



Article Using Real Time Measurements to Derive the Indoor and Outdoor Contributions of Submicron Particulate Species and Trace Gases

Evdokia Stratigou *^D, Sébastien Dusanter, Joel Brito, Emmanuel Tison and Véronique Riffault *^D

Centre for Energy and Environment, IMT Nord Europe, University of Lille, Institut Mines-Télécom, F-59000 Lille, France; sebastien.dusanter@imt-nord-europe.fr (S.D.); joel.brito@imt-nord-europe.fr (J.B.); emmanuel.tison@imt-nord-europe.fr (E.T.)

* Correspondence: evdokia.stratigou@imt-nord-europe.fr (E.S.); veronique.riffault@imt-nord-europe.fr (V.R.); Tel.: +33-7-6807-6942 (E.S.)

Abstract: The indoor environment is usually more polluted than outdoors due to emissions of gas and particle-phase pollutants from multiple sources, leading to their accumulation on top of the infiltration of outdoor pollution. While it is widely recognized that negative health effects arise from the exposure to outdoor air pollution, exposure to indoor pollutants also needs to be well assessed since we spend most of our time (~90%) breathing indoors. Indoor concentrations of pollutants are driven by physicochemical processes and chemical transformations taking place indoors, acting as sources and/or sinks. While these basic concepts are understood, assessing the contribution of each process is still challenging. In this study, we deployed online instrumentation in an unoccupied room to test a methodology for the apportionment of indoor and outdoor pollutant sources. This method was successfully applied to the apportionment of PM₁ and VOCs, however, there are limitations for reactive gases such as O₃. The results showed that this unoccupied indoor environment acts as a source of VOCs and contributes 87% on OVOCs and 6% on C_xH_y, while it acts as a sink for particles, likely due to losses through volatilization up to 60%.

Keywords: indoor air quality; indoor air chemistry; particulate matter (PM); volatile organic compounds (VOC); apportionment methodology

1. Introduction

The indoor environment is usually more polluted than just the infiltrated ambient air due to the presence of additional sources emitting pollutants such as volatile organic compounds (VOCs, a glossary can be found in the Supplementary Information) and particulate matter (PM) in a reduced volume [1]. Nowadays, as part of climate change mitigation measures, improving energy efficiency in the built environment leads to an increasing demand on reducing air exchange with outdoors, which in turn increases human exposure to indoor pollution [2]. While it has been recognized worldwide that negative health effects correlate with outdoor pollutant levels, we spend most of our time breathing indoors. In particular, in industrially developed countries, it is estimated that people spend on average 90–95% of their time inside buildings and an additional 5.5% inside vehicles [2,3]. According to the World Health Organization (WHO) [4], 3.8 million premature deaths worldwide were attributable to household air pollution in 2016, accounting for 7.7% of the global mortality. Thus, a good understanding of both the sources and the sinks of indoor pollutants is important to design strategies aiming at reducing personal exposure to harmful species.

Due to their numerous health impacts on the respiratory and cardiovascular systems, as well as their genotoxicity and carcinogenicity [5–8], several countries have established air quality standards on ambient mass concentrations of PM with an aerodynamic diameter lower than 10 μ m (PM₁₀, termed inhalable PM) or 2.5 μ m (PM_{2.5}, termed fine PM).



Citation: Stratigou, E.; Dusanter, S.; Brito, J.; Tison, E.; Riffault, V. Using Real Time Measurements to Derive the Indoor and Outdoor Contributions of Submicron Particulate Species and Trace Gases. *Toxics* **2022**, *10*, 161. https:// doi.org/10.3390/toxics10040161

Academic Editor: Yuan Cheng

Received: 8 March 2022 Accepted: 26 March 2022 Published: 29 March 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). PM of even smaller sizes (e.g., PM_1 with a diameter lower than 1 μ m) are of particular interest since they can deposit more efficiently in the lower respiratory tract and access the circulatory system, also impacting other organs [9,10]. Furthermore, smaller particles are typically observed at higher number concentrations and therefore exhibit an overall larger surface area, which can act as a medium for the adsorption of organic pollutant [11]. However, no official regulations exist nowadays for this fraction. Only a few studies have assessed the specific impact of PM_1 on human health [12–17]. For instance, it has been found that a daily increase of 10 μ g m⁻³ is significantly associated with a 0.6% increase in total mortality [12] as well as an increased risk of emergency hospital visits [13] or with at least one lower respiratory symptom [15,17]. On the other hand, a wide range of VOCs can cause allergies, mucous membrane irritation and/or inflammation, systemic effects, such as fatigue and difficulty in concentrating, respiratory problems such as asthma and wheezing, as well as toxic effects such as carcinogenicity [18–22]. Official regulations for ambient VOCs (outdoors) concerns only benzene [23]. However, guide values have been established in 2011 by the French law for formaldehyde and benzene for establishments open to the public [24]. Besides, the French Agency for Food, Environmental and Occupational Health & Safety (ANSES) also recommends guide values for indoor air concerning ten additional gaseous pollutants taking into account short- and long-term exposures [25]. Indoor concentrations of particles and gases are driven by complex physical and chemical processes acting as sources—of primary (direct emission) or secondary (products of chemical transformations) nature, and sinks. Primary pollutants include species originating either from outdoor due to air exchange or from indoors (emissions from building materials, consumer products, occupants' activities), while sinks can be attributed to air exchange and adsorption/deposition processes for gases/particles, respectively. Secondary pollutants originate from gas-to-particle conversion processes occurring in the gaseous phase or at the gas-surface interface. The extent to which specific pollutants observed indoors are the result of outdoor infiltration or indoor processes mentioned above is still difficult to quantify. Methodologies developed for source apportionment of indoor pollution are mainly based on complex source receptor or personal exposure modelling (i.e., Positive Matrix Factorization, chemical mass balance model, gas phase chemistry or kinetic process modelling) [26–35], which often require numerous assumptions (e.g., that highly correlated compounds come from the same source) [36] and/or sets of observations exhibiting a large temporal variability to be able to deconvolve the different sources.

The objective of this work was to assess a simpler methodology capable of apportioning indoor and outdoor contributions to the observed indoor concentrations of pollutants, including organic and inorganic particles and gases. The study was conducted in an unoccupied low energy building, where we combined high (time and mass) resolution measurements of PM₁ and trace gases. In the following, we present indoor and outdoor contributions derived from the proposed methodology for the pool of pollutants mentioned above, discuss physicochemical processes impacting the composition of ambient infiltrated air, and address limitations associated to this methodology.

2. Materials and Methods

2.1. Measurement Facility

The measurement facility is located at the Institute Mines Telecom Nord Europe on the Douai Campus, Northern France. This building is mainly made of wood with a footprint of 12 m² and a height of 2.4 m, leading to a surface to volume ratio of $0.4 \text{ m}^2/\text{m}^3$ (Supplementary Information, Figure S1). It is a so-called energy-efficient building with the French BBC label (Bâtiment Basse Consommation—France) [37]. The test room chosen for our study is equipped with a dual-flow ventilation system with heat recovery (KWL EC 60 Pro, HELIOS) providing an air flow rate of 17 m³ h⁻¹. This system is equipped with an aluminum plate heat exchanger (efficiency higher than 70%), two fans with energysaving EC motors for homogeneous air exchange and two efficient air filters (class G4). Ventilation was kept on during the whole measurement period. The room was kept unfurnished and unoccupied during the experiments to minimize resuspension and material sources, as well as direct emissions by human activities (cooking, cleaning, occupancy, etc.).

The spatial homogeneity of PM within the room was investigated using the 2 AERO-TRACK instruments setup at various locations and heights, showing relative differences lower than 12% for all measured size ranges (0.3–0.5, 0.5–1.0, 1.0–3.0, 3.0–5.0, 5.0–10.0 µm), which is comparable to the instrumental uncertainty. The air exchange rate (α) was measured according to the ASTM E741-00 method [38] using CO₂ (Air Liquide) as a gas tracer and found constant over time at a value of 0.54 ± 0.05 h⁻¹ (n = 9). Particle penetration (P) was considered to occur only through the ventilation system since the building is tightly sealed to meet low energy requirements and was found to be 0.75 ± 0.09 for the PM₁ fraction (Figure S2). Penetration factors for gases were not measured but were inferred from the analysis shown hereafter. More details can be found in a previous study from our group [39].

2.2. Sampling Setup, Instrumentation and Measurement Procedures

An intensive campaign took place from 7 to 19 December 2017 and included online measurements of chemically-resolved indoor and outdoor particle concentrations and trace gases. More specifically, PM measurements were sequentially performed indoors and outdoors using a Scanning Mobility Particle Sizer (SMPS, TSI model 3788, 10.2–414.2 nm), two optical particle counters (OPC, AEROTRAK 8220, five size bins from 0.3 to 5.0 μ m) and an HR-ToF-AMS (Aerodyne Research Inc., AMS hereafter, chemically-resolved non-refractory PM₁). In addition, trace gases were measured using a PTR-QiToFMS (Ionicon Analytik GmbH, Innsbruck, Austria, PTRMS hereafter) for VOCs, an NO₂ CAPS (Aerodyne Research Inc., Billerica, MA, United States) for NO₂ and an O342 analyzer (Environnement SA) for ozone. Table 1 summarizes the instrumentation used during the intensive campaign together with time resolution and limits of detection (LOD).

Sequential indoor and outdoor particle concentration measurements were conducted continuously using two inlets made of stainless-steel and a valve-switching device that alternated the sampling between outdoor and indoor air every 5 min, resulting in a measurement every 10 min for each environment; the sampling line was shared between the AMS, the SMPS and the CO_2 analyzer as shown in Figure 1 (blue line). A Nafion dryer was used to minimize moisture upstream the AMS and SMPS. The AMS measured the non-refractory PM_1 (NR-PM₁) [40,41] with a time resolution of 5 min including a 50-s flush of the inlet before acquiring data. Measurements were performed using 5 cycles of 10 s in MS mode (V-mode) and 20 s in PToF mode. Due to issues with ion optics, the W-mode data were not used. Blank measurements were performed each day for 10 min by guiding the ambient air through a HEPA particle filter. Figure S3, panel b in SI depicts the detailed setup.

Calibrations of the inlet flow, the particle size and NO₃ ionization efficiency (IE), as well as the relative ionization efficiencies (RIE) of nitrate, sulfate and ammonium for the AMS measurements were performed twice during the campaign, using pure ammonium nitrate (NH₄NO₃) and ammonium sulfate ((NH₄)₂SO₄) particles and the setup shown in Figure S3, panel a, following the procedures recommended by the manufacturer (e.g., [42]).

Instrument (Manufacturer)	Measured Parameters or Species	Time Resolution	LOD (3σ) n/a		Location
AEROTRAK Handheld Particle Counter 8220 (TSI)	Particle number concentrations (6 size bins: 0.3–5.0 μm)	50 s			IN, OUT
HR-ToF-AMS (Aerodyne Research)	Non-refractory PM ₁ : NO ₃ , SO ₄ , Cl, NH ₄ and Organics	- 5 min - -	Species *	V mode ($\mu g m^{-3}$)	IN, OUT
			Org	0.64	
			SO ₄	0.05	
			NO ₃	0.28	
			NH_4	0.37	
			Cl	0.19	
PTR—QiToFMS (Ionicon Analytik)	VOCs	10 s	<i>m/z</i> 79: <30 ppt <i>m/z</i> 205: <1 ppt		IN, OUT
CO ₂ analyzer (Horiba)	CO ₂	1 min	0.2 ppb		IN, OUT
NO ₂ CAPS (Aerodyne Research)	NO ₂	10 s	0.1 ppb		IN, OUT
O ₃ analyzer (Environnement SA)	O ₃	10 s	0.4 ppb		IN, OUT
CO ₂ , RH, T probes (Testo)	CO ₂ , Relative Humidity, Temperature	20 s	n/a		IN
TEOM FDMS (Thermo Fisher Scientific, Waltham, MA, USA)	PM ₁ mass concentrations	1 h	n/a		OUT
SMPS (TSI, model 3788)	Particle number concentrations (10.2–414.2 nm)	4.5 min	n/a		IN, OUT
Spectroradiometer (METCON)	Photolysis rate of NO ₂ (J _{NO2})	1 s	n/a		IN

Table 1. Summar	y of instrumentation,	time resolution and	limit of detection ((LOD).
-----------------	-----------------------	---------------------	----------------------	--------

 \ast LOD has been calculated as the 3σ of the daily 10-min blank measurements.

During this campaign, PTRMS measurements [43,44] were automatically switched between indoor and outdoor every 5 min using solenoid valves (SV). The zeros were performed automatically every 70 min for a duration of 20 min (a total of 90-min cycle). The measurement time resolution was set to 10 s for each environment including zeroing. Both the O₃ and NO₂ analyzers were connected to the same sampling system with the same measurement sequence and time resolution (Figure 1, red line). For this study, the PTRMS reactor was operated with a voltage of 960 V, a temperature of 70 °C, and a pressure of 3.8 mbar resulting in an electric field (E/N) of 137 Td. The two 15-m long sampling lines were made of $\frac{1}{4}$ " PFA tubing. A PFA filter, changed every week, was used at the entrance of the line to filter aerosols. Ambient air was sampled at a total flow rate of 10 sLPM, leading to a residence time lower than 2 s in the sampling line. Calibrations have been conducted before (n = 6), during (n = 1) and after the end of the intensive campaign (n = 1). Section 1 in SI describes in detail the calibration procedure.

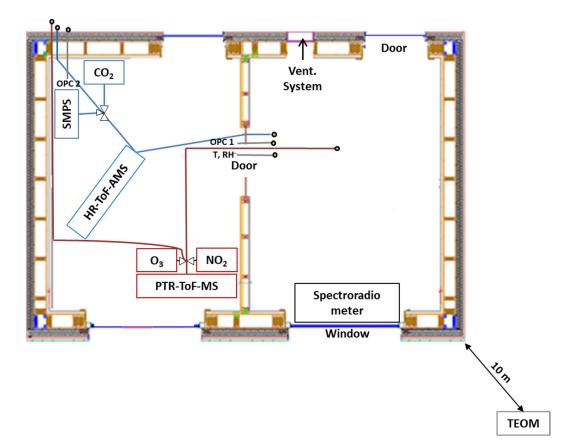


Figure 1. Schematic of the instrumentation room (left) and the experimental room (right).

2.3. Data Treatment for Online Mass Spectrometers

The AMS dataset was analyzed using Squirrel v.1.60 and Pika v.1.20 module in the Igor Pro version 6.37 (Wavemetrics Inc.) software.

Nitrate measured by AMS comes from NO⁺ and NO₂⁺ fragments. These fragments can originate from organic molecules such as RONO2 or inorganic molecules such as NH₄NO₃. According to Farmer et al., 2010 [45], the measured nitrate can be separated into these two types by applying the following equations. This separation is important, since it is expected that the two fragments have a different volatility with the inorganic fraction to be linked with NH₄NO₃ volatilization indoors [26]. The organic-bonded NO₃ fraction is given by:

$$pOrgNO_{3 frac} = \frac{\left(1 + R_{OrgNO_3}\right) \times (R_{measured} - R_{calib})}{\left(1 + R_{measured}\right) \times \left(R_{OrgNO_3} - R_{calib}\right)}$$
(1)

and the organic-bonded NO₃ mass concentration by:

$$\operatorname{orgNO}_{3} = \operatorname{pOrgNO}_{3 \operatorname{frac}} \times [\operatorname{NO}_{3}]$$
⁽²⁾

where $R_{measured}$ is the ratio between the signals NO_2^+ to NO^+ (or m/z46 and m/z30, respectively) and R_{calib} is the ratio observed during the calibration with ammonium nitrate (here 0.7). The value of R_{orgNO3} was set to be 0.1 according to other studies reporting measurements of organonitrates [45,46]. Moreover, this method is reliable for values of pOrgNO_{3 frac} > 0.15 and orgNO₃ > 0.1 µg m⁻³ [46]; thus, these thresholds were used to filter out the data (valid data: 77% indoors and 88% outdoors). Subsequently, the inorganic-bonded NO₃ (inorgNO₃) can be derived by subtracting orgNO₃ from the total NO₃ concentration.

Analysis of the PTRMS data was performed using PTRMS Viewer (v3.2.7). The first step consisted in recalibrating the mass scale of each acquired spectrum using the large

reference peaks at m/z21.022, 37.0275, 203.943 and 330.848 provided by H_3O^+ , $H_3O^+(H_2O)$ and the di-iodobenzene internal standard continuously injected in the PTRMS reactor through the built-in Permeation device for Mass Scale Calibration (PerMaSCal) feature. The second step required to generate a peak table containing all the peaks exhibiting an ambient signal statistically larger than the associated background signal. A total of 116 single peaks have been selected over the mass range 21–510 Th. The last step consisted in the deconvolution of all selected peaks using a multi-peak fitting tool. An additional step of filtering the extracted masses was necessary to remove ions generated in the ion source based on their mass, their behavior, and their temporal variability. The final dataset contains 91 ions attributed to ambient traces gases.

2.4. Indoor/Outdoor Apportionment Methodology

Each compound can be categorized as originating from indoor, outdoor, or both. In addition, when the outdoor environment is the main source, the indoor environment may either not significantly affect the amount of pollutant transmitted indoor or may act as a sink (through slow chemical processes or adsorption on surfaces).

For the analysis described below, measurements of PM_1 and trace gases were averaged over four hours to reduce the impact of the buffering time, which is defined here as the time needed for the indoor environment to respond to an abrupt change in concentration outdoors as previously discussed for the building used in this study [39].

In order to assess the contribution of indoor processes to the observed pollutant concentrations, we assumed perfect instantaneous mixing during air exchange and the invariability of air exchange, penetration and particle deposition throughout the 12-day monitoring period. Under these assumptions, the steady state indoor concentration of a pollutant is given by Equation (3):

$$C_{IN} = \frac{\alpha P C_{OUT}}{\alpha + K} + \frac{(\sum S)/V}{\alpha + K}$$
(3)

where V is the volume of the room (m³); C_{IN} and C_{OUT} , the indoor and outdoor mass concentrations of a pollutant, respectively; α , the air exchange rate (hr⁻¹); P, the penetration factor (dimensionless); K (hr⁻¹), either the deposition rate for particles or the first-order loss rate for the reaction of VOCs with gaseous oxidants (OH, O₃, NO₃); and Σ S an indoor source/sink term (mass unit per hour) accounting for adsorption/desorption processes leading to either net emission or sink, secondary formation in the gaseous phase (chemical transformations) and any other potential sources or sinks. For instance, if the analysis was focusing on gaseous formaldehyde, the source/sink term Σ S would account for direct emissions from building materials, the release from household chemicals and gas-phase formation from the oxidation of other VOCs.

The first term on the right-hand side of Equation (3) represents the contribution of species with an outdoor origin (Outdoor Contribution: OC), which results from outdoor pollutants partially penetrating indoors through the ventilation system or air leakage through cracks and openings, while the second term on the right-hand side represents the contribution of species with an indoor origin (Indoor Contribution: IC). The indoor concentration of a pollutant can therefore be expressed as follows:

$$C_{\rm IN} = OC + IC \text{ with } OC = F_{\rm INF} \times C_{\rm OUT}$$
(4)

The term F_{INF} ($F_{INF} = \frac{aP}{a+K}$) is the dimensionless infiltration factor, which represents the penetration factor for outdoor trace gases when chemical reaction rates are considered negligible compared to air exchange, and the fraction of particles that penetrates and remains suspended for outdoor particles. In the following, IC and OC are referred to as indoor and outdoor contributions, respectively.

For this study, infiltration factors were derived for all VOCs as the slope of a linear regression of measured indoor (C_{IN}) vs. outdoor (C_{OUT}) concentrations, similar to an

approach already applied for $PM_{2.5}$ [47]. However, the amplitude of ΣS in Equation (3) can slowly change over time due to changes in environmental conditions, such as temperature, relative humidity, solar irradiation, etc. This change can lead to some scatter in the data when C_{IN} is plotted versus C_{OUT} , and as a consequence to a bias in the retrieval of F_{INF} . In order to minimize this issue, scatter plots were color-coded by time to check whether a time-dependence was observed, and if so, the dataset was binned in time segments (two or more groups of points) where observations indicate that ΣS was not significantly changing. Individual scatter plots related to each group of points showed similar slopes and only the intercepts, which represent average indoor contributions, were different. In this case, a weighted-average value of F_{INF} was derived from the weighted slopes:

$$F_{\rm INF} = \frac{\sum b_i N_i}{\sum N_i}$$
(5)

where b_i are the slopes observed for different groups of points and N_i the number of data points.

As shown in Equation (4), the product between F_{INF} and the outdoor concentration of the targeted pollutant provides the contribution of the outdoor environment (OC) to the observed indoor concentration. The contribution of the indoor environment (IC) is given by the difference between C_{IN} and OC. Note that IC and OC are time-dependent and derived with a time resolution of 4 h (averaging time of the indoor/outdoor dataset as mentioned above).

This methodology is based on the assumption that the indoor reactivity of the targeted pollutants does not significantly impact their ambient concentration. Thus, the slope of the regression line between indoor and outdoor concentrations for a specific trace gas only depends on its penetration factor. However, the reaction rate of some reactive VOCs with gaseous oxidants (OH, O₃, NO₃) can compete with air exchange, leading to a breakdown of the assumption (Table S2 in SI). For instance, assuming OH and NO₃ indoor mixing ratios of 20 ppq and 1 ppt as observed during some studies [48–51] and a maximum O₃ mixing ratio of 20 ppb observed in this study, species such as isoprene and monoterpenes are found to break the assumption. This limitation also applies to ozone as discussed in the result section below. Other species including hydrocarbons such as aromatic and OVOCs, such as alcohols, aldehydes, ketones and carboxylic acids, are found to be less reactive, allowing the use of the methodology described above.

Finally, global 4-h time-resolved emission rates were calculated for each VOC considering their indoor contribution:

$$ER = \alpha \times IC \tag{6}$$

where α is the air exchange rate (hr⁻¹) and IC the time resolved indoor contribution (μ g m⁻³).

It is interesting to note that the use of the indoor contribution (IC) instead of (C_{IN} - C_{OUT}) for the calculation of emission rates as performed in other studies [52,53] is more accurate since the potential loss of VOCs in the ventilation system is then taken into account.

3. Results

This section first gives an overview of the indoor and outdoor composition and application of the described methodology to calculate the time-resolved IC, OC and ER for the different compounds (VOCs, inorganic gases, PM₁ species).

3.1. Indoor and Outdoor Composition

Meteorological data—During the campaign, the temperature ranged between -0.6 and 8.4 °C outdoors and from 8.3 to 14.1 °C indoors (no heating inside the facility), leading to a positive difference of 2.3–11.1 °C, while the relative humidity varied from 68–96% and 43–76% outdoors and indoors, respectively.

Gas phase compounds—Average concentrations for inorganic gases reached 9.3 ± 5.5 ppb and 13.1 ± 6.9 ppb for O₃; and 11.7 ± 6.9 ppb and 11.2 ± 7.9 ppb for NO₂

for indoors and outdoors, respectively. Gas phase measurements also included 91 quantified ions (Table S3 in SI reports the exact protonated masses detected by PTRMS and examples of VOCs monitored at these m/z) both indoors and outdoors, with the exception of 3 ions observed indoors that were below detection limit outdoors. These ions include 20 hydrocarbons and potential fragments of protonated alcohols (C_xH_y), and 60 OVOCs which were further separated into three categories given their oxidation state ($C_xH_yO-n = 27$, $C_xH_yO_2-n = 24$ and $C_xH_yO_{z\geq3}-n = 9$). From this dataset, 81 molecular formulas were successfully attributed to the detected ions from their exact masses, but 11 masses above 133 Th were not identified due to a lack of mass resolution and are referred to as "Others" hereafter.

Timeseries of meteorological parameters as well as C_xH_y and OVOCs compounds are shown in Figure 2 (panels a and b, respectively).

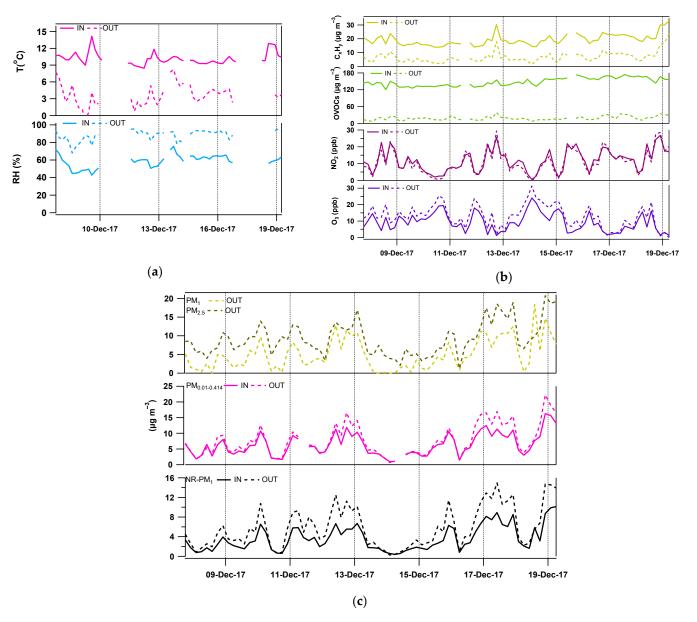


Figure 2. Timeseries of (**a**) meteorological parameters, (**b**) gas phase compounds, and (**c**, from top to bottom) particle mass concentrations monitored by TEOM-FDMS, SMPS and AMS indoors (line) and outdoors (dashed line).

On average, OVOCs contributed 88% to the total observed VOC concentration indoors, while only 69% outdoors. Conversely, hydrocarbons contributed 11% indoors compared to 29% outdoors (Figure 3a,b). The contribution of unidentified VOCs (categorized as Others) was lower than 3% for both indoors and outdoors. The total mass of VOCs increased from 26 to 167 μ g m⁻³ when ambient air was transferred indoors, a change which is mainly due to an increase of OVOCs by a factor of 8 and an increase of C_xH_y by a factor of 3. It should be noted that this comparison likely underestimates the contribution of hydrocarbons to the total pool of VOCs since the PTRMS technique is blind to alkanes, which have been observed at significant concentrations indoors [54,55]. More details for each species are given in Table S4.

Previous studies have also reported high concentrations of VOCs in various indoor environments (school classrooms, offices, houses/apartments) [28,32,52,56–69]. Interestingly, the concentrations observed in unoccupied buildings are elevated, often reaching similar or even higher levels than under occupied conditions. The wood structure of the facility does not lead to a significant increase of isoprene or monoterpenes compared to other buildings.

Figure 3 also reports the contribution of OVOCs outdoors (panel c) and indoors (panel d) grouped by category (C_xH_yO , $C_xH_yO_2$, $C_xH_yO_{z\geq 3}$). It is evident that while the most abundant compounds outdoors are those with one atom of oxygen, the most abundant compounds indoors contain two oxygen atoms, with the $C_xH_yO_2$ contribution to the total OVOCs more than doubling indoors. In the latter category, acetic ($C_2H_4O_2.H^+$) and formic ($CH_2O_2.H^+$) acids contribute 44% and 13%, respectively, to the total concentration of VOCs, while their respective contributions outdoors are 13% and 4%. These findings imply that there is a strong source of carboxylic acids indoors, possibly being emitted by building materials and/or produced by chemical transformations, accounting for most of the OVOC mass, in agreement with previous studies [56,70,71].

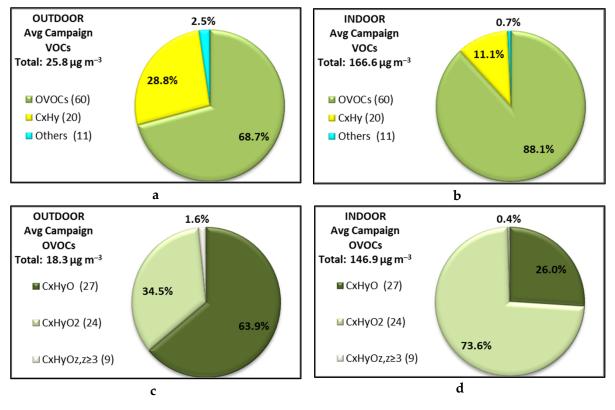


Figure 3. Cont.

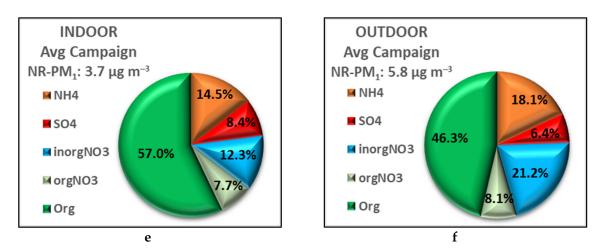


Figure 3. (**a**,**b**) Contributions of OVOCs (green), hydrocarbons and potential fragments of alcohols (C_xH_y , yellow) and others (light blue) to the observed pool of VOCs (**a**) outdoors and (**b**) indoors; (**c**,**d**) contributions of C_xH_yO (dark green), $C_xH_yO_2$ (light green) and $C_xH_yO_{z\geq3}$ (white) compounds to the total pool of observed OVOCs (**c**) outdoors and (**d**) indoors. Values in parenthesis indicate the number of measured species. (**e**,**f**) Contribution of inorganic (NH₄: orange, inorgNO₃: light blue, orgNO₃: light green, SO₄: red) and organic (green) fractions to the total mass of NR-PM₁ particles (**e**) indoors and (**f**) outdoors.

Particulate Matter—Concentrations of particles in the size range 10.2-414 nm (from SMPS measurements) ranged from 0.98–16.3 μ g m⁻³ and 0.70–22.2 μ g m⁻³ indoors and outdoors, respectively, showing a similar temporal variability in both environments, though in lower concentrations indoors by approximately 13%. Moreover, the NR-PM₁ indoor concentrations are usually lower than outdoor concentrations, showing mean Indoor-to-Outdoor ratios (I/O) below unity, in agreement with previous studies [27,72]. More specifically, Figure 3 (panels e and f) depicts the contribution of inorganic (NH_4 : orange, inorg NO_3 : light blue, SO_4 : red), organic (green) and org NO_3 (light green) fractions to the total mass. Total concentration of NR-PM₁ has decreased from 5.8 μ g m⁻³ outdoors to 3.7 μ g m⁻³ indoors (Figure 3e,f). The organic and sulfate contributions increased by approximately 10% and 2% from outdoors to indoors, while nitrate (mainly the inorganic bonded nitrate, inorgNO₃) and ammonium contributions decreased by 9% and 4%, respectively. Finally, particles measured by the optical counters (OPC, 0.3 to 10.0 µm) followed a similar behavior showing lower concentrations indoors compared to outdoors (I/O ratio 0.7 for the smaller fraction, down to 0.14 for larger particles). Descriptive statistics for each particle species are presented in Table S5.

Previous studies conducted in a classroom, a guest and residential house and a multiuse building [26,27,31,72] reported concentrations outdoors in the range of 0.86–4.0 μ g m⁻³, 0.44–3.2 μ g m⁻³, 1.35–7.2 μ g m⁻³, and 3.24–8.2 μ g m⁻³ for SO₄, NH₄, NO₃ and Org, respectively, while indoors the respective concentrations were 0.28–2.0 μ g m⁻³, 0.05–0.9 μ g m⁻³, 0.04–1.4 μ g m⁻³ and 1.14–4.4 μ g m⁻³, being similar to that observed in our study apart from SO₄ and Org outdoors which are slightly lower.

3.2. Contribution of the Indoor Environment to Observed Gaseous Pollutant Concentrations

The Indoor-to-Outdoor (I/O) ratio is only a primary indicator for the existence (or not) of indoor sources. In order to decouple the contributions of indoor and outdoor sources to observed indoor pollutant concentrations, the methodology described in Section 2.4 was applied using Equations (3) and (4).

The results of this study varied depending on the targeted pollutant. A compound exhibiting a large IC value (Figure 4, panel I) is categorized as a compound of indoor origin, while one originating from both indoors and outdoors will exhibit IC and OC values of the same magnitude (panel III). When the outdoor environment is the main source (panels II

and IV), the indoor environment might act as a sink due to chemical reactivity or adsorption on surfaces for gases (IC < 0; panel II) or might not significantly impact the pollutant levels (indoor and outdoor concentrations exhibiting close covariation; panel IV).

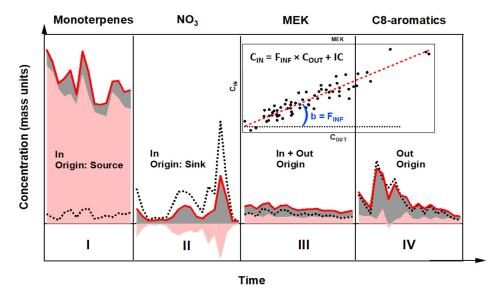


Figure 4. Schematic of possible indoor (IC; shaded pink area) and outdoor (OC; shaded grey area) contributions for four cases, calculated from indoor (red solid line) and outdoor (black dotted line) mass concentrations. Gas phase: panels I, III, IV. Particle phase: panel II. Inset plot: example of scatter plot for the retrieval of the infiltration factor (species: MEK).

VOCs—Figure 5 displays penetration factors for all monitored ions (blue), derived using the methodology described in Section 2.4 as a function of the four categories of VOCs defined above (C_xH_v , C_xH_vO , $C_xH_vO_2$ and $C_xH_vO_{z>3}$). For this analysis, only scatter plots of C_{IN} vs. C_{OUT} exhibiting statistically significant slopes, i.e., a 3 σ relative standard deviation (RSD) lower than 50%, were accepted. As can be observed from this figure, the penetration efficiency decreases from approximately 0.9 (C_xH_y) to 0.7 ($C_xH_yO_3$) as the number of oxygens in the molecule increases. This observation is likely due to a decrease in VOC volatility when the number of oxygen atoms increases, which translates into a higher "stickiness" on filters located in the ventilation system, especially for OVOCs in contact with humid surfaces since most of them are water soluble [73]. While three compounds or group of isomers ($C_{10}H_{16}$, $C_{9}H_{14}O$, $C_{7}H_{14}O_{2}$) seem to exhibit a penetration factor higher than unity, the lower bounds (considering an uncertainty at 3σ) are close to unity. As mentioned above, the use of a C_{IN}-C_{OUT} scatter plot may not reliably provide the penetration factor if the compound of interest exhibits a significant reactivity with ambient oxidants (competition with air exchange). It is interesting to note that $C_{10}H_{16}$ compounds are likely monoterpenes, for which the assumption of low reactivity indoors breaks down (Section 2.4). In addition, oxidation products from monoterpenes are expected to be observed as $C_9H_{14}O$ compounds (limonaketone, nopinone). The larger-than-expected penetration factors observed for C₉H₁₄O and C₇H₁₄O₂ compounds are therefore likely due to an impact of chemical processes on the slopes of the C_{IN}-C_{OUT} scatter plots.

Example of scatter plots between measured indoor and outdoor concentrations, colorcoded according to the date of the measurements, are shown in Figure 6 for selected species exhibiting different origins and behaviors (a: main indoor origin, b: both indoor and outdoor origins, c: main outdoor origin and indoor environment acting as a sink, d: main outdoor origin). For instance, the scatter plot for MEK (detected as $C_4H_8O.H^+$ in PTRMS) shown in panel b leads to a slope of 0.85 ± 0.05 (1σ), which represents the penetration factor for this compound, and a y-intercept of 0.27 ± 0.02 (1σ) µg m⁻³, which provides an average value of its IC over the whole campaign. As shown on panel a, a precise penetration factor cannot be derived for compounds exhibiting a large y-intercept such as acetic acid 3.0

2.7

2.4

2.1

1.8

1.5

1.2

0.9

0.6

0.3

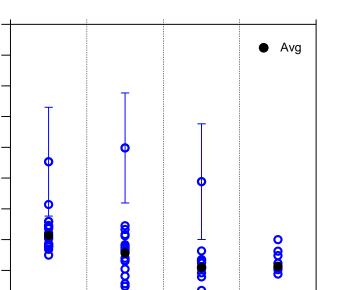
CxHy

N=20

CxHyO

N=27

Penetration factor



 $(C_2H_4O_2.H^+)$ since they show poor correlation with outdoor concentration and their main source is the indoor environment.

Figure 5. Penetration factors derived for different categories of VOCs. Average values are shown in black. Error bars represent 3σ. N is the number of species reported for each category.

CxHyOz (z>=3)

N=9

CxHyO2

N=24

Figure 6 also shows concentrations measured indoors (red line) and outdoors (black dashed line), together with the indoor (pink fill) and outdoor (grey fill) contributions for the selected compounds. In addition, the relative indoor contribution (RIC = IC/C_{IN} , purple dashed line), relative humidity (RH, blue dashed line) and temperature (T, pink dashed line) measured indoors are also plotted, together with the PTRMS detection limit (LOD, orange dashed line). As mentioned above, four main categories of species exhibiting different origins and behaviors can be discussed separately:

(i) A compound characterized by a much higher indoor than outdoor contribution (typically an average RIC over 70%) is categorized as a compound of indoor origin (Figure 6a). Acetic acid ($C_2H_4O_2.H^+$) is one such compounds exhibiting this behavior with an average indoor contribution of 70.8 µg m⁻³ (RIC of 97%).

(ii) A compound is categorized as originating from both indoors and outdoors (Figure 6b) when the average RIC is between 30% and 70%. $C_4H_8O.H^+$ (tentatively identified as methylethylketone) exhibits an average RIC of 47% (IC = 0.3 µg m⁻³), indicating a similar impact of both environments on indoor concentrations.

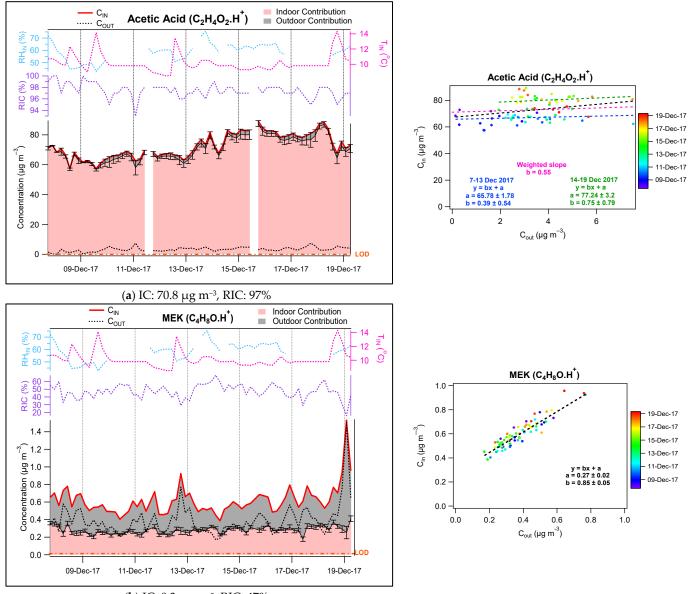
(iii, iv) Compounds originating mainly from outdoors (average IC below 30%; Figure 6c,d). For these species, indoor concentrations can be highly variable since it mainly depends on the OC and as a consequence on fast varying outdoor sources. This case can be further divided into two categories depending whether the indoor environment acts as a sink (IC < 0) or does not significantly impact the concentration of the infiltrated compound (IC \approx 0).

(iii) For instance, the IC for ethanol ($C_2H_5O.H^+$; Figure 6c) is negative most of the time, with approximately 32% of this compound lost, either through chemical processes (reaction with oxidants) or adsorption on surfaces. Since ethanol reacts slowly with oxidants but is known to stick easily on surfaces, adsorption processes are likely the cause for its indoor sink. Since the adsorption rate depends on the indoor VOC concentration, this process will likely lead to an underestimation of the penetration factor that is derived from the

C_{IN}-C_{OUT} scatter plot, which in turn may lead to an underestimation of the sink effect (overestimation of IC).

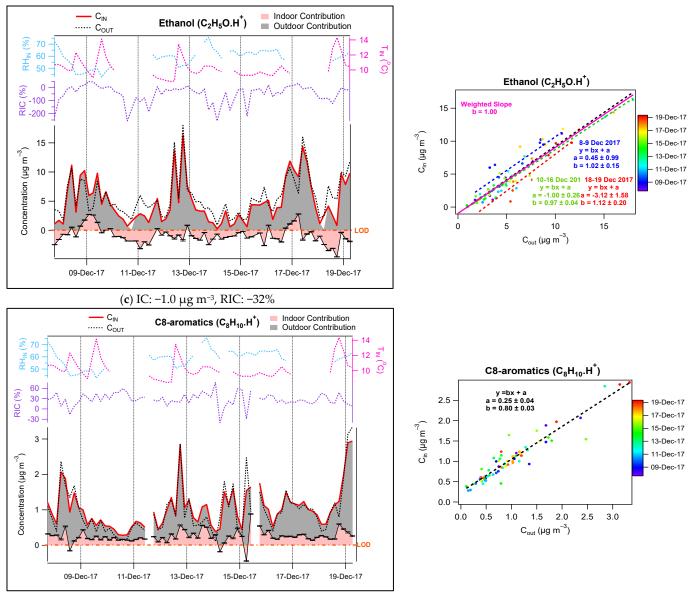
(iv) The example of C8-aromatics (C_8H_{10} .H⁺; Figure 6d) is also interesting because the time series of indoor and outdoor concentrations are very close and covary together over time, suggesting that these compounds are only of outdoor origin. However, the same figure also shows that the indoor environment contributes to approximately 30% of the indoor concentration. These contrasting observations are likely due to a compensation between a penetration factor lower than unity and the presence of small indoor emissions.

The average relative indoor (RIC, in pink) and outdoor (ROC, in grey) contributions for all compounds measured during the intensive campaign are displayed in Figure 7. They are also reported in Table S6, together with the slopes of C_{IN} - C_{OUT} scatter plots and absolute indoor contributions (IC).

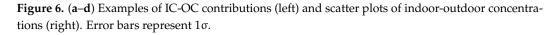


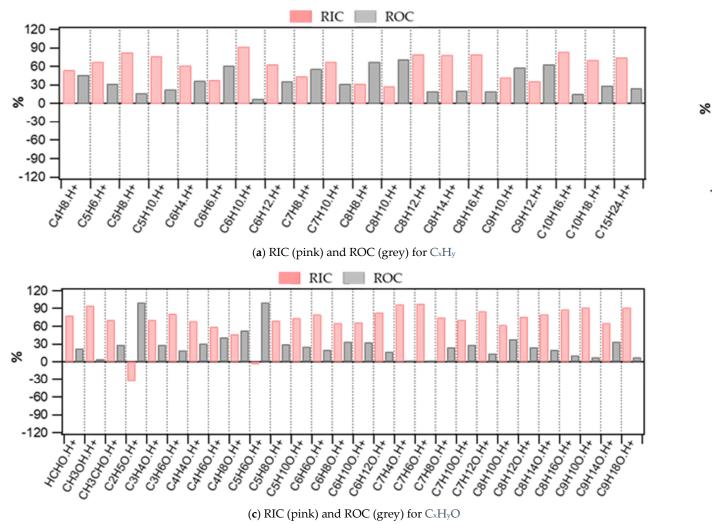
⁽b) IC: 0.3 μg m^-3, RIC: 47%

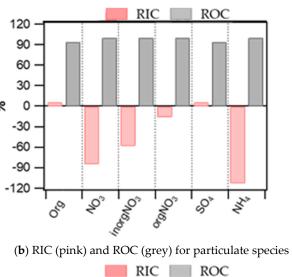
Figure 6. Cont.

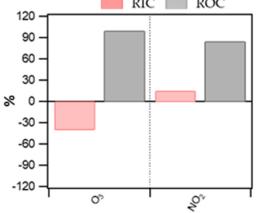


(d) IC: 0.25 μg m^-3, RIC: 28%



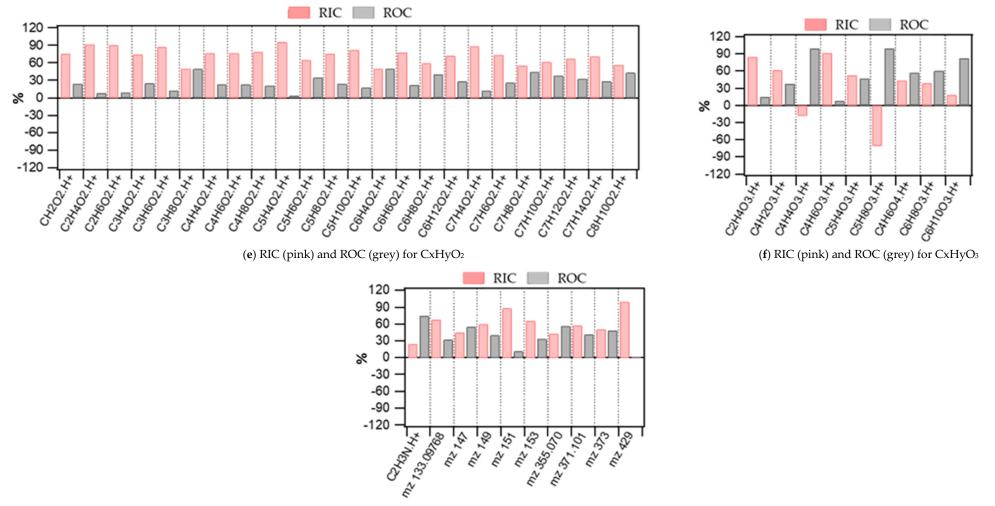






(d) RIC (pink) and ROC (grey) for inorganic gases

Figure 7. Cont.



(g) RIC (pink) and ROC (grey) for unidentified species (Others)

Figure 7. (a-g) Average relative indoor and outdoor contributions (RIC and ROC, respectively) for all compounds during the intensive campaign.

Considering all the ions detected by PTRMS, the majority of them (53%) originates from indoors (RIC > 70%), corresponding to 93% of the total indoor mass of measured VOCs. The other half of the compounds is divided between compounds which had the outdoor air as a main source (8% of the compounds with RIC < 30%), holding 4% of the total indoor mass, and compounds that had both outdoor and indoor contributions (39% of the compounds, 30% < RIC < 70%), holding 3% of the total indoor mass (Figure 8). As shown in Figure 8, a large mass contribution (68%) for species of indoor origin comes from compounds containing two atoms of oxygen, which likely include a large number of carboxylic acids. Indeed, formic acid ($CH_2OH.H^+$) and acetic acid ($C_2H_4O_2.H^+$) contribute 20% and 68%, respectively, to the total indoor concentration of this family. High concentrations of carboxylic acids have also been previously reported by Liu et al. [56] in a university classroom, indicating that formic acid is emitted by wood-based products or latex paint, which are two materials that were present in our building. In addition, large concentrations of OVOCs observed in our study (methanol) have also been observed in other types of buildings and are actually in the lower range of reported values [28,30]. These results tend to indicate that the building used in our study has a chemical signature similar to that of other buildings previously used to investigate indoor air quality.

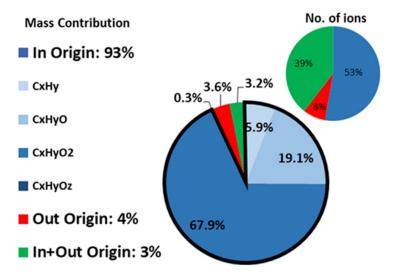
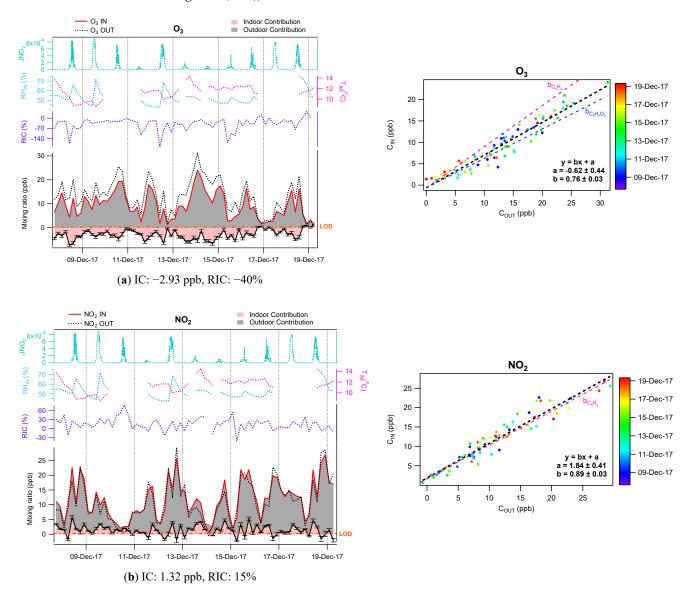


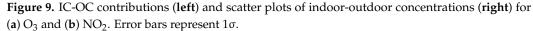
Figure 8. Origin of VOCs (grouped by category) observed indoors in terms of number of ions (small pie) and of mass concentration (large pie).

These results show that the indoor environment acts mainly as a source of VOCs by emitting these compounds continuously, even in the absence of occupants and furnishing. Average emission rates (ER; Equation (6)) were found to be 0.20, 0.19, 0.14, 0.03 and 0.04 μ g m⁻³ h⁻¹ for C_xH_y, C_xH_yO, C_xH_yO₂, C_xH_yO_{z≥3} and unidentified species, respectively. A few species have been observed to exhibit high emission rates amongst the abovementioned VOC categories. For instance, methanol (CH₃OH.H⁺) has also been reported in previous studies and attributed to emissions from wood decomposition [74]. Formic (CH₂OH.H⁺) and acetic acids (C₂H₄O₂.H⁺) are emitted as well by wood-based products and wood decomposition or latex paint [56,74]. C₅H₄O₂.H⁺ is tentatively identified as furfural, which has been reported in previous studies as emitted by wood products too [56,75]. Overall emission rates observed in this study are inside the observed range of previous studies [33,76].

Inorganic gases—Indoor and outdoor contributions were also inferred for both O_3 and NO_2 . The scatter plots between indoor and outdoor concentrations shown in Figure 9 show a small dispersion around the regression line, indicating some variability in the strength of their indoor sources and sinks, however not significantly larger than for VOCs reported in Figure 6. The penetration factors of approximately 0.9 and 0.7 observed for C_xH_y and $C_xH_yO_z$ species are also reported on these scatter plots for comparison. It can be

seen that the slope observed for NO₂ is very close to the penetration factor characteristic of C_xH_y species, which seems to indicate that the slope is not affected by significant indoor chemistry, while for O₃, the slope of the scatter plot is closer to that observed for $C_xH_yO_z$ species. The lower slope for O₃ could be either due to fast indoor reactions involving ozone or a loss of ozone into the ventilation system. In order to perform the IC-OC analysis for O₃, the penetration factor observed for C_xH_y species was used in the calculations assuming that O₃ is lost through indoor reactivity. This penetration factor has to be seen as an upper limit due to possible losses of O₃ in the ventilation unit, which in turn will lead to a lower limit (respectively, an upper limit) of IC if its sign is positive (source) (respectively, negative (sink)).





As can be seen in Figure 9, the indoor environment acts mainly as a sink for O_3 and as a small source for NO_2 , with RIC of -40% and +15%, respectively. Interestingly, both O_3 and NO_2 are known to be linked together through the O_3 - NO_x photostationary state (PSS) in the atmosphere (Reactions 7–9) [77].

$$O_3 + NO \rightarrow NO_2 + O_2 \tag{7}$$

$$NO_2 + h\nu \to NO + O(^3P) \tag{8}$$

$$O(^{3}P) + O_{2} + M \rightarrow O_{3} + M$$

$$\tag{9}$$

During daytime, this PSS is disturbed when ambient air is transferred indoors since the photolysis frequency of NO₂ is larger outdoors than indoors. When the air enters the room, the photolysis rate of NO₂ (Reaction 8) decreases and, as a result, the PSS switches towards the formation of NO₂. This effect will appear indoors as a source of NO₂ and a sink of O₃. According to reactions 7–9, the conversion between O₃ and NO₂ should be 1:1. However, the IC-OC analysis revealed a larger decrease of ozone compared to the increase of NO₂ indicating that a significant amount of O₃ is lost indoors, likely through its reaction with unsaturated species in the gas-phase and heterogeneous reactions on surfaces.

3.3. Contribution of the Indoor Environment to the Observed Concentrations of Submicron *Particulate Species*

The contribution of the indoor environment to PM species concentrations may arise from indoor emissions but also physicochemical processes, such as volatilization, condensation and chemical transformations, which can impact the slope of the C_{IN} - C_{OUT} scatter plot. The slope of the scatter plot provides in the case of particles the infiltration factor, which is different from the penetration factor since deposition can be significant. Unlike trace gases, the RIC in the case of particles was normalized to OC to avoid large uncertainties related to low concentrations that are sometimes observed indoors.

SO₄—For sulfate, a low level of dispersion around the regression line and an intercept close to zero is observed on the scatter plot shown in Figure 10 (panel a). The penetration factor derived for SO₄ from the scatter plot is 0.80, which is consistent with the penetration factor of 0.75 measured for PM₁ particles (Figure S2 in SI). This agreement is expected considering the absence of both (i) known sources of SO₄ indoors [27] and (ii) physicochemical processes that would significantly affect its concentration. As a consequence, the indoor contribution of sulfate was found to be close to zero (IC = $0.02 \ \mu g \ m^{-3}$, Figure 10b). Since the behavior of SO₄ only reflects the impact of mechanical losses, the infiltration factor determined for this species was used below to infer indoor contributions for other components of NR-PM₁ (NO₃, NH₄, Org) and to assess whether additional mass losses due to volatilization and other physicochemical transformations are occurring for these species (Figure S5 in SI).

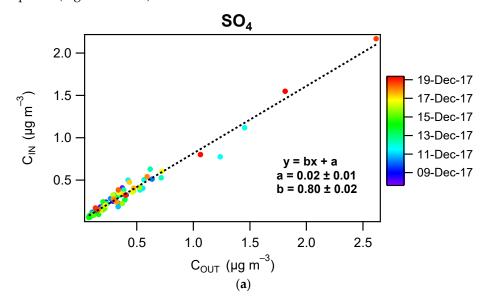


Figure 10. Cont.

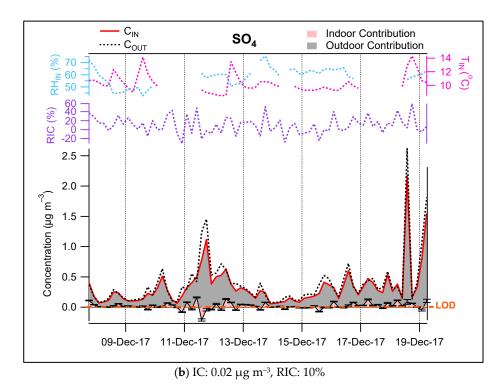


Figure 10. (**a**) Scatter plot of indoor-outdoor concentrations color-coded by date for NR-PM₁ SO₄, (**b**) IC-OC contribution. Error bars represent 1σ.

NH₄ and NO₃—The RIC of nitrate (orgNO₃ and inorgNO₃) and ammonium were found to be -19%, -57% and -47%, respectively (Figure S6 in SI), clearly showing that the indoor environment acts as a sink for both NH₄ and NO₃. This behavior is likely due to the volatilization of NH₄NO₃ when ambient air enters into a warmer environment. As shown in Figure 11, the average RIC of NH₄, inorgNO₃ and orgNO₃ decreased by 6%, 50% and 5%, respectively, as the difference in temperature between indoors and outdoors (Δ T) increased from at least 2 °C (<5 °C to >7 °C). These results indicate a higher volatilization rate for inorgNO₃ compared to orgNO₃ since the latter is less volatile than the inorganic fraction. This result, which has also been observed by Avery et al. [26], not only corroborates the IC/OC calculation method, but independently verifies as well the orgNO₃ estimation from AMS [45].

Organics—The average RIC for the organic fraction was found to be 3% ranging from -34% to 65% (Figure 12).

The RIC dependency on the indoor-outdoor temperature gradient showed a decrease from 13% (enrichment in organics) to -8% (depletion) for the total organic fraction when Δ T increases from <5 °C to >7 °C (Figure 13). According to Avery et al. [26], many families of organics and their gas-phase precursors are water soluble; thus, these species will be more likely lost when passing through the ventilation system which contains aqueous films. This is consistent with previous observations from this work for VOCs (Figure 5), where the penetration efficiency of the oxygenated VOCs decreased as the number of oxygens increased in the molecule.

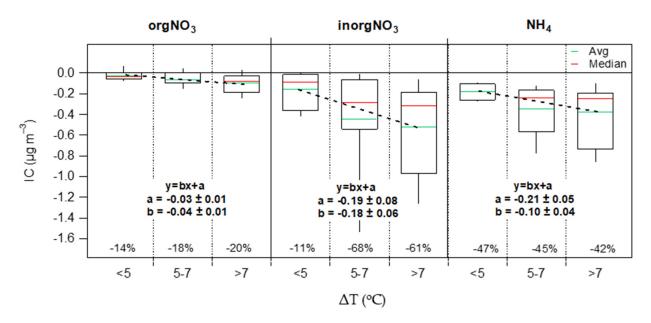


Figure 11. Indoor contribution of orgNO₃, inorgNO₃ and NH₄ as a function of indoor-outdoor temperature differences. The boxes and whiskers (**bottom** to **top**) represent the 10th and 25th percentiles, median (red line), and the 75th and 90th percentiles. Green lines represent average IC. Percentages correspond to the relative indoor contribution (RIC = IC/OC) for each bin.

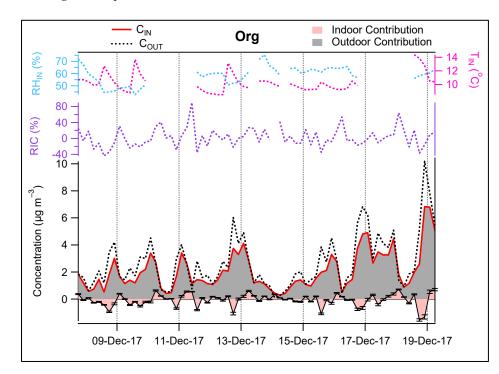


Figure 12. IC-OC contribution of Org. IC: $-0.03 \ \mu g \ m^{-3}$, RIC: 3%. Error bars represent 1σ .

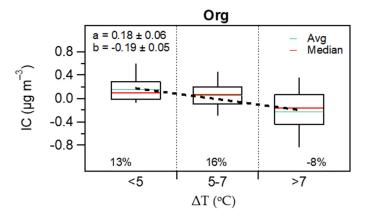


Figure 13. Indoor contributions of Organics as a function of indoor-outdoor temperature differences. The boxes and whiskers (bottom to top) represent the 10th and 25th percentiles, median (red line), and the 75th and 90th percentiles. Green lines represent average IC. Percentages correspond to the relative indoor contribution (RIC = IC/OC) for each bin.

3.4. Method Limitations

While the methodology described here performs well for the majority of the targeted pollutants, it is important to note that there are also some important limitations. A crucial step in the proposed methodology is to derive a penetration factor for each species of interest from the slope of a linear regression between its measured indoor and outdoor concentrations. The retrieval of the penetration factor can be biased by physicochemical processes operating indoors. Those include the reactivity with ambient oxidants (OH, O3 and NO3) for VOCs and physicochemical transformations for particles, if the rate of these processes compares with that of the air exchange. For example, it was found that oxidants such as O_3 , and reactive VOCs, such as isoprene and monoterpenes, cannot be reliably analyzed with the proposed methodology. A similar concern could be expressed for other VOCs exhibiting a significant reactivity with ambient oxidants, which unfortunately could not be investigated in this study since Time-of-Flight PTRMS measurements only provide information on the elemental composition of detected ions and no information on their chemical structure and reactivity. It is therefore recommended to carefully apply this methodology with online measurements from instruments that can provide molecular structure information (e.g., gas chromatography). For particles, this issue can be avoided when the chemical composition is monitored since the infiltration factor observed for non-reactive species such as SO₄ can be used for the analysis of all particulate species.

Furthermore, although this methodology can highlight trace gases lost indoors (negative IC due to adsorption processes), the strength of the sink will likely be underestimated due to the impact of the loss processes on the retrieval of the penetration factor, as discussed for ethanol.

Finally, this methodology is not considered applicable for compounds exhibiting low concentrations indoors, for instance, OVOCs potentially impacted by their formation from fast homogeneous or heterogeneous chemical reactions, or semi-volatile compounds impacted by a shift in gas/particle partitioning induced by temperature gradients from outdoors/indoors.

4. Conclusions

A simple apportionment methodology has been described in detail in this study and was applied to a vacant facility, typical of an energy-efficient building, to assess whether and how the unoccupied indoor environment affects the chemical composition of infiltrated ambient air. Concomitant indoor and outdoor time-resolved measurements of inorganic trace gases, VOCs and NR-PM₁ particles (including composition, i.e., SO₄, NH₄, inorgNO₃, orgNO₃ and Org) from online mass spectrometers were used to categorize the indoor/outdoor nature of the targeted pollutants. This methodology provided an apportionment of indoor and outdoor contributions to observe pollutant levels, identify whether the indoor environment acts as a source or a sink, and provide information on physicochemical processes operating indoors.

The results indicate that the indoor environment mainly acts as a source for VOCs, leading to a large enrichment in these species, by approximately a factor of 3 and 17 for OVOCs containing 1 and 2 atoms of oxygen, respectively. An increase in concentration was observed for nine C_xH_y , 18 C_xH_yO , 15 $C_xH_yO_2$ and two $C_xH_yO_z$ species, the indoor environment acting as a sink for only a few compounds, including ethanol and a few other OVOCs (corresponding to $C_5H_6O.H^+$, $C_4H_4O_3.H^+$ and $C_5H_8O_3.H^+$ ions in the PTRMS). Unsurprisingly, the indoor environment acts as a sink for O₃ due to its high reactivity with alkenes present in ambient air or adsorbed on surfaces. In contrast, the NR-PM₁ concentration is reduced by a factor of approximately 2 on average when ambient air is transferred indoors, with different reductions in SO₄ (factor of 1.2), NH₄ (1.9), NO₃ (2.3) and Org (1.3). This analysis also provided evidence that in addition to direct emissions, the unoccupied indoor environment impacts the chemical composition of infiltrated air through changes in the O₃-NO_x photostationary state and physicochemical processes associated with the volatilization of ammonium nitrate.

Supplementary Materials: The following supporting information can be downloaded at: https: //www.mdpi.com/article/10.3390/toxics10040161/s1, Glossary. Figure S1: Schematic of the measurement facility: (a) front side, (b) back side, (c) floor plan of the instrumentation room (left) and the experimental room (right). Figure S2 Average values of penetration factors during the intensive campaign as a function of particle size. Figure S3: Schematic of (a) AMS calibration unit and (b) AMS sampling configuration coupled with the CPC, SMPS and CO₂ analyzer. Figure S4: Schematic of the PTR-MS configuration coupled with the O₃ and NO₂ analyzers. Figure S5: Scatter plots of C_{IN} vs. C_{OUT} for NH₄, NO₃, and Org. Figure S6: IC-OC contributions (a) orgNO₃, (b) inorgNO₃ and (c) NH₄. Table S1: Compounds in the gas standard used for the transmission curve determination. Table S2: Calculations of VOC oxidation rates and comparison to the air exchange rate (a = 0.54 h^{-1}). Table S3: Exact protonated masses detected by PTRMS and examples of VOCs monitored at these *m/z*. Table S4: Descriptive statistics of compounds measured by PTRMS. Table S5: Descriptive statistics of measured particles (mass and number concentration). Table S6: Parameters from C_{IN} vs. C_{OUT} scatter plots, indoor (IC) and outdoor contributions, (OC), and emission rates (ER) for aerosols and trace gases.

Author Contributions: Conceptualization, S.D., V.R., E.S., J.B.; methodology, S.D., V.R.; validation, S.D., V.R., J.B.; formal analysis, E.S.; investigation, E.S., E.T., S.D., V.R.; resources, S.D., V.R.; writing—original draft preparation, E.S.; writing—review and editing, J.B., S.D., V.R.; visualization, E.S.; supervision, S.D., V.R.; project administration, V.R.; funding acquisition, S.D., V.R. All authors have read and agreed to the published version of the manuscript.

Funding: IMT Nord Europe acknowledges financial support from the CaPPA project, which is funded by the French National Research Agency (ANR) through the PIA (Programme d'Investissement d'Avenir) under contract ANR-11-LABX-0005-01, and the CLIMIBIO project, both financed by the Regional Council "Hauts-de-France" and the European Regional Development Fund (ERDF).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Acknowledgments: We thank Serge Russeil (IMT Nord Europe) who allowed us to use the energyefficient facility. ES's PhD grant was funded by Armines.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

References

- 1. Wallace, A. The Total Exposure Assessment Methodology (TEAM) Study: An Analysis of Exposures, Sources, and Risks Associated with Four Volatile Organic Chemicals. *J. Am. Coll. Toxicol.* **1989**, *8*, 883–895. [CrossRef]
- Abbatt, J.P.D.; Wang, C. The atmospheric chemistry of indoor environments. *Environ. Sci. Process. Impacts* 2020, 22, 25–48. [CrossRef]
- Klepeis, N.E.; Nelson, W.C.; Ott, W.R.; Robinson, J.P.; Tsang, A.M.; Switzer, P.; Behar, J.V.; Hern, S.C.; Engelmann, W.H. The National Human Activity Pattern Survey (NHAPS): A resource for assessing exposure to environmental pollutants. *J. Expo. Anal. Environ. Epidemiol.* 2001, *11*, 231–252. [CrossRef] [PubMed]
- 4. WHO. Burden of Disease from Household Air Pollution for 2016. WHO Regional Office for Europe. 2018. Available online: www.who.int (accessed on 1 December 2021).
- 5. Loomis, D.; Grosse, Y.; Lauby-Secretan, B.; El Ghissassi, F.; Bouvard, V.; Benbrahim-Tallaa, L.; Guha, N.; Baan, R.; Mattock, H.; Straif, K. The carcinogenicity of outdoor air pollution. *Lancet Oncol.* **2013**, *14*, 1262–1263. [CrossRef]
- 6. Kelly, F.J.; Fussell, J.C. Size, source and chemical composition as determinants of toxicity attributable to ambient particulate matter. *Atmos. Environ.* **2012**, *60*, 504–526. [CrossRef]
- Saleh, Y.; Antherieu, S.; Dusautoir, R.; Alleman, L.Y.; Sotty, J.; De Sousa, C.; Platel, A.; Perdrix, E.; Riffault, V.; Fronval, I.; et al. Exposure to Atmospheric Ultrafine Particles Induces Severe Lung Inflammatory Response and Tissue Remodeling in Mice. *Int. J. Environ. Res. Public Health* 2019, 16, 1210. [CrossRef] [PubMed]
- Leclercq, B.; Platel, A.; Antherieu, S.; Alleman, L.; Hardy, E.; Perdrix, E.; Grova, N.; Riffault, V.; Appenzeller, B.; Happillon, M.; et al. Genetic and epigenetic alterations in normal and sensitive COPD-diseased human bronchial epithelial cells repeatedly exposed to air pollution-derived PM 2.5. *Environ. Pollut.* 2017, 230, 163–177. [CrossRef] [PubMed]
- 9. Lanzinger, S.; Schneider, A.; Breitner, S.; Stafoggia, M.; Erzen, I.; Dostal, M.; Pastorkova, A.; Bastian, S.; Cyrys, J.; Zscheppang, A.; et al. Associations between ultrafine and fine particles and mortality in five central European cities—Results from the UFIREG study. *Environ. Int.* **2016**, *88*, 44–52. [CrossRef] [PubMed]
- Oberdörster, G.; Oberdörster, E.; Oberdörster, J. Nanotoxicology: An Emerging Discipline Evolving from Studies of Ultrafine Particles. Environ. Health Perspect. 2005, 113, 823–839. [CrossRef]
- 11. Pandis, S.N.; Skyllakou, K.; Florou, K.; Kostenidou, E.; Kaltsonoudis, C.; Hasa, E.; Presto, A.A. Urban particulate matter pollution: A tale of five cities. *Faraday Discuss.* **2016**, *189*, 277–290. [CrossRef]
- 12. Hu, K.; Guo, Y.; Hu, D.; Du, R.; Yang, X.; Zhong, J.; Fei, F.; Chen, F.; Chen, G.; Zhao, Q.; et al. Mortality burden attributable to PM1 in Zhejiang province, China. *Environ. Int.* 2018, *121*, 515–522. [CrossRef] [PubMed]
- Chen, G.; Li, S.; Zhang, Y.; Zhang, W.; Li, D.; Wei, X.; He, Y.; Bell, M.L.; Williams, G.; Marks, G.B.; et al. Effects of ambient PM 1 air pollution on daily emergency hospital visits in China: An epidemiological study. *Lancet Planet. Health* 2017, 1, e221–e229. [CrossRef]
- Lin, H.; Tao, J.; Du, Y.; Liu, T.; Qian, Z.; Tian, L.; Di, Q.; Rutherford, S.; Guo, L.; Zeng, W.; et al. Particle size and chemical constituents of ambient particulate pollution associated with cardiovascular mortality in Guangzhou, China. *Environ. Pollut.* 2016, 208, 758–766. [CrossRef] [PubMed]
- 15. Yu, O.; Sheppard, L.; Lumley, T.; Koenig, J.Q.; Shapiro, G.G. Effects of ambient air pollution on symptoms of asthma in Seattle-area children enrolled in the CAMP study. *Environ. Health Perspect.* **2000**, *108*, 1209–1214. [CrossRef] [PubMed]
- Zwozdziak, A.; Sówka, I.; Willak-Janc, E.; Zwozdziak, J.; Kwiecińska, K.; Balińska-Miśkiewicz, W. Influence of PM1 and PM2.5 on lung function parameters in healthy schoolchildren—A panel study. *Environ. Sci. Pollut. Res.* 2016, 23, 23892–23901. [CrossRef] [PubMed]
- 17. Mar, T.F.; Larson, T.V.; Stier, R.A.; Claiborn, C.; Koenig, J.Q. An Analysis of the Association Between Respiratory Symptoms in Subjects with Asthma and Daily Air Pollution in Spokane, Washington. *Inhal. Toxicol.* **2004**, *16*, 809–815. [CrossRef] [PubMed]
- Madureira, J.; Paciência, I.; Rufo, J.C.; Severo, M.; Ramos, E.; Barros, H.; Fernandes, E.D.O. Source apportionment of CO₂, PM10 and VOCs levels and health risk assessment in naturally ventilated primary schools in Porto, Portugal. *Build. Environ.* 2016, 96, 198–205. [CrossRef]
- 19. Bernstein, J.A.; Alexis, N.; Bacchus, H.; Bernstein, I.L.; Fritz, P.; Horner, E.; Li, N.; Mason, S.; Nel, A.; Oullette, J.; et al. The health effects of nonindustrial indoor air pollution. *J. Allergy Clin. Immunol.* **2008**, 121, 585–591. [CrossRef]
- 20. Wolkoff, P.; Wilkins, C.K.; Clausen, P.A.; Nielsen, G.D. Organic compounds in office environments-sensory irritation, odor, measurements and the role of reactive chemistry. *Indoor Air* 2005, *16*, 7–19. [CrossRef]
- 21. Zhang, J.F.; Smith, K.R. Indoor air pollution: A global health concern. Br. Med. Bull. 2003, 68, 209–225. [CrossRef]
- 22. Alford, K.L.; Kumar, N. Pulmonary Health Effects of Indoor Volatile Organic Compounds—A Meta-Analysis. *Int. J. Environ. Res. Public Health* **2021**, *18*, 1578. [CrossRef] [PubMed]
- 23. European Enviromental Agency. Air Quality in Europe. NO 28/2016. Available online: https://www.eea.europa.eu/publications/air-quality-in-europe-2016 (accessed on 1 December 2021).
- Service Public de la Diffusion du Droit. Décret n 2015-1000 du 17 Août 2015 Relatif aux Modalités de Surveillance de la Qualité de l'air Intérieur dans Certains Établissements Recevant du Public; 2015. Available online: https://www.legifrance.gouv.fr/loda/id/ JORFTEXT000031052712/2015-08-20#JORFTEXT000031052712 (accessed on 25 March 2022).

- Agence Nationale de Sécurité Sanitaire de L'alimentation. Valeurs Guides de Qualité d'Air Intérieur (VGAI). Available online: https://www.anses.fr/fr/content/valeurs-guides-de-qualit%C3%A9-d%E2%80%99air-int%C3%A9rieur-vgai (accessed on 25 March 2022).
- Avery, A.M.; Waring, M.S.; Decarlo, P.F. Seasonal variation in aerosol composition and concentration upon transport from the outdoor to indoor environment. *Environ. Sci. Process. Impacts* 2019, 21, 528–547. [CrossRef] [PubMed]
- 27. Johnson, A.M.; Waring, M.S.; DeCarlo, P.F. Real-time transformation of outdoor aerosol components upon transport indoors measured with aerosol mass spectrometry. *Indoor Air* 2017, 27, 230–240. [CrossRef]
- Bari, M.A.; Kindzierski, W.B.; Wheeler, A.J.; Héroux, M.È.; Wallace, L.A. Source apportionment of indoor and outdoor volatile organic compounds at homes in Edmonton, Canada. *Build. Environ.* 2015, 90, 114–124. [CrossRef]
- Bari, A.; Kindzierski, W.B.; Wallace, L.A.; Wheeler, A.J.; MacNeill, M.; Héroux, M.-È. Indoor and Outdoor Levels and Sources of Submicron Particles (PM1) at Homes in Edmonton, Canada. *Environ. Sci. Technol.* 2015, 49, 6419–6429. [CrossRef] [PubMed]
- Liu, S.; Li, R.; Wild, R.J.; Warneke, C.; de Gouw, J.; Brown, S.S.; Miller, S.L.; Luongo, J.C.; Jimenez, J.L.; Ziemann, P.J. Contribution of human-related sources to indoor volatile organic compounds in a university classroom. *Indoor Air* 2016, 26, 925–938. [CrossRef] [PubMed]
- Omelekhina, Y.; Eriksson, A.; Canonaco, F.; Prevot, A.S.H.; Nilsson, P.; Isaxon, C.; Pagels, J.; Wierzbicka, A. Cooking and electronic cigarettes leading to large differences between indoor and outdoor particle composition and concentration measured by aerosol mass spectrometry. *Environ. Sci. Process. Impacts* 2020, *22*, 1382–1396. [CrossRef]
- Plaisance, H.; Mocho, P.; Sauvat, N.; Vignau-Laulhere, J.; Raulin, K.; Desauziers, V. Using the chemical mass balance model to estimate VOC source contributions in newly built timber frame houses: A case study. *Environ. Sci. Pollut. Res.* 2017, 24, 24156–24166. [CrossRef]
- 33. Kopperud, R.J.; Ferro, A.R.; Hildemann, L.M. Outdoor Versus Indoor Contributions to Indoor Particulate Matter (PM) Determined by Mass Balance Methods. *J. Air Waste Manag. Assoc.* **2004**, *54*, 1188–1196. [CrossRef]
- 34. Carslaw, N. A new detailed chemical model for indoor air pollution. Atmos. Environ. 2007, 41, 1164–1179. [CrossRef]
- Shiraiwa, M.; Carslaw, N.; Tobias, D.J.; Waring, M.S.; Rim, D.; Morrison, G.; Lakey, P.S.J.; Kruza, M.; Von Domaros, M.; Cummings, B.E.; et al. Modelling consortium for chemistry of indoor environments (MOCCIE): Integrating chemical processes from molecular to room scales. *Environ. Sci. Process. Impacts* 2019, 21, 1240–1254. [CrossRef] [PubMed]
- 36. Chavent, M.; Guégan, H.; Kuentz, V.; Patouille, B.; Saracco, J. PCA- and PMF-based methodology for air pollution sources identification and apportionment. *Environmetrics* 2009, 20, 928–942. [CrossRef]
- 37. RT2012: Un saut énergétique pour les bâtiments neufs. Available online: https://www.ecologie.gouv.fr/reglementation-thermique-rt2012#:~{}:text=La%20RT2012%20permet%20de%20prendre,confondre%20avec%20l\T1\textquoteright%C3%A9 nergie%20finale (accessed on 25 March 2022).
- Guerrero, F. ASTM E 741—00: Standard Test Method for Determining Air Change in a Single Zone by Means of a Tracer Gas Dilution. 200. Available online: https://www.academia.edu/40099720/Standard_Test_Method_for_Determining_Air_Change_ in_a_Single_Zone_by_Means_of_a_Tracer_Gas_Dilution_1 (accessed on 25 March 2022).
- 39. Stratigou, E.; Dusanter, S.; Brito, J.; Riffault, V. Investigation of PM10, PM2.5, PM1 in an unoccupied airflow-controlled room: How reliable to neglect resuspension and assume unreactive particles? *Build. Environ.* **2020**, *186*, 107357. [CrossRef]
- DeCarlo, P.F.; Kimmel, J.R.; Trimborn, A.; Northway, M.J.; Jayne, J.T.; Aiken, A.C.; Gonin, M.; Fuhrer, K.; Horvath, T.; Docherty, K.S.; et al.Field-Deployable, High-Resolution, Time-of-Flight Aerosol Mass Spectrometer. *Anal. Chem.* 2006, 78, 8281–8289. [CrossRef]
- Canagaratna, M.; Jayne, J.; Jimenez, J.; Allan, J.; Alfarra, M.; Zhang, Q.; Onasch, T.; Drewnick, F.; Coe, H.; Middlebrook, A.; et al. Chemical and microphysical characterization of ambient aerosols with the aerodyne aerosol mass spectrometer. *Mass Spectrom. Rev.* 2007, 26, 185–222. [CrossRef]
- Crenn, V.; Fronval, I.; Petitprez, D.; Riffault, V. Fine particles sampled at an urban background site and an industrialized coastal site in Northern France—Part 1: Seasonal variations and chemical characterization. *Sci. Total Environ.* 2017, 578, 203–218. [CrossRef]
- Lindinger, W.; Hansel, A.; Jordan, A. On-line monitoring of volatile organic compounds at pptv levels by means of proton-transferreaction mass spectrometry (PTR-MS) medical applications, food control and environmental research. *Int. J. Mass Spectrom. Ion Process.* 1998, 173, 191–241. [CrossRef]
- 44. de Gouw, J.; Warneke, C.; Karl, T.; Eerdekens, G.; van der Veen, C.; Fall, R. Sensitivity and specificity of atmospheric trace gas detection by proton-transfer-reaction mass spectrometry. *Int. J. Mass Spectrom.* **2003**, 223–224, 365–382. [CrossRef]
- Farmer, D.K.; Matsunaga, A.; Docherty, K.S.; Surratt, J.D.; Seinfeld, J.H.; Ziemann, P.J.; Jimenez, J.L. Response of an aerosol mass spectrometer to organonitrates and organosulfates and implications for atmospheric chemistry. *Proc. Natl. Acad. Sci. USA* 2010, 107, 6670–6675. [CrossRef]
- Kiendler-Scharr, A.; Mensah, A.; Friese, E.; Topping, D.; Nemitz, E.; Prevot, A.S.H.; Äijälä, M.; Allan, J.; Canonaco, F.; Canagaratna, M.; et al. Ubiquity of organic nitrates from nighttime chemistry in the European submicron aerosol. *Geophys. Res. Lett.* 2016, 43, 7735–7744. [CrossRef]
- Polidori, A.; Turpin, B.; Meng, Q.Y.; Lee, J.H.; Weisel, C.; Morandi, M.; Colome, S.; Stock, T.; Winer, A.; Zhang, J.; et al. Fine organic particulate matter dominates indoor-generated PM2.5 in RIOPA homes. *J. Expo. Sci. Environ. Epidemiol.* 2006, 16, 321–331. [CrossRef] [PubMed]

- Arata, C.; Zarzana, K.J.; Misztal, P.K.; Liu, Y.; Brown, S.S.; Nazaroff, W.W.; Goldstein, A.H. Measurement of NO₃ and N₂O₅ in a Residential Kitchen. *Environ. Sci. Technol. Lett.* 2018, 5, 595–599. [CrossRef]
- Carslaw, N.; Shaw, D. Secondary product creation potential (SPCP): A metric for assessing the potential impact of indoor air pollution on human health. *Environ. Sci. Process. Impacts* 2019, 21, 1313–1322. [CrossRef] [PubMed]
- 50. Waring, M.S.; Wells, J.R. Volatile organic compound conversion by ozone, hydroxyl radicals, and nitrate radicals in residential indoor air: Magnitudes and impacts of oxidant sources. *Atmos. Environ.* **2015**, *106*, 382–391. [CrossRef]
- 51. Alvarez, E.G.; Amedro, D.; Afif, C.; Gligorovski, S.; Schoemaecker, C.; Fittschen, C.; Doussin, J.-F.; Wortham, H. Unexpectedly high indoor hydroxyl radical concentrations associated with nitrous acid. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 13294–13299. [CrossRef]
- 52. Hodgson, A.T.; Rudd, A.F.; Beal, D.; Chandra, S. Volatile Organic Compound Concentrations and Emission Rates in New Manufactured and Site-Built Houses. *Indoor Air* 2001, *10*, 178–192. [CrossRef]
- 53. Holøs, S.B.; Yang, A.; Lind, M.; Thunshelle, K.; Schild, P.; Mysen, M. VOC emission rates in newly built and renovated buildings, and the influence of ventilation—A review and meta-analysis. *Int. J. Vent.* **2018**, *18*, 153–166. [CrossRef]
- 54. Villanueva, F.; Tapia, A.; Lara, S.; Amo-Salas, M. Indoor and outdoor air concentrations of volatile organic compounds and NO2 in schools of urban, industrial and rural areas in Central-Southern Spain. *Sci. Total Environ.* **2018**, *622–623*, 222–235. [CrossRef]
- 55. Fortenberry, C.; Walker, M.; Dang, A.; Loka, A.; Date, G.; De Carvalho, K.C.; Morrison, G.; Williams, B. Analysis of indoor particles and gases and their evolution with natural ventilation. *Indoor Air* **2019**, *29*, 761–779. [CrossRef]
- 56. Liu, S.; Thompson, S.L.; Stark, H.; Ziemann, P.J.; Jimenez, J.L. Gas-Phase Carboxylic Acids in a University Classroom: Abundance, Variability, and Sources. *Environ. Sci. Technol.* **2017**, *51*, 5454–5463. [CrossRef]
- 57. Patkó, C.; Patko, I.; Pásztory, Z. Indoor Air Quality Testing in Low-Energy Wooden Houses: Measurement of Formaldehyde and VOC-s. *Acta Polytech. Hung.* **2013**, *10*, 105–116.
- 58. Derbez, M.; Berthineau, B.; Cochet, V.; Lethrosne, M.; Pignon, C.; Riberon, J.; Kirchner, S. Indoor air quality and comfort in seven newly built, energy-efficient houses in France. *Build. Environ.* 2014, 72, 173–187. [CrossRef]
- 59. Solomon, S.; Schade, G.; Kuttippurath, J.; Ladstätter-Weissenmayer, A.; Burrows, J.P. VOC Concentrations in an Indoor Workplace Environment of a University Building. *Indoor Built Environ.* **2008**, *17*, 260–268. [CrossRef]
- 60. Allen, J.G.; Macnaughton, P.; Satish, U.; Santanam, S.; Vallarino, J.; Spengler, J.D. Associations of Cognitive Function Scores with Carbon Dioxide, Ventilation, and Volatile Organic Compound Exposures in Office Workers: A Controlled Exposure Study of Green and Conventional Office Environments. *Environ. Health Perspect.* **2016**, *124*, 805–812. [CrossRef]
- 61. Shin, S.-H.; Jo, W.-K. Longitudinal variations in indoor VOC concentrations after moving into new apartments and indoor source characterization. *Environ. Sci. Pollut. Res.* 2013, 20, 3696–3707. [CrossRef]
- 62. Schoemaecker, C.; Verriele, M.; Hanoun, B.; Petitprez, D.; Leclerc, N.; Dusanter, S.; Le Calve, S.; Millet, M.; Bernhardt, P.; Mendez, M.; et al. *Characterization of the IAQ in Low Energy Public Buildings in France through a Dual Experiment and Modeling Approach*; ICPEES: Hiltigham, France, 2014.
- 63. Geiss, O.; Giannopoulos, G.; Tirendi, S.; Barrero-Moreno, J.; Larsen, B.R.; Kotzias, D. The AIRMEX study-VOC measurements in public buildings and schools/kindergartens in eleven European cities: Statistical analysis of the data. *Atmos. Environ.* 2011, 45, 3676–3684. [CrossRef]
- 64. Csobod, E.; Annesi-Maesano, I.; Carrer, P.; Kephalopoulos, S.; Madureira, J.; Rudnai, P.; De Oliveira, F.E.; Barrero, J.; Beregszászi, T.; Hyvärinen, A.; et al. SINPHONIE: Schools Indoor Pollution & Health Observatory Network in Europe; EUR—Scientific and Technical Research Reports; Publications Office of the European Union: Luxembourg, 2014.
- 65. Schlink, U.; Rehwagen, M.; Damm, M.; Richter, M.; Borte, M.; Herbarth, O. Seasonal cycle of indoor-VOCs: Comparison of apartments and cities. *Atmos. Environ.* 2004, *38*, 1181–1190. [CrossRef]
- 66. Tang, X.; Misztal, P.K.; Nazaroff, W.W.; Goldstein, A.H. Volatile Organic Compound Emissions from Humans Indoors. *Environ. Sci. Technol.* **2016**, *50*, 12686–12694. [CrossRef]
- 67. Duncan, S.M.; Tomaz, S.; Morrison, G.; Webb, M.; Atkin, J.M.; Surratt, J.D.; Turpin, B.J. Dynamics of Residential Water-Soluble Organic Gases: Insights into Sources and Sinks. *Environ. Sci. Technol.* **2019**, *53*, 1812–1821. [CrossRef]
- 68. Dodson, R.E.; Levy, J.I.; Spengler, J.D.; Shine, J.P.; Bennett, D.H. Influence of basements, garages, and common hallways on indoor residential volatile organic compound concentrations. *Atmos. Environ.* **2008**, *42*, 1569–1581. [CrossRef]
- 69. Jia, C.; Batterman, S.; Godwin, C.; Charles, S.M.; Chin, J.-Y. Sources and migration of volatile organic compounds in mixed-use buildings. *Indoor Air* **2010**, *20*, 357–369. [CrossRef] [PubMed]
- 70. Smedemark, S.H.; Ryhl-Svendsen, M.; Schieweck, A. Quantification of formic acid and acetic acid emissions from heritage collections under indoor room conditions. Part I: Laboratory and field measurements. *Heritage Sci.* 2020, *8*, 58. [CrossRef]
- 71. Zhang, J.; Wilson, E.W.; Lioy, P.J. Sources of organic acids in indoor air: A field study. *J. Expo. Anal. Environ. Epidemiol.* **1994**, *4*, 25–47. [PubMed]
- Talbot, N.; Kubelová, L.; Makeš, O.; Ondráček, J.; Cusack, M.; Schwarz, J.; Vodička, P.; Zíková, N.; Ždímal, V. Transformations of Aerosol Particles from an Outdoor to Indoor Environment. *Aerosol Air Qual. Res.* 2017, 17, 653–665. [CrossRef]
- 73. Duncan, S.M.; Sexton, K.G.; Turpin, B.J. Oxygenated VOCs, aqueous chemistry, and potential impacts on residential indoor air composition. *Indoor Air* 2017, 28, 198–212. [CrossRef] [PubMed]

- 74. Liu, Y.; Misztal, P.K.; Xiong, J.; Tian, Y.; Arata, C.; Weber, R.J.; Nazaroff, W.W.; Goldstein, A.H. Characterizing sources and emissions of volatile organic compounds in a northern California residence using space- and time-resolved measurements. *Indoor Air* **2019**, *29*, 630–644. [CrossRef] [PubMed]
- 75. Fischer, A.; Langer, S.; Ljungström, E. Chemistry and indoor air quality in a multi-storey wooden passive (low energy) building: Formation of peroxyacetyl nitrate. *Indoor Built Environ*. **2013**, *23*, 485–496. [CrossRef]
- 76. Offermann, F.J.; Maddalena, R.L.; Offermann, F.J.; Singer, B.C.; Willem, H. The impact of ventilation rate on the emission rates of volatile organic compounds in residences. In Proceedings of the Healthy Buildings 2012—10th International Conference, Brisbane, Australia, 8–12 July 2012.
- 77. Finlayson-Pitts, B.J.; Pitts, J.N., Jr. *Chemistry of the Upper and Lower Atmosphere Theory, Experiments, and Applications*; Elsevier: Amsterdam, The Netherlands, 2000.