

# Supplementary Material of Combinatorial Metabolic Engineering in *Saccharomyces cerevisiae* for the Enhanced Production of the FPP-Derived Sesquiterpene Germacrene

a)

LPP1_WT	ATGATCTCTGTATGGCGGATGAGAACATAAGGAGTATTAAAGCTATACTACTTCAG
LPP1_delta	ATGATCTCTGTATGGCGGATGAGAACATAAGGAGTATTAAAGCTATACTACTTCAG
	*****
LPP1_WT	TACATGATAATTGGTCTATGTACGATATTATTCCCTATTGGAGATATCCCTGGTACCT
LPP1_delta	TACATGATAATTGGTCTATGTACGATATTATTCCCTATTGGAGATATCCCTGGTACCT
	*****
LPP1_WT	AGGGGCCAACATCGAATTAGTCTTGATGACCCCAGTATATCAAAACGTTATGTACCT
LPP1_delta	AGGGGCCAACATCGAATTAGTCTTGATGACCCCAGTATATCAAAACGTTATGTACCT
	*****
LPP1_WT	AACGAACTCGTGGCCCACTAGAATGTTGATTGAGTGGACTGAGTAACATGGTC
LPP1_delta	AACGAACTCGTGGCCCACTAGAATGTTGATTGAGTGGACTGAGTAACATGGTC
	*****
LPP1_WT	GTCTTCTGGACCTGCATGTTGACAAGGACTTACTGAAGAAGAATAGAGTA-AAGAGACT
LPP1_delta	GTCTTCTGGACCTGCATGTTGACAAGGACTTACTGAAGAAGAATAGAGTA-TAAGAGACT
	*****
LPP1_WT	AAGAGAGAGGCCGGACCGAATCTGAACGATTTCACCTCATGCATACTAGCATTCTATG
LPP1_delta	AAGAGAGAGGCCGGACCGAATCTGAACGATTTCACCTCATGCATACTAGCATTCTATG
	*****
LPP1_WT	TCTGATGCTGATTATAAGCATAAATGCTGCCCTAACAGGCGCTTAAAGTTGATTATAGG
LPP1_delta	TCTGATGCTGATTATAAGCATAAATGCTGCCCTAACAGGCGCTTAAAGTTGATTATAGG
	*****
LPP1_WT	AAACTTGAGGCCTGACTTGTGATAGATGTACCTGACCTCCAAAGATGAGTGATTTC
LPP1_delta	AAACTTGAGGCCTGACTTGTGATAGATGTACCTGACCTCCAAAGATGAGTGATTTC
	*****
LPP1_WT	AGATTCTTGGTTGGCTTGGACATTGCAAGCAGACTAACAAATGGATTCTATACGA
LPP1_delta	AGATTCTTGGTTGGCTTGGACATTGCAAGCAGACTAACAAATGGATTCTATACGA
	*****

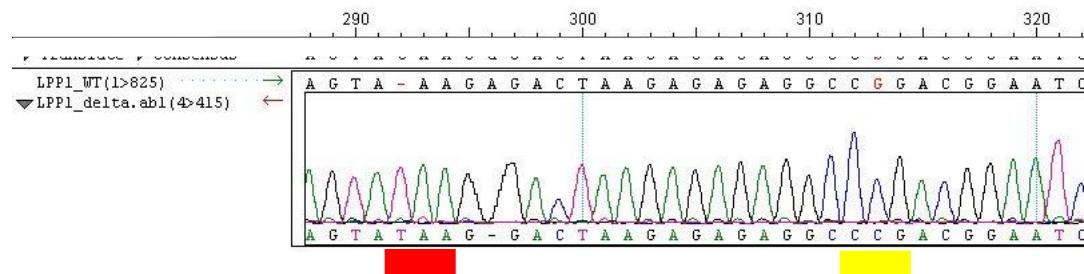
LPP1\_WT AGGCTTAAAAAGCACTCCAAGCGGACATTCAAGTTCATAGTCAGTACCATGGGCTTTAC  
 LPP1\_delta AGGCTTAAAAAGCACTCCAAGCGGACATTCAAGTTCATAGTCAGTACCATGGGCTTTAC  
 \*\*\*\*\*

LPP1\_WT ATATCTTGCAAAGGGTTTCACCACACGCAATAACAAGAAGTTGCATTGGTGCCCTT  
 LPP1\_delta ATATCTTGCAAAGGGTTTCACCACACGCAATAACAAGAAGTTGCATTGGTGCCCTT  
 \*\*\*\*\*

LPP1\_WT ATTAGCTCTAGTAGTAATGGTTCAAGGGTTATCGATCACAGACATCATTGGTACGATGT  
 LPP1\_delta ATTAGCTCTAGTAGTAATGGTTCAAGGGTTATCGATCACAGACATCATTGGTACGATGT  
 \*\*\*\*\*

LPP1\_WT TGTCTCTGGAGCTGTTCTAGCATTAGTCATTATTGGTCTGGAAATGGACATTAC  
 LPP1\_delta TGTCTCTGGAGCTGTTCTAGCATTAGTCATTATTGGTCTGGAAATGGACATTAC  
 \*\*\*\*\*

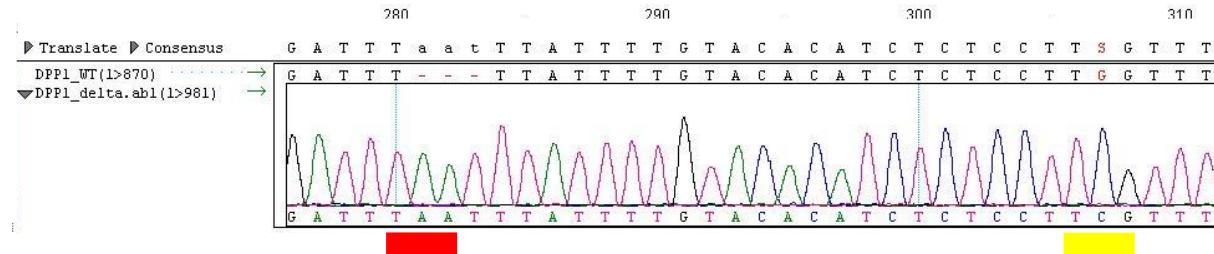
LPP1\_WT AAACTTGGCGAAAAGAGACATACTCCTTCACCGGTTAGTGTAG  
 LPP1\_delta AAACTTGGCGAAAAGAGACATACTCCTTCACCGGTTAGTGTAG  
 \*\*\*\*\*



**b)**

DPP1_WT	ATGAACAGAGTTCGTTATTAAAACGCCTTCAACATAGGGCGAAATGGAGATTAGAA
DPP_delta	ATGAACAGAGTTCGTTATTAAAACGCCTTCAACATAGGGCGAAATGGAGATTAGAA
*****	
DPP1_WT	GATGTCTTTGCTCATTATCATGATACTCTTAACACTACCCAGTGTATTACCAACAACCG
DPP_delta	GATGTCTTTGCTCATTATCATGATACTCTTAACACTACCCAGTGTATTACCAACAACCG
*****	
DPP1_WT	TTCGAACGTCAGTTACATTAACGATCTCACTATATCGCATCCTATGCGACAACGTGAA
DPP_delta	TTCGAACGTCAGTTACATTAACGATCTCACTATATCGCATCCTATGCGACAACGTGAA
*****	
DPP1_WT	CGTGTAAATAACAACATGTTGTTGTTATAGTTGCGGCCATCTTAACCATTG
DPP_delta	CGTGTAAATAACAACATGTTGTTGTTATAGTTGCGGCCATCTTAACCATTG
*****	
DPP1_WT	ATAATTGGTTCCATTTGGCCGATAGAACAGACATTGATT <del>TTT</del> TTGTACACATCT
DPP_delta	ATAATTGGTTCCATTTGGCCGATAGAACAGACATTGATT <del>TA</del> TTTGTACACATCT
*****	
DPP1_WT	CTCCT <del>TGG</del> TTTACTCGCTTGGTCAGTACGAGTTCTTACAAACTCATCAAGAAT
DPP_delta	CTCCT <del>TCG</del> TTTACTCGCTTGGTCAGTACGAGTTCTTACAAACTCATCAAGAAT
*****	
DPP1_WT	TGGATTGGAAGACTAAGACCAGATTCTAGATCGTGCAACCTGTTGAAGGCTTGCA
DPP_delta	TGGATTGGAAGACTAAGACCAGATTCTAGATCGTGCAACCTGTTGAAGGCTTGCA
*****	
DPP1_WT	TTGGACACTTATTACTGCAAAAGATGTTGACTGACTAAGAACGACGTCGTTG
DPP_delta	TTGGACACTTATTACTGCAAAAGATGTTGACTGACTAAGAACGACGTCGTTG
*****	
DPP1_WT	GATGGGTTTAGGACAACCTCGTCAGTCATTCAAGTGAAGCTTGAGGACTGGTTAT
DPP_delta	GATGGGTTTAGGACAACCTCGTCAGTCATTCAAGTGAAGCTTGAGGACTGGTTAT
*****	
DPP1_WT	TTGTACTTCTGGCTATGTGGCAACTTTGACTGAATCACCGTTGATGCCTTATGGAGA
DPP_delta	TTGTACTTCTGGCTATGTGGCAACTTTGACTGAATCACCGTTGATGCCTTATGGAGA
*****	

DPP1_WT	AAAATGGTGGCCTTCTACCACTGTTAGGAGCTGCACTAATTGCTCTATCCAGAACTCAA
DPP_delta	AAAATGGTGGCCTTCTACCACTGTTAGGAGCTGCACTAATTGCTCTATCCAGAACTCAA
	*****
DPP1_WT	GATTACAGACATCTCGTCGATGTAATTAGGGTCTATGTTGGGTTATATAATGGCA
DPP_delta	GATTACAGACATCTCGTCGATGTAATTAGGGTCTATGTTGGGTTATATAATGGCA
	*****
DPP1_WT	CACCTTTCTACAGAAGAACATTCCCACCCATTGATGATCCTCTTCCGTTCAAACCATTG
DPP_delta	CACCTTTCTACAGAAGAACATTCCCACCCATTGATGATCCTCTTCCGTTCAAACCATTG
	*****
DPP1_WT	ATGGACGATTAGATGTCACCCCTGGAGGAAGCAGTCACCCATCAGAGGATCCGGATGAG
DPP_delta	ATGGACGATTAGATGTCACCCCTGGAGGAAGCAGTCACCCATCAGAGGATCCGGATGAG
	*****
DPP1_WT	GAATTACATCCTTGTCCGATGAAGGTATGTAA
DPP_delta	GAATTACATCCTTGTCCGATGAAGGTATGTAA
	*****



**Figure S1.** Sequence alignments and sequencing data corresponding to the *LPP1*- and *DPP1*-knockout strains. **a)** Sequence alignment and corresponding sequence data of the *S. cerevisiae* *LPP1* wildtype-allele (*LPP1\_WT*) and *LPP1* knockout-allele (*LPP1\_delta*) resulting from the integration of a premature stop codon. **b)** Sequence alignment and corresponding sequence data of the *S. cerevisiae* *DPP1* wildtype-allele (*DPP1\_WT*) and *DPP1* knockout-allele (*DPP1\_delta*) resulting from the integration of a premature stop codon. Red boxes indicate the integrated stop codon; yellow boxes indicate the modified PAM; underlined sequences correspond to the presented sequencing results; alignments of sequences were generated using the Clustal MUSCLE algorithm (<http://www.ebi.ac.uk/Tools/msa/muscle/>); Sequencing data were analysed using Lasergene SeqMan Pro (DNASTAR).

**Table S1.** Oligonucleotides used to generate the guide RNA expression cassette in pML104 plasmids or for stop codon integration. Italic letters indicate 20mer guide sequences; bold letters indicate the integrated stop codon; italic and bold letters indicate the modified protospacer-adjacent motif (PAM) to prevent further Cas9 activity.

Oligonucleotide Name	Sequence (5' to 3')	Used for
<i>ROX1</i> -Guide RNA	GAT CTG AGG GGT CGA GTT AGC CCT GTT TTA GAG CTA G	ligation in pML104
<i>P<sub>erg9</sub></i> -Guide RNA	GAT CCG TGT TCT CGC TGC TCG <b>TTT</b> GTT TTA GAG CTA G	ligation in pML104
<i>LPP1</i> -Guide RNA	GAT CAG ACT AAG AGA GAG GCC GGA GTT TTA GAG CTA G	ligation in pML104
<i>DPP1</i> -Guide RNA	GAT CAT <b>TTT</b> GTA CAC ATC TCT CCT GTT TTA GAG CTA G	ligation in pML104
<i>LPP1</i> -Stop	TTT GAC AAG GAC TTA CTG AAG AAG AAT AGA GTA <b>TAA</b> GAG ACT AAG AGA GAG GCC CGA CGG AAT CTC GAA CGA <b>TTT</b> T	integration of stop codon
<i>DPP1</i> -Stop	ATT TTG GCC GAT AGA AGA CAT TTG ATT <b>TAA</b> TTT ATT TTG TAC ACA TCT CTC CTT CGT TTA TCA CTC GCT TGG TTC AG	integration of stop codon

**Table S2.** Yeast strains used to enhance the metabolic flux towards sesquiterpenoid FPP and the corresponding sterol- and farnesol/germacrene-A measurements. Values are represented in g/g CDW.

Yeast Strain	Short Name Yeast Strain	Initial pH	CuSO <sub>4</sub> (μM)	Squalene	2,3-Oxidosqualene	lanosterol	Ergosterol	Trans-Trans-Farnesol	Germacrene-A
WT		6.5	0	0.0032 ( $\pm 0.000877$ )	0.000015 ( $\pm 0.000003$ )	0.00424 ( $\pm 0.000691$ )	0.018667 ( $\pm 0.001036$ )	-	-
WT	ST0	6.5	0	0.004219 ( $\pm 0.00099$ )	0.000011 ( $\pm 0.000002$ )	0.007663 ( $\pm 0.001554$ )	0.020636 ( $\pm 0.002723$ )	0.000716 ( $\pm 0.000023$ )	0.000004 ( $\pm 0.000006$ )
<i>rox1::P<sub>GAL1</sub>-tHMGR P<sub>GAL10</sub>-ERG13</i>	ST1	6.5	0	0.150902 ( $\pm 0.044394$ )	0.000096 ( $\pm 0.000031$ )	0.127558 ( $\pm 0.043748$ )	0.082939 ( $\pm 0.02365$ )	-	-
<i>rox1::P<sub>GAL1</sub>-tHMGR P<sub>GAL10</sub>-ERG13 P<sub>GAL1</sub>-TkGAS2</i>	ST3	6.5	0	0.077391 ( $\pm 0.011625$ )	0.000063 ( $\pm 0.000008$ )	0.029367 ( $\pm 0.004729$ )	0.016594 ( $\pm 0.002407$ )	0.003338 ( $\pm 0.000284$ )	0.000393 ( $\pm 0.000038$ )
<i>rox1::P<sub>GAL1</sub>-tHMGR P<sub>GAL10</sub>-ERG13 P<sub>erg9Δ::P<sub>CTR3</sub></sub></i>	ST2	6.5	0	0.100485 ( $\pm 0.011249$ )	0.000109 ( $\pm 0.00003$ )	0.028349 ( $\pm 0.003271$ )	0.038247 ( $\pm 0.004749$ )	-	-
<i>rox1::P<sub>GAL1</sub>-tHMGR P<sub>GAL10</sub>-ERG13 P<sub>erg9Δ::P<sub>CTR3</sub></sub></i>	ST2	6.5	150	0.001521 ( $\pm 0.000204$ )	0.000782 ( $\pm 0.000056$ )	0.003958 ( $\pm 0.000602$ )	0.024172 ( $\pm 0.002494$ )	-	-
<i>rox1::P<sub>GAL1</sub>-tHMGR P<sub>GAL10</sub>-ERG13 P<sub>erg9Δ::P<sub>CTR3</sub></sub></i>	ST2	6.5	375	0.001337 ( $\pm 0.00023$ )	0.001044 ( $\pm 0.000239$ )	0.005181 ( $\pm 0.001151$ )	0.022969 ( $\pm 0.002797$ )	-	-
<i>rox1::P<sub>GAL1</sub>-tHMGR P<sub>GAL10</sub>-ERG13 P<sub>erg9Δ::P<sub>CTR3</sub></sub></i>	ST4	6.5	150	0.001268 ( $\pm 0.000067$ )	0.000387 ( $\pm 0.000029$ )	0.004861 ( $\pm 0.0001324$ )	0.011888 ( $\pm 0.001612$ )	0.122461 ( $\pm 0.003535$ )	0.001097 ( $\pm 0.000044$ )
<i>rox1::P<sub>GAL1</sub>-tHMGR P<sub>GAL10</sub>-ERG13 P<sub>erg9Δ::P<sub>CTR3</sub></sub></i>	ST5	4.5	15	0.002755 ( $\pm 0.000917$ )	0.003491 ( $\pm 0.000512$ )	0.007104 ( $\pm 0.001601$ )	0.03149 ( $\pm 0.005712$ )	0.11631 ( $\pm 0.003323$ )	0.007454 ( $\pm 0.000317$ )
<i>rox1::P<sub>GAL1</sub>-tHMGR P<sub>GAL10</sub>-ERG13 P<sub>erg9Δ::P<sub>CTR3</sub></sub></i>	ST5	6.5	150	0.001315 ( $\pm 0.000284$ )	0.001117 ( $\pm 0.000549$ )	0.003273 ( $\pm 0.000744$ )	0.013671 ( $\pm 0.001356$ )	0.111314 ( $\pm 0.003375$ )	0.00414 ( $\pm 0.000015$ )
<i>rox1::P<sub>GAL1</sub>-tHMGR P<sub>GAL10</sub>-ERG13 P<sub>GAL1-TkGAS2</sub></i>	ST5	6.5	150	0.041856 ( $\pm 0.005932$ )	0.000837 ( $\pm 0.000153$ )	0.02078 ( $\pm 0.003467$ )	0.038452 ( $\pm 0.004871$ )	0.032799 ( $\pm 0.009024$ )	0.003457 ( $\pm 0.001213$ )
<i>rox1::P<sub>GAL1</sub>-tHMGR P<sub>GAL10</sub>-ERG13 P<sub>erg9Δ::P<sub>CTR3</sub></sub></i>	ST6	8.5	150	0.00102 ( $\pm 0.000091$ )	0.000732 ( $\pm 0.000117$ )	0.003713 ( $\pm 0.002224$ )	0.009354 ( $\pm 0.003776$ )	0.091141 ( $\pm 0.002983$ )	0.000922 ( $\pm 0.000069$ )
<i>rox1::P<sub>GAL1</sub>-tHMGR P<sub>GAL10</sub>-ERG13 P<sub>erg9Δ::P<sub>CTR3</sub></sub></i>	ST7	6.5	150	0.000961 ( $\pm 0.000032$ )	0.000545 ( $\pm 0.0003278$ )	0.002599 ( $\pm 0.001472$ )	0.013053 ( $\pm 0.003278$ )	0.048922 ( $\pm 0.005419$ )	0.000743 ( $\pm 0.000307$ )



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