

# **Supplementary Materials for**

## **Bioprospection of Tenellins Produced by the Entomopathogenic Fungus *Beauveria neobassiana***

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## Biofilm inhibition assay

Biofilm inhibition assays were conducted according to the methodology previously described (Soliga *et al.*, 2021), targeting the inhibition of biofilm formation in *Staphylococcus aureus* (DSM 1104).

*S. aureus* were cultured in LB and CASO medium at 37 °C for 18 h, respectively, in a shaker at 120 rpm after being thawed from a -20 °C stock solution. The culture's optical density at 600 nm (OD<sub>600</sub>) was measured and adjusted to a 0.1 McFarland standard in M63 minimal medium supplemented with necessary nutrients and CASO medium with 4% glucose for *S. aureus*, respectively. Microtiter plates were loaded with 150 µL of the bacterial suspension containing serially diluted test compounds (7.8 -125 µg/mL) and incubated for 24 h at 37 °C. The evaluation of biofilm inhibition was performed by staining the biofilm with 0.1% crystal violet (CV). After staining, the plates were washed, and the biofilm dissolved in 95% ethanol. The absorbance of the dissolved biofilm was measured at 530 nm using a plate reader. Negative control involved methanol (2.5%) while microporenic acid A at a concentration range of 7.8 to 125 µg/mL were used as the positive control for *S. aureus* (Chepkirui *et al.*, 2018). All experiments were performed three times, and the standard deviation (SD) was maintained at or below 15% for *S. aureus*.

Differences between samples and the control group were determined by a two-tailed Student's *t*-test. Statistical significance was defined as  $p < 0.01$ . Analysis was carried out using GraphPad Prism 9<sup>®</sup> (GraphPad Software, San Diego, CA, USA).

## References

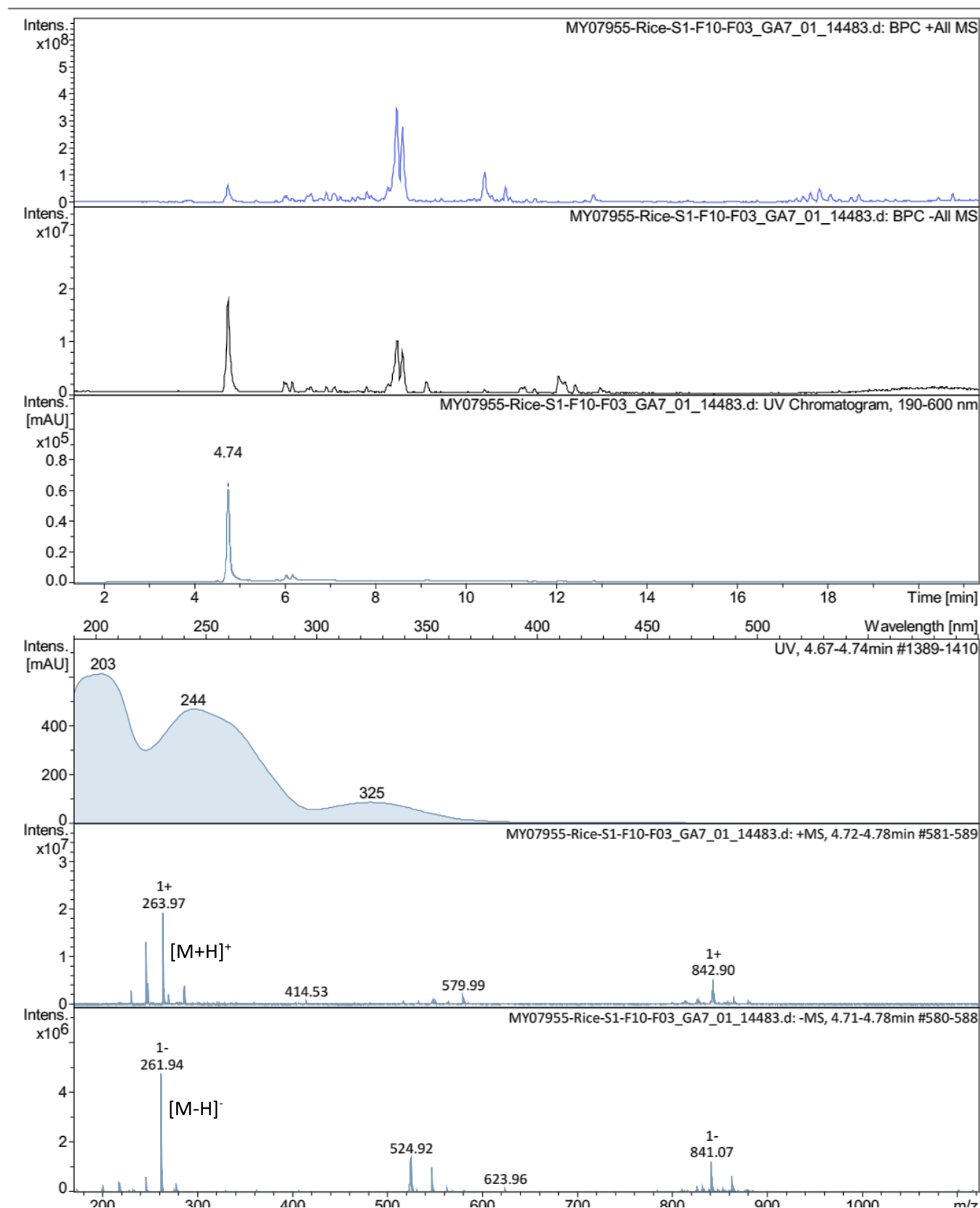
Chepkirui, C.; Yuyama, K.T.; Wanga, L.A.; Decock, C.; Matasyoh, J.C.; Abraham, W.R.; Stadler, M. Microporenic acids A-G, biofilm inhibitors, and antimicrobial agents from the Basidiomycete *Microporus* Species. *J. Nat. Prod.* **2018**, *81*, 778-784.

Soliga, KJ.; Bär, SI.; Oberhuber, N.; Zeng, H.; Schrey, H.; Schobert, R. Synthesis and bioactivity of ancorinoside B, A marine diglycosyl tetramic acid. *Mar. Drugs*. **2021**, *19*, 583.

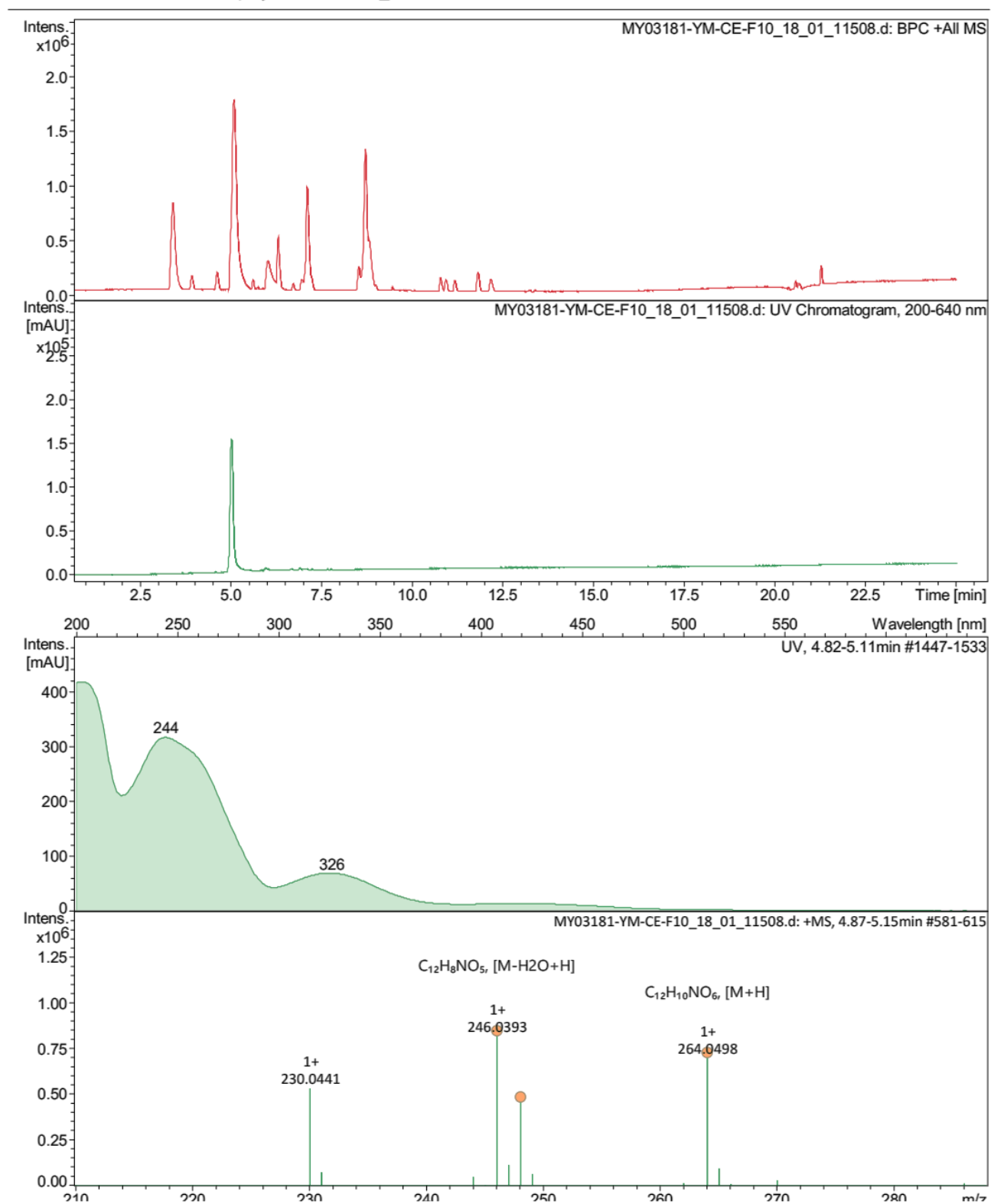
**Table S1.** Inhibition of biofilm formation of *S. aureus* by compounds **1-6** at different concentrations.

Tested organism	Conc. (µg/ml)	Biofilm inhibition (% ±SD)						Microporenic acid A (MAA)
		1	2	3	4	5	6	
<i>S. aureus</i> (DSM 1104)	125	74 ± 5	84 ± 17	83 ± 6	78 ± 5	86 ± 1	64 ± 14	86 ± 0
	62.5	63 ± 16	83 ± 9	83 ± 6	80 ± 4	85 ± 2	48 ± 5	84 ± 1
	31.25	46 ± 9	80 ± 10	79 ± 8	73 ± 15	78 ± 5	-	85 ± 1
	15.62	-	56 ± 8	52 ± 6	51 ± 19	47 ± 12	-	70 ± 13
	7.8	-	53 ± 7	37 ± 7	36 ± 13	-	-	41 ± 9

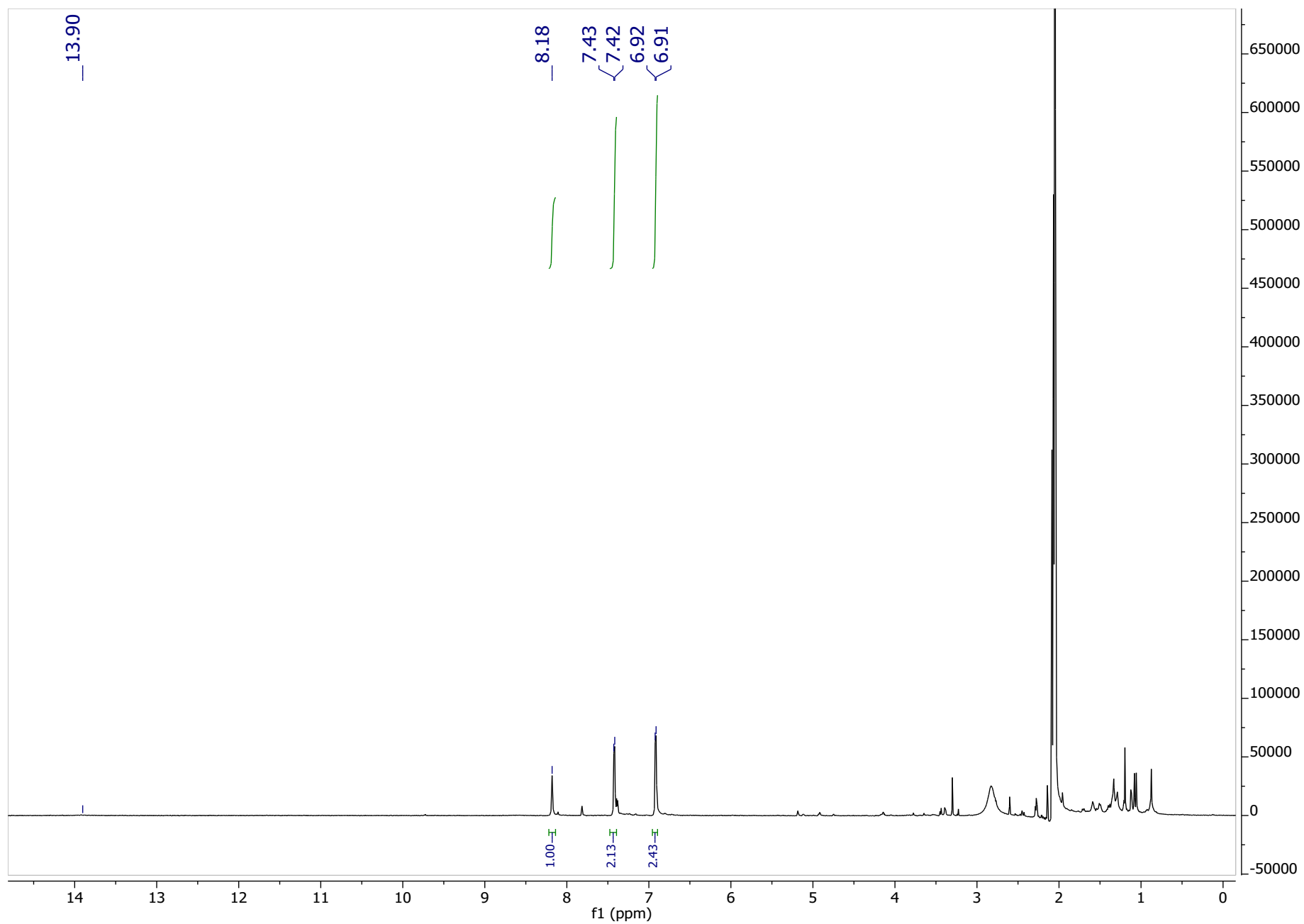




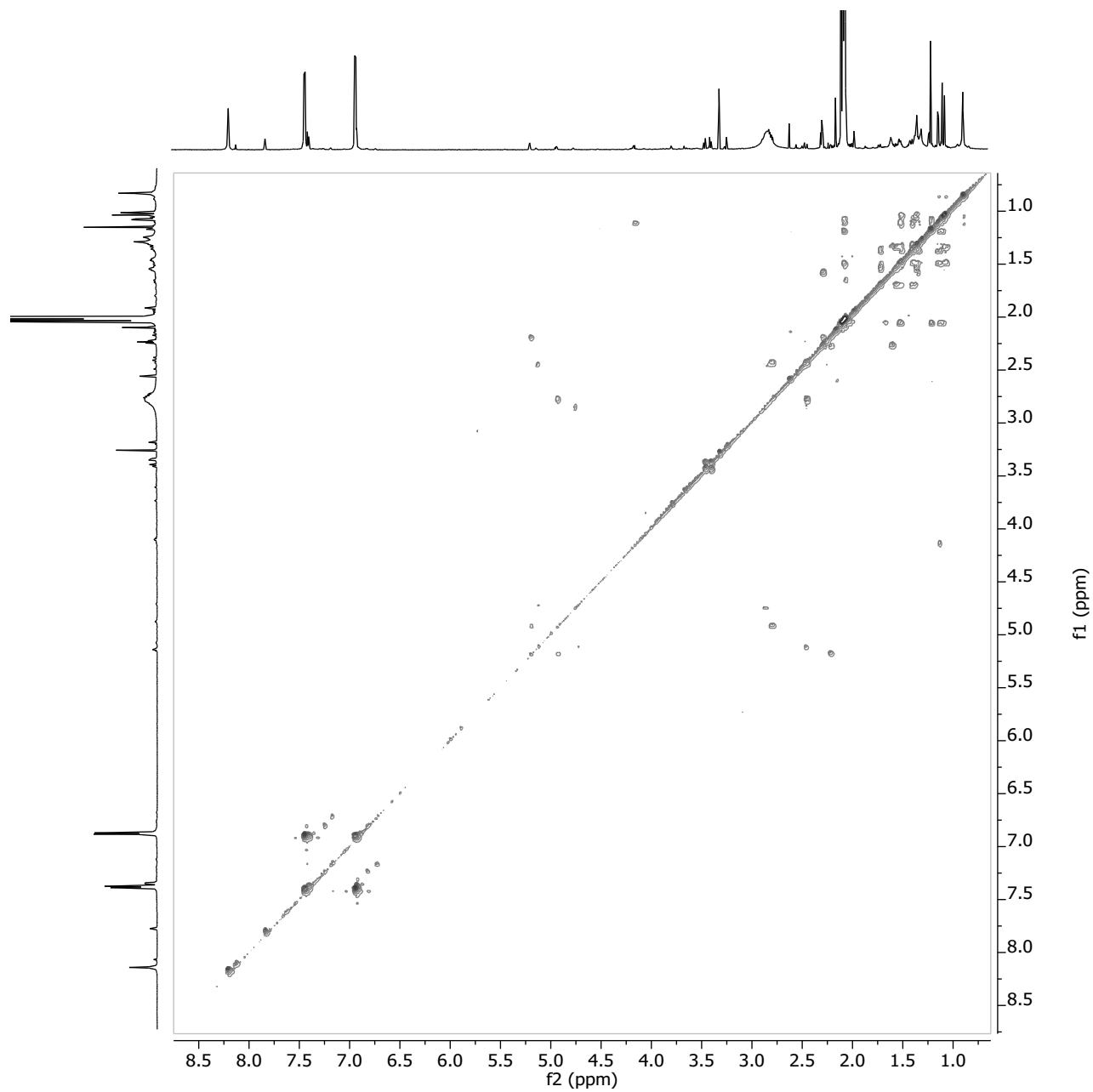
**Figure S1. LRESIMS of 1.**



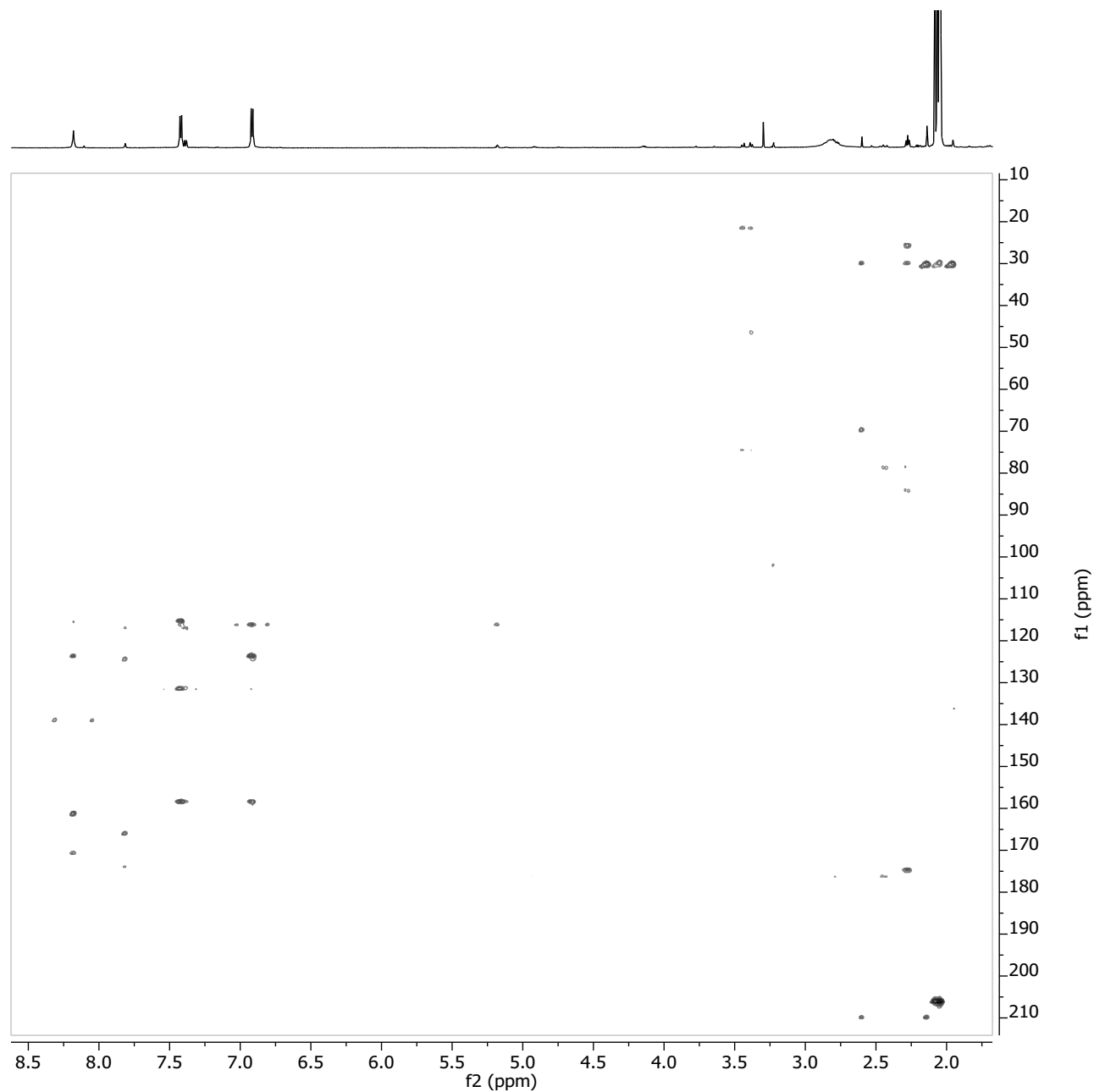
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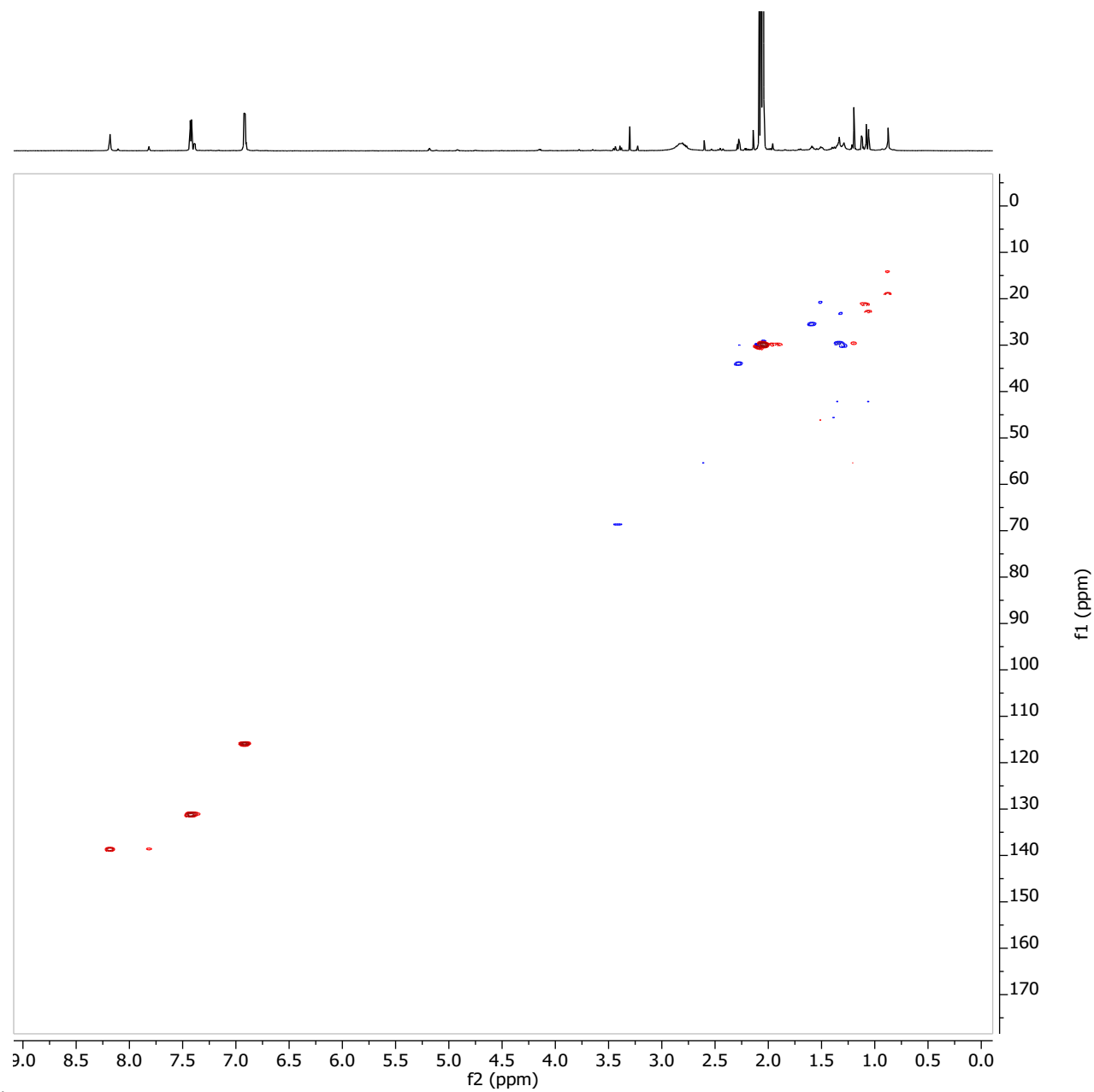
**Figure S3.** <sup>1</sup>H NMR spectrum of **1** in acetone-*d*<sub>6</sub> at 500 MHz.



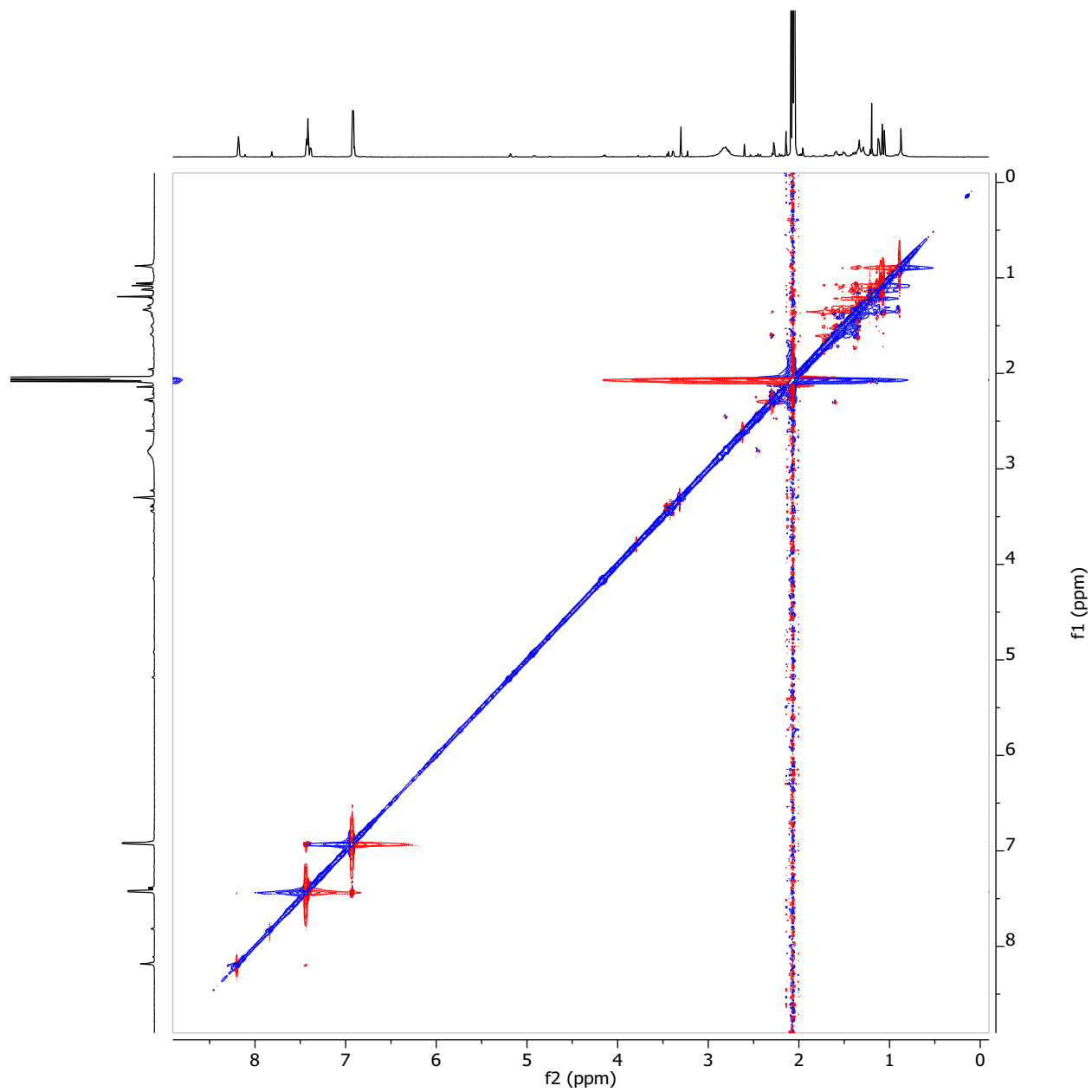
**Figure S4.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of **1** in acetone- $d_6$  at 500 MHz.



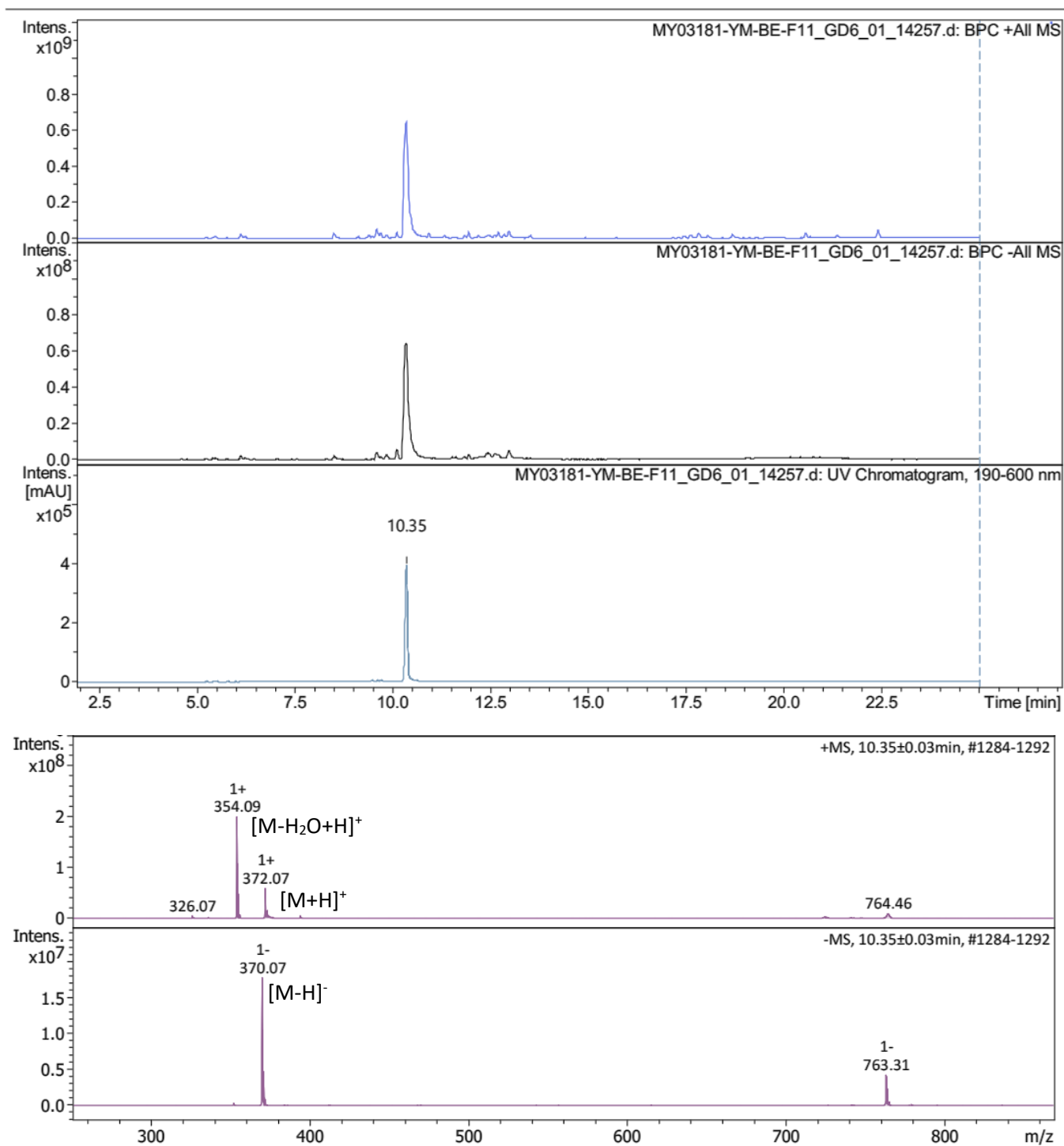
**Figure S5.** HMBC spectrum of **1** in acetone- $d_6$  at 500 MHz.



**Figure S6.** HSQC spectrum of **1** in acetone- $d_6$  at 500 MHz.



**Figure S7.** ROESY spectrum of **1** in acetone- $d_6$  at 700 MHz.



**Figure S8. LRESIMS of 2.**



## Generic Display Report

### Analysis Info

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Acquisition Date 28.05.2023 07:28:36

Operator ate06

Instrument maXis

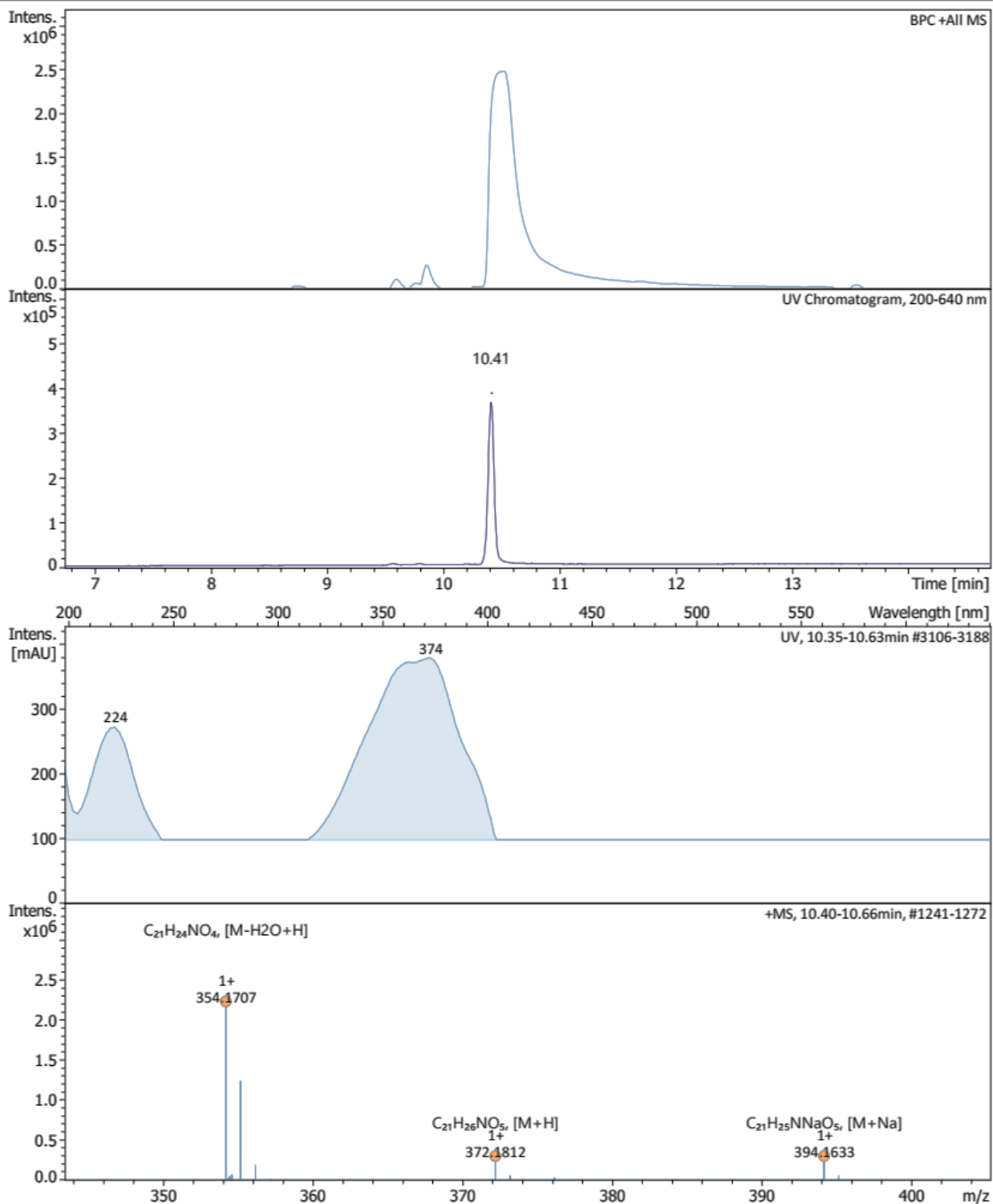
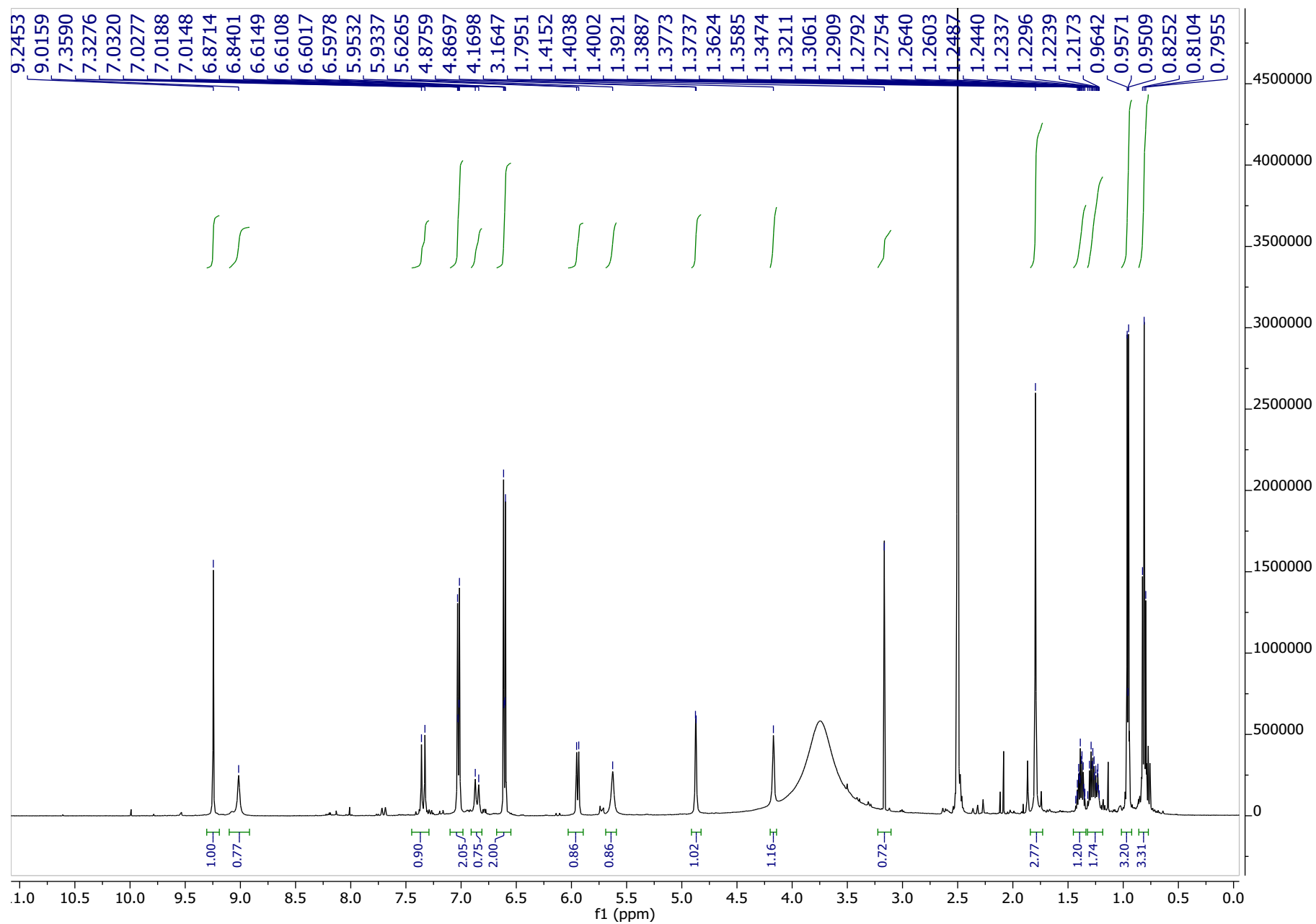
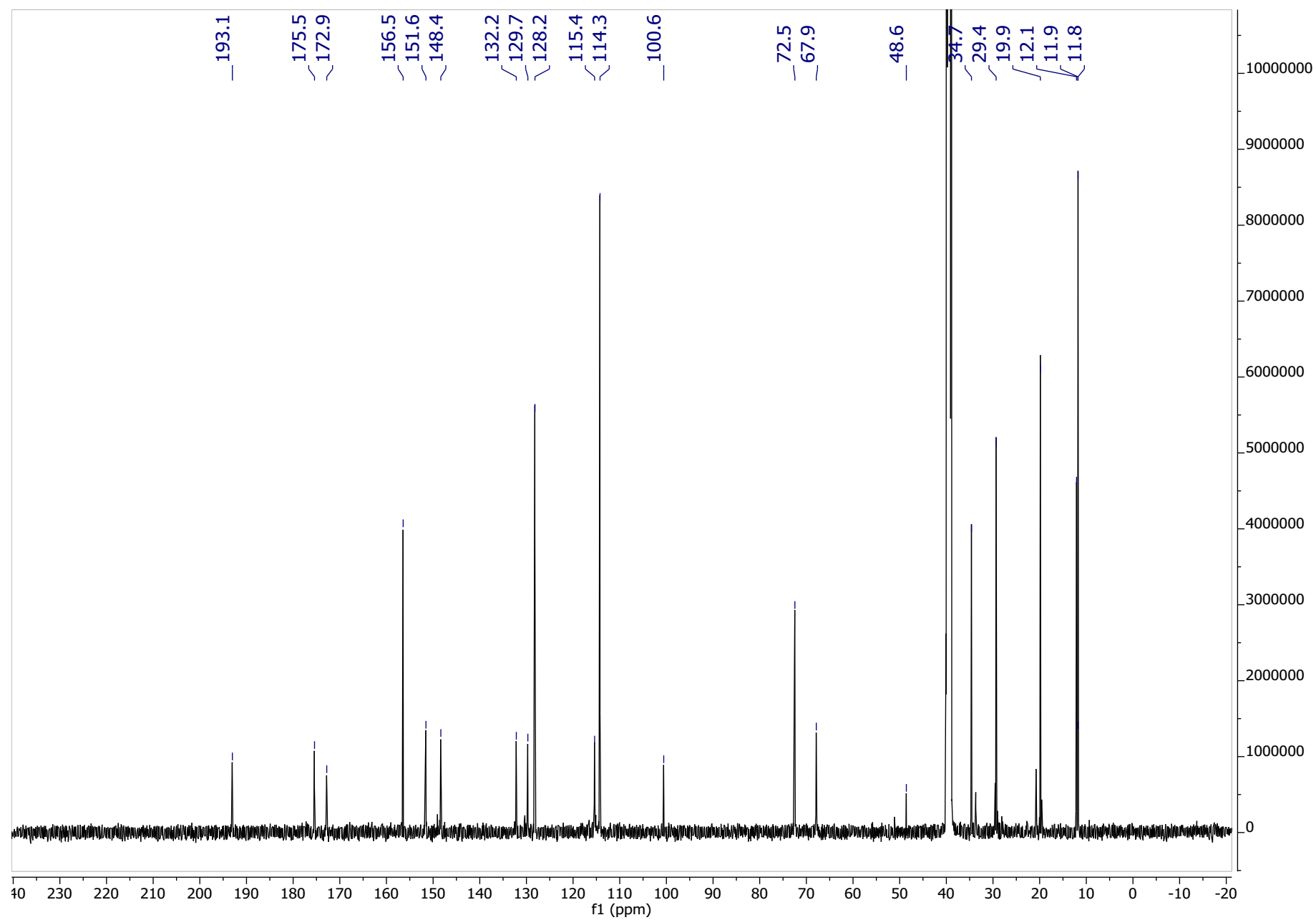


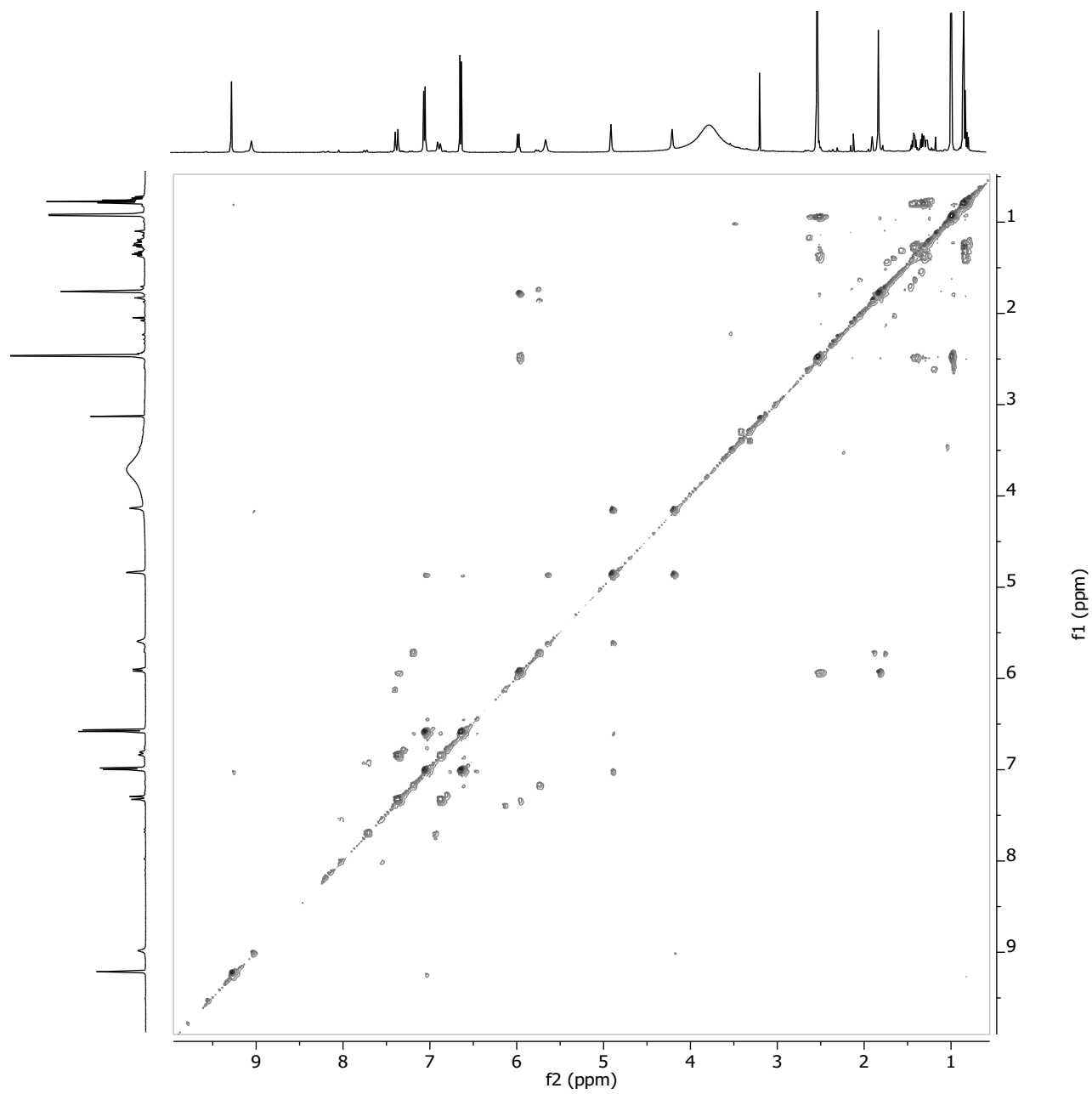
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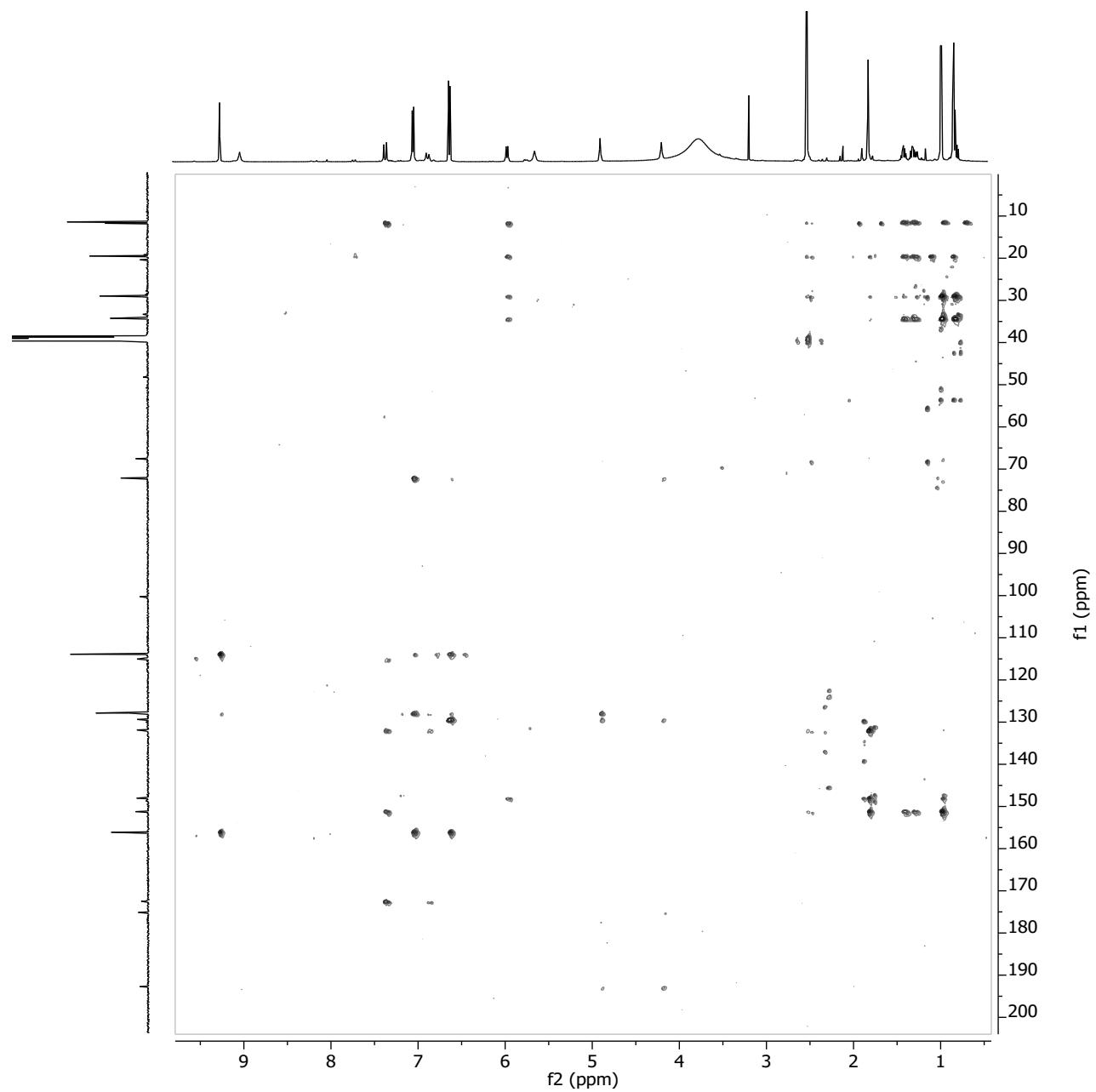
**Figure S10.**  $^1\text{H}$  NMR spectrum of **2** in  $\text{DMSO-}d_6$  at 500 MHz.



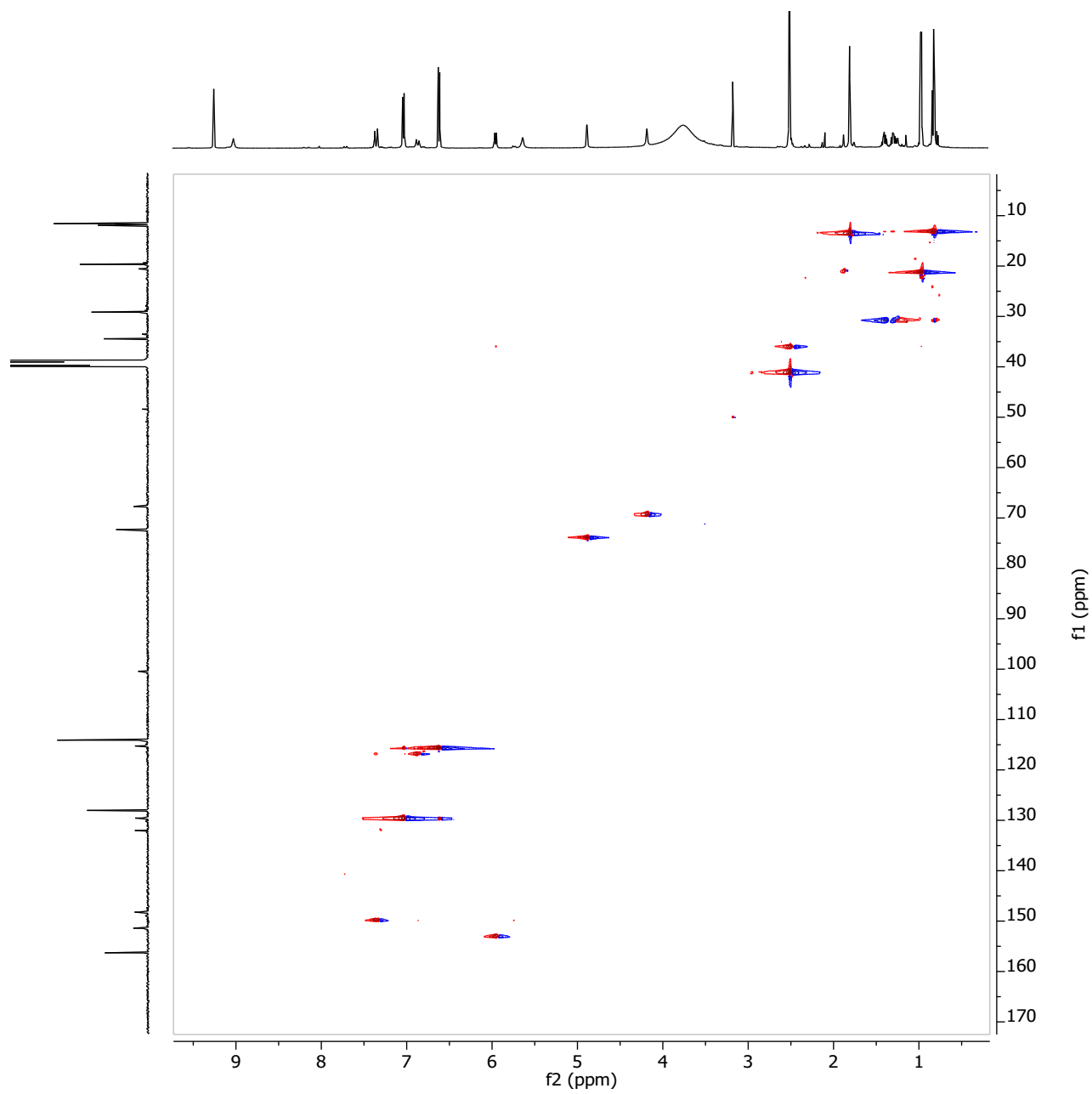
**Figure S11.** <sup>1</sup>H NMR spectrum of **2** in DMSO-*d*<sub>6</sub> at 125 MHz.



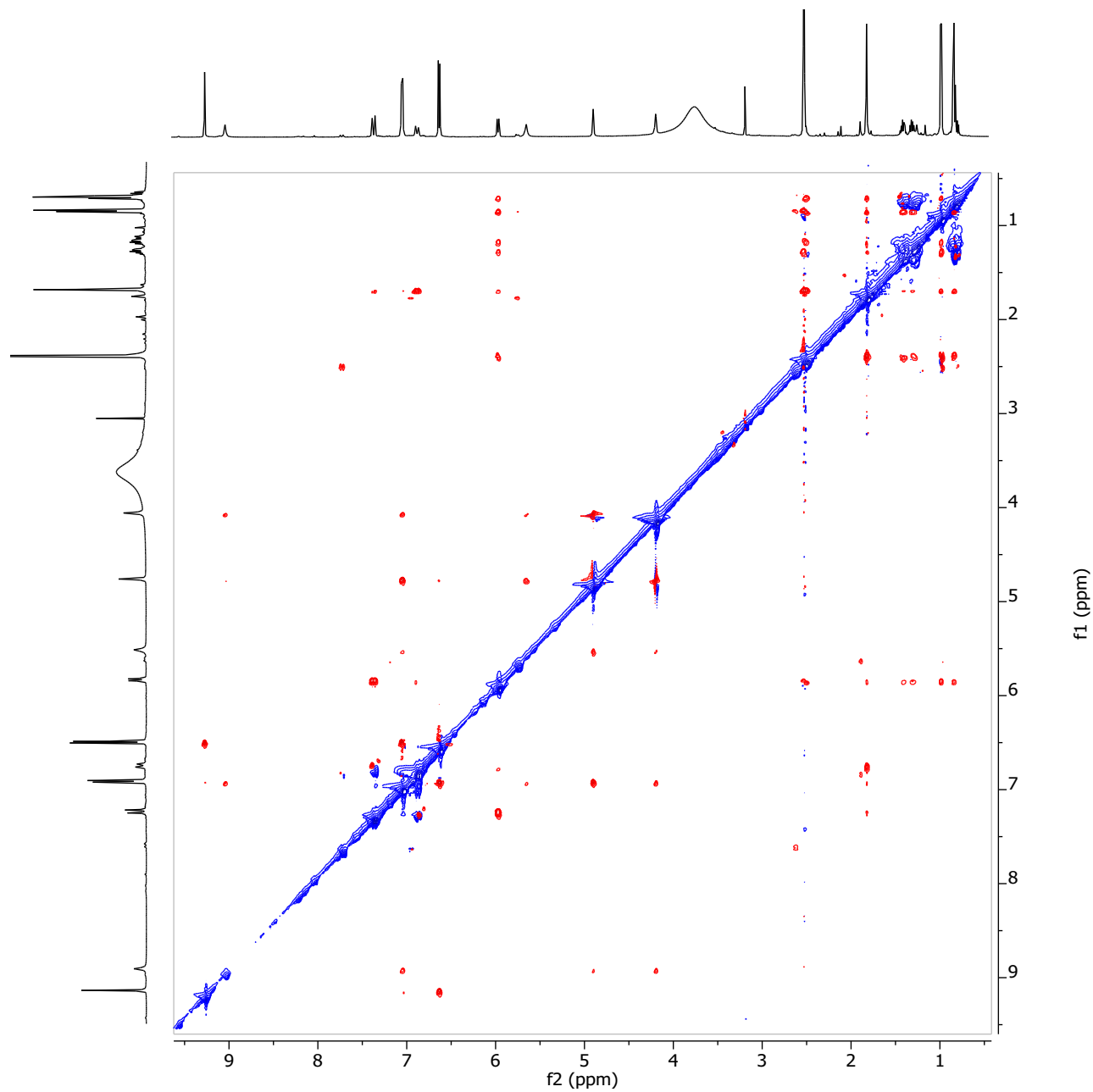
**Figure S12.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of **2** in  $\text{DMSO-}d_6$  at 500 MHz.



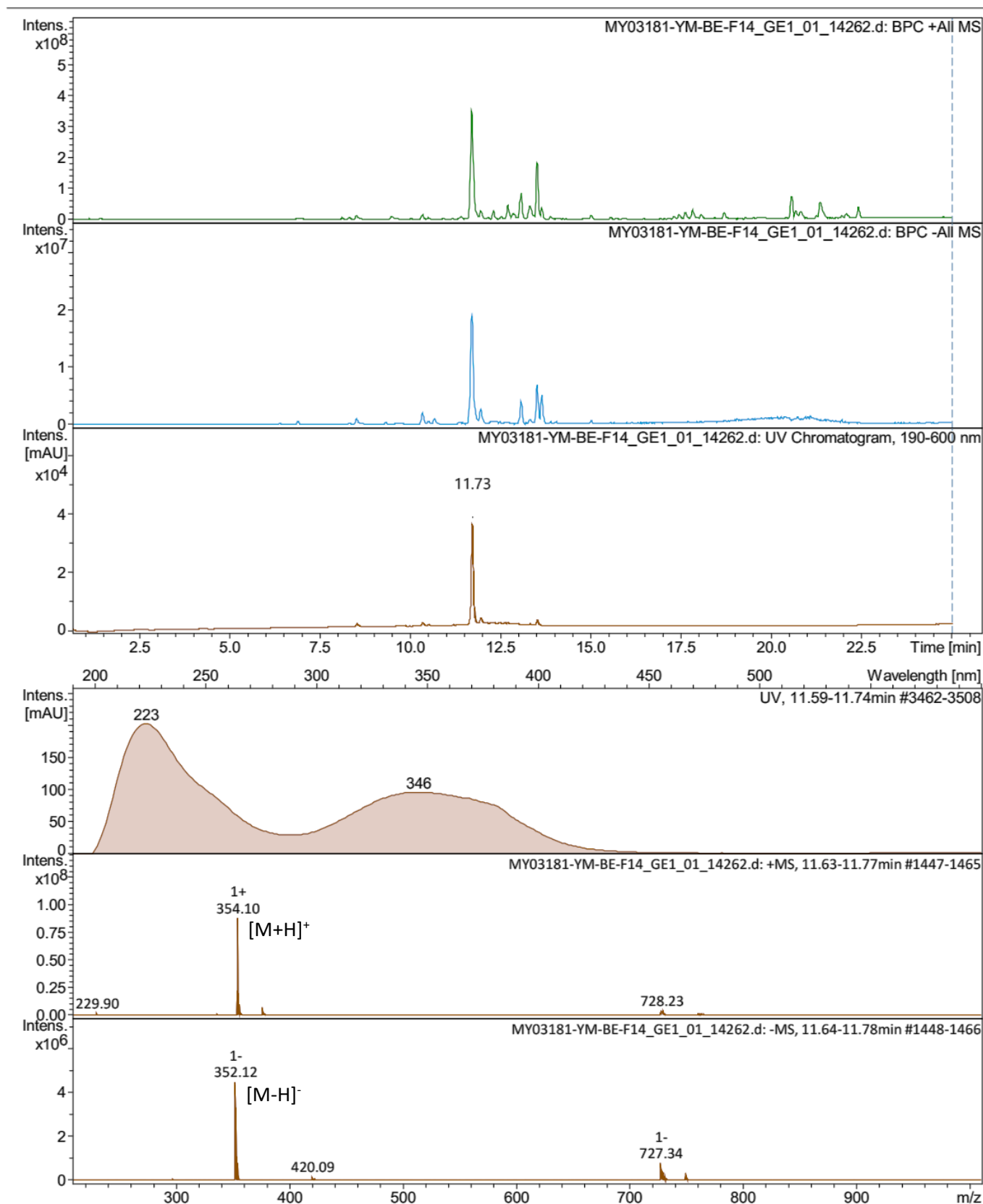
**Figure S13.** HMBC spectrum of **2** in DMSO-*d*<sub>6</sub> at 500 MHz.



**Figure S14.** HSQC spectrum of **2** in DMSO-*d*<sub>6</sub> at 500 MHz.



**Figure S15.** ROESY spectrum of **2** in DMSO-*d*<sub>6</sub> at 700 MHz.



**Figure S16.** LRESIMS of **3**.



## Generic Display Report

### Analysis Info

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Operator ate06

Instrument maXis

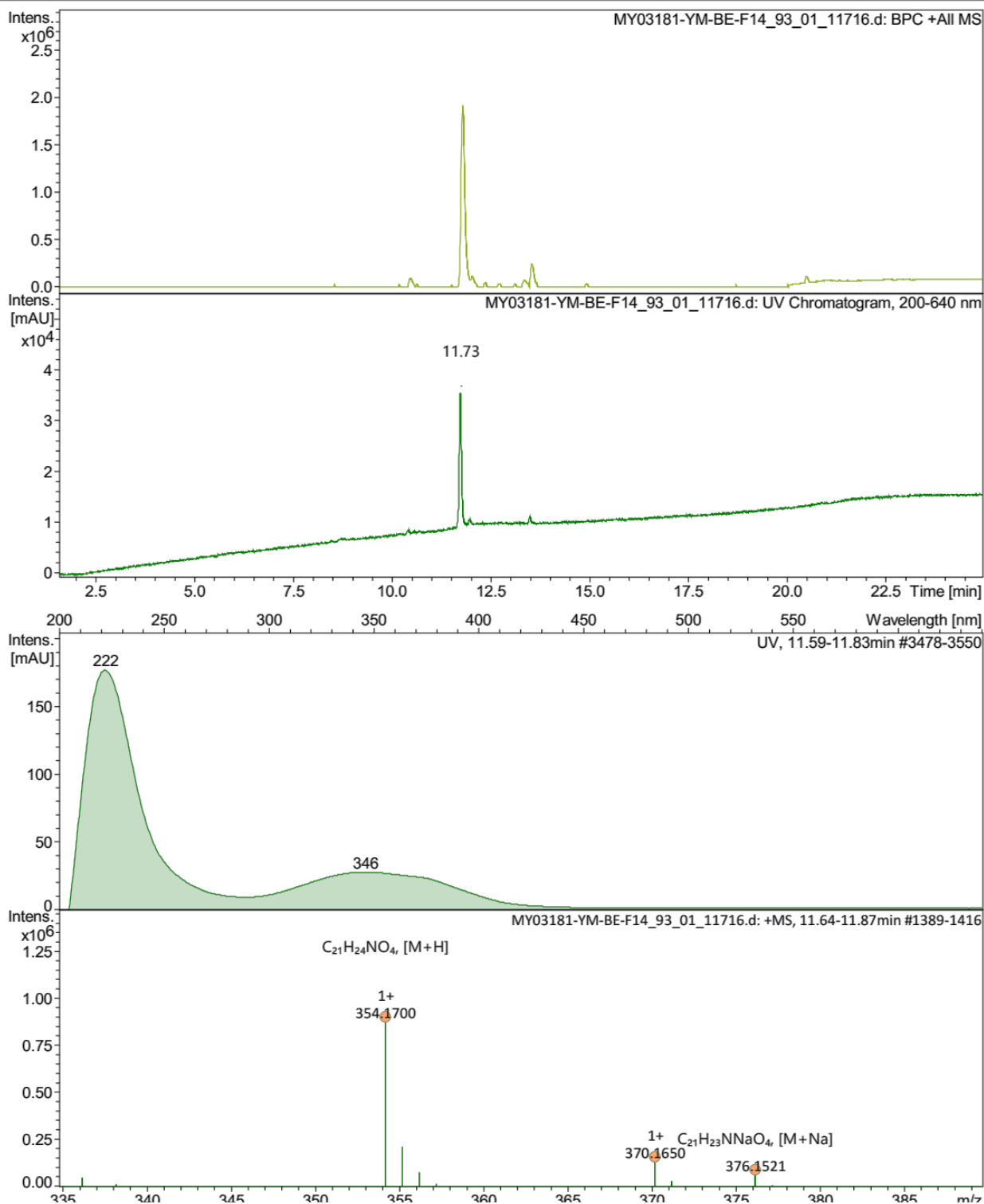
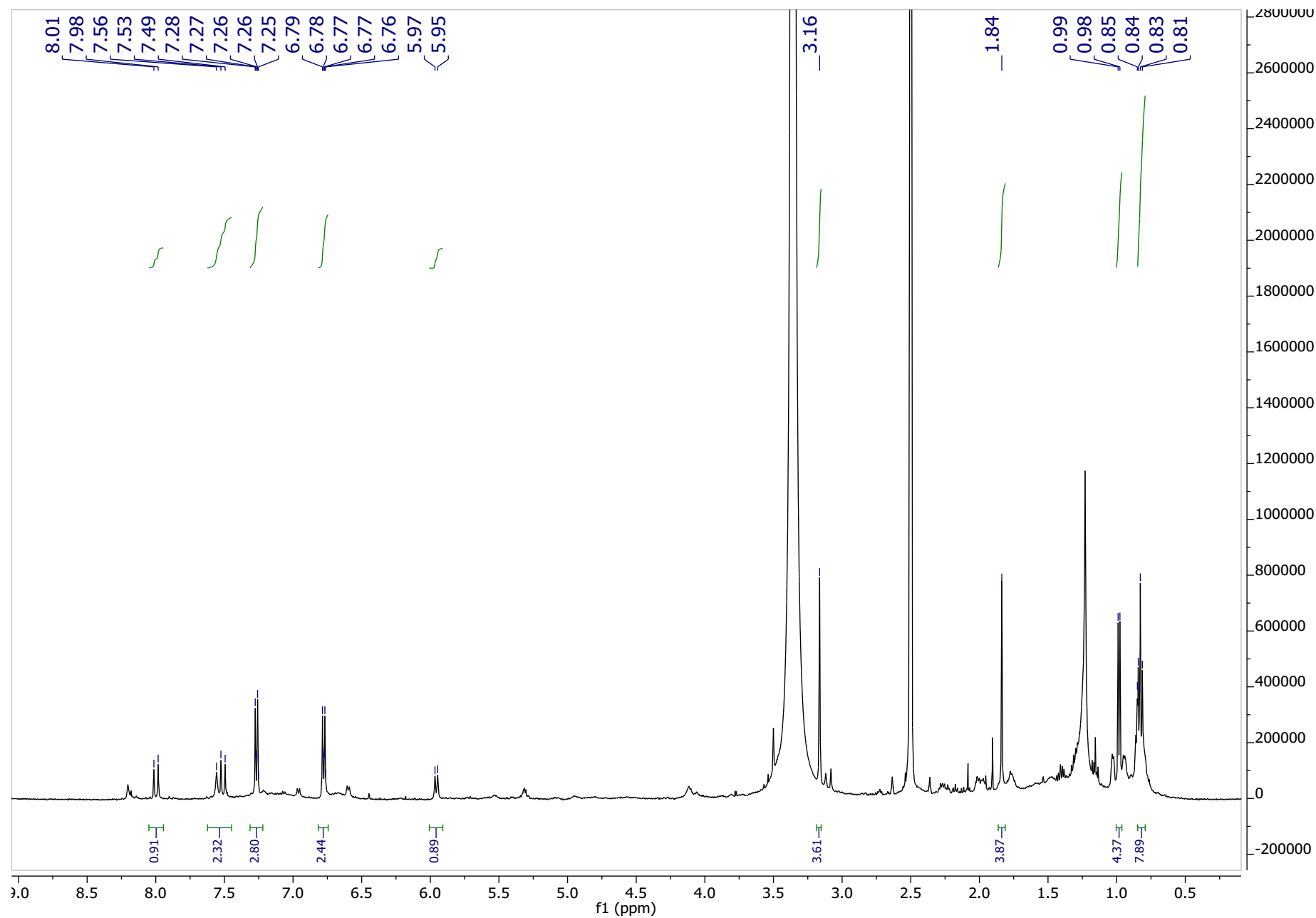
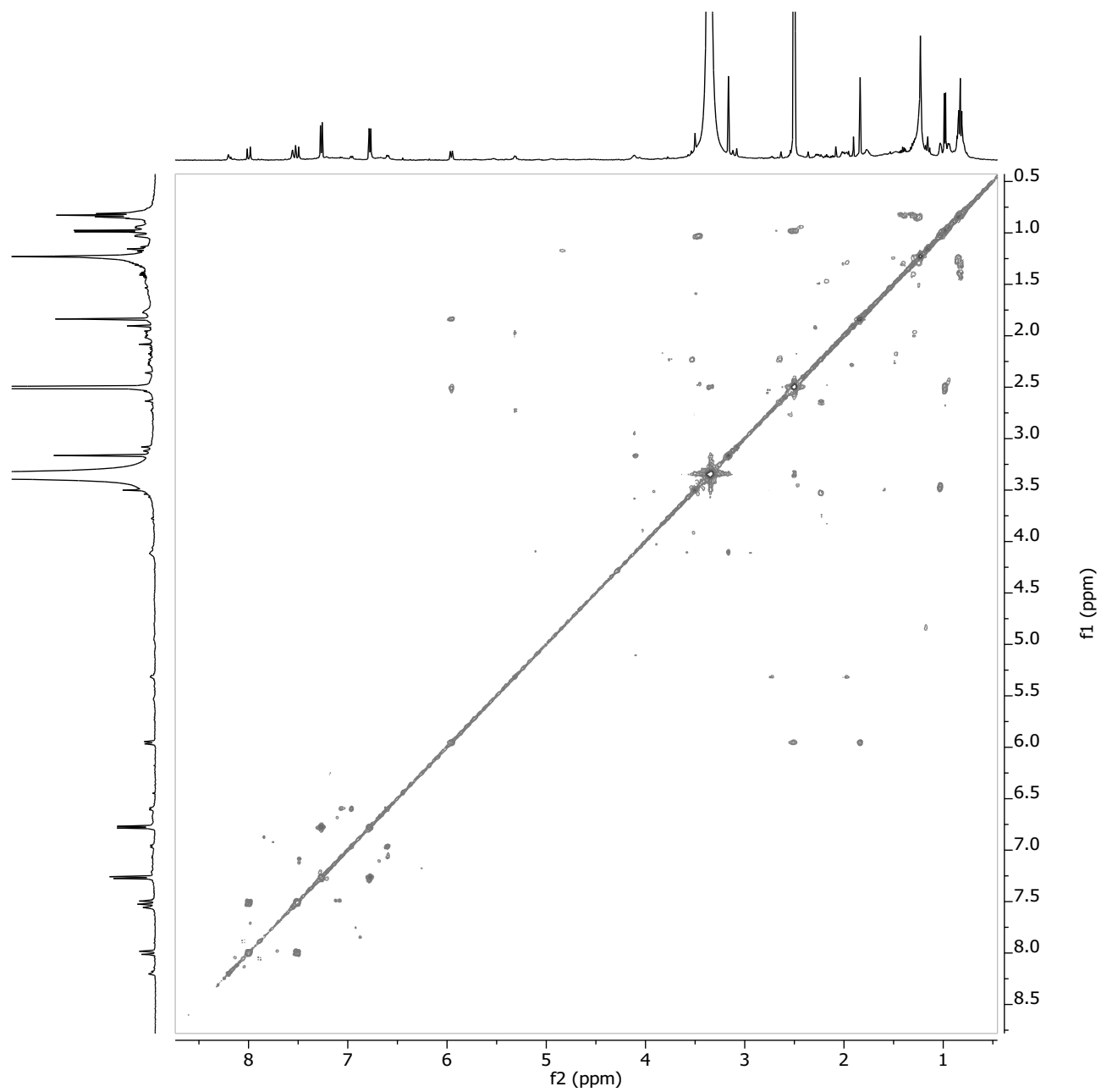


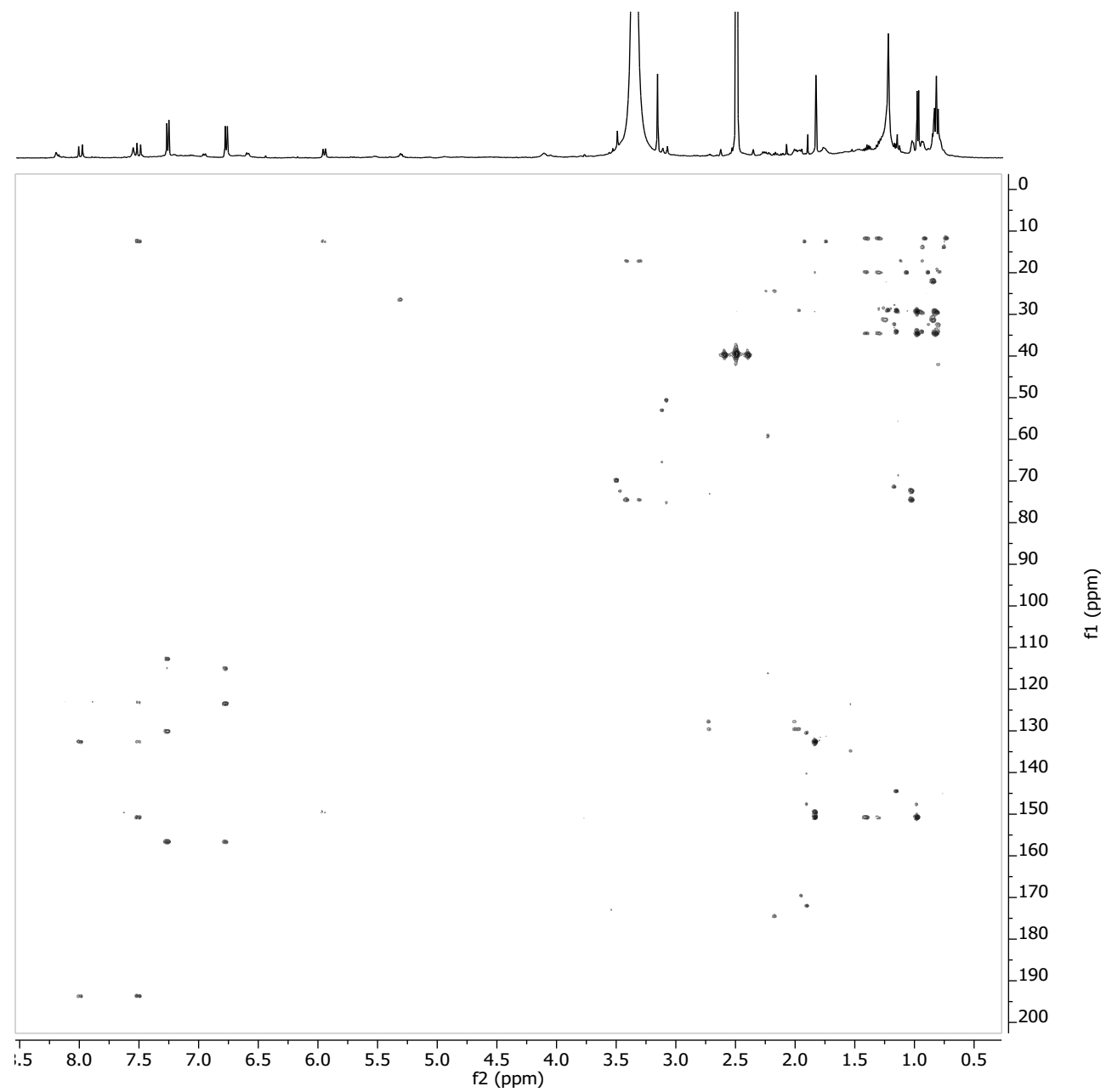
Figure S17. HRESIMS of 3.



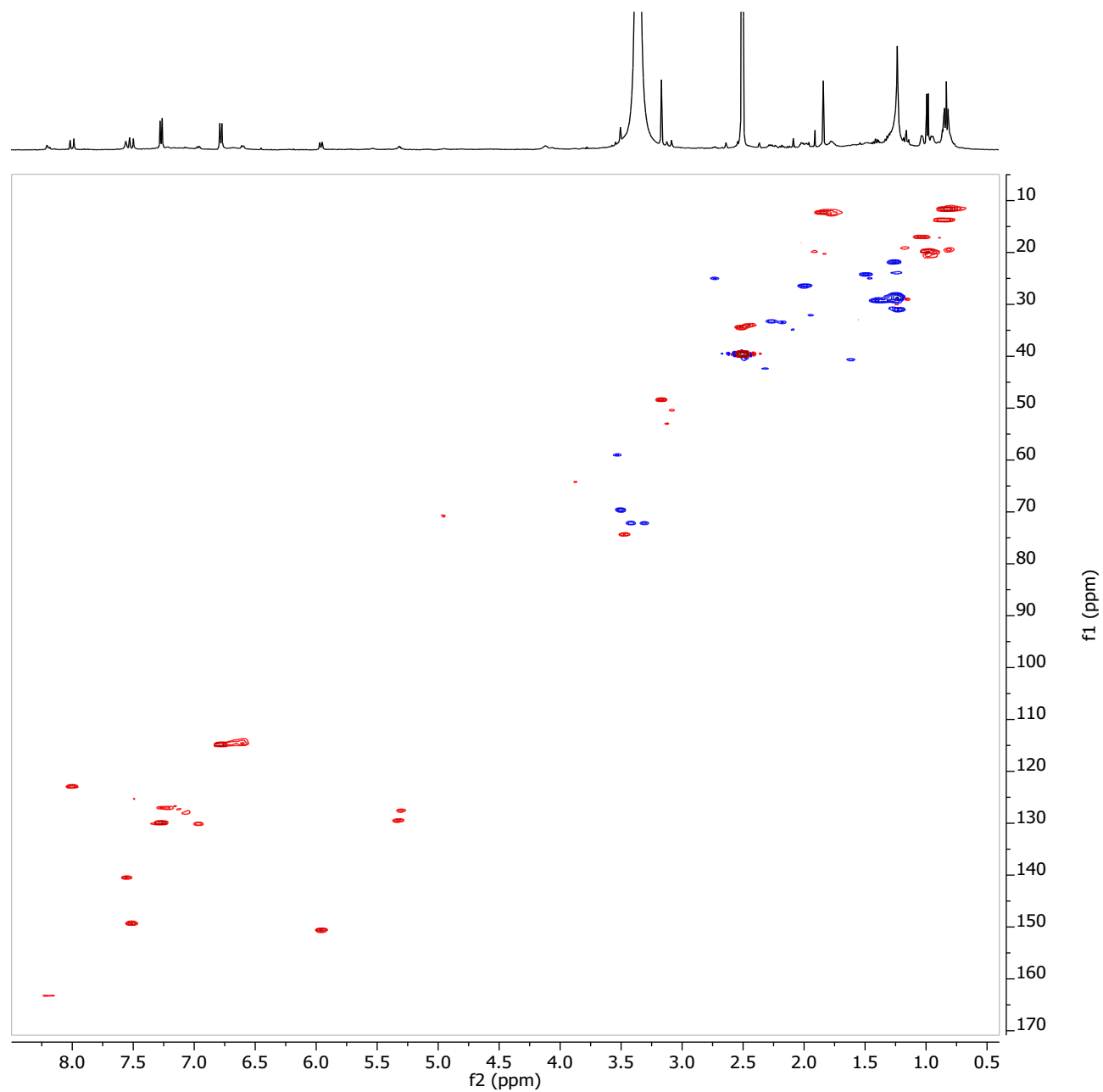
**Figure S18.**  $^1\text{H}$  NMR spectrum of **3** in  $\text{DMSO-}d_6$  at 500 MHz.



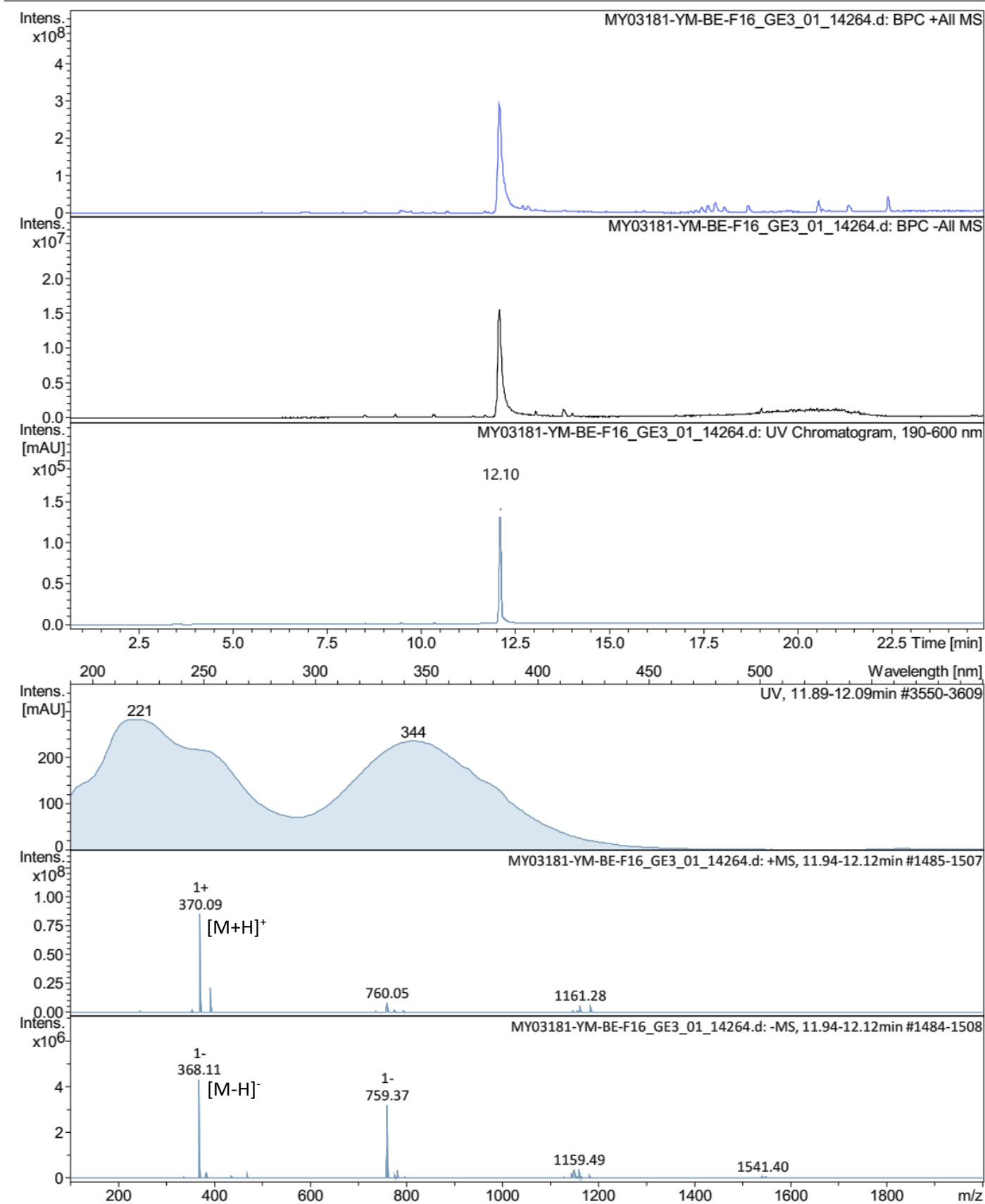
**Figure S19.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of **3** in  $\text{DMSO}-d_6$  at 500 MHz.



**Figure S20.** HMBC spectrum of **3** in DMSO-*d*<sub>6</sub> at 500 MHz.



**Figure S21.** HSQC spectrum of **3** in  $\text{DMSO-}d_6$  at 500 MHz.



**Figure S22. LRESIMS of 4.**

## Generic Display Report

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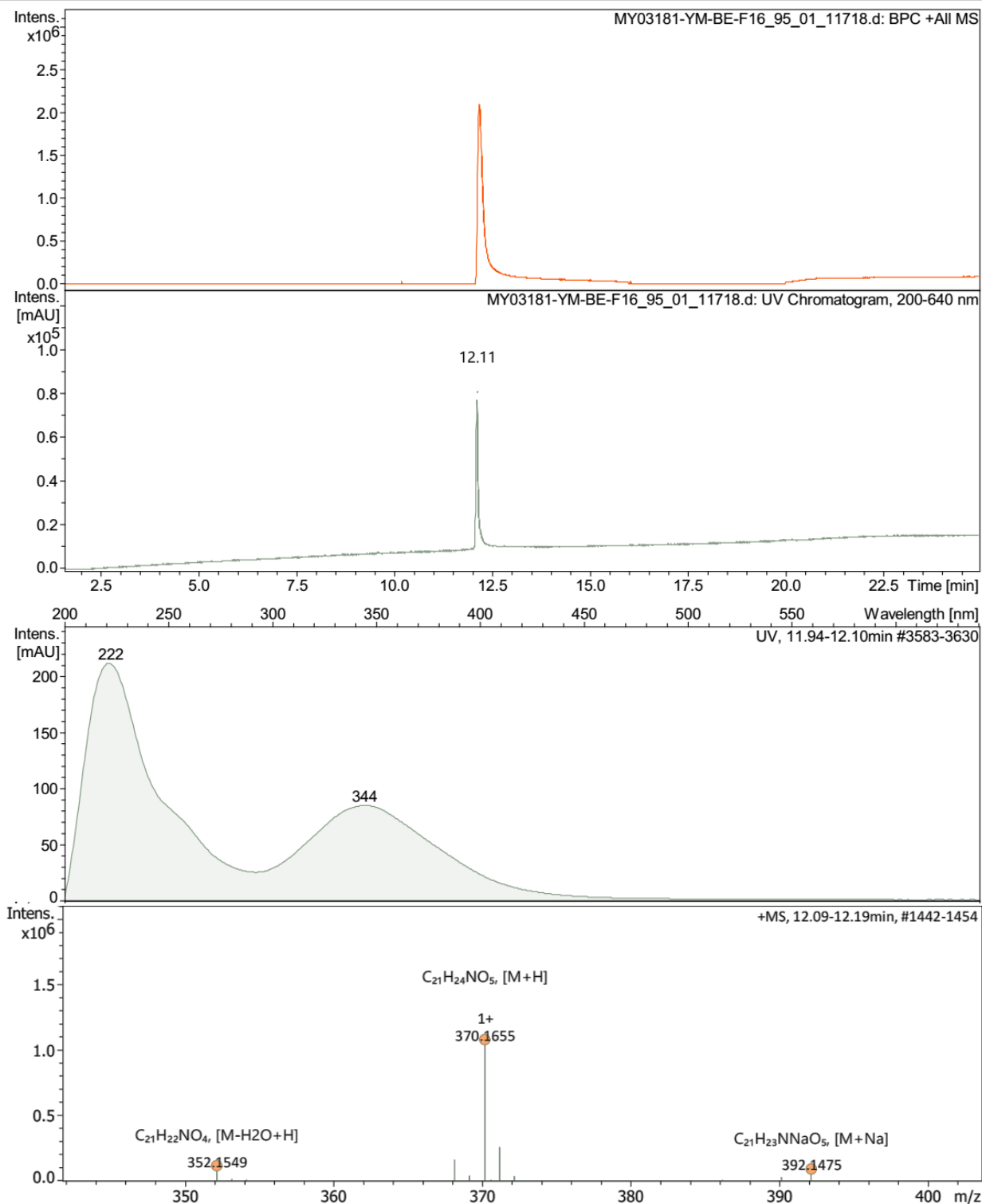
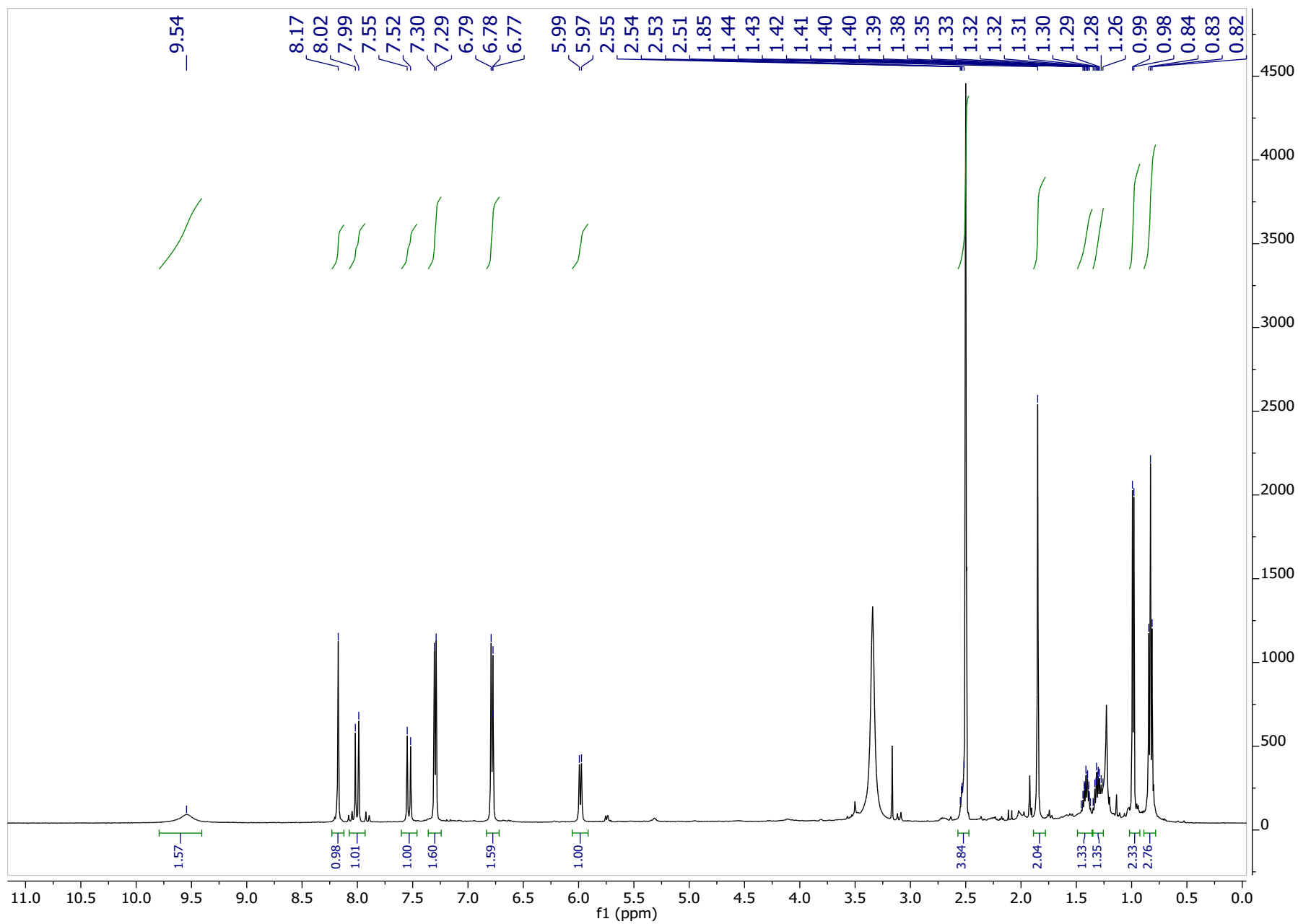
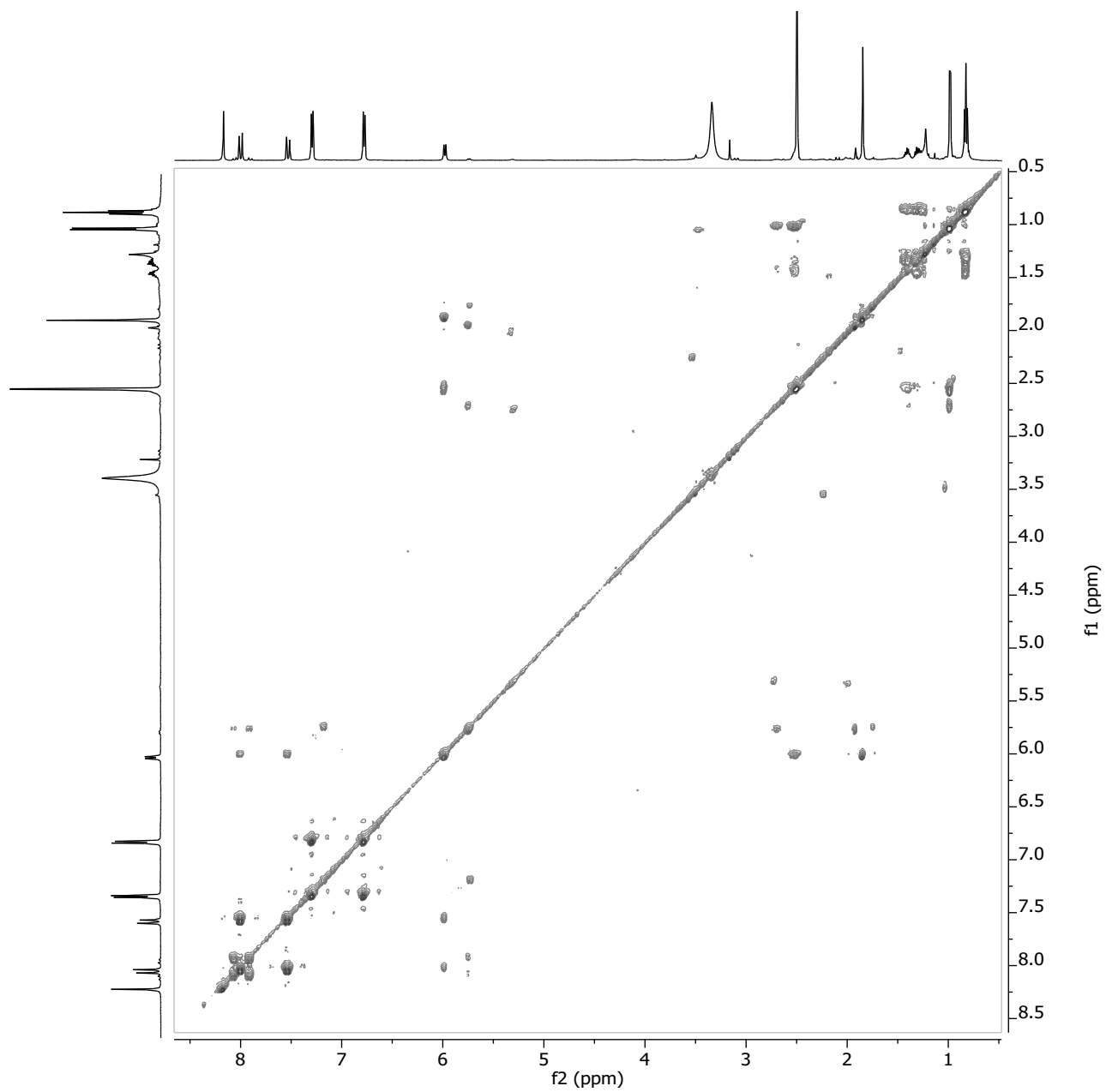


Figure S23. HRESIMS of 4.

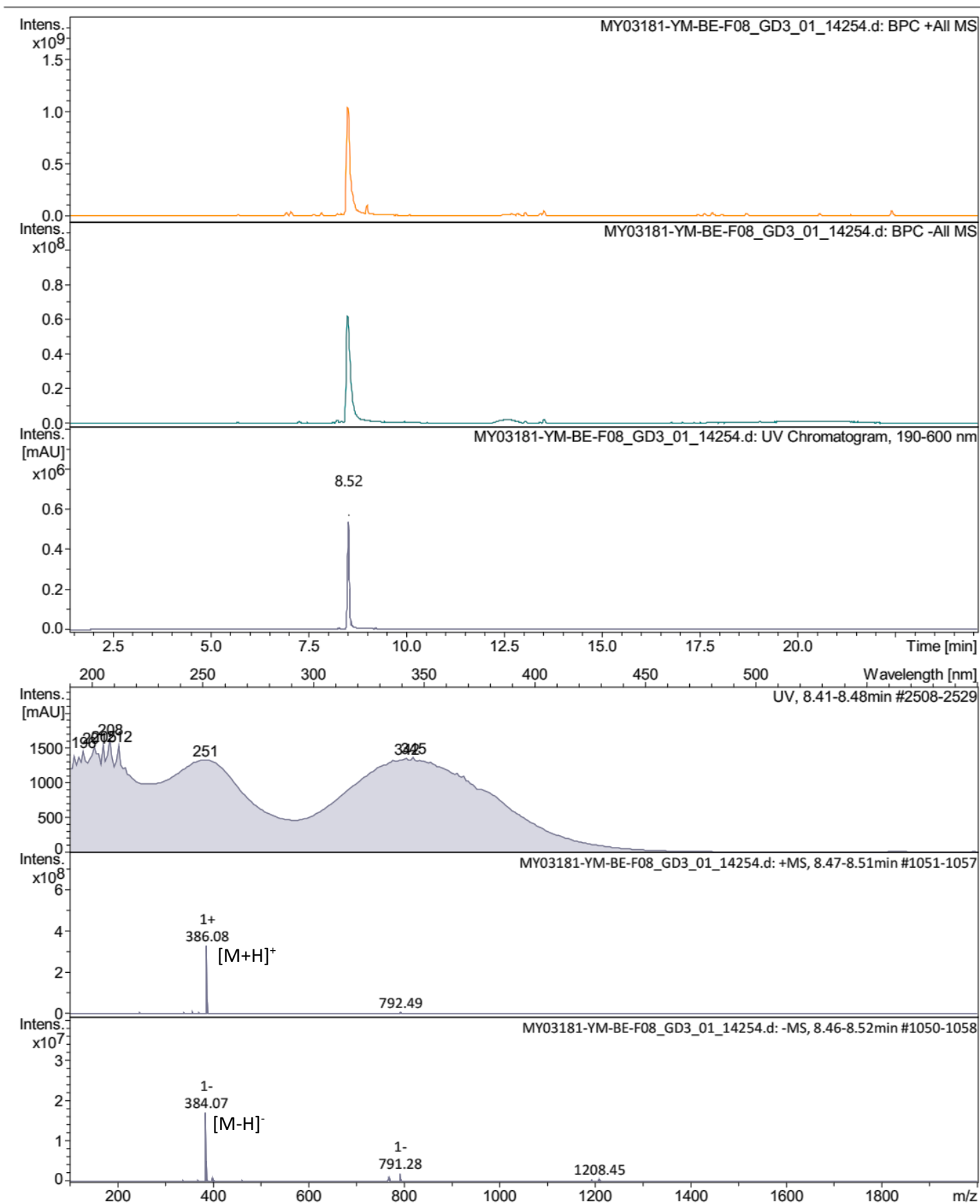


**Figure S24.**  $^1\text{H}$  NMR spectrum of **4** in  $\text{DMSO}-d_6$  at 500 MHz.





**Figure S25.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of **4** in  $\text{DMSO}-d_6$  at 500 MHz.



**Figure S26. LRESIMS of 5.**

## Generic Display Report

### Analysis Info

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Instrument maXis

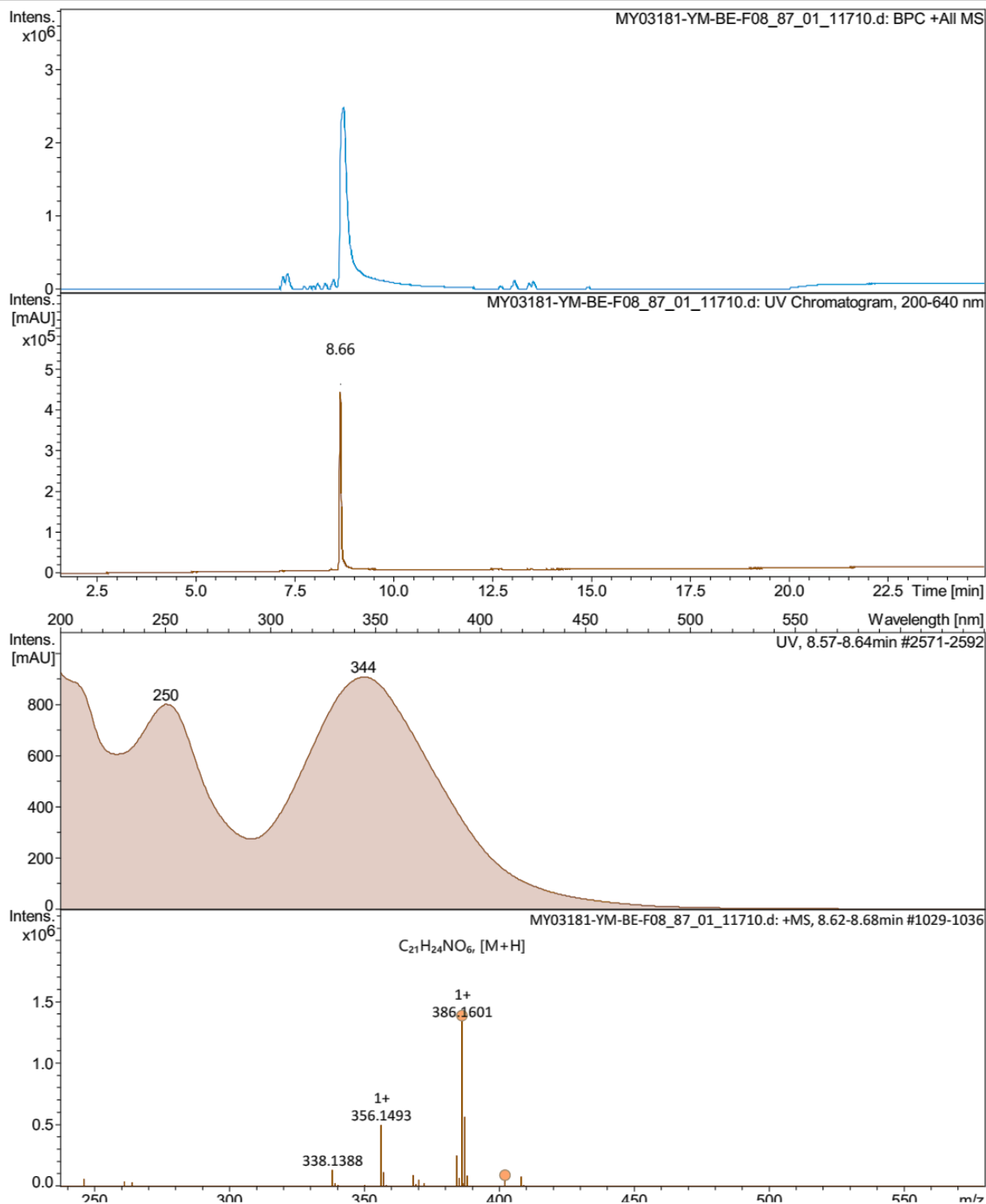
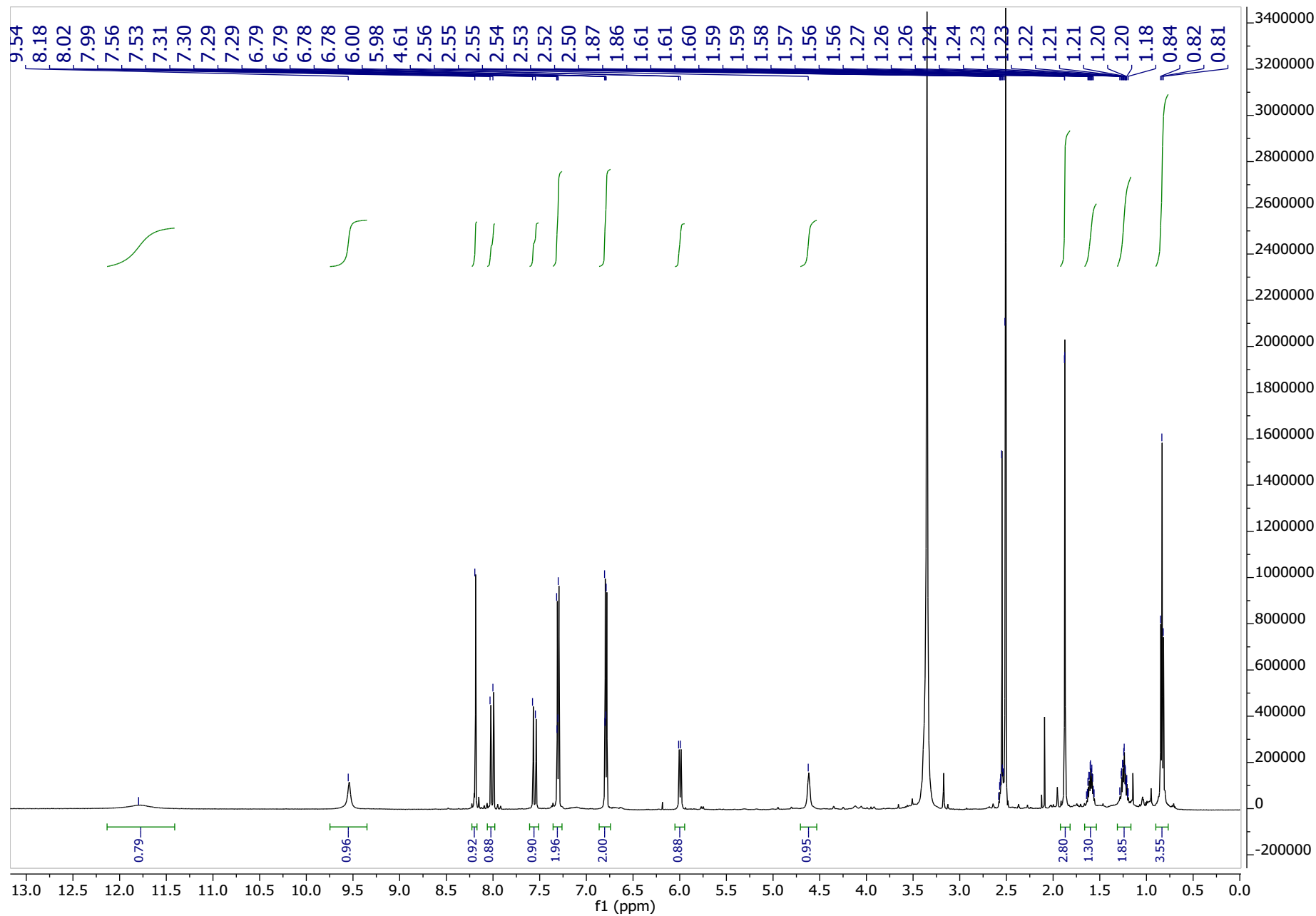
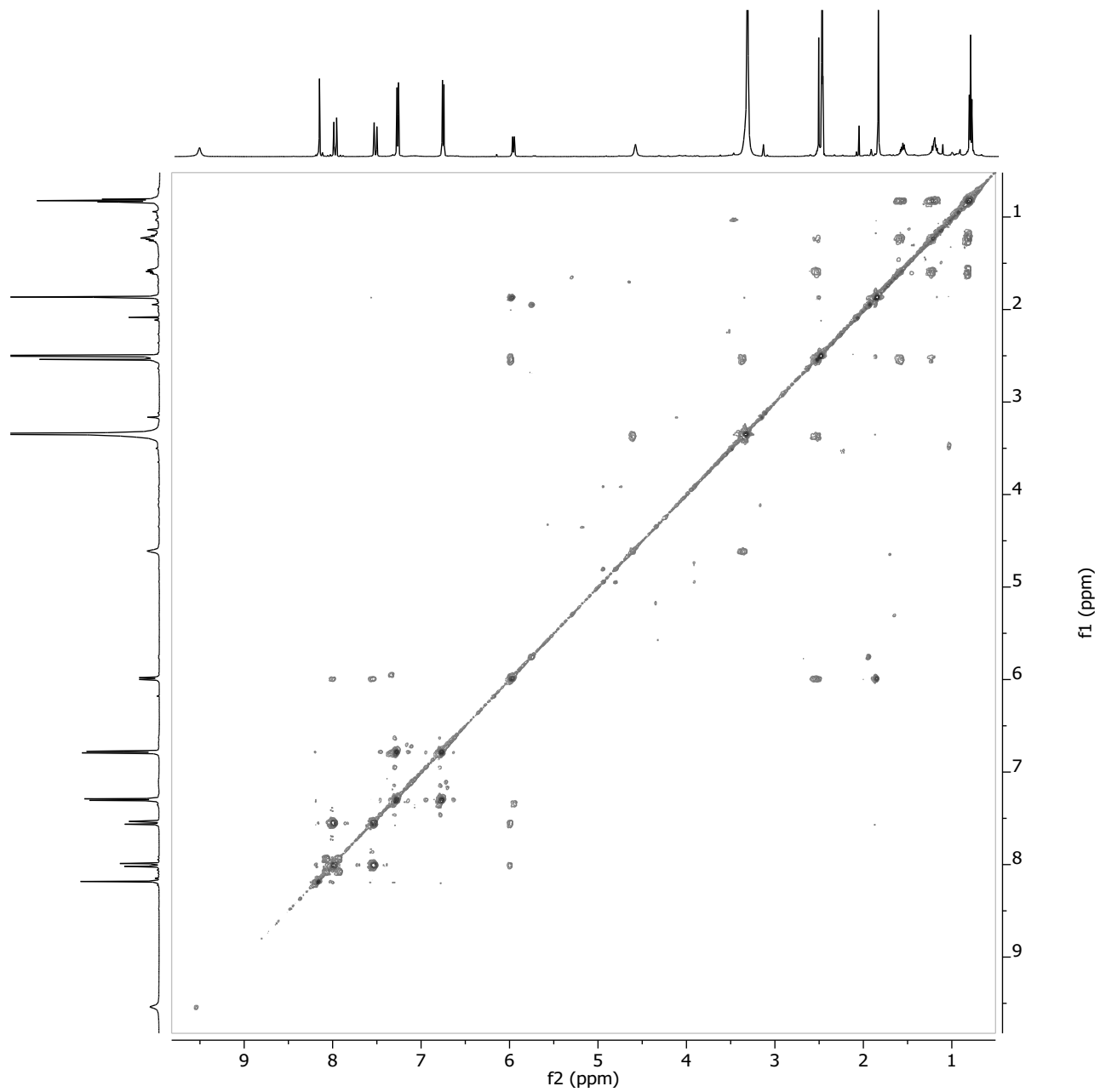


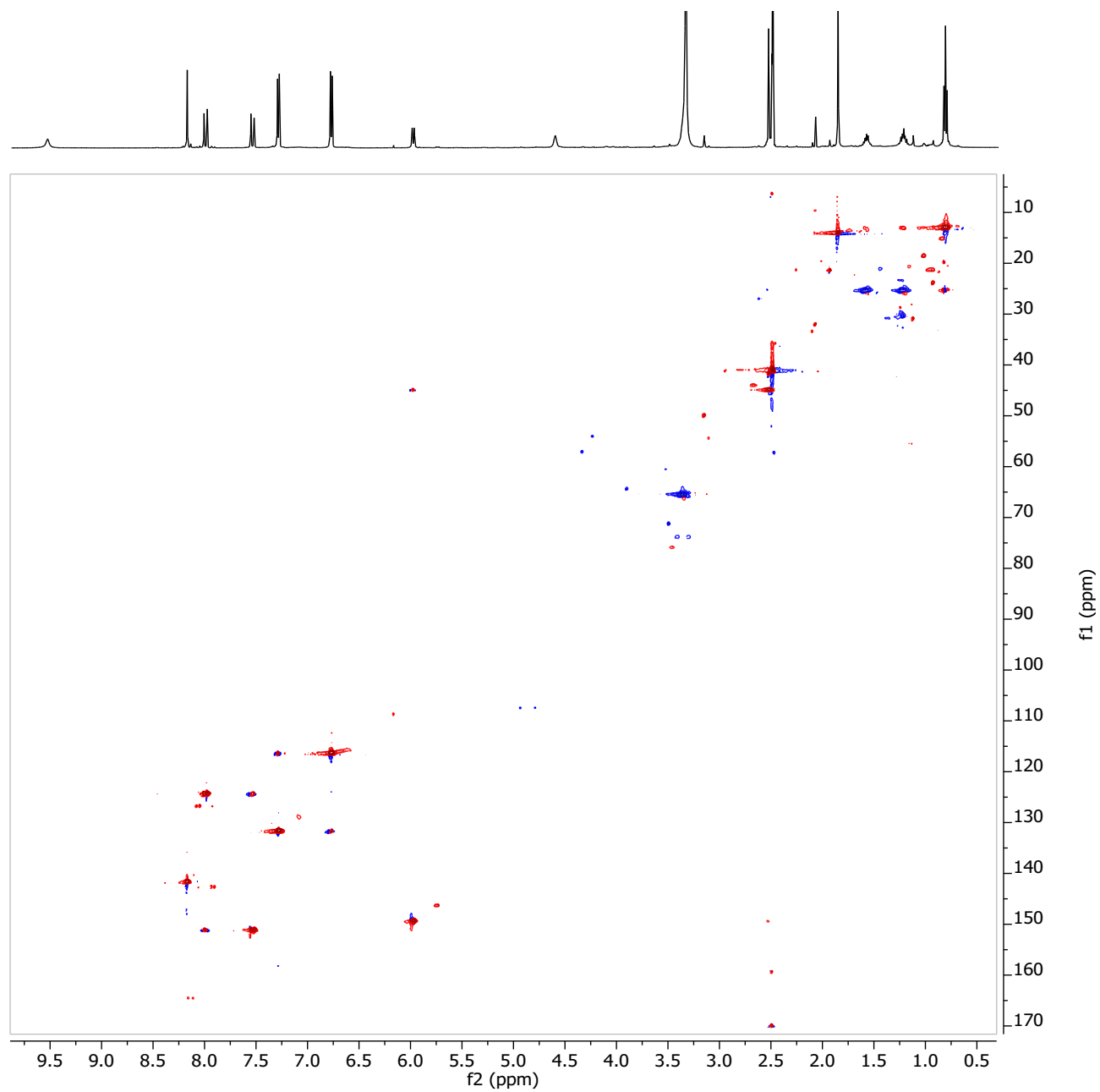
Figure S27. HRESIMS of 5.



**Figure S28.**  $^1\text{H}$  NMR spectrum of **5** in  $\text{DMSO}-d_6$  at 500 MHz.



**Figure S29.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of **5** in  $\text{DMSO}-d_6$  at 500 MHz.



**Figure S30.** HSQC spectrum of **5** in DMSO-*d*<sub>6</sub> at 500 MHz.

# Generic Display Report

## Analysis Info

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Instrument maXis

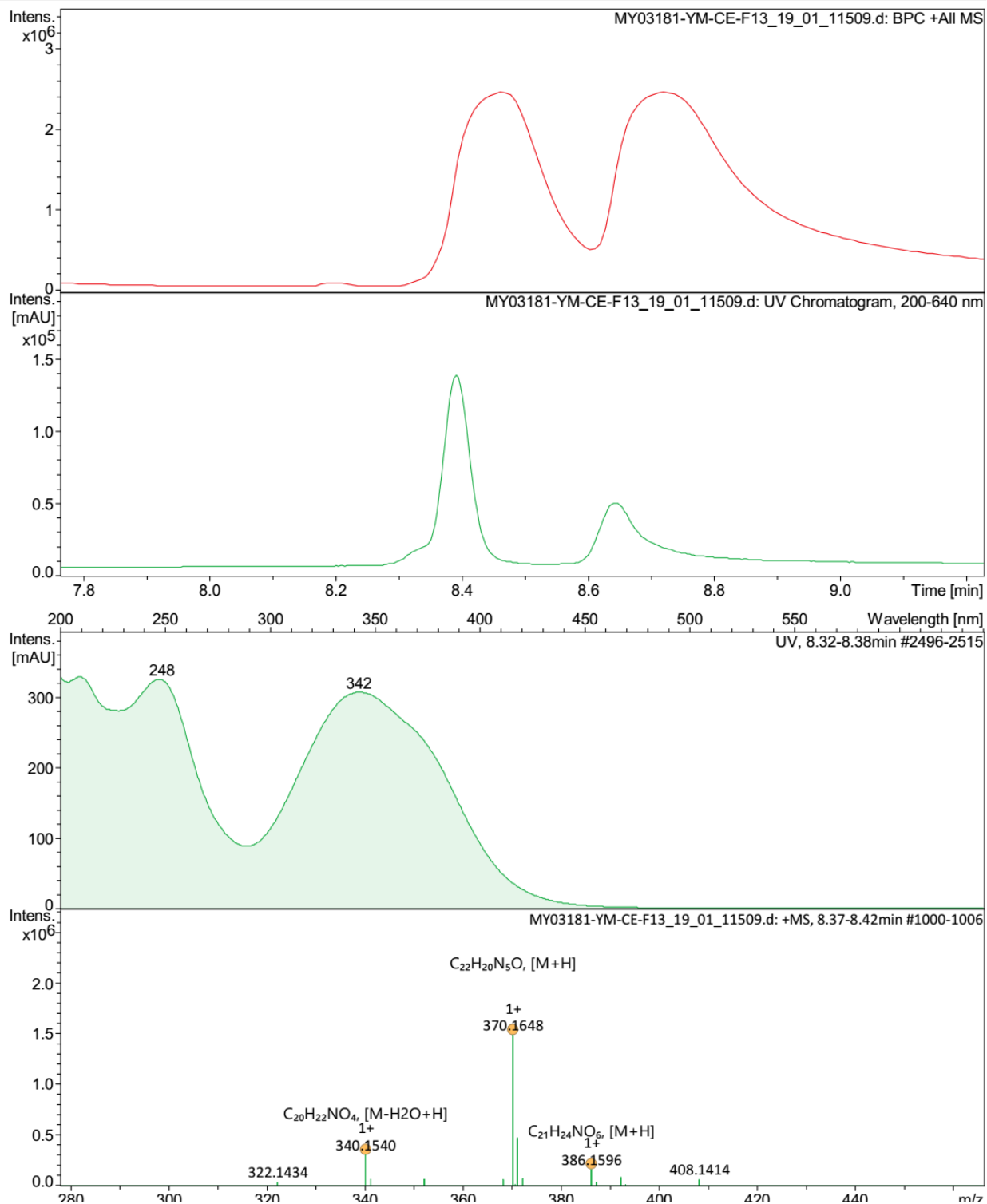
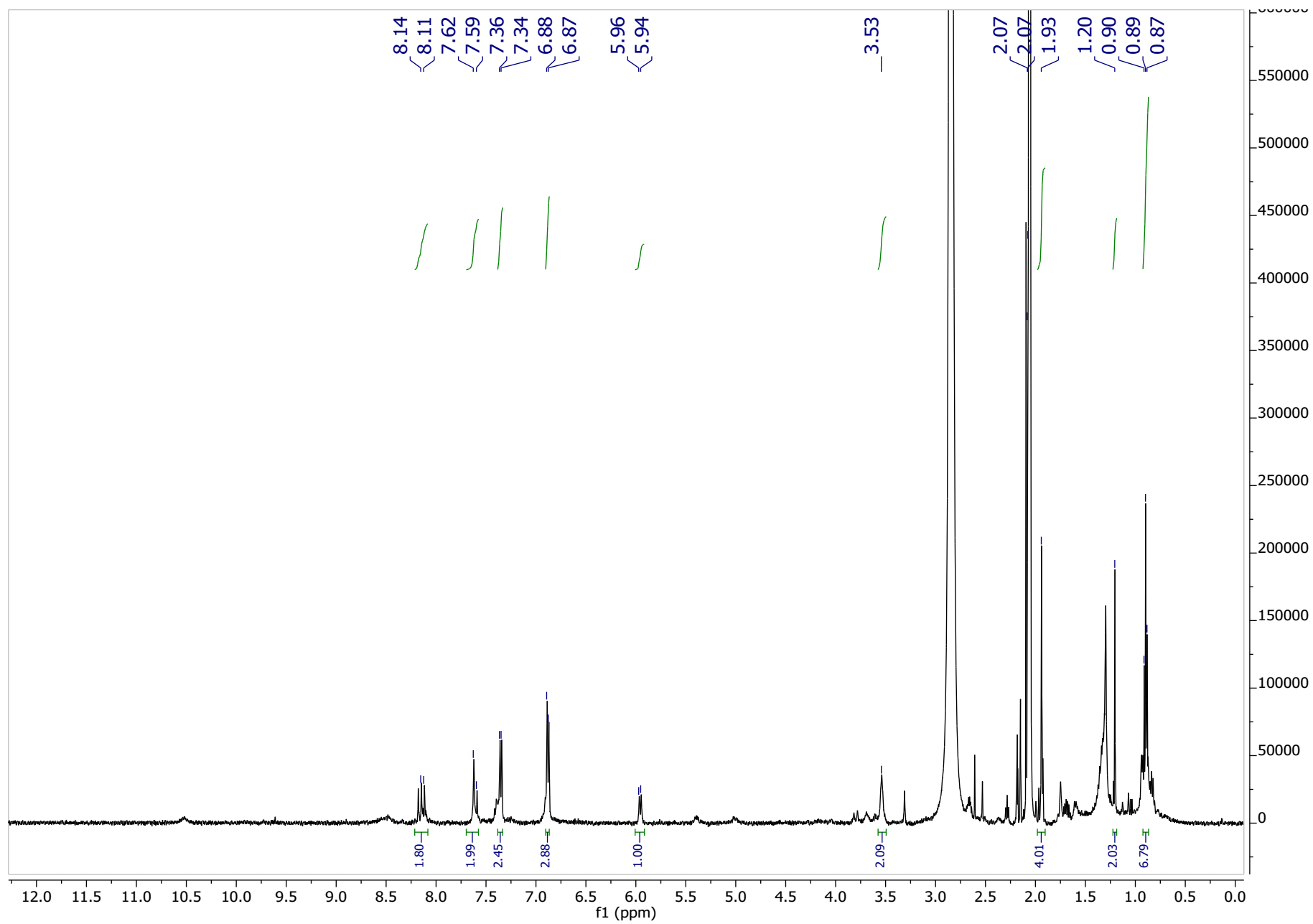
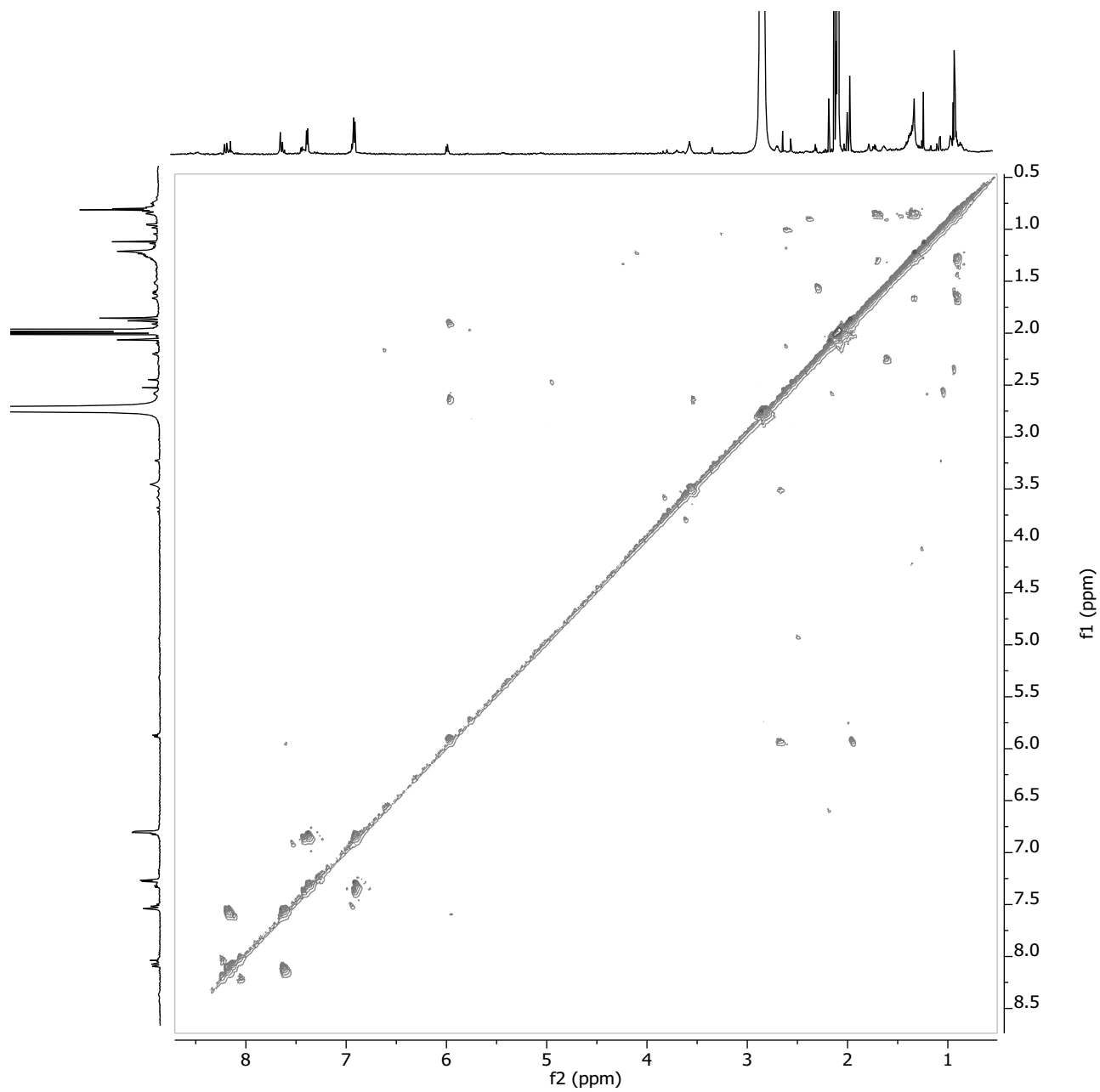


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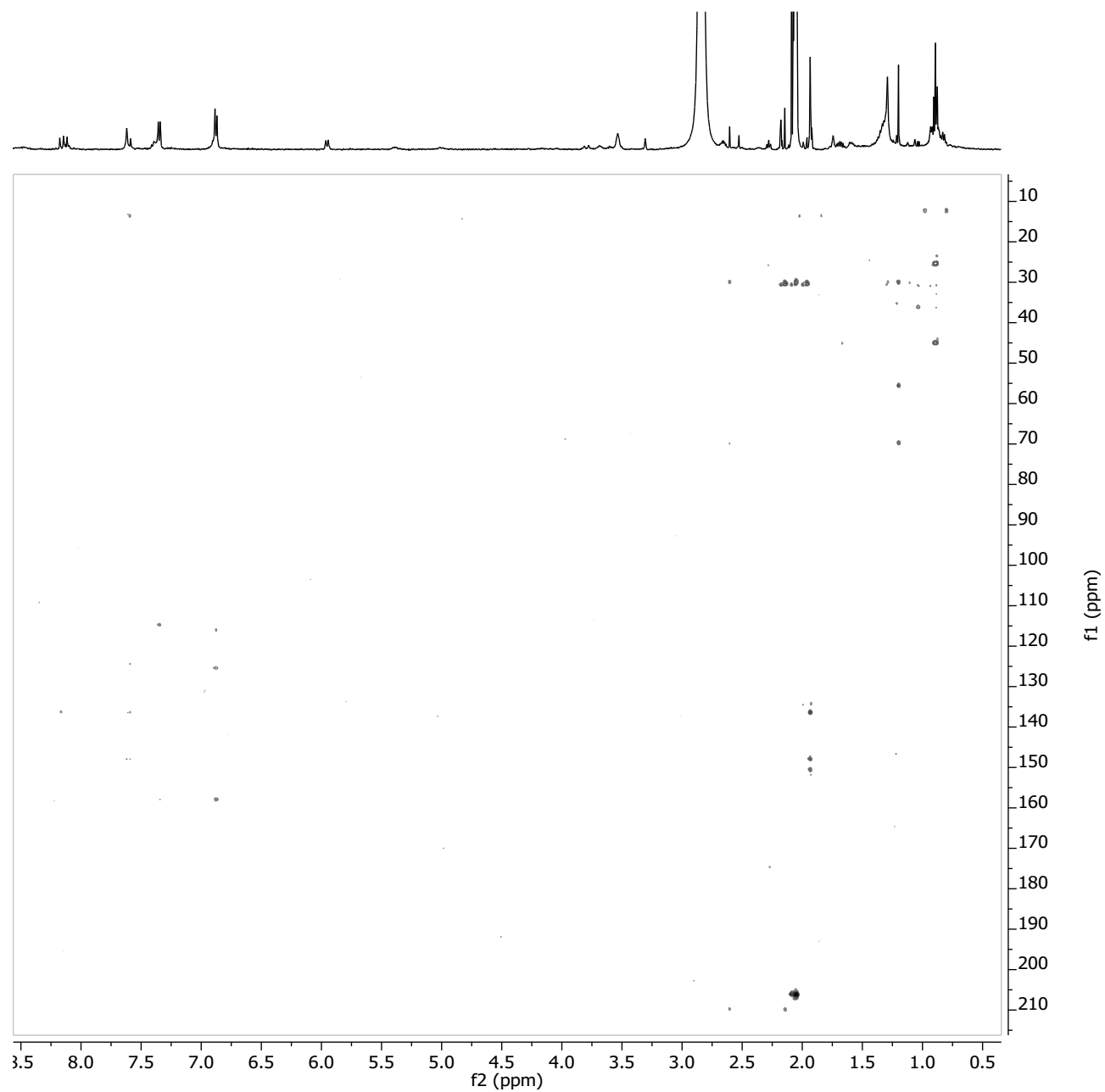


**Figure S32.**  $^1\text{H}$  NMR spectrum of **6** in  $\text{DMSO}-d_6$  at 500 MHz.

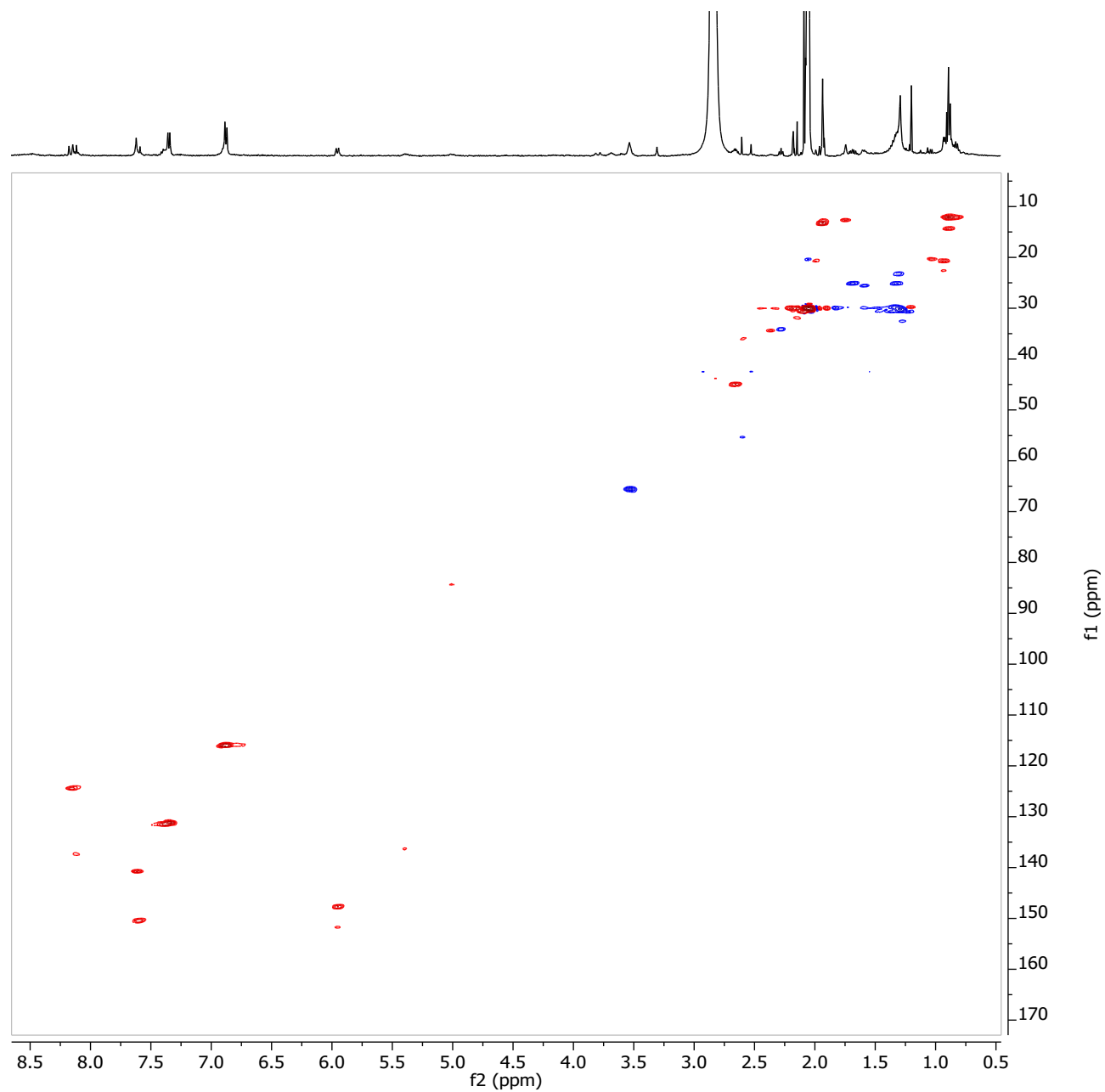




**Figure S33.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum of **6** in  $\text{DMSO}-d_6$  at 500 MHz.



**Figure S34.** HMBC spectrum of **6** in DMSO-*d*<sub>6</sub> at 500 MHz.



**Figure S35.** HSQC spectrum of **6** in DMSO- $d_6$  at 500 MHz.