

# Glycosylphosphatidylinositol Anchors from Galactomannan and GPI-Anchored Protein Are Synthesized by Distinct Pathways in *Aspergillus fumigatus*

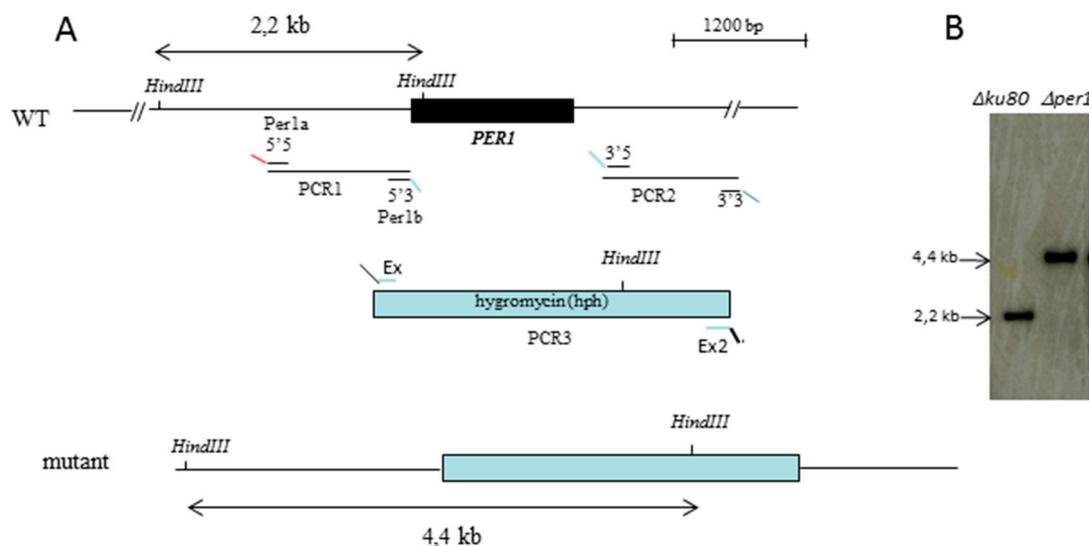
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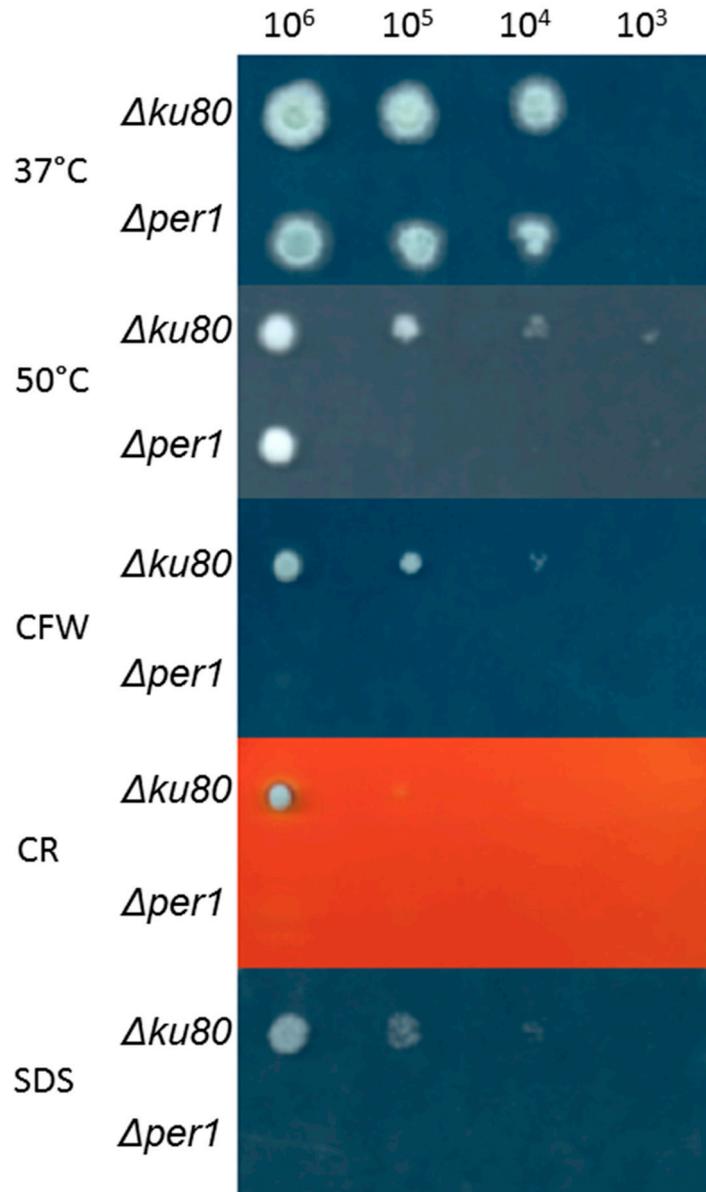
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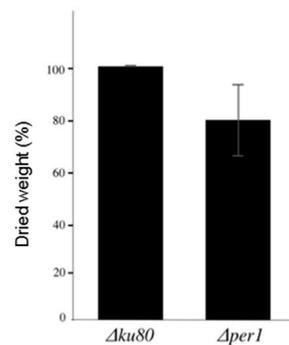
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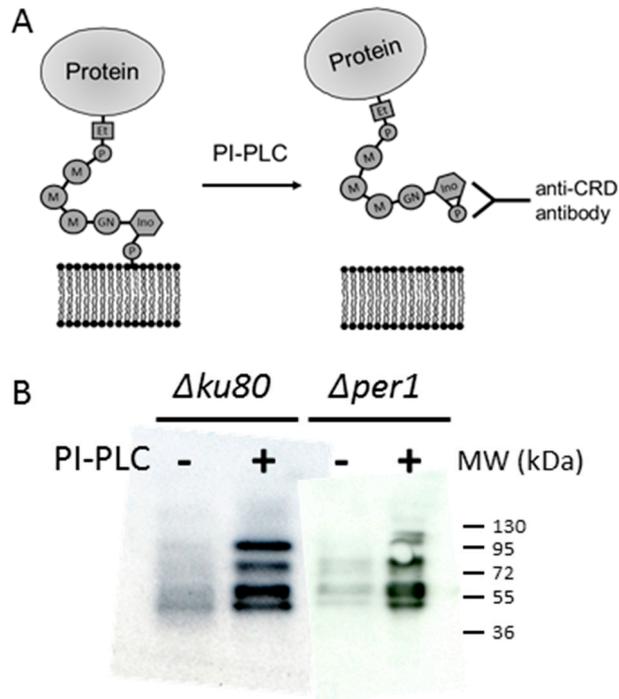
**Figure S1.** (A) Construction of  $\Delta per1$  mutant by PCR fusion. (B) Southern blot analysis: Genomic DNA has been digested by *HindIII* and hybridized with PCR1 probe amplified with primers Per1a-Per1b.



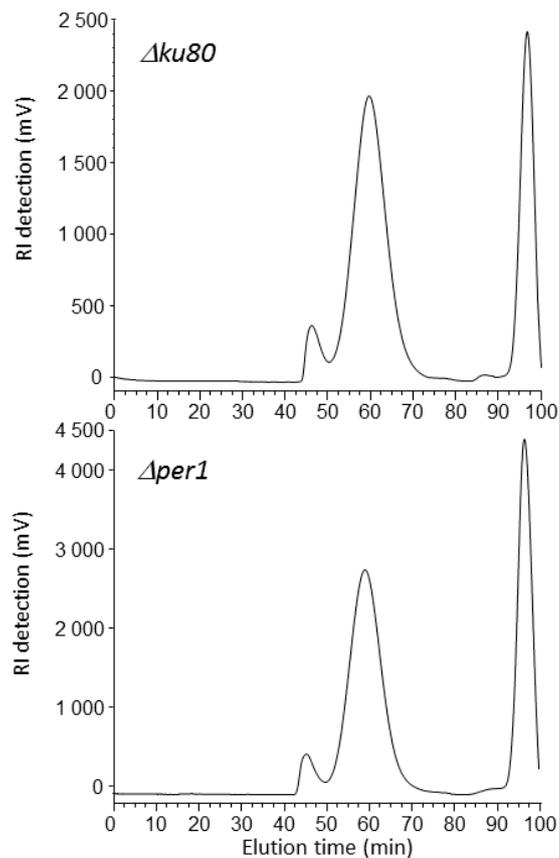
**Figure S2.** Growth of PER1 deletion mutant strain on solid minimum medium. Radial growth of the parental strain and PER1 deletion mutant strain on malt agar medium (48–72 h at 37 °C or 50 °C) with or without calcofluor white (CFW, 40 μg/mL), Congo red (CR, 50 μg/mL), SDS (0.01%).



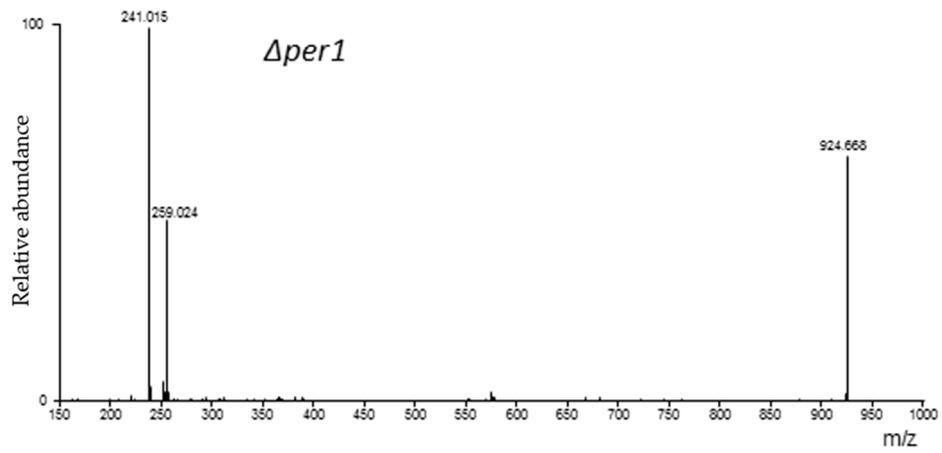
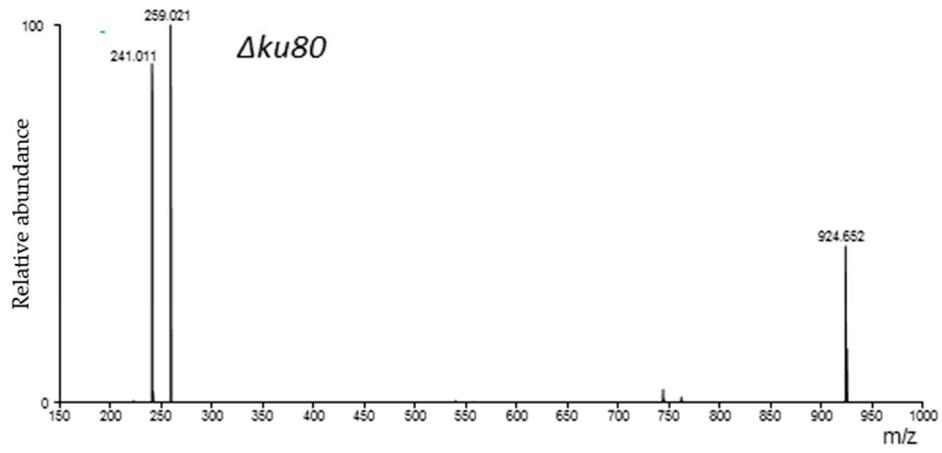
**Figure S3.** Growth of PER1 deletion mutant strain on Liquid Sabouraud medium. Growth was estimated as the measure of the dried weight of biomass.



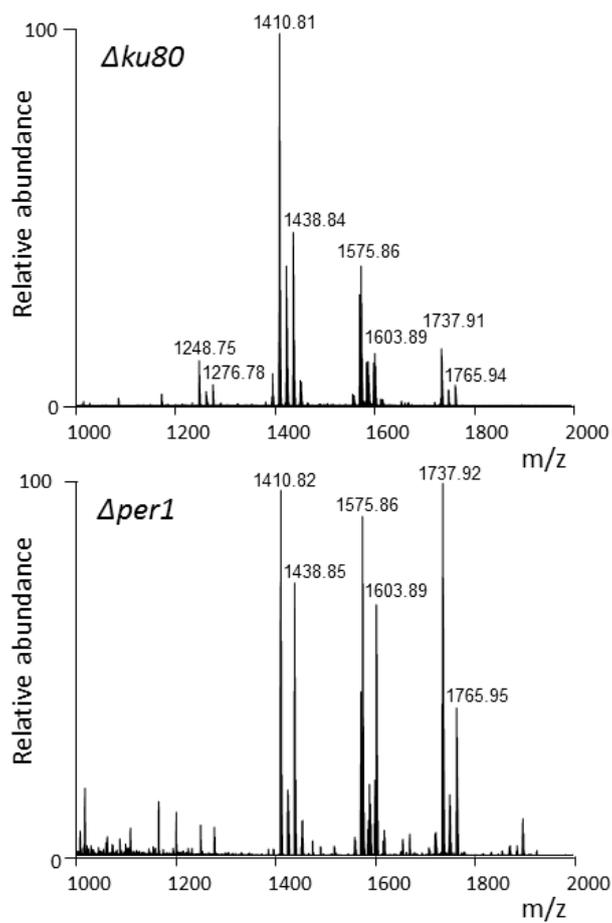
**Figure S4.** Detection of purified GPI-Aps. **(A)** Scheme of detection using a PI-phospholipase C and an anti-CRD antibody. **(B)** Western blot of GPI-APs fraction purified from parental ( $\Delta ku80$ ) and  $\Delta per1$  mutant strains.



**Figure S5.** Gel filtration chromatography on Superdex 75 of purified LGM fraction from  $\Delta ku80$  and  $\Delta per1$  strains. Prior to the analysis, LGM was submitted to a nitrous deamination. Products were detected by a RI detector.



**Figure S6.** MS-MS spectra of the ion  $m/z$  924.650 of the lipid anchor of LGM isolated from the parental strain ( $\Delta ku80$ ) and from the mutant strain ( $\Delta per1$ ).



**Figure S7.** MS spectra of GIPC fraction isolated from parental ( $\Delta ku80$ ) and  $\Delta per1$  mutant strains. The MS data of GIPC fractions from  $\Delta ku80$  and  $\Delta per1$  were similar with the presence of characteristic ions [M-H]<sup>-</sup> at m/z 1410.8, 1572.8, 1575.9 and 1737.9 corresponding to the presence of 3 or 4 hexose residues with or without a choline-phosphate group linked to a IPC [38].

**Table S1.** Primers used in the paper.

Name	Sequence
Primers used to construct the <i>per1Δ::HYG</i> deletion cassette	
AFUB_006580-3'3	GCGGATAACAATTTACACACAGGAAACAGCGATTATGGCTCGCAGTGACC
AFUB_006580-3'5	CTCCTTCAATATCATCTTCTGTCTCCAACACGCGTTCCCAGAATGATGTCTCAAGCCGC
AFUB_006580-5'5	GTAACGCCAGGGTTTTCCCAGTCACGACGCTCATGAATGCCTTAGCACGG
AFUB_006580-5'3	ATCCACTTAACGTTACTGAAATCTCCTTACCATGGTGCTAATGGAGTGG
AFUB_006580-Ex	ATCCACTTAACGTTACTGAAATCTCCTTACCATGGTGCTAATGGAGTGG
AFUB_006580-Ex2	AACGAAGTGTCAGCATCGAGAG
AFUB_006580-MKRr	GGAACGCGTGTGGAGACAGAAGATGATATTGAAGGAG
AFUB_006580-MKRf	GAAGGAGATTCAGTAACGTTAAGTGGAT
Primers used to construct the pNE478 plasmid	
GFPf	GAAGGAGATTCAGTAACGTTAAGTGGATATGGTGAGCAAGGGCGAGGA
GFPPr	AGATCTGGATCCTTTACTTGTACAGCTCGTCC
HygF	AGATCTGTCCAATTGCTTCCGATCTGG
HygR	GTTGGAGACAGAAGATGATATTGAAGGAGCGCGGCCGCGATGAATGTGTCTCTGTAGGC
Primers used to do the DIG probe	
Per1a	CATGAATGCCTTAGCACGG
Per1b	CACTCCATTAGCACCATG

**Table S2.** Global sugar composition of cell wall AI and AS fractions (%).

	%	%
Alkali-insoluble fraction	<i>Δku80</i>	<i>Δper1</i>
Mannose	12.16 +/- 2.19	7.1 +/- 1.01
Glucose	48.81 +/- 3.25	48.17 +/- 2.26
Galactose	10.96 +/- 1.6	8.52 +/- 0.92
GlcNac	27.71 +/- 1.2	36.2 +/- 4.17
GalNAc	0.35 +/- 0.07	0
Alkali-soluble fraction		
Mannose	3.41 +/- 1.21	3.44 +/- 1.59
Glucose	81.5 +/- 3.48	82.6 +/- 4.74
Galactose	9.3 +/- 1.93	7.91 +/- 3.77
GlcNac	0	0
GalNAc	5.8 +/- 1.14	6.05 +/- 0.64