/As ratio	[135]
Cu solution (0, 1.5, 3.5, 5.5, 7.5 mM CuSO4)	Rhizophagus intraradices and Rhizophagus etunicatum	Increased biomass, sugar, proline, and antioxidant enzymes	[137]

Table 4. Application of arbuscular mycorrhizal fungi on heavy metal stress alleviation.

These results can be related to chlorophyll content and Cu translocation pattern as seen in arsenic supplementation. Further, AM fungus and host plant interaction show shooting up of protein synthesis in elucidating oxidative enzyme levels under metal stress. The results of the findings escalate the idea that involvement of stress proteins like phytochelatins and metallothioneines will have possible roles in mechanistic protection [138,139]. At the same time, copper has its ill-effects in the metabolic hindrance of cellular activities and protein synthesis machinery which ultimately affects the total protein content in non-mycorrhizal inoculated tomato plants providing a clue to search for deeper insights for the assessment and confirmation. With this notion, it was evident that soil pollution with toxic heavy metals showed a considerable rise in enzymatic levels involved in antioxidant properties viz, APX and GUPX [139]. Thus, it is evident that the review addresses the physiology, inoculation efficiency, phenomenal interactions mediating the plant-microbe association for beneficial aspects of the involvement of AM fungi in combating a variety of abiotic stress modes and effective research perspectives in mitigating the stress and abatement strategies like sequestration, oxidative stress quenching, stress proteins involvement, and metal chelating proteins along with other essential modalities. Thus, the review compiles rationally the approaches to tackle the abiotic stress arresting by AM fungi and other molecular plant strategies in the apprehension of stress and establishment of sustainable plant growth not only for tomato but also other horticultural crop plants.

8. Concluding Remarks and Future Prospects

The stress biology of tomato has been vehemently researched worldwide to substantiate the increasing population need for copious food. This comprehensive analysis demonstrated the molecular mechanisms underlying abiotic stress physiology and combating strategies abated by AMF. However, photosynthetic efficiency (Pn, Gs, Tr, Fv/Fm), root morphology, hydraulic conductivity, elicitors' modality in arresting stress mechanisms remains to be explored still. The practice of genetic engineering and microbial biotechnology resulted in more investigation on development of plant tolerance possible, even when conventional breeding reached its limits. Most recently, it was found that clustered regularly interspaced short palindromic repeats (CRISPR)/CRISPR-associated protein 9 (Cas9) systems are significant ameliorators in the regime of biotic and abiotic stress and plant-microbe interactions. Genome editing provides target sequence tailored modifications through agricultural trait improvement. Moreover, transcriptome analysis for stress management will provide deeper insights for symbiotic management for multiple stress modalities. In the present scenario, this review will allow agricultural scientists to explore the potentials of AMF, and further, it will act as a launchpad for effective sustainable agricultural practices.

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References

- Odriozola-Serrano, I.; Soliva-Fortuny, R.; Hernández-Jover, T.; Martín-Belloso, O. Carotenoid and phenolic profile of tomato juices processed by high intensity pulsed electric fields compared with conventional thermal treatments. *Food Chem.* 2009, 112, 258–266, doi:10.1016/j.foodchem.2008.05.087.
- Agarwal, S.; Rao, A.V. Tomato lycopene and its role in human health and chronic diseases. *Can. Med. Assoc. J.* 2000, 163, 739– 744.
- 3. Rao, A.V.; Agarwal, S. Role of Antioxidant Lycopene in Cancer and Heart Disease. J. Am. Coll. Nutr. 2000, 19, 563–569, doi:10.1080/07315724.2000.10718953.
- Toor, R.K.; Lister, C.; Savage, G. Antioxidant activities of New Zealand-grown tomatoes. Int. J. Food Sci. Nutr. 2005, 56, 597–605, doi:10.1080/09637480500490400.
- Schüβler, A.; Schwarzott, D.; Walker, C. A new fungal phylum, the *Glomeromycota*: Phylogeny and evolution. *Mycol. Res.* 2001, 105, 1413–1421, doi:10.1017/s0953756201005196.
- Bonfante, P.; Genre, A. Mechanisms underlying beneficial plant–fungus interactions in mycorrhizal symbiosis. *Nat. Commun.* 2010, *1*, 48, doi:10.1038/ncomms1046.
- Redecker, D.; Schüssler, A.; Stockinger, H.; Stürmer, S.L.; Morton, J.B.; Walker, C. An evidence-based consensus for the classification of arbuscular mycorrhizal fungi (*Glomeromycota*). *Mycorrhiza* 2013, 23, 515–531, doi:10.1007/s00572-013-0486-y.
- 8. Smith, S.E.; Read, D.J. Mycorrhizal Symbiosis, 3rd ed.; Academic Press: New York, NY, USA, 2008.
- 9. Parniske, M. Arbuscular mycorrhiza: The mother of plant root endosymbioses. *Nat. Rev. Genet.* 2008, *6*, 763–775, doi:10.1038/nrmicro1987.
- Bonfante, P.; Genre, A. Plants and arbuscular mycorrhizal fungi: An evolutionary-developmental perspective. *Trends Plant Sci.* 2008, 13, 492–498, doi:10.1016/j.tplants.2008.07.001.
- 11. Bago, B.; Pfeffer, P.E.; Shachar-Hill, Y. Carbon Metabolism and Transport in Arbuscular Mycorrhizas. *Plant Physiol.* **2000**, *124*, 949–958, doi:10.1104/pp.124.3.949.
- 12. Jiang, Y.; Wang, W.; Xie, Q.; Liu, N.; Liu, L.; Wang, D.; Zhang, X.; Yang, C.; Chen, X.; Tang, D.; et al. Plants transfer lipids to sustain colonization by mutualistic mycorrhizal and parasitic fungi. *Science* **2017**, *356*, 1172–1175, doi:10.1126/science.aam9970.
- 13. Gianinazzi, S.; Gollotte, A.; Binet, M.N.; van Tuinen, D.; Redecker, D.; Wipf, D. Agroecology: The key role of arbuscular mycorrhizas in ecosystem services. *Mycorrhiza* **2010**, *20*, 519–530.
- 14. Baligar, V.C.; Fageria, N.K.; He, Z.L. Nutrient use efficiency in plants. Commun. Soil Sci. Plant Anal. 2001, 32, 921–950, doi:10.1081/css-100104098.
- 15. Giovannetti, M.; Avio, L.; Barale, R.; Ceccarelli, N.; Cristofani, R.; Iezzi, A.; Mignolli, F.; Picciarelli, P.; Pinto, B.; Reali, D.; et al. Nutraceutical value and safety of tomato fruits produced by mycorrhizal plants. *Br. J. Nutr.* **2011**, *107*, 242–251, doi:10.1017/s000711451100290x.
- 16. Conversa, G.; Lazzizera, C.; Bonasia, A.; Elia, A. Yield and phosphorus uptake of a processing tomato crop grown at different phosphorus levels in a calcareous soil as affected by mycorrhizal inoculation under field conditions. *Biol. Fertil. Soils* **2013**, *49*, 691–703, doi:10.1007/s00374-012-0757-3.
- 17. Poulton, J.L.; Bryla, D.; Koide, R.T.; Stephenson, A.G. Mycorrhizal infection and high soil phosphorus improve vegetative growth and the female and male functions in tomato. *New Phytol.* **2002**, *154*, 255–264, doi:10.1046/j.1469-8137.2002.00366.x.
- Colla, G.; Rouphael, Y.; Di Mattia, E.; El-Nakhel, C.; Cardarelli, M. Co-inoculation of *Glomus intraradices* and *Trichoderma atro*viride acts as a biostimulant to promote growth, yield and nutrient uptake of vegetable crops. J. Sci. Food Agric. 2015, 95, 1706– 1715.

- 19. Kowalska, I.; Konieczny, A.; Gąstoł, M.; Sady, W.; Hanus-Fajerska, E. Effect of mycorrhiza and phosphorus content in nutrient solution on the yield and nutritional status of tomato plants grown on rockwool or coconut coir. *Agric. Food Sci.* **2015**, *24*, 39–51, doi:10.23986/afsci.47204.
- Hart, M.; Ehret, D.L.; Krumbein, A.; Leung, C.; Murch, S.; Turi, C.; Franken, P. Inoculation with arbuscular mycorrhizal fungi improves the nutritional value of tomatoes. *Mycorrhiza* 2015, 25, 359–376, doi:10.1007/s00572-014-0617-0.
- Ortas, I. The effect of mycorrhizal fungal inoculation on plant yield, nutrient uptake and inoculation effectiveness under longterm field conditions. *Field Crop. Res.* 2012, 125, 35–48, doi:10.1016/j.fcr.2011.08.005.
- Sardans, J.; Peñuelas, J.; Ogaya, R. Drought's impact on Ca, Fe, Mg, Mo and S concentration and accumulation patterns in the plants and soil of a Mediterranean evergreen Quercus ilex forest. *Biogeochemistry* 2007, *87*, 49–69, doi:10.1007/s10533-007-9167-2.
- 23. Yang, Y.; Chen, Y.; Li. W. Arbuscular mycorrhizal fungi infection in desert riparian forest and its environmental im-plications: a case study in the lower reach of Tarim River. *Prog. Nat. Sci.* **2008**, *18*, 983–991.
- 24. Aroca, R. (Ed.). Plant Responses to Drought Stress: from Morphological to Molecular Features; Springer-Verlag: Berlin/Heidelberg, Germany, 2012.
- 25. Augé, R.M. Water relations, drought and vesicular-arbuscular mycorrhizal symbiosis. Mycorrhiza 2001, 11, 3–42.
- 26. Augé, R.M. Arbuscular mycorrhizae and soil/plant water relations. Can. J. Soil Sci. 2004, 84, 373–381.
- 27. Porcel, R.; Ruiz-Lozano, J.M. Arbuscular mycorrhizal influence on leaf water potential, solute accumulation and oxidative stress in soybean plants subjected to drought stress. *J. Exp. Bot.* **2004**, *55*, 1743–1750.
- Duc, N.H.; Csintalan, Z.; Posta, K. Arbuscular mycorrhizal fungi mitigate negative effects of combined drought and heat stress on tomato plants. *Plant Physiol. Biochem.* 2018, 132, 297–307, doi:10.1016/j.plaphy.2018.09.011.
- Chitarra, W.; Pagliarani, C.; Maserti, B.; Lumini, E.; Siciliano, I.; Cascone, P.; Schubert, A.; Gambino, G.; Balestrini, R.; Guerrieri, E. Insights on the Impact of Arbuscular Mycorrhizal Symbiosis On Tomato Tolerance to Water Stress. *Plant Physiol.* 2016, 171, 1009–1023, doi:10.1104/pp.16.00307.
- Nuruddin, M.M.; Madramootoo, C.A.; Dodds, G.T. Effects of Water Stress at Different Growth Stages on Greenhouse Tomato Yield and Quality. *HortScience* 2003, 38, 1389–1393, doi:10.21273/hortsci.38.7.1389.
- Sánchez-Rodríguez, E.; Moreno, D.A.; Ferreres, F.; Rubio-Wilhelmi, M.D.M.; Ruiz, J.M. Differential responses of five cherry tomato varieties to water stress: Changes on phenolic metabolites and related enzymes. *Phytochemistry* 2011, 72, 723–729, doi:10.1016/j.phytochem.2011.02.011.
- 32. Giannakoula, A.; Ilias, I. The effect of water stress and salinity on growth and physiology of tomato (*Lycopersicon esculentum* Mil.). *Arch. Biol. Sci.* **2013**, *65*, 611–620, doi:10.2298/abs1302611g.
- Sacco, A.; Greco, B.; Di Matteo, A.; De Stefano, R.; Barone, A. Evaluation of Tomato Genetic Resources for Response to Water Deficit. Am. J. Plant Sci. 2013, 4, 131–145, doi:10.4236/ajps.2013.412a3016.
- Dell'Amico, J.; Torrecillas, A.; Rodríguez, P.; Morte, A.; Sánchez-Blanco, M.J. Responses of tomato plants associated with the arbuscular mycorrhizal fungus *Glomus clarum* during drought and recovery. *J. Agric. Sci.* 2002, 138, 387–393, doi:10.1017/s0021859602002101.
- 35. Subramanian, K.S.; Santhanakrishnan, P.; Balasubramanian, P. Responses of field grown tomato plants to arbuscular mycorrhizal fungal colonization under varying intensities of drought stress. *Sci. Hortic. Amst.* **2006**, *107*, 245–253.
- Aroca, R.; Del Mar Alguacil, M.; Vernieri, P.; Ruiz-Lozano, J.M. Plant responses to drought stress and exogenous ABA application are modulated differently by mycorrhization in tomato and an ABA-deficient mutant (Sitiens). *Microb. Ecol.* 2008, *56*, 704– 719.
- 37. Wang, B.; Yao, Z.; Zhao, S.; Guo, K.; Sun, J.; Zhang, J. Arbuscular mycorrhizal fungal application to improve growth and tolerance of processing tomato (*Lycopersicon esculentum* Miller) under drought stress. J. Food Agric. Environ. 2014, 12, 452–457.
- Ruiz-Lozano, J.M.; Aroca, R.; Zamarreño, Á.M.; Molina, S.; Andreo-Jiménez, B.; Porcel, R.; García-Mina, J.M.; Ruyter-Spira, C.; López-Ráez, J.A. Arbuscular mycorrhizal symbiosis induces strigolactone biosynthesis under drought and improves drought tolerance in lettuce and tomato. *Plant Cell Environ*. 2016, 39, 441–452.
- 39. Augé, R.M.; Toler, H.D.; Saxton, A.M. Arbuscular mycorrhizal symbiosis alters stomatal conductance of host plants more under drought than under amply watered conditions: A meta-analysis. *Mycorrhiza* **2015**, *25*, 13–24, doi:10.1007/s00572-014-0585-4.
- Kim, K.Y.; Jordan, D.; McDonald, G.A. Effect of phosphate-solubilizing bacteria and vesicular-arbuscular mycorrhizae on tomato growth and soil microbial activity. *Biol. Fertil. Soils* 1997, 26, 79–87, doi:10.1007/s003740050347.
- 41. Rivero, J.; Gamir, J.; Aroca, R.; Pozo, M.J.; Flors, V. Metabolic transition in mycorrhizal tomato roots. *Front. Microbiol.* **2015**, *6*, 598.
- 42. Xu, L.; Li, T.; Wu, Z.; Feng, H.; Yu, M.; Zhang, X.; Chen, B. Arbuscular mycorrhiza enhances drought tolerance of tomato plants by regulating the 14-3-3 genes in the ABA signaling pathway. *Appl. Soil Ecol.* **2018**, *125*, 213–221, doi:10.1016/j.apsoil.2018.01.012.
- Rivero, J.; Álvarez, D.; Flors, V.; Azcón-Aguilar, C.; Pozo, M.J. Root metabolic plasticity underlies functional diversity in mycorrhiza-enhanced stress tolerance in tomato. *New Phytol.* 2018, 220, 1322–1336.
- 44. Mannino, G.; Nerva, L.; Gritli, T.; Novero, M.; Fiorilli, V.; Bacem, M.; Bertea, C.M.; Lumini, E.; Chitarra, W.; Balestrini, R. Effects of Different Microbial Inocula on Tomato Tolerance to Water Deficit. *Agronomy* **2020**, *10*, 170, doi:10.3390/agronomy10020170.
- Volpe, V.; Chitarra, W.; Cascone, P.; Volpe, M.G.; Bartolini, P.; Moneti, G.; Pieraccini, G.; Di Serio, C.; Maserti, B.; Guerrieri, E.; et al. The Association With Two Different Arbuscular Mycorrhizal Fungi Differently Affects Water Stress Tolerance in Tomato. *Front. Plant Sci.* 2018, *9*, 1480, doi:10.3389/fpls.2018.01480.

- 46. Berruti, A.; Borriello, R.; Della Beffa, M.T.; Scariot, V.; Bianciotto, V. Application of nonspecific commercial AMF inocula results in poor mycorrhization in *Camellia japonica* L. *Symbiosis* **2013**, *61*, 63–76, doi:10.1007/s13199-013-0258-7.
- 47. Berruti, A.; Lumini, E.; Balestrini, R.; Bianciotto, V. Arbuscular Mycorrhizal Fungi as Natural Biofertilizers: Let's Benefit from Past Successes. *Front. Microbiol.* **2016**, *6*, 1559, doi:10.3389/fmicb.2015.01559.
- 48. Smith, S.E.; Smith, F.A. Fresh perspectives on the roles of arbuscular mycorrhizal fungi in plant nutrition and growth. *Mycologia* **2012**, *104*, 1–13, doi:10.3852/11-229.
- 49. Bucher, M. Functional biology of plant phosphate uptake at root and mycorrhiza interfaces. *New Phytol.* 2006, 173, 11–26, doi:10.1111/j.1469-8137.2006.01935.x.
- Javot, H.; Penmetsa, R.V.; Terzaghi, N.; Cook, D.R.; Harrison, M.J. A *Medicago truncatula* phosphate transporter indispensable for the arbuscular mycorrhizal symbiosis. *Proc. Natl. Acad. Sci. USA* 2007, 104, 1720–1725, doi:10.1073/pnas.0608136104.
- 51. Smith, F.A.; Smith, S.E. What is the significance of the arbuscular mycorrhizal colonisation of many economically important crop plants? *Plant Soil* **2011**, *348*, 63–79, doi:10.1007/s11104-011-0865-0.
- Chen A., Chen X., Wang H., Liao D., Gu M., Qu H. Genome-wide investigation and expression analysis suggest diverse roles and genetic redundancy of *Pht1* family genes in response to Pi deficiency in tomato. *BMC Plant Biol.* 2014, 14, 61.
- Nagy, R.; Drissner, D.; Amrhein, N.; Jakobsen, I.; Bucher, M. Mycorrhizal phosphate uptake pathway in tomato is phosphorusrepressible and transcriptionally regulated. *New Phytol.* 2009, 181, 950–959.
- Nagy, R.; Karandashov, V.; Chague, V.; Kalinkevich, K.; Tamasloukht, M.; Xu, G.; Jakobsen, I.; Levy, A.A.; Amrhein, N.; Bucher, M. The characterization of novel mycorrhiza-specific phosphate transporters from *Lycopersicon esculentum* and *Solanum tuberosum* uncovers functional redundancy in symbiotic phosphate transport in solanaceous species. *Plant J.* 2005, 42, 236–250, doi:10.1111/j.1365-313x.2005.02364.x.
- Balestrini, R.; Gómez-Ariza, J.; Lanfranco, L.; Bonfante, P. Laser Microdissection Reveals That Transcripts for Five Plant and One Fungal Phosphate Transporter Genes Are Contemporaneously Present in Arbusculated Cells. *Mol. Plant-Microbe Interact.* 2007, 20, 1055–1062, doi:10.1094/mpmi-20-9-1055.
- Balestrini, R.; Rosso, L.C.; Veronico, P.; Melillo, M.T.; De Luca, F.; Fanelli, E.; Colagiero, M.; Di Fossalunga, A.S.; Ciancio, A.; Pentimone, I. Transcriptomic Responses to Water Deficit and Nematode Infection in Mycorrhizal Tomato Roots. *Front. Microbiol.* 2019, 10, 1807, doi:10.3389/fmicb.2019.01807.
- 57. Handa, Y.; Nishide, H.; Takeda, N.; Suzuki, Y.; Kawaguchi, M.; Saito, K.RNA-seq transcriptional profiling of an arbuscular mycorrhiza provides insights into regulated and coordinated gene expression in *Lotus japonicus* and *Rhizophagus irregularis*. *Plant Cell Physiol.* **2015**, *8*, 1490–1511.
- Xu, J.; Xing, X.-J.; Tian, Y.-S.; Peng, R.-H.; Xue, Y.; Zhao, W.; Yao, Q.-H. Transgenic Arabidopsis Plants Expressing Tomato Glutathione S-Transferase Showed Enhanced Resistance to Salt and Drought Stress. *PLoS ONE* 2015, 10, e0136960, doi:10.1371/journal.pone.0136960.
- Ruiz-Lozano, J.M. Arbuscular mycorrhizal symbiosis and alleviation of osmotic stress. New perspectives for molecular studies. *Mycorrhiza* 2003, 13, 309–317, doi:10.1007/s00572-003-0237-6.
- 60. Pedranzani, H.E.; Rodríguez-Rivera, M.; Gutierrez, M.I.; Porcel, R.; Hause, B.; Ruizlozano, J.M. Arbuscular mycorrhizal symbiosis regulates physiology and performance of *Digitaria eriantha* plants subjected to abiotic stresses by modulating antioxidant and jasmonate levels. *Mycorrhiza* **2016**, *26*, 141–152, doi:10.1007/s00572-015-0653-4.
- Davies, F.T., Jr.; Svenson, S.E.; Cole, J.C.; Phavaphutanon, L.; Duray, S.A.; Olalde-Portugal, V.; Meier, C.E.; Bo, S.H. Non-nutritional stress acclimation of mycorrhizal woody plants exposed to drought. *Tree Physiol.* 1996, 16, 985–993.
- 62. Rodríguez, P.; Dell'Amico, J.; Morales, D.; Blanco, M.J.S.; Alarcón, J.J. Effects of salinity on growth, shoot water relations and root hydraulic conductivity in tomato plants. *J. Agric. Sci.* **1997**, *128*, 439–444, doi:10.1017/s0021859697004309.
- 63. Bárzana, G.; Aroca, R.; Bienert, G.P.; Chaumont, F.; Ruiz-Lozano, J.M. New Insights into the Regulation of Aquaporins by the Arbuscular Mycorrhizal Symbiosis in Maize Plants Under Drought Stress and Possible Implications for Plant Performance. *Mol. Plant-Microbe Interact.* **2014**, *27*, 349–363, doi:10.1094/mpmi-09-13-0268-r.
- 64. Quiroga, G.; Erice, G.; Aroca, R.; Chaumont, F.; Ruiz-Lozano, J.M. Enhanced drought stress tolerance by the arbuscular mycorrhizal symbiosis in a drought-sensitive maize cultivar is related to a broader and differential regulation of host plant aquaporins than in a drought-tolerant cultivar. *Front. Plant Sci.* **2017**, *8*, 1056.
- Herrera-Medina, M.J.; Steinkellner, S.; Vierheilig, H.; Bote, J.A.O.; Garrido, J.M.G. Abscisic acid determines arbuscule development and functionality in the tomato arbuscular mycorrhiza. *New Phytol.* 2007, 175, 554–564, doi:10.1111/j.1469-8137.2007.02107.x.
- 66. Li, W.; Wang, T.; Zhang, Y.; Li, Y. Overexpression of soybean miR172c confers tolerance to water deficit and salt stress, but increases ABA sensitivity in transgenic *Arabidopsis thaliana*. *J. Exp. Bot.* **2016**, *67*, 175–194, doi:10.1093/jxb/erv450.
- 67. Thompson, A.J.; Jackson, A.C.; Symonds, R.C.; Mulholland, B.J.; Dadswell, A.R.; Blake, P.S.; Burbidge, A.; Taylor, I.B. Ectopic expression of a tomato 9-cis-epoxycarotenoid dioxygenase gene causes over-production of abscisic acid. *Plant J.* **2000**, *23*, 363–374, doi:10.1046/j.1365-313x.2000.00789.x.
- Martín-Rodríguez, J.A.; León-Morcillo, R.; Vierheilig, H.; Ocampo Bote, J.A.; Ludwig-Müllerm, J.; García-Garrido, J.M. Mycorrhization of the notabilis and sitiens tomato mutants in relation to abscisic acid and ethylene contents. J. Plant Physiol. 2010, 167, 606–613.
- 69. Wasternack, C.; Hause, B. Jasmonates: Biosynthesis, perception, signal transduction and action in plant stress response, growth and development. An update to the 2007 review in Annals of Botany. *Ann. Bot.* **2013**, *111*, 1021–1058, doi:10.1093/aob/mct067.

- Bucher, M.; Hause, B.; Krajinski, F.; Küster, H. Through the doors of perception to function in arbuscular mycorrhizal symbioses. *New Phytol.* 2014, 204, 833–840.
- Lim, C.W.; Baek, W.; Jung, J.; Kim, J.-H.; Lee, S.C. Function of ABA in Stomatal Defense against Biotic and Drought Stresses. *Int. J. Mol. Sci.* 2015, *16*, 15251–15270, doi:10.3390/ijms160715251.
- Abbaspour, H.; Saeid-Sar, S.; Afshari, H.; Abdel-Wahhab, M.A. Tolerance of mycorrhiza infected Pistachio (Pistacia vera L.) seedlings to drought stress under glasshouse conditions. J. Plant Physiol. 2012, 169, 704–709.
- Yooyongwech, S.; Phaukinsang, N.; Cha-Um, S.; Supaibulwatana, K. Arbuscular mycorrhiza improved growth performance in Macadamia tetraphylla L. grown under water deficit stress involves soluble sugar and proline accumulation. Plant Growth Regul. 2012, 69, 285–293, doi:10.1007/s10725-012-9771-6.
- Zou, Y.N.; Wu, Q.S.; Huang, Y.M.; Ni, Q.D.; He, X.H. Mycorrhizal-mediated lower proline accumulation in Poncirus trifoliate under water deficit derives from the integration of inhibition of proline synthesis with increase of proline degradation. *PLoS* ONE 2013, 8, e80568.
- Maurel, C.; Boursiac, Y.; Luu, D.-T.; Santoni, V.; Shahzad, Z.; Verdoucq, L. Aquaporins in Plants. *Physiol. Rev.* 2015, 95, 1321– 1358, doi:10.1152/physrev.00008.2015.
- Reuscher, S.; Akiyama, M.; Mori, C.; Aoki, K.; Shibata, D.; Shiratake, K. Genome-Wide Identification and Expression Analysis of Aquaporins in Tomato. *PLoS ONE* 2013, *8*, e79052, doi:10.1371/journal.pone.0079052.
- 77. Munns, R.; Tester, M. Mechanisms of salinity tolerance. Annu. Rev. Plant Biol. 2008, 59, 651–681, doi:10.1146/annurev.ar-plant.59.032607.092911.
- 78. Munns, R.; Gilliham, M. Salinity tolerance of crops–what is the cost? New Phytol. 2015, 208, 668–673.
- Yurtseven, E.; Kesmez, G.D.; Unlukara, A. The effects of water salinity and potassium levels on yield, fruit quality and water consumption of a native central Anatolian tomato species (Lycopersicon Esculentum). *Agric. Water Manag.* 2005, *78*, 128–135.
- He, Z.; He, C.; Zhang, Z.; Zou, Z.; Wang, H. Changes of antioxidative enzymes and cell membrane osmosis in tomato colonized by arbuscular mycorrhizae under NaCl stress. *Colloids Surfaces B Biointerfaces* 2007, 59, 128–133, doi:10.1016/j.colsurfb.2007.04.023.
- 81. Tanveer, K.; Gilani, S.; Hussain, Z.; Ishaq, R.; Adeel, M.; Ilyas, N. Effect of salt stress on tomato plant and the role of calcium. *J. Plant Nutr.* **2019**, *43*, 28–35, doi:10.1080/01904167.2019.1659324.
- Hajiboland, R.; Aliasgharzadeh, N.; Laiegh, S.F.; Poschenrieder, C. Colonization with arbuscular mycorrhizal fungi improves salinity tolerance of tomato (*Solanum lycopersicum* L.) plants. *Plant Soil* 2010, 331, 313–327, doi:10.1007/s11104-009-0255-z.
- Santander, C.; Sanhueza, M.; Olave, J.; Borie, F.; Valentine, A.; Cornejo, P. Arbuscular Mycorrhizal Colonization Promotes the Tolerance to Salt Stress in Lettuce Plants through an Efficient Modification of Ionic Balance. *J. Soil Sci. Plant Nutr.* 2019, 19, 321– 331, doi:10.1007/s42729-019-00032-z.
- 84. Juniper, S.; Abbott, L. Vesicular-arbuscular mycorrhizas and soil salinity. Mycorrhiza 1993, 4, 45–57, doi:10.1007/bf00204058.
- 85. Ebrahim, M.K.; Saleem, A.-R. Alleviating salt stress in tomato inoculated with mycorrhizae: Photosynthetic performance and enzymatic antioxidants. *J. Taibah Univ. Sci.* **2017**, *11*, 850–860, doi:10.1016/j.jtusci.2017.02.002.
- Al-Karaki, G.N. Growth of mycorrhizal tomato and mineral acquisition under salt stress. *Mycorrhiza* 2000, 10, 51–54, doi:10.1007/s005720000055.
- Al-Karaki, G.N.; Hammad, R.; Rusan, M. Response of two tomato cultivars differing in salt tolerance to inoculation with mycorrhizal fungi under salt stress. *Mycorrhiza* 2001, *11*, 43–47, doi:10.1007/s005720100098.
- Sellitto, V.M.; Golubkina, N.A.; Pietrantonio, L.; Cozzolino, E.; Cuciniello, A.; Cenvinzo, V.; Florin, I.; Caruso, G. Tomato Yield, Quality, Mineral Composition and Antioxidants as Affected by Beneficial Microorganisms Under Soil Salinity Induced by Balanced Nutrient Solutions. *Agriculture* 2019, *9*, 110, doi:10.3390/agriculture9050110.
- Latef, A.A.H.A.; Chaoxing, H. Effect of arbuscular mycorrhizal fungi on growth, mineral nutrition, antioxidant enzymes activity and fruit yield of tomato grown under salinity stress. *Sci. Hortic.* 2011, 127, 228–233, doi:10.1016/j.scienta.2010.09.020.
- Balliu, A.; Sallaku, G.; Rewald, B. AMF Inoculation Enhances Growth and Improves the Nutrient Uptake Rates of Transplanted, Salt-Stressed Tomato Seedlings. *Sustainability* 2015, 7, 15967–15981, doi:10.3390/su71215799.
- Kaya, C.; Ashraf, M.; Sonmez, O.; Aydemir, S.; Tuna, L.A.; Cullu, A.M. The influence of arbuscular mycorrhizal colonisation on key growth parameters and fruit yield of pepper plants grown at high salinity. *Sci. Hort.* 2009, *121*, 1–6.
- 92. Ruiz-Lozano, J.M.; Azcón, R. Symbiotic efficiency and infectivity of an autochthonous arbuscular mycorrhizal Glomus sp. from saline soils and *Glomus deserticola* under salinity. *Mycorrhiza* **2000**, *10*, 137–143, doi:10.1007/s005720000075.
- 93. Porcel, R.; Aroca, R.; Ruiz-Lozano, J.M. Salinity stress alleviation using arbuscular mycorrhizal fungi. A review. *Agron. Sustain. Dev.* **2012**, *32*, 181–200, doi:10.1007/s13593-011-0029-x.
- 94. Cantrell, I.C.; Linderman, R.G. Preinoculation of lettuce and onion with VA mycorrhizal fungi reduces deleterious effects of soil salinity. *Plant Soil* **2001**, 233, 269–281, doi:10.1023/a:1010564013601.
- Feng, G.; Li, X.L.; Zhang, F.S.; Li, S.X. Effect of phosphorus and arbuscular mycorrhizal fungus on response of maize plant to saline environment. J. Plant Res. Environ. 2000, 9, 22–26.
- 96. Sannazzaro, A.I.; Echeverria, M.; Alberto, E.O.; Ruiz, O.A.; Menendez, A.B. Modulation of polyamine balance in *Lotus glaber* by salinity and arbuscular mycorrhiza. *Plant Physiol. Biochem.* **2007**, *45*, 39–46.
- Evelin, H.; Kapoor, R.; Giri, B. Arbuscular mycorrhizal fungi in alleviation of salt stress: A review. Ann. Bot. 2009, 104, 1263– 1280, doi:10.1093/aob/mcp251.

- Evelin, H.; Giri, B.; Kapoor, R. Contribution of *Glomus intraradices* inoculation to nutrient acquisition and mitigation of ionic imbalance in NaCl-stressed *Trigonella foenum-graecum*. *Mycorrhiza* 2012, 22, 203–217.
- Kong, L.; Gong, X.; Zhang, X.; Zhang, W.; Sun, J.; Chen, B. Effects of arbuscular mycorrhizal fungi on photosynthesis, ion balance of tomato plants under saline-alkali soil condition. J. Plant Nutr. 2020, 43, 682–698, doi:10.1080/01904167.2019.1701029.
- Dasgan, H.; Aktas, H.; Abak, K.; Cakmak, I. Determination of screening techniques to salinity tolerance in tomatoes and investigation of genotype responses. *Plant Sci.* 2002, 163, 695–703, doi:10.1016/s0168-9452(02)00091-2.
- 101. Khalloufi, M.; Martínez-Andújar, C.; Lachaâl, M.; Karray-Bouraoui, N.; Pérez-Alfocea, F.; Albacete, A. The interaction between foliar GA3 application and arbuscular mycorrhizal fungi inoculation improves growth in salinized tomato (*Solanum lycopersicum* L.) plants by modifying the hormonal balance. *J. Plant Physiol.* **2017**, 214, 134–144.
- Augã, R.M.; Toler, H.D.; Saxton, A.M.; Augé, R.M. Arbuscular mycorrhizal symbiosis and osmotic adjustment in response to NaCl stress: A meta-analysis. Front. Plant Sci. 2014, 5, 562, doi:10.3389/fpls.2014.00562.
- Ouziad, F.; Wilde, P.; Schmelzer, E.; Hildebrandt, U.; Bothe, H. Analysis of expression of aquaporins and Na+/H+ transporters in tomato colonized by arbuscular mycorrhizal fungi and affected by salt stress. *Environ. Exp. Bot.* 2006, 57, 177–186, doi:10.1016/j.envexpbot.2005.05.011.
- Birhane, E.; Sterck, F.J.; Fetene, M.; Bongers, F.; Kuyper, T.W. Arbuscular mycorrhizal fungi enhance photosynthesis, water use efficiency, and growth of frankincense seedlings under pulsed water availability conditions. *Oecologia* 2012, 169, 895–904, doi:10.1007/s00442-012-2258-3.
- Chandrasekaran, M.; Chanratana, M.; Kim, K.; Seshadri, S.; Sa, T. Impact of Arbuscular Mycorrhizal Fungi on Photosynthesis, Water Status, and Gas Exchange of Plants Under Salt Stress–A Meta-Analysis. *Front. Plant Sci.* 2019, 10, 457, doi:10.3389/fpls.2019.00457.
- Lin, J.; Wang, Y.; Sun, S.; Mu, C.; Yan, X. Effects of arbuscular mycorrhizal fungi on the growth, photosynthesis and photosynthetic pigments of *Leymus chinensis* seedlings under salt-alkali stress and nitrogen deposition. *Sci. Total Environ.* 2017, 576, 234–241.
- 107. Riemann, M.; Dhakarey, R.; Hazman, M.; Miro, B.; Kohli, A.; Nick, P. Exploring jasmonates in the hormonal network of drought and salinity responses. *Front. Plant Sci.* 2015, *6*, 1077.
- Chen, L.; Ren, Y.; Zhang, Y.; Xu, J.; Sun, F.; Zhang, Z.; Wang, Y. Genome-wide identification and expression analysis of heatresponsive and novel microRNAs in *Populus tomentosa*. *Gene* 2012, 504, 160–165, doi:10.1016/j.gene.2012.05.034.
- Aroca, R.; Vernieri, P.; Irigoyen, J.J.; Sancher-Diaz, M.; Tognoni, F.; Pardosso, A.Involvement of abscisic acid in leaf and root of maize (*Zea mays* L.) in avoiding chilling induced water stress. *Plant Sci.* 2003, 165, 671–679.
- 110. Zhu, X.-C.; Song, F.-B.; Xu, H.-W. Arbuscular mycorrhizae improve low temperature stress in maize via alterations in host water status and photosynthesis. *Plant Soil* **2010**, *331*, 129–137, doi:10.1007/s11104-009-0239-z.
- 111. Apel, K.; Hirt, H. Reactive oxygen species: Metabolism, oxidative stress, and signal transduction. *Annu. Rev. Plant Biol.* **2004**, *55*, 373–399, doi:10.1146/annurev.arplant.55.031903.141701.
- 112. Foolad, M.; Lin, G. Relationship between Cold Tolerance during Seed Germination and Vegetative Growth in Tomato: Germplasm Evaluation. J. Am. Soc. Hortic. Sci. 2000, 125, 679–683, doi:10.21273/jashs.125.6.679.
- 113. Ghanbari, F.; Sayyari, M. Controlled drought stress affects the chilling-hardening capacity of tomato seedlings as indicated by changes in phenol metabolisms, antioxidant enzymes activity, osmolytes concentration and abscisic acid accumulation. *Sci. Hortic.* 2018, 229, 167–174, doi:10.1016/j.scienta.2017.10.009.
- 114. Caradonia, F.; Francia, E.; Morcia, C.; Ghizzoni, R.; Moulin, L.; Terzi, V.; Ronga, D. Arbuscular Mycorrhizal Fungi and Plant Growth Promoting Rhizobacteria Avoid Processing Tomato Leaf Damage during Chilling Stress. *Agronomy* **2019**, *9*, 299, doi:10.3390/agronomy9060299.
- 115. Liu, A.; Chen, S.; Wang, M.; Liu, D.; Chang, R.; Wang, Z.; Lin, X.; Bai, B.; Ahammed, G.J. Arbuscular Mycorrhizal Fungus Alleviates Chilling Stress by Boosting Redox Poise and Antioxidant Potential of Tomato Seedlings. *J. Plant Growth Regul.* 2016, 35, 109–120, doi:10.1007/s00344-015-9511-z.
- Charest, C.; Dalpe, Y.; Brown, A. The effect of vesicular arbuscular mycorrhizae and chilling on two hybrids of *Zea mays* L. *Mycorrhiza* 1993, 4, 89–92.
- Latef, A.A.H.A.; Chaoxing, H. Arbuscular mycorrhizal influence on growth, photosynthetic pigments, osmotic adjustment and oxidative stress in tomato plants subjected to low temperature stress. *Acta Physiol. Plant.* 2011, 33, 1217–1225, doi:10.1007/s11738-010-0650-3.
- 118. Paradis, R.; Dalpe, Y.; Charest, C. The combined effect of arbuscular mycorrhizas and short-term cold exposure on wheat. *New Phytol.* **1995**, *129*, 637–642.
- 119. Wu, Q.-S.; Xia, R.-X.; Zou, Y.-N. Reactive oxygen metabolism in mycorrhizal and non-mycorrhizal citrus (*Poncirus trifoliata*) seedlings subjected to water stress. J. Plant Physiol. **2006**, 163, 1101–1110, doi:10.1016/j.jplph.2005.09.001.
- Haddidi, I.; Duc, N.; Tonk, S.; Rápó, E.; Posta, K. Defense Enzymes in Mycorrhizal Tomato Plants Exposed to Combined Drought and Heat Stresses. *Agronomy* 2020, *10*, 1657, doi:10.3390/agronomy10111657.
- 121. Clemens, S. Toxic metal accumulation, responses to exposure and mechanisms of tolerance in plants. *Biochimie* 2006, 88, 1707–1719, doi:10.1016/j.biochi.2006.07.003.
- 122. Cabral, L.; Soares, C.R.F.S.; Giachini, A.J.; Siqueira, J.O. Arbuscular mycorrhizal fungi in phytoremediation of contaminated areas by trace elements: Mechanisms and major benefits of their applications. *World J. Microbiol. Biotechnol.* 2015, 31, 1655–1664, doi:10.1007/s11274-015-1918-y.

- 123. Hildebrandt, U.; Regvar, M.; Bothe, H. Arbuscular mycorrhiza and heavy metal tolerance. *Phytochemistry* **2007**, *68*, 139–146, doi:10.1016/j.phytochem.2006.09.023.
- Jeffries, P.; Gianinazzi, S.; Perotto, S.; Turnau, K.; Barea, J.-M. The contribution of arbuscular mycorrhizal fungi in sustainable maintenance of plant health and soil fertility. *Biol. Fertil. Soils* 2003, *37*, 1–16, doi:10.1007/s00374-002-0546-5.
- 125. Zhang, X.; Chen, B.; Ohtomo, R. Mycorrhizal effects on growth, P uptake and Cd tolerance of the host plant vary among different AM fungal species. *Soil Sci. Plant Nutr.* 2015, *61*, 359–368, doi:10.1080/00380768.2014.985578.
- Nayuki, K.; Chen, B.; Ohtomo, R.; Kuga, Y. Cellular Imaging of Cadmium in Resin Sections of Arbuscular Mycorrhizas Using Synchrotron Micro X-ray Fluorescence. *Microbes Environ.* 2014, 29, 60–66, doi:10.1264/jsme2.me13093.
- 127. Göhre, V.; Paszkowski, U. Contribution of the arbuscular mycorrhizal symbiosis to heavy metal phytoremediation. *Planta* **2006**, 223, 1115–1122, doi:10.1007/s00425-006-0225-0.
- 128. Fernández-Fuego, D.; Bertrand, A.; González, A. Metal accumulation and detoxification mechanisms in mycorrhizal *Betula pubescens*. *Environ*. *Pollut*. **2017**, 231, 1153–1162, doi:10.1016/j.envpol.2017.07.072.
- 129. Hashem, A.; Abd Allah, E.F.; Alqarawi, A.A.; Al Huqail, A.A.; Egamberdieva, D.; Wirth, S. Alleviation of cadmium stress in *Solanum lycopersicum* L. by arbuscular mycorrhizal fungi via induction of acquired systemic tolerance. *Saudi J. Biol. Sci.* 2016, 23, 272–281.
- Jing, D.; Fei-bo, W.; Guo-ping, Z. Effect of cadmium on growth and photosynthesis of tomato seedlings. J. Zhejiang Univ. Sci. 2005, 6, 974.
- Hayat, S.; Hasan, S.A.; Ahmad, A. Growth, nitrate reductase activity and antioxidant system in cadmium stressed tomato (*Ly-copersicon esculentum Mill*) cultivars. *Biotech. Agron. Soc. Environ.* 2011, 15, 401–414.
- 132. Li, Y.; Zeng, J.; Wang, S.; Lin, Q.; Ruan, D.; Chi, H.; Zheng, M.; Chao, Y.; Qiu, R.; Yang, Y. Effects of cadmium-resistant plant growth-promoting rhizobacteria and *Funneliformis mosseae* on the cadmium tolerance of tomato (*Lycopersicon esculentum* L.). *Int. J. Phytoremediation* 2019, 22, 451–458, doi:10.1080/15226514.2019.1671796.
- 133. Ogundola, L. Effects of pre- and post-transplant inoculation with *Glomus mosseae* on heavy metal (cadmium) absorption by potted tomato plants. *Middle East J. Sci. Res.* **2006**, *1*, 16–22.
- 134. Kumar, P.; Lucini, L.; Rouphael, Y.; Cardarelli, M.; Kalunke, R.M.; Colla, G. Insight into the role of grafting and arbuscular mycorrhiza on cadmium stress tolerance in tomato. *Front. Plant Sci.* **2015**, *6*, 477.
- 135. Liu, Y.; Zhu, Y.; Chen, B.; Christie, P.; Li, X. Yield and arsenate uptake of arbuscular mycorrhizal tomato colonized by *Glomus mosseae* BEG167 in As spiked soil under glasshouse conditions. *Environ. Int.* **2005**, *31*, 867–873, doi:10.1016/j.envint.2005.05.041.
- Elahi, F.E.; Aminuzzaman, F.M.; Mridha, M.A.U.; Begum, B.; Harun, A.K.M.Y. AMF inoculation reduced arsenic toxicity and increased growth, nutrient uptake and chlorophyll content of tomato grown in arsenic amended soil. *Adv. Environ. Biol.* 2010, 4, 194–200.
- 137. Malekzadeh, P.; Farshian, S.H.; Ordubadi, B. Interaction of arbuscular mycorrhizal fungus (*Glomus intraradices* and *Glomus etunicatum*) with tomato plants grown under copper toxicity. *Afr. J. Biotech.* **2012**, *11*, 10555–10567.
- Burleigh, S.H.; Kristensen, B.K.; Bechmann, I.E. A plasma membrane zinc transporter from *Medicago truncatula* is up-regulated in roots by Zn fertilization, yet down-regulated by arbuscular mycorrhizal colonization. *Plant Mol. Biol.* 2003, *52*, 1077–1088, doi:10.1023/a:1025479701246.
- Tong, Y.; Kneer, R.; Zhu, Y. Vacuolar compartmentalization: A second-generation approach to engineering plants for phytoremediation. *Trends Plant Sci.* 2004, 9, 7–9.