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Assessment of Vitality, Blood Profile, and Degree of Meconium Staining on the Skin in Newborn Dogs According to Its Birth Weight

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Citation: Lezama-García, K.; Martínez-Burnes, J.;

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Assessment of Vitality, Blood Profile, and Degree of Meconium Staining on the Skin in Newborn Dogs According to Its Birth Weight. *Vet. Sci.* **2023**, *10*, 453. <https://doi.org/10.3390/vetsci10070453>

Academic Editor: Camila

Infantosi Vannucchi

Received: 5 June 2023

Revised: 5 July 2023

Accepted: 6 July 2023

Published: 11 July 2023



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Simple Summary: Preventing neonatal mortality is a critical aspect of canine perinatology. Among the leading causes of mortality, a low vitality score at birth, hypothermia, hypoxia, and hypoglycemia can be mentioned. This study aimed to assess vitality, blood values, and the degree of meconium staining on newborns' skin concerning their birth weight. It is concluded that larger newborns tend to present greater problems in surviving.

Abstract: Neonatal mortality in dogs reaches up to 40%. Due to the high rates, promptly detecting the causes and preventing newborns from dying are extremely important. Vitality evaluation, blood parameters, and the degree of meconium staining on the skin are valuable resources in canine perinatology. In this study, 435 puppies from 85 bitches close to parturition were recruited and divided into four quartiles according to the puppy's birth weight: Q₁ (127–200 g) n = 110 puppies, Q₂ (201–269 g) n = 108 puppies, Q₃ (270–388 g) n = 108 puppies, and Q₄ (389–464 g) n = 109 puppies. This experimental article aimed to report the effect of birth weight on the blood profile variables, the vitality of newborn puppies, and the meconium staining degree, integrating these three aspects. It was concluded that the weight of newborns was correlated with the degree of meconium staining, presenting more cases of severe meconium staining in the puppies of the highest birth weight group. The weight of the newborns was correlated with a higher number of stillbirths and alterations in the blood variables, showing the most severe cases of metabolic acidosis, hypoxia, and hypoglycemia in the puppies of the Q₄ quartile. On the contrary, no statistically significant correlations were found between the weight of newborns and vitality. Nevertheless, the analysis of the results showed that the most vigorous puppies were found at Q₁; however, at minute 60 after birth (AB), all the puppies in the four quartiles standardized their vitality scores.

Keywords: whelping; animal perinatology; puppy welfare; newborn puppy; vitality; meconium staining; blood biomarkers

1. Introduction

During whelping, dogs' mortality rates can reach up to 40% [1]. For this reason, these high rates worry dog owners and breeders [1,2] since one in ten puppies can die before

60 days of age [3]. In addition, newborn puppies are highly immature, making them extremely vulnerable [4,5]. Perinatal death could occur before parturition when the fetus is forming in the uterus, during expulsion, immediately after birth, or during the first weeks of life [2,6,7], especially on the first seven days [8,9], but during birth is when the majority of stillbirths occur [10].

In dogs, similarly to humans, pigs, and cattle [11], puppies with less birth weight are more likely to die. For example, in piglets, this risk is a dozen times more likely than in animals with normal weight at birth [11].

One factor observed to affect newborns' adaptation to extrauterine life negatively is asphyxia [12,13], including both their vitality and viability. This, in turn, can delay the newborn reaching the dam's teat [8,14,15]. There have been some reports in dogs that have presented a certain level of transitory asphyxiation during parturition. In a certain way, this can become normal during parturition, generating in puppies transitory acidosis and hypercapnia [16–18]. The gas exchange could be altered if these conditions continue [19], decreasing respiration rates and generating metabolic acidosis in newborns [20]. Fortunately, blood gases can be evaluated through the umbilical cord in other species, such as humans. In this way, we can obtain valuable information regarding the neonatal acid–base status [21,22]. Therefore, gasometry is currently an important tool for assessing the health status of newborns [8,23]. In small species, the assessment of umbilical blood parameters is limited. However, it is a technique that has been applied to newborn piglets to evaluate concentrations of pH, partial pressure of oxygen (pO_2), partial pressure of carbon dioxide (pCO_2), glucose, lactate, hematocrit, sodium, potassium, and ionized calcium [24].

Newborn mortality risk also includes the proportion of liveborn (LB) vs. stillbirth (SB) puppies and their viability [25,26], with hypoxia being a determinant factor that can alter a newborn's blood profile [27–29].

On the other hand, meconium staining of the skin at birth and aspiration of meconium reflect dystocia processes with severe intrauterine hypoxia [30]. Newborns exposed to meconium aspiration develop Meconium Aspiration Syndrome (MAS) [31]. MAS increases neonatal mortality due to hypoxemia, acidosis, respiratory distress [31–33], and pulmonary edema due to the proinflammatory mediators contained in meconium [34]. MAS has been reported to occur in diverse species; for example, in puppies, the reported mortality from MAS can reach 1–3% [35]. Various articles explain the pathophysiology of MAS; however, Swarman et al. [36], Martínez-Burnes et al. [31], and Mota-Rojas et al. [33] describe the association between MAS, airway obstruction, and fetal hypoxia, this being one of the most critical factors that can cause a loss of vitality in newborns.

Neonatal vitality refers to the capacity of newborns to respond to parturition stress. Vitality is evaluated by the Apgar scoring system adapted for human and animal newborns [2,37]. According to Randall [38], low viability scores in piglets are related to low birth weights and hypercapnia and, as stated by Zaleski and Hacker [39] and De Roth and Downie [40], are also positively associated with pH and negatively with PCO_2 .

Although studies in dogs assessing the effect of the dam's weight on the puppy's birth weight, litter size, vitality, survival [1,41,42], and the occurrence of asphyxia and newborn hematological values have been evaluated [27], the effect of the birth weight on the blood profile, meconium staining degree, and vitality has not been comprehensively studied. Therefore, this study aimed to evaluate the effect of birth weight on blood profile variables, the vitality of newborn puppies, and the meconium staining degree, integrating the three aspects. We hypothesized that bigger newborns would have lower vitality scores, significant blood profile alterations, and more cases of meconium staining than smaller puppies.

2. Materials and Methods

2.1. Facilities

This study was developed in the facilities of 10 veterinary hospitals in the municipality of Campeche, Campeche State, Mexico, where there is a tropical climate with a temperature

between 36 and 40 °C. To carry out this study, the tutors of pregnant bitches were asked for their collaboration. The bitches were given medical attention and monitoring from day 25 of gestation until 48 after the puppy's birth. All births took place in the clinics. Once the probable date of parturition was calculated, some bitches stayed sheltered in the clinics, or the guardians took them to the clinics when they began to notice changes in the bitches' behavior. The tutors did not participate during the whelping; only veterinary staff attended and monitored it.

2.2. Study Population

Four hundred thirty-five puppies from eighty-five parturient bitches were recruited and divided into four groups classified in quartiles, following Mugnier et al. [43] and Tesi et al.'s [44] method. The first quartile (Q_1) represents the lowest 25% of registered values, the second quartile (Q_2) represents 25–50%, the third quartile (Q_3) represents 50–75%, and the fourth quartile represents 75–100% (Q_4). Animals in group Q_1 were considered low-weight, while those belonging to Q_4 were considered high-weight puppies. This classification is based on the great variety of dog breeds, ranging from Chihuahuas weighing 500 g as adults to mastiffs weighing 100 kg [45]. Within the breeds included in this study, we can mention Chihuahua, Yorkshire Terrier, Poodle, Scottish Terrier, Cocker Spaniel, Standard Schnauzer, German Shepherd, Labrador, Golden Retriever, Great Dane, and Belgian Shepherd. Quartiles were calculated at the puppy level with this formula: $Q_a = Li ((aN/4 + Fi-1)/Fi) Ai$, where Li is the lower limit of the class where the quartile is located, N is the sum of the absolute frequencies, $Fi-1$ is the accumulated frequency of the previous class, and Ai is the amplitude of the class, that is, the number of values contained in the interval. The groups were Q_1 (127–200 g) $n = 110$ puppies, Q_2 (201–269 g) $n = 108$ puppies, Q_3 (270–388 g) $n = 108$ puppies, and Q_4 (389–464 g) $n = 109$ puppies.

The inclusion criteria were the same used in previous studies by Reyes-Sotelo et al. [27,28] and Lezama-García et al., 2022. Mota-Rojas et al.'s [29,46] classification of type I and type II stillbirths in piglets was used to define which could be considered for the study and which could not. According to Mota-Rojas et al. [30,46], stillbirths can be classified into two types: type I, also known as prepartum or antepartum deaths, includes fetuses that die before the end of gestation, usually of infectious causes and have a rather characteristic hemorrhagic and edematous appearance with a grayish-brown discoloration; type II stillbirths, which are also referred to as intrapartum deaths, can die during parturition typically from intrauterine asphyxia and rarely from infectious diseases, and they have the same appearance of their normal littermates, but they do not breathe. Type I stillbirths were excluded from the study, and only type II stillbirths were included, classified by necropsy.

Puppies were weighed using a digital scale from Salter Weight Tronix Ltd., West Bromwich, UK, immediately after the dam stopped licking and cleaning their amniotic fluids and placental membranes.

2.3. Blood Sampling

Blood samples were taken by a veterinarian when the bitch finished removing the chorioallantoic membranes. When necessary, an assistant held the puppy in a supine position and exposed the abdominal region. When the puppies allowed it, they were only placed in lateral decubitus on an adult diaper to perform the sampling. Venous blood from the umbilical cord (0.3 mL) was obtained with a tuberculin syringe and a lithium heparin-impregnated needle. All samples (150 μ L) were processed using a GEM Premier[®] critical blood variable analyzer (Instrumentation Laboratory Diagnostics, Lexington, KY, USA/Milano, Italy). The metabolites analyzed were glucose (mg/dL), lactate (mg/dL), blood gases pCO_2 (mmHg), pO_2 (mmHg) pH, HCO_3^- (mmol/L), EB (mEq/L), Ca^{++} (mmol/L), and hematocrit (Htc %). All profiles were tested for each LB and type II SB pup.

2.4. Meconium Staining Degree on Skin

To assess the degree of meconium staining in the skin of the puppies, they were divided according to the methodology of Mota-Rojas et al. [30] into absent, mild, moderate, and severe (Figure 1). It is important to emphasize that the process of resuscitation of the newborns by the mother was not interrupted since the puppies were taken just after the bitch finished removing the chorioallantoic membranes and as soon as possible to prevent the bitch from licking the meconium stain; once the stain was impregnated in the white towel, the puppy was returned to the dam so that she could continue with the licking and maternal care.

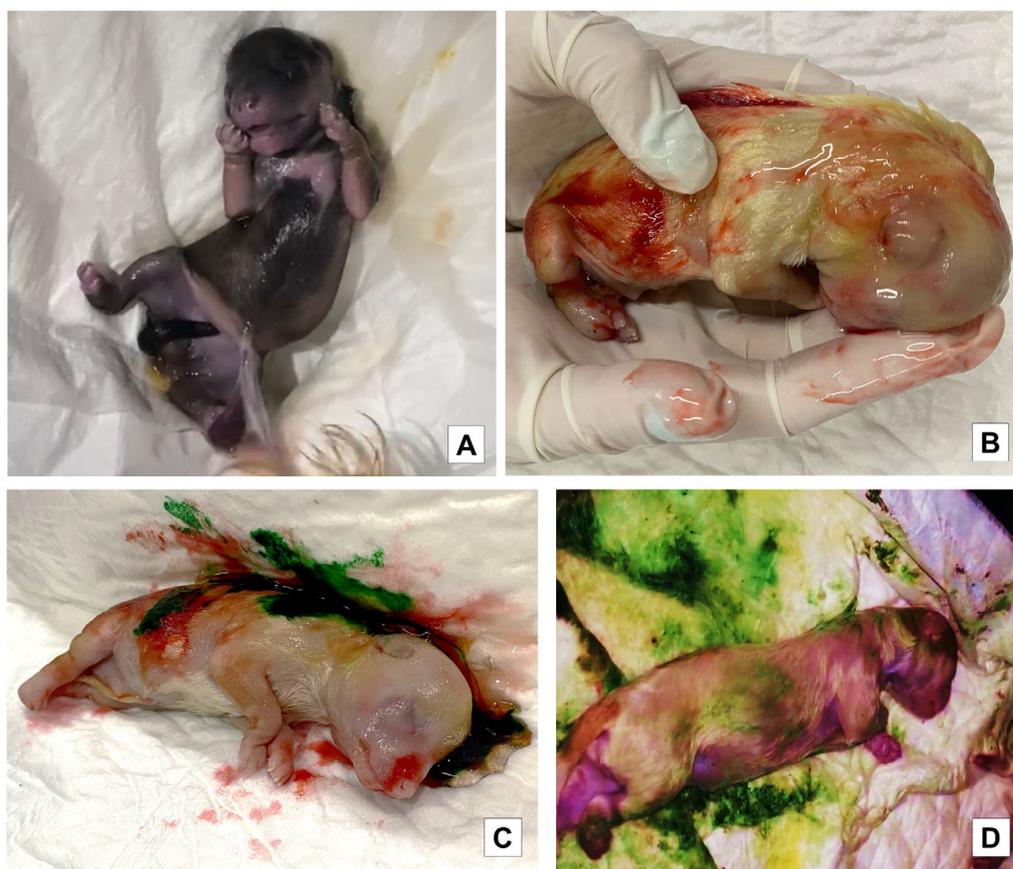


Figure 1. Different degrees of meconium staining on the skin in liveborn (LB) puppies. (A). Liveborn (LB) with amniotic fluid and no meconium staining. (B). Liveborn (LB) with mild meconium staining. (C). Liveborn (LB) with moderate meconium staining. (D). Liveborn (LB) with severe meconium staining.

The puppies born free of meconium staining on the skin were considered absent of staining, a mild degree was considered when the body was covered less than 30% of its surface, a moderate degree was considered by 30 to 60%, and a severe degree was considered when its body was covered over 60%. A white human bed diaper was used to identify the degree of meconium staining in the dark-colored animals, pressing it on the newborn body to more clearly observe the impregnated color (Figures 2 and 3). It is worth mentioning that the puppies were removed from the mother as soon as possible at birth to prevent the bitch from licking the meconium stain; once the stain was impregnated in the white towel, the puppy was returned to the dam so that she could continue with the licking and maternal care.



Figure 2. Methodology used to evaluate the degree of meconium staining in puppies. (A): The pup was gently picked up before the bitch began to lick it to prevent her from removing meconium staining if it was present. (B,C): The puppy was placed in a white diaper and surrounded with it to impregnate the meconium staining.



Figure 3. Different degrees of meconium staining. Different degrees of meconium staining were observed using the methodology described in the previous figure. (A): Mild. (B): Moderate. (C): Severe.

2.5. Vitality Score

For the evaluation of vitality in the newborns, the Veronesi [1] scale was used with Randall's [38] adaptation of the Apgar score for human newborns and modified by Mota-Rojas et al. [47]. The variables measured in the first minute after birth were respiratory effort (no crying/< 6 respiratory rates (rr), mild crying/6 to 15 rr, and crying/>15 rr); motility (flaccid, some flexions, and active motion); heart rate (beats per min): <180, between 180 and 220, and >220; mucus color (cyanotic, pale, and pink). Also, the meconium staining on the skin was classified as severe, moderate, mild, or absent based on the methodology used before in pigs by Mota-Rojas et al. [30,47]. The score for vitality was 0 (the least favorable)

to 2 (the most favorable), and a global score ranging from 1 to 10 was obtained for each newborn puppy (Figure 4). A vitality score of 6 or less was considered as failure.

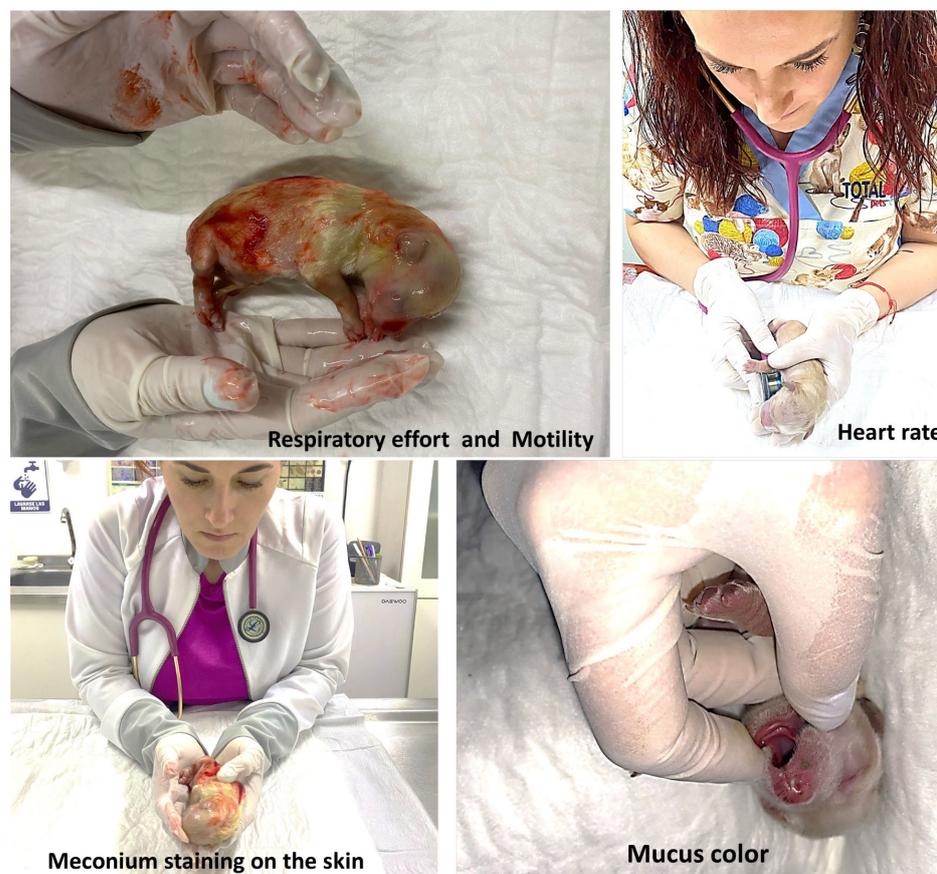


Figure 4. Evaluation of vitality score in first minute after birth.

2.6. Statistical Analysis

Statistical analyses were performed in R version 4.2.2 (R Core Team, Vienna, Austria) using the packages “moments”, “ggpubr”, “stats”, “emmeans”, and “multcompView”. The significance level was set at $p < 0.05$.

The effect of puppy weight on blood profile variables was analyzed using separate one-way ANOVA with the four puppy weight groups (quartiles) as categorical predictors. The normality of blood-profile-dependent variables was assessed through a visual inspection of histograms, Q-Q plots, and the skewness and kurtosis of data. Post hoc pairwise comparisons between quartiles were carried out using Tukey HSD tests.

The effect of puppy weight on newborn puppy vitality scores at different time points (1 min AB, 5 min AB, 60 min AB) was evaluated using a linear mixed model with puppy vitality score set as the response variable. Puppy weight groups (quartiles) and timepoint were fitted as fixed effects, and puppy ID was set as a random effect to account for the nonindependence of puppies across time points. The normality and homoscedasticity of model residuals were assessed with a visual inspection of Q-Q plots and residuals vs. predicted values plots. Post hoc pairwise comparisons between groups and time points were conducted using Tukey HSD tests.

Differences in the proportion of stillbirths grouped according to birth weight (quartiles) and the presence of meconium stain on their skin were assessed using a chi-square test.

2.7. Ethical Statement

The Ph.D. Program in Biological and Health Science Academic Committee approved the present study (CBS.114.19). The care and use of animals was performed according

to guidelines for the ethical use of animals in applied ethology studies [48]. The owners provided informed consent.

3. Results

A one-way ANOVA was used to determine significant differences between puppy weight quartiles in pH ($F(3) = 102.70$, $p < 0.001$), $p\text{CO}_2$ ($F(3) = 30.97$, $p < 0.001$), $p\text{O}_2$ ($F(3) = 54.53$, $p < 0.001$), glucose ($F(3) = 4.581$, $p = 0.004$), Ca^{++} ($F(3) = 67.36$, $p < 0.001$), lactate ($F(3) = 48.27$, $p < 0.001$), hematocrit ($F(3) = 51.98$, $p < 0.001$), and HCO_3^- ($F(3) = 40.28$, $p < 0.001$). The post hoc Tukey HSD pairwise comparisons between quartiles for the blood profile variables of puppies born alive are shown in Table 1.

Table 1. Mean and standard error of blood profile values from puppies born alive and classified according to their birth weight.

Blood Trait	Q ₁ n = 110	Q ₂ n = 108	Q ₃ n = 108	Q ₄ n = 109
pH	7.49 ± 0.0126 ^a	7.24 ± 0.0136 ^b	7.25 ± 0.0116 ^b	7.24 ± 0.0125 ^b
$p\text{CO}_2$ (mmHg)	46.9 ± 0.879 ^a	56.5 ± 1.41 ^b	58.8 ± 1.46 ^b	64.0 ± 1.42 ^c
$p\text{O}_2$ (mmHg)	18.0 ± 0.325 ^a	14.4 ± 0.323 ^b	14.0 ± 0.302 ^b	12.5 ± 0.306 ^c
Glucose (mg/dL)	93.7 ± 1.83 ^a	92.7 ± 2.44 ^a	93.4 ± 2.76 ^a	81.9 ± 2.99 ^b
Ca^{++} (mmol/L)	1.59 ± 0.0196 ^a	1.90 ± 0.0206 ^a	1.90 ± 0.0200 ^a	1.94 ± 0.0208 ^b
Lactate (mmol/L)	4.74 ± 0.226 ^a	6.86 ± 0.198 ^{a,b}	7.54 ± 0.216 ^b	8.29 ± 0.252 ^c
Hematocrit (%)	45.3 ± 0.408 ^a	48.7 ± 0.348 ^b	50.0 ± 0.404 ^b	51.8 ± 0.316 ^c
HCO_3^- (mmol/L)	21.7 ± 0.357 ^a	19.0 ± 0.322 ^b	17.8 ± 0.305 ^{b,c}	17.0 ± 0.321 ^c

(One-way ANOVA.) Q₁, 127–200 g; Q₂, 201 g–269 g; Q₃, 270 g–388 g; Q₄, 389 g–464 g. ^{a,b,c} Different superscripts between columns indicate statistically significant differences between quartiles with $p < 0.05$.

Table 1 shows how some blood values in the puppies born alive presented marked variations between Q₁ and Q₄, such as PCO_2 , with the highest significant difference between these quartiles. If we compare the blood values obtained in Q₁ with Q₄, the biggest changes were reflected in Q₄: PO_2 decreased by 5.5 mmHg, glucose decreased by 11.8 mg/dL, hematocrit increased by 6.5%, and HCO_3^- decreased by 4.7 mmol/L. Lactate showed significant changes between Q₁ and Q₄, doubling its value in the quartile of puppies with the highest weight. All these findings can be assumed as indicators of stress to which larger puppies were subjected, which can be evidenced by hypercapnia, hypoxia, hypoglycemia, and marked metabolic acidosis, as well as polycythemia, hypercalcemia, and marked hyperlactatemia.

A statistically significant difference between puppy weight quartiles in the blood profile variables $p\text{CO}_2$ ($F(3) = 10.74$, $p < 0.001$), $p\text{O}_2$ ($F(3) = 12.04$, $p < 0.001$), lactate ($F(3) = 11.88$, $p < 0.001$), and HCO_3^- ($F(3) = 7.316$, $p < 0.001$) were found. Unlike these results, pH ($F(3) = 2.217$, $p = 0.0943$), glucose ($F(3) = 0.177$, $p = 0.912$), Ca^{++} ($F(3) = 2.073$, $p = 0.112$), and hematocrit ($F(3) = 0.672$, $p = 0.572$) did not show a significant difference between puppy weight quartiles. The post hoc Tukey HSD pairwise comparisons between quartiles for the blood profile variables of the stillbirth puppies are shown in Table 2.

Table 2. Mean and standard error of blood profile values in stillbirth puppies classified according to their birth weight.

Blood Trait	Q ₁ n = 100	Q ₂ n = 108	Q ₃ n = 108	Q ₄ n = 109
pH	6.82 ± 0.0479 ^a	6.80 ± 0.0449 ^a	6.73 ± 0.0325 ^a	6.82 ± 0.0176 ^a
$p\text{CO}_2$ (mmHg)	82.5 ± 2.37 ^a	83.2 ± 1.34 ^{ab}	91.7 ± 1.48 ^c	95.1 ± 1.36 ^{b,c}
$p\text{O}_2$ (mmHg)	9.24 ± 0.531 ^a	8.2 ± 0.932 ^a	5.71 ± 0.486 ^b	4.34 ± 0.386 ^b
Glucose (mg/dL)	41.0 ± 6.06 ^a	40.0 ± 4.34 ^a	37.7 ± 2.10 ^a	37.7 ± 2.26 ^a
Ca^{++} (mmol/L)	2.12 ± 0.0630 ^a	2.12 ± 0.0650 ^a	2.26 ± 0.0474 ^a	2.17 ± 0.0216 ^a
Lactate (mmol/L)	12.3 ± 0.682 ^a	12.1 ± 0.480 ^a	14.3 ± 0.340 ^b	14.7 ± 0.226 ^b
Hematocrit (%)	59.9 ± 1.01 ^a	60.5 ± 0.423 ^a	59.3 ± 0.725 ^a	59.1 ± 0.585 ^a
HCO_3^- (mmol/L)	12.8 ± 0.685 ^a	12.8 ± 0.685 ^a	13.2 ± 0.571 ^{ab}	10.6 ± 0.343 ^b

(One-way ANOVA.) Q₁, 127–200 g; Q₂, 201 g–269 g; Q₃, 270 g–388 g; Q₄, 389 g–464 g. ^{a,b,c} Different superscripts between columns indicate statistically significant differences between quartiles with $p < 0.05$.

Table 2 reports the blood profile alterations between the four quartiles in the SB puppies, and as shown in Table 1 for LB, many parameter alterations were observed. However, in this case, all the blood profile indicators in SB showed acid–base imbalance, manifested with a pH below 7 that denotes severe metabolic acidosis. In the same way, there was hypercapnia in the four quartiles, which was more marked in Q₄. Also, there were marked decreases in glucose, with values ranging from 37.7 mg/dL (Q₄) to 41 mg/dL (Q₁), and a notable increase in Ca⁺⁺ in all quartiles regardless of weight. In addition, lactate showed extremely high values—practically tripled if we compare them with the lactate values for LBs (LB 8.29 mmol/L and SB 14.7 mmol/L). Elevated hematocrit again denoted splenic contraction because of the release of catecholamines, and HCO₃ was found in values decreased to half of those presented in this parameter in LBs due to the marked metabolic acidosis that the newborns experienced until death.

A linear mixed model (LMM) revealed that newborn puppies’ vitality score was significantly affected by the interaction between puppy weight group (quartile) and timepoint (F(6,722) = 5.652, *p* < 0.001). The post hoc Tukey HSD pairwise comparisons between quartiles and time after birth (AB) are reported in Table 3.

Table 3. Estimated marginal mean and standard error of vitality score of newborn puppies evaluated at different times and classified according to their birth weight.

Time	Q ₁ n = 110	Q ₂ n = 108	Q ₃ n = 108	Q ₄ n = 109
Minute 1 AB	7.28 ± 0.173 ^{a,1}	6.80 ± 0.180 ^{a,1}	6.62 ± 0.187 ^{a,b,1}	5.77 ± 0.207 ^{b,1}
Minute 5 AB	7.64 ± 0.173 ^{a,1,2}	7.38 ± 0.180 ^{a,2}	7.28 ± 0.187 ^{a,2}	6.81 ± 0.207 ^{a,2}
Minute 60 AB	7.95 ± 0.173 ^{a,2}	7.73 ± 0.180 ^{a,2}	7.68 ± 0.187 ^{a,2}	7.48 ± 0.207 ^{a,3}

(Linear mixed model.) Q₁, 127–200 g; Q₂, 201 g–269 g; Q₃, 270 g–388 g; Q₄, 389 g–464 g. AB: After birth. ^{a,b} Different superscripts between columns indicate statistically significant differences between quartiles with *p* < 0.05. ^{1,2,3} Different numbers among rows indicate statistically significant differences between times of evaluation.

In Table 3, we can see that Q₁ puppies were the most vigorous, and Q₄ had the lowest scores in terms of vitality. However, after 60 min AB, the vitality began to become uniform, showing an increase in the vitality score to almost double in Q₄ (from 5.77 min 1 AB to 7.48 min 60 AB), with statistically significant differences ceasing to exist between the four quartiles 60 min AB.

The results of the two-way ANOVAs for blood profile variables showed that the interaction between birth weight quartiles and puppy vitality score groups significantly affected the levels of pCO₂ (F(6) = 3.389, *p* = 0.003), pO₂ (F(6) = 3.539, *p* = 0.002), Ca⁺⁺ (F(6) = 2.738, *p* = 0.013), lactate (F(6) = 3.091, *p* = 0.006), hematocrit (F(6) = 6.628, *p* < 0.001), and HCO₃ (F(6) = 6.628, *p* < 0.001) in the newborn puppies. Furthermore, pH and glucose levels differed significantly between puppy weight quartiles (pH: F(3) = 184.034, *p* < 0.001; glucose: F(3) = 8.686, *p* < 0.001) and across puppies classified according to vitality score (pH: (F(2) = 140.863, *p* < 0.001; F(2) = 159.742, *p* < 0.001). The post hoc Tukey HSD pairwise comparisons between puppy weight quartiles and vitality score groups are reported in Table 4.

A statistically significant effect on vitality score for all blood values on puppies classified according to weight was observed (Table 4). Glucose and pH had no interactions, while pCO₂, pO₂, Ca⁺⁺, lactate, hematocrit, and HCO₃ had interactions and were significantly affected. Also, in all quartiles, the values that were most affected in puppies with failed vitality were pCO₂ (increased), pO₂ (decreased), lactate (doubled), glucose (halved), and HCO₃ (almost halved). The results demonstrate how the puppies that presented the lowest vitality scale (failed) fought to survive, thereby causing extreme metabolic acidosis because of the activation of all the compensatory factors the newborn uses to survive.

Table 4. Mean and standard error of blood profile values in newborn puppies born alive classified according to their vitality score (failed, medium, and high) and birth weight.

Blood Trait	V 0–5 (Failed) <i>n</i> = 102	V 6–7 (Medium) <i>n</i> = 105	V 8–10 (High) <i>n</i> = 158
Q ₁			
pH	7.29 ± 0.0207 ^{a,1}	7.50 ± 0.0197 ^{b,1}	7.55 ± 0.0108 ^{b,1}
pCO ₂ (mmHg)	62.3 ± 2.30 ^{a,1}	42.5 ± 1.07 ^{b,1}	43.8 ± 0.362 ^{b,1}
pO ₂ (mmHg)	13.7 ± 0.578 ^{a,1}	18.7 ± 0.697 ^{b,1}	19.0 ± 0.306 ^{b,1}
Glucose (mg/dL)	62.8 ± 4.73 ^{a,1}	99.4 ± 2.55 ^{b,1}	101.0 ± 0.943 ^{b,1}
Ca ⁺⁺ (mmol/L)	1.92 ± 0.0519 ^{a,1}	1.56 ± 0.0259 ^{b,1}	1.51 ± 0.0108 ^{b,1}
Lactate (mmol/L)	8.09 ± 0.639 ^{a,1}	4.25 ± 0.325 ^{b,1}	3.91 ± 0.163 ^{b,1}
Hematocrit (%)	52.2 ± 0.858 ^{a,1}	43.6 ± 0.555 ^{b,1}	43.8 ± 0.273 ^{b,1}
HCO ₃ ⁻ (mmol/L)	15.1 ± 0.665 ^{a,1}	22.2 ± 0.328 ^{b,1}	23.6 ± 0.189 ^{b,1}
Q ₂			
pH	7.09 ± 0.0214 ^{a,2}	7.27 ± 0.0159 ^{b,2}	7.31 ± 0.0196 ^{b,2}
pCO ₂ (mmHg)	75.3 ± 2.83 ^{a,2}	53.9 ± 1.84 ^{b,2}	48.6 ± 0.447 ^{b,1,2}
pO ₂ (mmHg)	11.0 ± 0.502 ^{a,2}	14.4 ± 0.443 ^{b,2}	16.4 ± 0.370 ^{c,2}
Glucose (mg/dL)	65.3 ± 4.67 ^{a,1}	95.1 ± 3.21 ^{b,1}	106.0 ± 2.59 ^{b,1}
Ca ⁺⁺ (mmol/L)	2.08 ± 0.0500 ^{a,1}	1.87 ± 0.0294 ^{b,2}	1.82 ± 0.0231 ^{b,2}
Lactate (mmol/L)	9.08 ± 0.416 ^{a,1,2}	6.96 ± 0.249 ^{b,2}	5.50 ± 0.145 ^{c,2}
Hematocrit (%)	51.5 ± 0.871 ^{a,1}	47.9 ± 0.519 ^{b,2}	47.8 ± 0.380 ^{b,2}
HCO ₃ ⁻ (mmol/L)	14.6 ± 0.608 ^{a,1}	19.8 ± 0.385 ^{b,2}	20.6 ± 0.213 ^{b,2}
Q ₃			
pH	7.14 ± 0.0183 ^{a,1,2}	7.26 ± 0.0164 ^{b,2}	7.32 ± 0.0122 ^{b,2}
pCO ₂ (mmHg)	71.0 ± 2.70 ^{a,1}	57.1 ± 2.65 ^{b,2}	52.1 ± 1.28 ^{b,2}
pO ₂ (mmHg)	11.3 ± 0.472 ^{a,1,2}	14.2 ± 0.429 ^{b,2}	15.6 ± 0.371 ^{b,2}
Glucose (mg/dL)	71.3 ± 5.24 ^{a,1}	92.7 ± 4.69 ^{b,1}	108.0 ± 2.44 ^{c,1}
Ca ⁺⁺ (mmol/L)	2.05 ± 0.0427 ^{a,1,2}	1.89 ± 0.0336 ^{b,2}	1.81 ± 0.0171 ^{b,2}
Lactate (mmol/L)	9.75 ± 0.226 ^{a,2}	7.16 ± 0.377 ^{b,2}	6.36 ± 0.212 ^{b,2}
Hematocrit (%)	52.2 ± 0.976 ^{a,1}	48.9 ± 0.538 ^{b,2}	49.2 ± 0.498 ^{b,2,3}
HCO ₃ ⁻ (mmol/L)	14.4 ± 0.382 ^{a,1}	18.6 ± 0.505 ^{b,2,3}	19.5 ± 0.204 ^{b,2}
Q ₄			
pH	7.17 ± 0.0150 ^{a,2}	7.28 ± 0.0147 ^{b,2}	7.33 ± 0.0169 ^{b,2}
pCO ₂ (mmHg)	71.4 ± 1.49 ^{a,2}	61.8 ± 3.13 ^{b,2}	51.9 ± 0.782 ^{b,2}
pO ₂ (mmHg)	11.2 ± 0.395 ^{a,2}	14.4 ± 0.507 ^{b,2}	13.3 ± 0.471 ^{a,b,3}
Glucose (mg/dL)	62.6 ± 1.82 ^{a,1}	89.8 ± 6.21 ^{b,1}	111.0 ± 1.96 ^{c,1}
Ca ⁺⁺ (mmol/L)	2.06 ± 0.0178 ^{a,2}	1.90 ± 0.0448 ^{b,2}	1.76 ± 0.0193 ^{b,2}
Lactate (mmol/L)	9.68 ± 0.228 ^{a,2}	8.26 ± 0.428 ^{a,2}	5.75 ± 0.226 ^{b,2}
Hematocrit (%)	53.1 ± 0.369 ^{a,1}	50.1 ± 0.546 ^{b,2}	50.6 ± 0.593 ^{a,b,3}
HCO ₃ ⁻ (mmol/L)	15.4 ± 0.349 ^{a,1}	17.2 ± 0.690 ^{a,3}	19.7 ± 0.230 ^{b,2}

(Two-way ANOVA.) Q₁, 127–200 g; Q₂, 201 g–269 g; Q₃, 270 g–388 g; Q₄, 389 g–464 g. V: Vitality. ^{a,b,c} Different superscripts between columns indicate statistically significant differences between vitality score and quartiles with *p* < 0.05. ^{1,2,3} Different numbers among rows indicate statistically significant differences between blood values on different quartiles.

All blood profile variables maintained a significant interaction between birth weight and the variation in the degree of meconium staining (pH: F(9) = 4.538, *p* < 0.001; pCO₂: F(9) = 4.349, *p* < 0.001; pO₂: F(9) = 3.287, *p* < 0.001; glucose: F(9) = 5.318, *p* < 0.001; Ca⁺⁺: F(9) = 4.792, *p* < 0.001; lactate: F(9) = 3.751, *p* < 0.001; hematocrit: F(9) = 5.599, *p* < 0.001; and HCO₃: F(9) = 6.013, *p* < 0.001). The post hoc Tukey HSD pairwise comparisons are shown in Table 5.

Table 5. Effect of the meconium staining degree on blood profile values of newborn puppies classified according to birth weight.

Blood Trait	Absent	Mild	Moderate	Severe
Q ₁				
pH	7.54 ± 0.0098 ^{a,1}	7.47 ± 0.0323 ^{a,1}	7.11 ± 0.0919 ^{b,1}	7.21 ± 0.0476 ^{b,1}
pCO ₂ (mmHg)	43.5 ± 0.378 ^{a,1}	45.4 ± 2.45 ^{a,1}	69.8 ± 2.45 ^{b,1}	67.9 ± 3.79 ^{b,1}
pO ₂ (mmHg)	18.9 ± 0.314 ^{a,1}	18.5 ± 0.666 ^{a,1}	11.8 ± 0.869 ^{b,1}	12.2 ± 0.482 ^{b,1}
Glucose (mg/dL)	101.00 ± 0.877 ^{a,1}	95.4 ± 4.70 ^{a,1}	55.0 ± 5.59 ^{b,1}	51.0 ± 3.76 ^{b,1}
Ca ⁺⁺ (mmol/L)	1.51 ± 0.0102 ^{a,1}	1.60 ± 0.0485 ^{a,1}	2.04 ± 0.0371 ^{b,1}	1.98 ± 0.0659 ^{b,1}
Lactate (mmol/L)	3.87 ± 0.145 ^{a,1}	4.96 ± 0.604 ^{a,1}	8.95 ± 0.736 ^{b,1,2}	10.1 ± 0.749 ^{b,1}
Hematocrit (%)	43.8 ± 0.259 ^{a,1}	44.9 ± 1.49 ^{a,1}	55.0 ± 1.21 ^{b,1}	54.0 ± 1.15 ^{b,1}
HCO ₃ ⁻ (mmol/L)	23.3 ± 0.189 ^{a,1}	21.9 ± 0.468 ^{a,1}	14.1 ± 0.435 ^{b,1}	13.5 ± 0.402 ^{b,1}
Q ₂				
pH	7.29 ± 0.0154 ^{a,2}	7.26 ± 0.0224 ^{a,1,2}	7.14 ± 0.0280 ^{a,1}	6.90 ± 0.0380 ^{b,2}
pCO ₂ (mmHg)	49.6 ± 0.774 ^{a,1,2}	60.5 ± 3.56 ^{a,b,1,2}	68.2 ± 4.42 ^{b,1}	82.6 ± 1.31 ^{c,2}
pO ₂ (mmHg)	15.5 ± 0.371 ^{a,2}	13.8 ± 0.824 ^{a,2}	12.5 ± 0.484 ^{a,b,1}	9.04 ± 0.637 ^{b,1,2}
Glucose (mg/dL)	103.0 ± 1.99 ^{a,1}	87.6 ± 5.28 ^{a,b,1}	76.0 ± 6.78 ^{b,1}	44.3 ± 3.75 ^{c,1}
Ca ⁺⁺ (mmol/L)	1.82 ± 0.0176 ^{a,2}	1.95 ± 0.0453 ^{a,b,2}	1.99 ± 0.0560 ^{b,1}	2.15 ± 0.0595 ^{b,1}
Lactate (mmol/L)	6.02 ± 0.184 ^{a,2}	7.74 ± 0.373 ^{a,1,2}	8.18 ± 0.596 ^{a,1}	11.0 ± 0.470 ^{b,1}
Hematocrit (%)	48.2 ± 0.435 ^{a,2}	49.0 ± 1.20 ^{a,1,2}	52.3 ± 1.14 ^{a,b,1}	54.2 ± 1.44 ^{b,1}
HCO ₃ ⁻ (mmol/L)	20.6 ± 0.247 ^{a,2}	19.2 ± 0.469 ^{a,1,2}	14.8 ± 0.562 ^{b,1}	13.1 ± 0.380 ^{b,1}
Q ₃				
pH	7.28 ± 0.0185 ^{a,2}	7.14 ± 0.0614 ^{a,b,2,3}	7.05 ± 0.0704 ^{b,1}	7.00 ± 0.0409 ^{b,2}
pCO ₂ (mmHg)	53.9 ± 1.58 ^{a,2}	64.9 ± 5.03 ^{a,b,2}	77.2 ± 4.34 ^{b,1}	77.7 ± 2.92 ^{b,1,2}
pO ₂ (mmHg)	14.9 ± 0.405 ^{a,2,3}	12.8 ± 0.976 ^{a,b,2}	9.75 ± 1.01 ^{b,1}	9.42 ± 0.709 ^{b,1,2}
Glucose (mg/dL)	105.0 ± 2.79 ^{a,1}	74.9 ± 7.15 ^{b,1,2}	63.4 ± 8.55 ^{b,1}	58.7 ± 4.92 ^{b,1}
Ca ⁺⁺ (mmol/L)	1.83 ± 0.0219 ^{a,2}	1.98 ± 0.0630 ^{a,b,2}	2.10 ± 0.0688 ^{b,1}	2.12 ± 0.0378 ^{b,1}
Lactate (mmol/L)	6.51 ± 0.258 ^{a,2}	9.46 ± 0.899 ^{b,2,3}	10.8 ± 0.832 ^{b,1,2}	11.4 ± 0.399 ^{b,1}
Hematocrit (%)	49.7 ± 0.445 ^{a,2}	51.2 ± 1.60 ^{a,b,2,3}	53.6 ± 1.89 ^{a,b,1}	54.2 ± 0.941 ^{b,1}
HCO ₃ ⁻ (mmol/L)	19.5 ± 0.207 ^{a,2}	17.8 ± 0.712 ^{a,2,3}	14.2 ± 0.526 ^{b,1}	13.2 ± 0.273 ^{b,1}
Q ₄				
pH	7.29 ± 0.0275 ^{a,2}	7.00 ± 0.0670 ^{b,3}	7.08 ± 0.0445 ^{b,1}	7.02 ± 0.0283 ^{b,2}
pCO ₂ (mmHg)	55.3 ± 2.13 ^{a,2}	83.5 ± 3.65 ^{b,3}	76.2 ± 3.05 ^{b,1}	83.1 ± 2.43 ^{b,2}
pO ₂ (mmHg)	12.7 ± 0.614 ^{a,3}	8.12 ± 1.13 ^{b,3}	10.0 ± 0.854 ^{a,b,1}	8.10 ± 0.731 ^{b,2}
Glucose (mg/dL)	103.0 ± 4.41 ^{a,1}	49.8 ± 4.14 ^{b,2}	61.2 ± 4.79 ^{b,1}	51.9 ± 3.05 ^{b,1}
Ca ⁺⁺ (mmol/L)	1.78 ± 0.0218 ^{a,2}	2.08 ± 0.0288 ^{b,2}	2.07 ± 0.0320 ^{b,1}	2.13 ± 0.0212 ^{b,1}
Lactate (mmol/L)	6.75 ± 0.443 ^{a,2}	11.9 ± 0.880 ^{b,3}	11.1 ± 0.630 ^{b,2}	12.0 ± 0.436 ^{b,1}
Hematocrit (%)	51.3 ± 0.689 ^{a,2}	56.7 ± 1.50 ^{b,3}	54.8 ± 0.970 ^{a,b,1}	55.0 ± 0.558 ^{b,1}
HCO ₃ ⁻ (mmol/L)	19.1 ± 0.422 ^{a,2}	15.6 ± 0.970 ^{b,3}	12.8 ± 0.565 ^{c,1}	13.2 ± 0.514 ^{b,c,1}

(Two-way ANOVA.) Q₁, 127–200 g; Q₂, 201 g–269 g; Q₃, 270 g–388 g; Q₄, 389 g–464 g. ^{a,b,c} Different superscripts between columns indicate statistically significant differences between the degree of meconium staining and quartiles with *p* < 0.05. ^{1,2,3} Different numbers among rows indicate statistically significant differences between blood values on different quartiles.

Table 5 shows that puppies with a higher degree of meconium staining presented low pH values. pCO₂ and Ca⁺⁺ increased in severe meconium staining, and pO₂ decreased. These findings suggest that as the degree of meconium staining increased, more significant blood profile alterations were observed regardless of weight or quartile. Ca⁺⁺, pCO₂, hematocrit, and lactate values increased directly in severe meconium staining. On the contrary, in severe meconium staining, the pH, pO₂, glucose, and HCO₃ decreases were inversely proportional.

A chi-square test was performed to determine whether the proportion of stillbirths differed between puppies classified according to birth weight groups and the presence of meconium staining. The proportion of stillbirth puppies significantly differed between

puppy weight quartiles and degrees of meconium staining ($\chi^2 = 89.475$, $df = 15$, $p < 0.001$). The number and percentage of liveborn (LB) and stillbirths (SBs) can be observed in Table 6.

Table 6. Number and percentage of liveborn (LB) and stillbirth (SB) puppies according to their weight and the presence of meconium staining on the skin: absent, mild, moderate, and severe, and significant statistical differences.

Staining Degree	Q ₁ n = 110		Q ₂ n = 108		Q ₃ n = 108		Q ₄ n = 109	
	LB (%)	SB (%)	LB (%)	SB (%)	LB (%)	SB (%)	LB (%)	SB (%)
Absent	77 (70) ^{a,1}	0 ^{a,1}	58 (53) ^{a,1}	3 (2.7) ^{a,1}	51 (47.2) ^{a,1}	2 (1.8) ^{a,1}	26 (23.8) ^{a,1}	2 (1.8) ^{a,1}
Mild	12 (10.9) ^{a,1,2}	0 ^{a,1,2}	14 (12.9) ^{a,1}	0 ^{a,1}	11 (10.1) ^{a,1}	3 (2.7) ^{a,1}	6 (5.5) ^{a,1}	6 (5.5) ^{a,1}
Moderate	6 (5.4) ^{a,2}	3 (2.7) ^{a,2}	12 (11.1) ^{a,1}	2 (1.8) ^{a,1}	10 (9.2) ^{a,1}	5 (4.6) ^{a,1}	18 (16.5) ^{a,1}	10 (9.1) ^{a,1}
Severe	10 (9) ^{a,1,2}	2 (1.8) ^{a,1,2}	12 (11.1) ^{a,1}	7 (6.4) ^{a,1}	18 (16.6) ^{a,1}	8 (7.4) ^{a,1}	23 (21.1) ^{a,1}	18 (16.5) ^{a,1}

(X^2 test). Q₁, 127–200 g; Q₂, 201 g–269 g; Q₃, 270 g–388 g; Q₄, 389 g–464 g. LB: liveborn; SB: stillbirth. ^a Different superscript between columns indicates statistically significant differences between quartiles and meconium staining degree with $p < 0.05$. ^{1,2} Different numbers among rows indicate statistically significant differences between the degree of meconium staining in liveborn and stillbirth puppies.

Table 6 shows that regardless of weight or quartile, the degree of meconium staining significantly affected the number of SBs, which was greater in severe meconium staining and lower or even null in absent meconium staining degree.

The correlation coefficients for the blood profile values, the degree of meconium staining, and the vitality score are reported in Table 7, Table 8, and Table 9, respectively.

Table 7. Significant correlations between puppies’ weight and blood profile values.

Variables	Correlation Coefficient (rho)	p-Value
Q ₁		
pH	−0.125	0.193
pCO ₂ (mmHg)	0.273	0.004 *
pO ₂ (mmHg)	−0.137	0.154
Glucose (mg/dL)	−0.165	0.085
Ca ⁺⁺ (mmol/L)	0.197	0.039
Lactate (mmol/L)	0.138	0.151
Hematocrit (%)	0.119	0.217
HCO ₃ [−] (mmol/L)	−0.109	0.257
Q ₂		
pH	0.178	0.065
pCO ₂ (mmHg)	−0.205	0.034 *
PO ₂ (mmHg)	0.168	0.082
Glucose (mg/dL)	0.324	<0.001 *
Ca ⁺⁺ (mmol/L)	−0.038	0.695
Lactate (mmol/L)	−0.214	0.026 *
Hematocrit (%)	−0.266	0.005 *
HCO ₃ [−] (mmol/L)	0.224	0.019 *
Q ₃		
pH	0.167	0.084
pCO ₂ (mmHg)	−0.006	0.954
pO ₂ (mmHg)	0.012	0.900
Glucose (mg/dL)	0.304	0.001 *
Ca ⁺⁺ (mmol/L)	−0.213	0.027 *
Lactate (mmol/L)	0.063	0.516
Hematocrit (%)	0.017	0.861
HCO ₃ [−] (mmol/L)	0.039	0.686
Q ₄		
pH	−0.587	<0.001 *
pCO ₂ (mmHg)	0.621	<0.001 *
PO ₂ (mmHg)	−0.512	<0.001 *
Glucose (mg/dL)	−0.579	<0.001 *
Ca ⁺⁺ (mmol/L)	0.412	<0.001 *
Lactate (mmol/L)	0.603	<0.001 *
Hematocrit (%)	0.532	<0.001 *
HCO ₃ [−] (mmol/L)	−0.541	<0.001 *

Spearman’s rank correlation coefficients and their statistical significance between puppies’ weight and their blood profile values. * Indicates significant statistical differences.

Table 8. Significant correlations between puppies' weight and meconium staining degree. The meconium staining degree has been recoded to numerical values. Higher the stain degree = higher number. Absent = 1, mild = 2, moderate = 3, and severe = 4.

Variables	Correlation Coefficient (<i>r</i>)	<i>p</i> -Value
All puppies (quartiles)	0.3319875	<i>p</i> < 0.001
Q ₁	0.111	0.248
Q ₂	−0.200	0.038 *
Q ₃	0.023	0.811
Q ₄	0.214	0.025 *

Spearman's rank correlation coefficients and their statistical significance between puppies' weight and their meconium staining degree. * Indicates significant statistical differences.

Table 9. Significant correlations between puppies' weight and its vitality.

Variables	Correlation Coefficient (<i>r</i>)	<i>p</i> -Value
	Q ₁	
Vitality Score min 1	0.0644	0.514
Vitality Score min 5	0.116	0.238
Vitality Score min 60	0.0325	0.742
	Q ₂	
Vitality Score min 1	0.0587	0.568
Vitality Score min 5	0.148	0.148
Vitality Score min 60	0.174	0.0882
	Q ₃	
Vitality Score min 1	−0.0323	0.762
Vitality Score min 5	0.0430	0.687
Vitality Score min 60	0.143	0.179
	Q ₄	
Vitality Score min 1	0.0912	0.443
Vitality Score min 5	0.0822	0.489
Vitality Score min 60	0.0887	0.456

Spearman's rank correlation coefficients and their statistical significance between puppies' weight and their vitality.

Table 7 shows that in Q₁, weight significantly affected pCO₂, with a positive correlation of 0.273 (*p* = 0.004). In Q₂, there was a significant effect of weight on pCO₂, lactate, glucose, hematocrit, and HCO₃. Q₃ had a positive correlation with glucose (*p* < 0.001) and calcium (*p* = 0.027). In the case of Q₄, there were significantly high positive and negative correlations, for all the variables with statistically significant levels of *p* < 0.001, so it can be concluded that having more weight causes high positive correlations with all blood variables. We have marked with an asterisk the values in which there were statistically significant correlations.

Table 8 reports that birth weight was correlated with the degree of meconium staining in Q₂, with $r^2 = -0.200$ and *p* = 0.038, and in Q₄, with $r^2 = 0.214$ and *p* = 0.025, so it can be stated that birth weight was correlated with meconium staining in Q₂ and Q₄. We have marked with an asterisk the values in which there were statistically significant correlations.

Table 9 shows that no statistically significant correlations were found in any of the groups or at any time due to the effect of birth weight and vitality.

4. Discussion

The results show that birth weight influences the thermoregulation, hemodynamic changes, and degree of meconium staining in newborn dogs, as reported in previous studies performed by the authors [4,49].

4.1. Blood Profile Values

The most notable blood profile changes were associations between Q₄ puppies (heavier newborns) with hypoxia, hypoglycemia, hypercalcemia, hyperlactatemia, and marked metabolic acidosis.

The plasma levels of HCO_3^- observed in this study were 21.7 mmol/L for Q₁ LB, 12.8 mmol/L for Q₁ SBs, 17 mmol/L for Q₄ LB, and 10.6 mmol/L for Q₄ SB. These values could demonstrate severe pulmonary hypoxia and elevated lactic acid production because when its production is greatly increased, the organism activates the use of HCO_3^- as a buffer. However, because of the increased consumption of HCO_3^- , newborns experience mixed (respiratory and metabolic) acidosis and metabolic deficiencies due to these anaerobic and respiratory processes that ultimately lead to hypoxia [50].

Variations in the balance of glucose levels during the neonatal stage depend on endogenous production and glucose metabolism, resulting in hyper- or hypoglycemia. According to Vannucchi et al. [51], hypoglycemia is the leading cause of neonatal mortality because a rapid depletion of energy resources can cause acute hypothermia. In other words, the rapid reduction in fetal circulation at delivery caused by uterine contractions and the pressure they can exert on the fetal umbilical cord is associated with rapid liver glycogen depletion and decreased glucose homeostasis [52]. In our study, within these adverse circumstances, we can mention that for Q₄, the whelping usually took longer because they are breeds that give birth to litters that are larger in number and size, which can prolong the expulsion interval. The significant differences in blood glucose concentration in the quartiles (62.8 mg/dL in Q₁; 65.3 mg/dL in Q₂; 71.3 mg/dL in Q₃, and 62.6 mg/dL in Q₄) could be associated with neonatal stress and the consequent release of catecholamine and promotion of liver glycogenolysis [53,54].

The reduction in the oxygen supply observed in the present study was also reported by Mota-Rojas et al. [46,47] in piglets, in which umbilical cord hypoxia leads to increased pCO₂ due to respiratory acidosis, acting as a risk for prenatal mortality. Similarly, in another study by Massip [16] in fetal sheep near term, the animals showed a compensatory mechanism to respiratory acidosis by increasing pCO₂. The contrary is observed during metabolic acidosis, leading to alveolar hyperventilation secondary to increased H⁺ ions in the plasma and a decrease in pCO₂.

These mentioned findings coincide with what was found in our study, where once again, the quartile of the heaviest puppies (Q₄) developed severe hypercapnia, ranging from 64 mmHg in LB to 95.1 mmHg in SB, and this could be explained by the expectation that during birth, there may be variations in cardiorespiratory parameters. Tachycardia, hypercapnia, and respiratory acidosis may arise immediately after birth. According to Alonso-Spilsbury et al. [55] and Vannucchi et al. [51], physiological hypoxia triggers an increase in respiratory rate and its consequent apnea. The important variations in pH in all quartiles, especially in Q₄ (LB 7.24; SB 6.82), show that bicarbonate and pCO₂ indicate neonatal acidosis and possible fetal hypoxic stress [56].

4.2. Vitality Score

Puppies of Q₄ had the lowest vitality rating of all the quartiles at minute 1 AB. This could be associated with some factors, such as the expulsion interval, since in smaller puppies, it is faster than in larger ones. Therefore, unlike larger puppies, which take longer to be expelled, the smaller ones do not consume energy reserves as much. Hence, the energy consumption in larger puppies to try to compensate for the stress of hypoxia is more significant in the expulsion phase. These biggest newborns have an increased risk of intrauterine asphyxia, affecting their survival chances [26].

Birth weight can also predispose to asphyxia. For example, in humans, low-birth-weight individuals have poor vitality scores [14]. Similarly, Okere et al. [57] found that piglet viability score was highly correlated with their weight ($r = 0.66$). In contrast to these studies, we found that the biggest puppies (Q₄) had low vitality scores at the first minute AB, but their score improved at minute 60 AB. Our results are similar to the ones reported by Trujillo et al. [58], who concluded that heavy piglets had scores ≤ 5 and showed asphyxia signs. A possible explanation could be that heavier piglets had greater difficulty passing through the birth canal, affecting the health and vitality of the newborns [4,59].

Additionally, Veronesi et al. [60] mention that small-sized puppies may have the highest levels of distress but higher chances of survival when compared to large-sized animals.

Vitality also influences the capability of a newborn to stand and ingest colostrum, limiting the absorption of nutrients and energy obtention [61], parameters that are important in the postpartum period because the disposition of energy resources, such as brown adipose tissue and glucose, differs between low- and high-birth-weight animals. Moreover, blood oxygenation, particularly oxygen saturation (SpO₂), has shown a negative correlation with birth weight, meaning that heavier newborns have lower SpO₂, and might influence the time to receive oxygen [62].

Surprisingly, as the minutes passed, it was possible to observe that the time factor favored Q₄ because the consumption of colostrum probably matched these puppies, and because at minute 60 AB, almost always, all newborns have already consumed colostrum, in addition to the fact that larger puppies will have more suction force and, therefore, consume more colostrum. This could be sustained with a study by Mila et al. [42], who mentioned that glucose in the colostrum is essential to newborn puppies because only 1.3% of the body fat is available. On the contrary, the colostrum of small-breed bitches (less than 10 kg) provides 10% more energy than the colostrum of large-breed females [3].

In the same way that Herpin et al. [53] and Trujillo-Ortega et al. [58,63] reported, newborn puppies that failed the vitality scale in our study showed a blood rise in pCO₂ and lactic acid, with a low pO₂ in plasma, finding that would confirm that these newborns suffered early tissue hypoxia.

Correlations between birth weight and vitality did not show a statistically significant difference, which can be explained by the fact that, even though the puppies in Q₄ showed failed vitality scores (5.77) at minute 1 AB, they managed to reach high vitality scores (7.48) at minute 60 AB, practically matching the vitality scores of the puppies of the other quartiles (Q₁, Q₂, and Q₃).

4.3. Meconium Staining Degree

Studies performed on piglets suggest that low-weight newborns have a higher frequency of intrapartum mortality [38,64,65]. In contrast, human babies with higher birth weights appear to have a higher predisposition to hypoxia and meconium aspiration [66,67]. This association could be related to parturition problems due to the size and weight of the newborn rather than problems with the umbilical cord. This also could explain why heavier intrapartum piglets showed more severe meconium staining in a study by Mota-Rojas et al. [30].

The present results show that 16.5% of stillbirths had severe meconium staining of the skin in Q₄ puppies. Most stillbirths had mild, moderate, and severe meconium staining (6, 10, and 18 puppies, respectively), suggesting that careful observation is required to assess intrapartum anoxia based on epidermal staining alone correctly. In other words, the highest percentages of stillbirths occurred in Q₄, with 33.02% (36 SB puppies from 109 in total), of which 5.5% were mild meconium staining, 9.1% were moderate, and 16.5% were severe.

The present study found an association between low blood pH values and severe meconium staining, regardless of birthweight, coinciding with previous studies where meconium staining demonstrated the ability to lead to fetal hypoxia and, consequently, acidosis [68]. The presence of meconium can obstruct the airways, leading to acute hypoxemia, hypercapnia, and metabolic acidosis due to tissular anaerobic metabolism that causes a progressive decrease in pulmonary blood flow, exacerbating hypoxia and acidosis [69]. A similar result was reported in humans, in whom 81.82% of newborns that presented thick meconium-stained fluid and low Apgar scores (around 6.45) had blood pH below 7.5 [70]. Therefore, observing meconium-stained puppies could indicate physiological alterations that might not always be evaluated during whelping but could be inferred to adopt a proper perinatal control of stained puppies.

One of the main limitations of this study was that the monitoring of the animals was carried out in 10 different clinics or hospitals to have a larger number of samples. For this

reason, factors such as standardizing the temperatures where the births took place were not possible, but to standardize the areas where the dams gave birth, foam mats and adult bed cover diapers were used. However, parturitions were carried out in Campeche, Mexico, where there is a tropical climate, so situations such as hypothermia due to environmental factors were not observed. As it is a species where the breeds have a great diversity of weights and sizes, the authors decided to classify the animals according to their weight and not their breed. Therefore, another limitation could be that some breeds could have more intense maternal care than others or have longer parturitions due to the size of the litter or newborns.

5. Conclusions

In the present study, we could observe that the weight of newborns was correlated with the degree of meconium staining, presenting more cases of severe meconium staining in the puppies of the highest-birth-weight group. In addition, the weight of the newborns was correlated with the alterations in the blood variables, showing the most severe cases of metabolic acidosis, hypoxia, hypoglycemia, and higher stillbirths in the puppies of the Q₄ quartile. On the contrary, no statistically significant correlations were found between the weight of newborns and vitality. However, it could be observed throughout the analysis of the results that the most vigorous puppies were found in Q₁. However, at minute 60 AB, all the puppies in the four quartiles standardized their vitality scores. According to this, our hypothesis that bigger newborns would have lower scores in vitality, more blood profile alterations, and more cases of meconium staining than smaller puppies was confirmed.

One of the main perspectives that we were able to elucidate with this study, and according to Groppetti et al. [71] and Forsberg [72], is that the use of both fetal and uterine monitoring before, during, and after parturition could be a valuable tool to help predict cases of hypoxia, asphyxia, dystocia, and uterine inertia, which could lead to prolonged deliveries with complications and, therefore, an increase in the mortality rate as well as a decrease in the vitality of newborns [73–76].

Author Contributions: All authors contributed to the conceptualization, writing, reading, and approval of the final manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: The study was conducted according to the guidelines of the Declaration of Helsinki. The Ph.D. Program in Biological and Health Science Academic Committee approved this project with the approval number CBS.114.19. The animals included in the present study were treated gently and were not stressed.

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

1. Veronesi, M. Assessment of canine neonatal viability—the Apgar score. *Reprod. Domest. Anim.* **2016**, *51*, 46–50. [[CrossRef](#)]
2. Veronesi, M.C.; Panzani, S.; Faustini, M.; Rota, A. An Apgar scoring system for routine assessment of newborn puppy viability and short-term survival prognosis. *Theriogenology* **2009**, *72*, 401–407. [[CrossRef](#)] [[PubMed](#)]
3. Chastant-Maillard, S.; Guillemot, C.; Feugier, A.; Mariani, C.; Grellet, A.; Mila, H. Reproductive performance and pre-weaning mortality: Preliminary analysis of 27,221 purebred female dogs and 204,537 puppies in France. *Reprod. Domest. Anim.* **2017**, *52*, 158–162. [[CrossRef](#)] [[PubMed](#)]
4. Lezama-García, K.; Martínez-Burnes, J.; Marcet-Rius, M.; Gazzano, A.; Olmos-Hernández, A.; Mora-Medina, P.; Domínguez-Oliva, A.; Pereira, A.M.F.; Hernández-Ávalos, I.; Baqueiro-Espinosa, U.; et al. Is the weight of the newborn puppy related to its thermal balance? *Animals* **2022**, *12*, 3536. [[CrossRef](#)]
5. Veronesi, M.C.; Fusi, J. Neonatal severity indicators in dogs. *Rev. Bras. Reprodução Anim.* **2021**, *45*, 525–532. [[CrossRef](#)]

6. Kirkden, R.D.; Broom, D.M.; Andersen, I.L. Invited review: Piglet mortality: Management solutions. *J. Anim. Sci.* **2013**, *91*, 3361–3389. [[CrossRef](#)]
7. Tønnessen, R.; Borge, K.S.; Nødtvedt, A.; Indrebø, A. Canine perinatal mortality: A cohort study of 224 breeds. *Theriogenology* **2012**, *77*, 1788–1801. [[CrossRef](#)] [[PubMed](#)]
8. Vassalo, F.G.; Simões, C.R.B.; Sudano, M.J.; Prestes, N.C.; Lopes, M.D.; Chiacchio, S.B.; Lourenço, M.L.G. topics in the routine assessment of newborn puppy viability. *Top. Companion Anim. Med.* **2015**, *30*, 16–21. [[CrossRef](#)] [[PubMed](#)]
9. Fusi, J.; Faustini, M.; Bolis, B.; Veronesi, M.C. Apgar score or birthweight in Chihuahua dogs born by elective Caesarean section: Which is the best predictor of the survival at 24 h after birth? *Acta Vet. Scand.* **2020**, *62*, 39. [[CrossRef](#)]
10. Gill, M.A. Perinatal and late neonatal mortality in the dog. *J. Am. Vet. Med. Assoc.* **2001**, *230*, 1463–1464.
11. Fix, J. Relationship of Piglet Birth Weight with Growth, Efficiency, Composition, and Mortality. Ph.D. Thesis, University of North Carolina State, Charlotte, NC, USA, 2010.
12. Manani, M.; Jegatheesan, P.; DeSandre, G.; Song, D.; Showalter, L.; Govindaswami, B. Elimination of admission hypothermia in preterm very low-birth-weight infants by standardization of delivery room management. *Perm. J.* **2013**, *17*, 8–13. [[CrossRef](#)]
13. Pereira, K.H.N.P.; Fuchs, K.M.; Corrêa, J.V.; Chiacchio, S.B.; Lourenço, M.L.G. Neonatology: Topics on Puppies and Kittens Neonatal Management to Improve Neonatal Outcome. *Animals* **2022**, *12*, 3426. [[CrossRef](#)]
14. Cavaliere, T.A. From fetus to neonate: A sensational journey. *Newborn Infant Nurs. Rev.* **2016**, *16*, 43–47. [[CrossRef](#)]
15. Mota-Rojas, D.; Villanueva-García, D.; Hernández-González, R.; Roldan-Santiago, P.; Martínez-Rodríguez, R.; Mora-Medina, P.; Trujillo-Ortega, M.E. Assessment of the vitality of the newborn: An overview. *Sci. Res. Essays* **2012**, *7*, 712–718. [[CrossRef](#)]
16. Massip, A. Relationship between pH, plasma cortisol and glucose concentrations in the calf at birth. *Br. Vet. J.* **1980**, *136*, 597–601. [[CrossRef](#)]
17. Wiberg, N.; Källén, K.; Olofsson, P. Physiological development of a mixed metabolic and respiratory umbilical cord blood acidemia with advancing gestational age. *Early Hum. Dev.* **2006**, *82*, 583–589. [[CrossRef](#)]
18. Nobre Pacífico Pereira, K.H.; Hibarú, V.Y.; Fuchs, K.M.; Cruz dos Santos Correia, L.E.; Lopes, M.D.; Ferreira, J.C.P.; Ferreira de Souza, F.; Machado, L.H.D.A.; Chiacchio, S.B.; Gomes Lourenço, M.L. Use of cardiac troponin I (cTnI) levels to diagnose severe hypoxia and myocardial injury induced by perinatal asphyxia in neonatal dogs. *Theriogenology* **2022**, *180*, 146–153. [[CrossRef](#)]
19. Bleul, U.; Lejeune, B.; Schwantag, S.; Kähn, W. Blood gas and acid-base analysis of arterial blood in 57 newborn calves. *Vet. Rec.* **2007**, *161*, 688–691. [[CrossRef](#)] [[PubMed](#)]
20. Andres, R.L.R.L.; Saade, G.; Gilstrap, L.C.C.; Wilkins, I.; Witlin, A.; Zlatnik, F.; Hankins, G.V.V. Association between umbilical blood gas parameters and neonatal morbidity and death in neonates with pathologic fetal acidemia. *Am. J. Obstet. Gynecol.* **1999**, *181*, 867–871. [[CrossRef](#)] [[PubMed](#)]
21. Armstrong, L.; Stenson, B.J. Use of umbilical cord blood gas analysis in the assessment of the newborn. *Arch. Dis. Child.-Fetal Neonatal Ed.* **2007**, *92*, F430–F434. [[CrossRef](#)]
22. Rodríguez Fernández, V.; López Ramón y Cajal, C.N.; Marín Ortiz, E.; Couceiro Naveira, E. Intrapartum and perinatal results associated with different degrees of staining of meconium stained amniotic fluid. *Eur. J. Obstet. Gynecol. Reprod. Biol.* **2018**, *228*, 65–70. [[CrossRef](#)]
23. Antończyk, A.; Kielbowicz, Z.; Nizański, W.; Ochota, M. Comparison of 2 anesthetic protocols and surgical timing during cesarean section on neonatal vitality and umbilical cord blood parameters. *BMC Vet. Res.* **2023**, *19*, 48. [[CrossRef](#)]
24. Rootwelt, V.; Reksen, O.; Farstad, W.; Framstad, T. Associations between intrapartum death and piglet, placental, and umbilical characteristics. *J. Anim. Sci.* **2012**, *90*, 4289–4296. [[CrossRef](#)] [[PubMed](#)]
25. Le Cozler, Y.; Guyomarc’h, C.; Pichodo, X.; Quinio, P.-Y.Y.; Pellois, H.H. Factors associated with stillborn and mummified piglets in high-prolific sows. *Anim. Res.* **2002**, *51*, 261–268. [[CrossRef](#)]
26. Lucia, T.; Corrêa, M.N.; Deschamps, J.C.; Bianchi, I.; Donin, M.A.; Machado, A.C.; Meincke, W.; Matheus, J.E.M. Risk factors for stillbirths in two swine farms in the south of Brazil. *Prev. Vet. Med.* **2002**, *53*, 285–292. [[CrossRef](#)] [[PubMed](#)]
27. Reyes-Sotelo, B.; Mota-Rojas, D.; Mora-Medina, P.; Ogi, A.; Mariti, C.; Olmos-Hernández, A.; Martínez-Burnes, J.; Hernández-Ávalos, I.; Sánchez-Millán, J.; Gazzano, A. Blood biomarker profile alterations in newborn canines: Effect of the mother’s weight. *Animals* **2021**, *11*, 2307. [[CrossRef](#)] [[PubMed](#)]
28. Reyes-Sotelo, B.; Ogi, A.; Mora-Medina, P.; Mariti, C.; Olmos-Hernández, A.; Hernández-Ávalos, I.; Domínguez-Oliva, A.; Rosas, M.E.; Verduzco-Mendoza, A.; Gazzano, A. Early blood analysis and gas exchange monitoring in the canine neonate: Effect of dam’s size and birth order. *Animals* **2022**, *12*, 1508. [[CrossRef](#)]
29. Ogi, A.; Naef, V.; Santorelli, F.M.; Mariti, C.; Gazzano, A. Oxytocin receptor gene polymorphism in lactating dogs. *Animals* **2021**, *11*, 3099. [[CrossRef](#)]
30. Mota-Rojas, D.; Martínez-Burnes, J.; Alonso-Spilsbury, M.L.; Lopez, A.; Ramirez-Necochea, R.; Trujillo-Ortega, M.E.; Medina-Hernandez, F.J.; de la Cruz, N.I.; Albores-Torres, V.; Loredo-Osti, J. Meconium staining of the skin and meconium aspiration in porcine intrapartum stillbirths. *Livest. Sci.* **2006**, *102*, 155–162. [[CrossRef](#)]
31. Martínez-Burnes, J.; Mota-Rojas, D.; Villanueva-García, D.; Ibarra-Ríos, D.; Lezama-García, K.; Barrios-García, H.; López-Mayagoitia, A. Meconium aspiration syndrome in mammals. *CAB Rev.* **2019**, *14*, 1–11. [[CrossRef](#)]
32. Chiruvolu, A.; Miklis, K.K.; Chen, E.; Petrey, B.; Desai, S. Delivery room management of meconium-stained newborns and respiratory support. *Pediatrics* **2018**, *142*, e20181485. [[CrossRef](#)]

33. Mota-Rojas, D.; Villanueva-García, D.; Mota-Reyes, A.; Orihuela, A.; Hernández-Ávalos, I.; Domínguez-Oliva, A.; Casas-Alvarado, A.; Flores-Padilla, K.; Jacome-Romero, J.; Martínez-Burnes, J. Meconium aspiration syndrome in animal models: Inflammatory process, apoptosis, and surfactant inactivation. *Animals* **2022**, *12*, 3310. [[CrossRef](#)] [[PubMed](#)]
34. Monfredini, C.; Cavallin, F.; Villani, P.E.; Paterlini, G.; Allais, B.; Trevisanuto, D. Meconium aspiration syndrome: A narrative review. *Children* **2021**, *8*, 230. [[CrossRef](#)]
35. Kirimi, E.; Tuncer, O.; Kösem, M.; Ceylan, E.; Tas, A.; Tasal, I.; Balahoroğlu, R.; Caksen, H.; Balahoroğlu, R.; Caksen, H. The effects of prednisolone and serum malondialdehyde levels in puppies with experimentally induced meconium aspiration syndrome. *J. Int. Med. Res.* **2003**, *31*, 113–122. [[CrossRef](#)] [[PubMed](#)]
36. Swarnam, K.; Soraisham, A.S.; Sivanandan, S. Advances in the management of meconium aspiration syndrome. *Int. J. Pediatr.* **2012**, *2012*, 1–7. [[CrossRef](#)] [[PubMed](#)]
37. Castagnetti, C.; Pirrone, A.; Mariella, J.; Mari, G. Venous blood lactate evaluation in equine neonatal intensive care. *Theriogenology* **2010**, *73*, 343–357. [[CrossRef](#)]
38. Randall, G.C. The relationship of arterial blood pH and pCO₂ to the viability of the newborn piglet. *Can. J. Comp. Med. Rev. Can. Med. Comp.* **1971**, *35*, 141–146.
39. Zaleski, H.M.; Hacker, R.R. Comparison of viability scoring and blood gas analysis as measures of piglet viability. *Can. J. Anim. Sci.* **1993**, *73*, 649–653. [[CrossRef](#)]
40. De Roth, L.; Downie, H.G. Evaluation of viability of neonatal swine. *Can. Vet. J. Rev. Vet. Can.* **1976**, *17*, 275–279.
41. Groppetti, D.; Ravasio, G.; Bronzo, V.; Pecile, A. The role of birth weight on litter size and mortality within 24 h of life in purebred dogs: What aspects are involved? *Anim. Reprod. Sci.* **2015**, *163*, 112–119. [[CrossRef](#)]
42. Mila, H.; Grellet, A.; Feugier, A.; Chastant-Maillard, S. Differential impact of birth weight and early growth on neonatal mortality in puppies. *J. Anim. Sci.* **2015**, *93*, 4436–4442. [[CrossRef](#)] [[PubMed](#)]
43. Mugnier, A.; Mila, H.; Guiraud, F.; Brévaux, J.; Lecarpentier, M.; Martinez, C.; Mariani, C.; Adib-Lesaux, A.; Chastant-Maillard, S.; Saegerman, C.; et al. Birth weight as a risk factor for neonatal mortality: Breed-specific approach to identify at-risk puppies. *Prev. Vet. Med.* **2019**, *171*, 104746. [[CrossRef](#)] [[PubMed](#)]
44. Tesi, M.; Miragliotta, V.; Scala, L.; Aronica, E.; Lazzarini, G.; Fanelli, D.; Abramo, F.; Rota, A. Relationship between placental characteristics and puppies' birth weight in toy and small sized dog breeds. *Theriogenology* **2020**, *141*, 1–8. [[CrossRef](#)] [[PubMed](#)]
45. Boyko, A.R.; Quignon, P.; Li, L.; Schoenebeck, J.J.; Degenhardt, J.D.; Lohmueller, K.E.; Zhao, K.; Brisbin, A.; Parker, H.G.; VonHoldt, B.M.; et al. A simple genetic architecture underlies morphological variation in dogs. *PLoS Biol.* **2010**, *8*, e1000451. [[CrossRef](#)]
46. Mota-Rojas, D.; Nava-Ocampo, A.A.; Trujillo, M.E.; Velázquez-Armenta, Y.; Ramírez-Necoechea, R.; Martínez-Burnes, J.; Alonso-Spilsbury, A. Dose minimization study of oxytocin in early labor in sows: Uterine activity and fetal outcome. *Reprod. Toxicol.* **2005**, *20*, 255–259. [[CrossRef](#)]
47. Mota-Rojas, D.; Rosales, A.M.; Trujillo, M.E.; Orozco, H.; Ramírez, R.; Alonso-Spilsbury, M. The effects of vetrabutrin chlorhydrate and oxytocin on stillbirth rate and asphyxia in swine. *Theriogenology* **2005**, *64*, 1889–1897. [[CrossRef](#)]
48. Sherwin, C.M.; Christiansen, S.B.; Duncan, I.J.; Erhard, H.W.; Lay, D.C.; Mench, J.A.; O'Connor, C.E.; Petherick, J.C. Guidelines for the ethical use of animals in applied ethology studies. *Appl. Anim. Behav. Sci.* **2003**, *81*, 291–305. [[CrossRef](#)]
49. Lezama-García, K.; Martínez-Burnes, J.; Pérez-Jiménez, J.C.; Domínguez-Oliva, A.; Mora-Medina, P.; Olmos-Hernández, A.; Hernández-Ávalos, I.; Mota-Rojas, D. Relation between the dam's weight on superficial temperature of her puppies at different stages of the post-partum. *Vet. Sci.* **2022**, *9*, 673. [[CrossRef](#)]
50. Manning, M. Electrolyte disorders. *Vet. Clin. N. Am.-Small Anim. Pract.* **2001**, *17*, 8–13. [[CrossRef](#)]
51. Vannucchi, C.L.; Rodrigues, J.A.; Silva, L.C.G.; Lúcio, C.F.; Veiga, G.A.L. A clinical and hemogasometric survey of neonatal lambs. *Small Rumin. Res.* **2012**, *108*, 107–112. [[CrossRef](#)]
52. Nowak, R.; Poindron, P. From birth to colostrum: Early steps leading to lamb survival. *Reprod. Nutr. Dev.* **2006**, *46*, 431–446. [[CrossRef](#)] [[PubMed](#)]
53. Herpin, P.; Le Dividich, J.; Hulin, J.C.; Fillaut, M.; De Marco, F.; Bertin, R. Effects of the level of asphyxia during delivery on viability at birth and early postnatal vitality of newborn pigs. *J. Anim. Sci.* **1996**, *74*, 2067. [[CrossRef](#)] [[PubMed](#)]
54. Tuchscherer, M.; Puppe, B.; Tuchscherer, A.; Tiemann, U. Early identification of neonates at risk: Traits of newborn piglets with respect to survival. *Theriogenology* **2000**, *54*, 371–388. [[CrossRef](#)]
55. Alonso-Spilsbury, M.; Mota-Rojas, D.; Villanueva-García, D.; Martínez-Burnes, J.; Orozco, H.; Ramírez-Necoechea, R.; Mayagoitia, A.L.; Trujillo, M.E. Perinatal asphyxia pathophysiology in pig and human: A review. *Anim. Reprod. Sci.* **2005**, *90*, 1–30. [[CrossRef](#)]
56. Zamora-Moran, E. Metabolic blood gas disorders. In *Acid-Base and Electrolyte Handbook for Veterinary Technicians*; Randels-Thorp, A., Liss, D., Eds.; John Wiley & Sons: Chichester, UK, 2017; pp. 121–135.
57. Okere, C.; Hacker, R.R.; Werchola, G. Relationships between serum IGF-I concentrations and piglet development or neonatal viability following porcine somatotropin (pST) and insulin administration to gestating gilts. *Theriogenology* **1997**, *47*, 1403–1412. [[CrossRef](#)] [[PubMed](#)]
58. Trujillo-Ortega, M.E.; Mota-Rojas, D.; Juárez, O.; Villanueva-García, D.; Roldán-Santiago, P.; Becerril-Herrera, M.; Hernández-González, R.; Mora-Medina, P.; Alonso-Spilsbury, M.; Rosales, A.M.; et al. Porcine neonates failing vitality score: Physio-metabolic profile and latency to the first teat contact. *Czech J. Anim. Sci.* **2011**, *56*, 499–508. [[CrossRef](#)]

59. Schrank, M.; Mollo, A.; Contiero, B.; Romagnoli, S. Bodyweight at birth and growth rate during the neonatal period in three canine breeds. *Animals* **2019**, *10*, 8. [[CrossRef](#)]
60. Veronesi, M.C.; Faustini, M.; Probo, M.; Rota, A.; Fusi, J. Refining the APGAR score cutoff values and viability classes according to breed body size in newborn dogs. *Animals* **2022**, *12*, 1664. [[CrossRef](#)]
61. Mota-Rojas, D.; Wang, D.D.-H.; Titto, C.G.; Martínez-Burnes, J.; Villanueva-García, D.; Lezama, K.; Domínguez, A.; Hernández-Avalos, I.; Mora-Medina, P.; Verduzco, A.; et al. Neonatal infrared thermography images in the hypothermic ruminant model: Anatomical-morphological-physiological aspects and mechanisms for thermoregulation. *Front. Vet. Sci.* **2022**, *9*, 63205. [[CrossRef](#)]
62. Røsvik, A.; Øymar, K.; Kvaløy, J.T.; Berget, M. Oxygen saturation in healthy newborns; influence of birth weight and mode of delivery. *J. Perinat. Med.* **2009**, *37*, 403–406. [[CrossRef](#)]
63. Trujillo-Ortega, M.E.; Mota-Rojas, D.; Olmos-Hernández, A.; Alonso-Spilsbury, M.; González, M.; Orozco, H.; Ramírez-Necoechea, R.; Nava-Ocampo, A.A. A study of piglets born by spontaneous parturition under uncontrolled conditions: Could this be a naturalistic model for the study of intrapartum asphyxia? *Acta Biomed.* **2007**, *78*, 29–35.
64. Marchant, J.; Rudd, A.; Mendl, M.; Broom, D.M.; Meredith, M.J.; Corning, S.; Simmins, P.H. Timing and causes of piglet mortality in alternative and conventional farrowing systems. *Vet. Rec.* **2000**, *147*, 209–214. [[CrossRef](#)]
65. Stanton, H.C.; Carroll, J.K. Potential mechanisms responsible for prenatal and perinatal mortality or low viability of swine. *J. Anim. Sci.* **1974**, *38*, 1037–1044. [[CrossRef](#)]
66. Cleary, G.M.; Wiswell, T.E. Meconium-stained amniotic fluid and the meconium aspiration syndrome. *Pediatr. Clin. N. Am.* **1998**, *45*, 511–529. [[CrossRef](#)] [[PubMed](#)]
67. Wiswell, T.E.; Bent, R.C. Meconium staining and the Meconium Aspiration Syndrome: Unresolved issues. *Pediatr. Clin. N. Am.* **1993**, *40*, 955–981. [[CrossRef](#)]
68. Mitchell, S.; Chandharan, E. Meconium-stained amniotic fluid. *Obstet. Gynaecol. Reprod. Med.* **2018**, *28*, 120–124. [[CrossRef](#)]
69. Olicker, A.L.; Raffay, T.M.; Ryan, R.M. Neonatal respiratory distress secondary to Meconium Aspiration Syndrome. *Children* **2021**, *8*, 246. [[CrossRef](#)] [[PubMed](#)]
70. Kima, V.; Roy, I.; Saikia, N. Correlation of umbilical cord arterial blood pH with meconium-stained liquor. *Indian J. Obstet. Gynecol. Res.* **2023**, *10*, 27–31. [[CrossRef](#)]
71. Groppetti, D.; Pecile, A.; Del Carro, A.P.; Copley, K.; Minero, M.; Cremonesi, F. Evaluation of newborn canine viability by means of umbilical vein lactate measurement, apgar score and uterine tocodynamometry. *Theriogenology* **2010**, *74*, 1187–1196. [[CrossRef](#)]
72. Forsberg, C.L. Pregnancy diagnosis, normal pregnancy and parturition in the bitch. In *BSAVA Manual of Canine and Feline Reproduction and Neonatology*; England, G.C., von Heimendahl, A., Eds.; British Small Animal Veterinary Association: Gloucester, UK, 2010; pp. 89–97.
73. Bienboire-Frosini, C.; Muns, R.; Marcet-Rius, M.; Gazzano, A.; Villanueva-García, D.; Martínez-Burnes, J.; Domínguez-Oliva, A.; Lezama-García, K.; Casas-Alvarado, A.; Mota-Rojas, D. Vitality in Newborn Farm Animals: Adverse Factors, Physiological Responses, Pharmacological Therapies, and Physical Methods to Increase Neonate Vigor. *Animals* **2023**, *13*, 1542. [[CrossRef](#)]
74. Uchanska, O.; Ochota, M.; Eberhardt, M.; Nizanski, W. Dead or Alive? A Review of Perinatal Factors That Determine Canine Neonatal Viability. *Animals* **2022**, *12*, 1402. [[CrossRef](#)] [[PubMed](#)]
75. Szenci, O. Importance of Monitoring Fetal and Neonatal Vitality in Bovine Practices. *Animals* **2023**, *13*, 1081. [[CrossRef](#)] [[PubMed](#)]
76. Plavec, T.; Knific, T.; Slapšak, A.; Raspor, S.; Lukanc, B.; Pipan, M.Z. Canine Neonatal Assessment by Vitality Score, Amniotic Fluid, Urine, and Umbilical Cord Blood Analysis of Glucose, Lactate, and Cortisol: Possible Influence of Parturition Type? *Animals* **2022**, *12*, 1247. [[CrossRef](#)] [[PubMed](#)]

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