

Table S1: Scoring of user needs versus real components for MBR for process development Configuration I

Needs	Σ BioS			Σ TS						Σ IS		
	BioS entrapment			TS transport			TS containment			IS online data		
	Suspension ¹	Porous microcarrier ²	Hollow-fiber ²	Peristaltic pump	Syringe Pump ³	Electroosmotic pump ⁴	Plastic tube array ⁷	Steel microwell ⁷	Plastic microreactor ⁷	ISM ⁸	Capacitance ⁹	Online HPLC
MBR allows cell density of 0.1 10 ⁶ – 100 10 ⁶ cells/mL	•••	••	••	•••	••	•	•••	•••	•••	•••	•	••
MBR shall remain sterile for at least 30 days experimentation	•••	•	••	•••	•••	••	•••	•••	•••	••	••	••
Chamber of MBR shall not have any gradients	•••	••	•	•••	•••	••	•	••	•••	•••	•	•••
Mixing efficiency of MBR should reproduce large scale	•••	•	•	•••	•••	•••	•	••	•••	••	••	•••
MBR shall allow continuous and fed-batch operation	•••	•••	•	•••	•••	•	•	•	•••	•••	•••	•••
MBR should have as many real-time sensors as possible	•••	•	•	•••	•••	•••	•	•	••	••	•	•
MBR shall be easy to scale-out for high-throughput tests	•••	••	•••	•••	•••	•••	•••	••	••	•	•	•
MBR shall be convenient to operate for lab technicians	•••	•	•	•••	•••	•••	•••	•••	•••	•••	••	••
Fabrication cost of MBR shall be as low as possible	•••	•	•	•	•	•••	•••	•	••	•	••	•
TOTAL SCORE	27	14	13	25	24	20	19	18	24	20	15	20

Comments to table: ¹ Suspension culture usually preferred [63]. ² Common alternatives are microcarrier cultures and hollow-fibers [63]. ³ Syringe pump for precision pumping [64]. ⁴ Electroosmotic pump delivering an oscillating flow [65]. ⁷ Examples of MBR vessels are found in current commercial devices [19,20]. ⁸ In situ microscopy for real-time monitoring [66]. ⁹ For example, dielectric probes [67].

Table S2: Scoring of user needs versus real components for Heart-on-a-Chip Configuration I

	Σ BioS			Σ TS						Σ IS		
Needs	BioS _{entrapment}			TS _{transport}			TS _{containment}			IS _{beating rate}		
	Cardiac Bodies ¹	3D-membrane	3D-matrigel	Gravity transport ²	Syringe Injection ³	Electroosotic pump ⁴	PDMS 10-well array ⁵	PE 96-microwell ⁶	PDMS Microcolumn ⁷	ISM ⁸	HCS ⁹	MEA ¹⁰
HoC device allows co-cultures of cardiac cells	•••	••	••	•••	•••	••	•••	••	••	•••	•••	•
HoC device shall remain sterile for > 2 weeks experimentation	••	••	••	•••	•	•••	••	••	••	•••	•••	•
Clusters of cardiac cells shall be confined in HoC device chamber	•••	•	••	•••	•	•••	•••	•••	•••	•••	•••	•••
Cell number in HoC shall equal <i>in vivo</i> heart tissue equivalent	•••	••	••	•••	•••	•••	•••	•	•	•••	•••	•••
HoC shall allow continuous perfusion of culture media	•••	••	•	•••	•	•••	•••	••	••	•••	•••	••
HoC shall have confocal imaging optics and inline measurement for monitoring beating rate and troponin release	•	••	•••	•••	•••	•••	•••	•	•	-	•••	-
HoC shall be easy to scale-out for high-throughput testing to at least ten units	•••	•	••	•••	•••	•	••	•••	••	•	•••	••
HoC device shall be convenient to operate for lab technicians	•••	••	••	•••	••	••	••	•••	••	•••	•••	•••
Fabrication cost of HoC shall be as low as possible	•••	••	••	•••	•••	•••	•••	•••	•••	•••	•	••
TOTAL SCORE	24	16	18	27	20	23	24	21	19	22	25	17

Comments to table: ¹ Co-culture of cell types prepared in spheroids (cardiac bodies) as described in [72]. ² Rocker used to transport liquid in linear channel [86].

³Syringe pump for precision pumping [81]. ⁴Electroosmotic pump delivering an oscillating flow [65]. ⁶MIMETAS system [88]. ⁷ PDMS microfluidic channel packed with entrapped cells [87]. ⁸ In situ microscopy for real-time monitoring [94]. ⁹High content screening with e.g. Perkin Elmer OPERA system [89]. ¹⁰Microelectroarray for cardiac cells [84].