

Review

# Synthesis of Magnetic Metal-Organic Frame Material and Its Application in Food Sample Preparation

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Received: 30 September 2020; Accepted: 3 November 2020; Published: 6 November 2020



**Abstract:** A variety of contaminants in food is an important aspect affecting food safety. Due to the presence of its trace amounts and the complexity of food matrix, it is very difficult to effectively separate and accurately detect them. The magnetic metal-organic framework (MMOF) composites with different structures and functions provide a new choice for the purification of food matrix and enrichment of trace targets, thus providing a new direction for the development of new technologies in food safety detection with high sensitivity and efficiency. The MOF materials composed of inorganic subunits and organic ligands have the advantages of regular pore structure, large specific surface area and good stability, which have been thoroughly studied in the pretreatment of complex food samples. MMOF materials combined different MOF materials with various magnetic nanoparticles, adding magnetic characteristics to the advantages of MOF materials, which are in terms of material selectivity, biocompatibility, easy operation and repeatability. Combined with solid phase extraction (SPE) technique, MMOF materials have been widely used in the food pretreatment. This article introduced the new preparation strategies of different MMOF materials, systematically summarizes their applications as SPE adsorbents in the pretreatment of food contaminants and analyzes and prospects their future application prospects and development directions.

**Keywords:** magnetic metal-organic frame material; food contaminants; sample preparation; synthesis and application

## 1. Introduction

Food is the material basis for human survival, and food safety issues are a great concern of the international community. The existence of food contaminants such as residues of pesticide and veterinary drugs, toxins, illegal additives, heavy metal ions etc., is an important aspect causing the problem of food safety and has attracted wide attention from researchers [1–3]. Therefore, various strategies for the detection of contaminants in food have also been developed [4,5]. So far, the analysis strategies based on high-precision analytical instruments are still the main detection method of food pollutants because of the merits of high accuracy and sensitivity [6–8]. At the same time, some rapid detection methods and devices have been developed to realize convenient, rapid, low-cost and real-time analysis of food contaminants [9–11]. Because the food matrices are complex and contaminants are usually trace present, one of the key problems to limit the detection of food contaminants is sample pretreatment [12]. It is necessary not only to isolate

analytes from the sample as thoroughly as possible, but also to enrich them before detection and quantification, while isolated analytes from the primary matrix remove the impact of interfering substances. Besides, foods belong to the fast-moving consumer product and have high requirement on detection speed or throughput. Therefore, while perfecting the detection technology, it is extremely urgent to develop accurate efficient and fast sample pretreatment techniques for food matrices [13,14]. An ideal food sample pretreatment technology not only requires high sensitivity, selectivity and reliability, but also has fast detection speed and convenient operation. This is a very critical step in the process of target analysis and one of the important research directions in the field of analytical chemistry [15].

After several years of development, the technology of solid-phase extraction (SPE) with simple operation, short pretreatment time, low consumption of organic solvents and high recovery of analyte has gradually become the most commonly used pretreatment technology for food complex matrices [16–18]. Varieties of SPE adsorbing materials can not only be customized according to the target substance, but also can match with different analysis equipment, which meets the detection requirements of high selectivity, high specificity and high throughput [19,20]. And many different nanomaterials have been studied and reported in the preparation of SPE adsorbent [21–23]. Among them, magnetic SPE, as a new type of pretreatment process, uses functionalized magnetic materials as adsorbents, and under the action of an external magnetic field, these magnetic adsorbents can be efficiently separated from the matrix samples [24–26]. The magnetic SPE (MSPE) procedure has a series of advantages such as simple operation, low solvent consumption, and high enrichment efficiency, which was widely used in the fields of food analysis, biomedicine and environmental monitoring [27–29]. Therefore, the synthesis and performance research of various MSPE materials has become a research hotspot in recent years [30,31].

Metal-organic framework (MOF) materials are a kind of crystal materials with three-dimensional network structure formed by self-assembly of inorganic subunits and organic ligands through coordination bonds. The inorganic subunits of MOFs usually include metal clusters, metal ions, or central chains [32–34]. The diversity and arrangement of metal elements and organic ligands can form different frameworks and pore structures, making the MOFs have different adsorption, optical and electromagnetic properties [35–38]. These characteristics have made MOFs materials widely concerned and used in the fields of sample pretreatment, chemical catalysis, drug delivery [39–42]. One of the research hotspots on MOFs is combining with various functional nano- and micro-particles to prepare highly porous nanocomposites [43–45]. These materials have the merits of nanomaterials while retaining the special properties and structure of MOF, thus expanding the application range of MOF materials. Among them, magnetic MOF (MMOFs) materials introduce magnetic nanoparticles (MNPs) to the surface or pores of MOFs, which not only maintains the characteristics of large specific surface area, strong adsorption capacity, chemical selectivity of MOFs, but also gives the material magnetic properties [46,47]. This kind of composite overcomes the shortcomings of MOFs which are easy to collapse under acid or alkaline conditions, thus increases the machinability and operability of MOFs, which has expanded the application range of MOFs in actual sample pretreatment [48,49]. As a new type of functional material, MMOFs materials have been widely used as SPE sorbent for the pretreatment of complex matrix, especially for the enrichment and separation of trace contaminants in food samples [50–52].

Therefore, this article summarizes the different preparation strategies of MMOFs currently used as adsorbents for food sample pretreatment, reviews the application of MMOFs in the detection and control of food contaminants, and looks forward to the future development direction and application prospects of MMOFs in the field of food analysis.

## 2. Synthesis Strategies of MMOFs Composite

MMOFs composite have become a burgeoning star in separation science [53,54]. The synthesis of MMOFs materials usually used MOFs as the precursors, and introduced or combined with MNPs (alloys, ferrites, metal oxides, etc.) by physical or chemical means. This not only maintain the

advantages of MOFs (porous, highly modifiable and high specific surface area), but also endow them with magnetic separation characteristics. Furthermore, regular pore structure of MOFs is beneficial to mass transfer and diffusion and offer confinement effects to avoid aggregation of analytes. These properties make MMOFs very suitable for the pretreatment and purification of food matrices [55–58]. At present, various MMOFs composites with different properties have been synthesized using different strategies for food analysis [59–61] (Table 1).

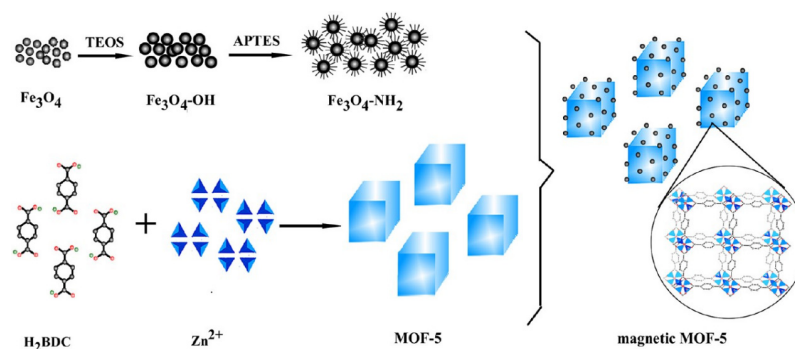
**Table 1.** Comparison of different strategies for MMOFs preparation.

Methods	Advantages	Disadvantages
Hybrid preparation method	Simple; suitable for most MMOFs	Easy to fall off
In-situ growth of MOFs on MNPs	Simple; synthesis at room temperature	Direct nucleation and growth of MOFs in solution
In-situ growth of MNPs on MOFs	More complete MOF structure; good adsorption property; remarkable magnetic response	Complex preparation process
Self-sacrificial template method	Core-shell structures; easy to operate; synthesis under mild conditions	Need a suitable precursor
Emulsion template method	Avoid embedding MNPs in MOFs tunnels; obtaining macroporous polymers	Require large amount of inorganic solvent; complex preparation process
LbL self-assembly method	Core-shell structure; precisely control the thickness and properties of MMOFs; synthesis at room temperature	Long preparation time Lack of self-assembled MOFs ligands
Dry gel conversion method	Avoid MNPs embedding in MOFs tunnels; reduce the loss of organic solvents	Not easily controlled uniformity of MOF growth
Mechanical grinding method	Synthesis at room temperature	Limited types of MOFs ligands; uneven MMOFs performance

### 2.1. Hybrid Preparation Method

Hybrid preparation method is a process of directly mixing pre-prepared MNPs with MOFs, obtaining MMOFs composites through the ultrasonic or high-temperature polymerization procedures [62–64]. The MNPs can be modified on the surface of MOFs by electrostatic interaction or chemical bonding. The structure and performance of the prepared MMOFs can also be controlled by adjusting the interactions between MNPs and MOFs. The MNPs on the MOFs surface make the prepared MMOFs usually have better magnetic response [65,66]. Jiang et al., mixed cysteine functionalized  $\text{Fe}_3\text{O}_4$  NPs with MIL-101(Fe) together under ultrasonication and obtained the  $\text{Fe}_3\text{O}_4/\text{MIL-101(Fe)}$  composite through electrostatic interaction [67]. It is worth noting that the  $\text{Fe}_3\text{O}_4/\text{MIL-101(Fe)}$  composite is more susceptible to magnetic fields and easier to isolate from aqueous solution, and exhibits much higher catalytic activity than the single component ( $\text{Fe}_3\text{O}_4$  NPs and MIL-101) for the synthesis of 2,3-diaminophenazine due to the synergistic peroxidase-like activity between the multiple active centers. A facile and efficient strategy about the synthesis of hybrid magnetic MOF-5 was illustrated in Figure 1 [68]. The pre-prepared amino-functionalized  $\text{Fe}_3\text{O}_4$  NPs were introduced on the surface of MOF-5 via chemical bonding to improve the stability and structure uniformity of the hybrid microcrystals, which allows for facile withdrawal of the porous materials by magnetic decantation. Peña-Méndez et al., prepared one MMOFs  $\text{Fe}_3\text{O}_4@(\text{Fe}-(\text{benzene-1,3,5-tricarboxylic acid}))$  using the hybrid preparation method [69]. The obtained MMOFs was demonstrated to have high specific surface area ( $803.62 \text{ m}^2/\text{g}$ ), good stability, strong magnetic response characteristics, which was further applied as a solid adsorbent of MSPE for food analysis.

Strictly speaking, this hybrid preparation method can prepare any required MMOFs with strong applicability. However, since the combination of MNPs and MOFs is not strong enough, causing easily separate from each other. Moreover, it is usually necessary to pre-prepare excessive amounts of MOFs during the preparation process, resulting in higher production costs [70].



**Figure 1.** Schematic of the fabrication process of the hybrid magnetic MOF-5 using amino-functionalized  $\text{Fe}_3\text{O}_4$  NPs [68]. Copyright Analytical Chemistry, 2013.

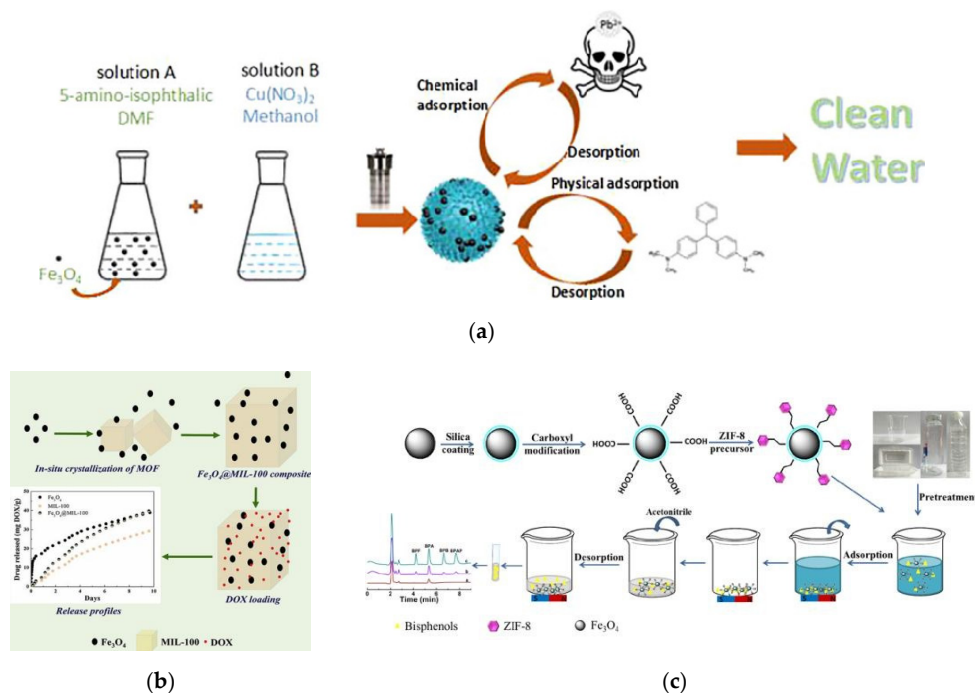
## 2.2. In-Situ Growth Method

### 2.2.1. In-Situ Growth of MOFs on MNPs

In-situ growth of MOFs on MNPs, as its name suggests, the MOFs are grown in-situ on the surface of the MNPs added to the MOF precursor solution under ultrasonic or hydrothermal conditions. This method guarantees the structural integrity of the MMOFs to the greatest extent, so the obtained MMOFs materials have good adsorption performance [71,72]. Figure 2a has shown the synthesis process of a Cu-MOFs/ $\text{Fe}_3\text{O}_4$  composite by in-situ growth of Cu-MOFs with doping  $\text{Fe}_3\text{O}_4$  NPs. The adsorption capacities of the Cu-MOFs/ $\text{Fe}_3\text{O}_4$  composite were found to be 113.67 mg/g for malachite green and 219.00 mg/g for  $\text{Pb}^{2+}$ , respectively, which are significantly higher than other reported materials [73]. Different amounts of  $\text{Fe}_3\text{O}_4$  NPs used would obviously affect the performance of in-situ synthesized MMOFs (Figure 2b) [74]. The obtained  $\text{Fe}_3\text{O}_4$ @MIL-100 composites were observed to have higher capacity in the application of loading the drug doxorubicin with the porous carriers. The highest loading capacity (about 19% mass) was observed for MIL-100 composites containing about 16% mass of  $\text{Fe}_3\text{O}_4$  particles.

The in-situ growth of MOFs on MNPs is relatively simple, which can be completed in one step. This preparation process can be carried out at room temperature, which eliminates the need for complex carbonization processes at high temperatures, greatly reducing the preparation difficulty and shortening the preparation time, so it is very suitable for industrial production [76,77]. Figure 2c illustrated the in-situ growth of zeolitic imidazolate framework (ZIF)-8 on the surface of  $\text{Fe}_3\text{O}_4$ @ $\text{SiO}_2$  microspheres at room temperature and its application. This  $\text{Fe}_3\text{O}_4$ @ $\text{SiO}_2$ /ZIF-8 material with large surface area and high super paramagnetism has great adsorption capacity to the analyte bisphenol A (a typical toxic chemical substance) [75].

However, in in-situ growth of MOF on MNPs, it is difficult to prevent the direct nucleation and growth of MOFs in solution, and part of the MNPs is embedded in the pores of the MOFs, resulting in a reduction in specific surface area [78,79]. Additionally, a large number of unreacted magnetic nanomaterials existing in the solution after reaction has made the subsequent separation steps cumbersome and require a large number of organic solvents [80].

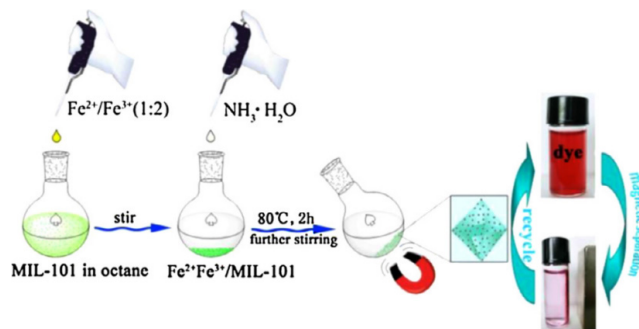


**Figure 2.** (a) Synthesis process of a Cu-MOFs/Fe<sub>3</sub>O<sub>4</sub> composite by in-situ growth of Cu-MOFs doping with Fe<sub>3</sub>O<sub>4</sub> NPs [73]. Copyright Colloids & Surfaces A, 2017. (b) In-situ synthesis of Fe<sub>3</sub>O<sub>4</sub>@MIL-100 doping with different amounts of Fe<sub>3</sub>O<sub>4</sub> NPs [74]. Copyright Microporous & Mesoporous Materials, 2018. (c) In-situ growth of ZIF-8 on the surface of Fe<sub>3</sub>O<sub>4</sub>@SiO<sub>2</sub> microspheres and its application [75]. Copyright New Journal of Chemistry, 2020.

### 2.2.2. In-Situ Growth of MNPs on MOFs

In this method, the prepared MOFs are dispersed in a mixture containing reagents for preparing MNPs. The growth of MNPs is carried out at the presence of MOFs, and MNPs is embedded on the surface of MOFs. The free MNPs can be separated by centrifugation, and the MMOFs and MOFs are separated under an external magnetic field.

Figure 3 shows the fabrication process of the magnetic hybrid Fe<sub>3</sub>O<sub>4</sub>/MIL-101 composite via in-situ growth of MNPs approach [81]. The solutions of MIL-101 and Fe<sup>3+</sup>/Fe<sup>2+</sup> (2/1) were first mixed in n-octane, and then synthesized Fe<sub>3</sub>O<sub>4</sub> NPs on MIL-101 crystal surface using ammonia solution as a precipitant. Characterizations of the synthesized Fe<sub>3</sub>O<sub>4</sub>/MIL-101 composite showed that the Fe<sub>3</sub>O<sub>4</sub> NPs were uniformly coated on the outer-surface of MIL-101. This composite has a larger surface area, pore volume and higher positive charge than MIL-101, making it an excellent candidate for the adsorption and removal of anionic dyes from aqueous solutions.



**Figure 3.** Schematic of the fabrication process of Fe<sub>3</sub>O<sub>4</sub>/MIL-101 [81]. Copyright Journal of the Taiwan Institute of Chemical Engineers, 2015.



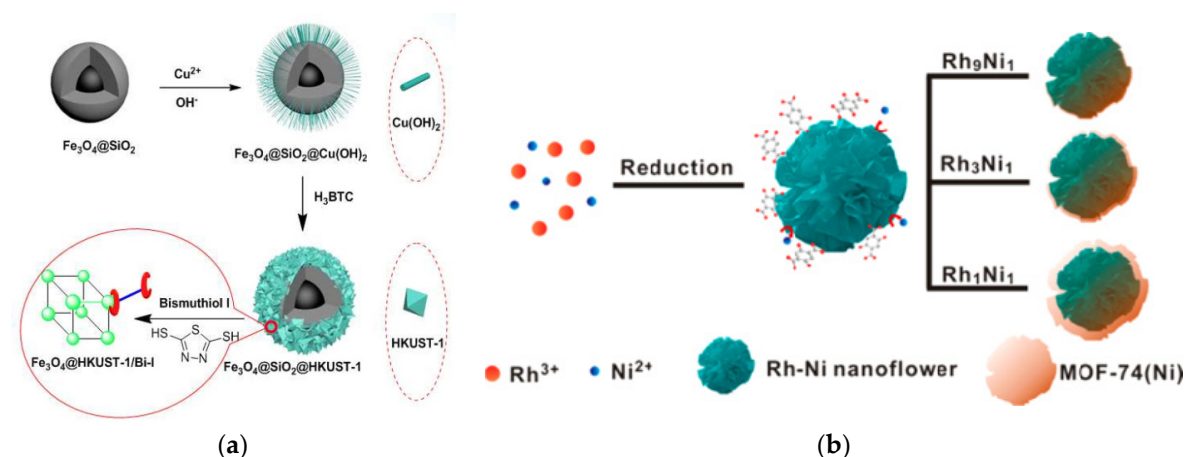
The MMOFs prepared by in-situ growth of MNPs method have very more complete MOF structure, good adsorption property and magnetic response. However, there are some technical defects. A large number of free MNPs in the solution makes the subsequent separation steps complicated compared to other methods [82,83]. Before adding MNPs, the stability of MOFs should be strictly controlled to avoid collapse of MOFs skeleton. These problems need to be solved in the follow-up research.

### 2.3. Template-Directed Method

Template-directed method is an important strategy for preparing nanocomposite materials, and the most extensive method in the research of nanomaterials in recent years [84,85]. This method uses the materials with stable structure and controllable morphology as template and deposits the target material into the pores or surface of the template by physical or chemical methods. After removing the template, a nanocomposite material with standardized morphology and size of the template can be obtained [86–88]. The template-directed synthesis method can design the texture and structure of the template according to the morphology and performance requirements of the synthetic material to meet actual needs [89,90].

#### 2.3.1. Self-Sacrificial Template Method

In self-sacrificial template method, MMOFs composites were prepared at room temperature by using metal oxide as metal source with MNPs pre-packaged in it [91,92]. The morphology and properties of the MMOFs can be controlled effectively [93,94]. Huang et al., have coated  $\text{Fe}_3\text{O}_4@\text{SiO}_2$  with  $\text{Cu}(\text{OH})_2$  as the self-template, and converted  $\text{Cu}(\text{OH})_2$  into HKUST-1 using ethanol and water as the solvent at room temperature (Figure 4a). The obtained MMOFs composite  $\text{Fe}_3\text{O}_4@\text{SiO}_2@\text{HKUST-1}$  was further applied as MSPE adsorbent to establish a sensitive, selective, simple-operated MSPE method for  $\text{Hg}^{2+}$  detection in water [95].



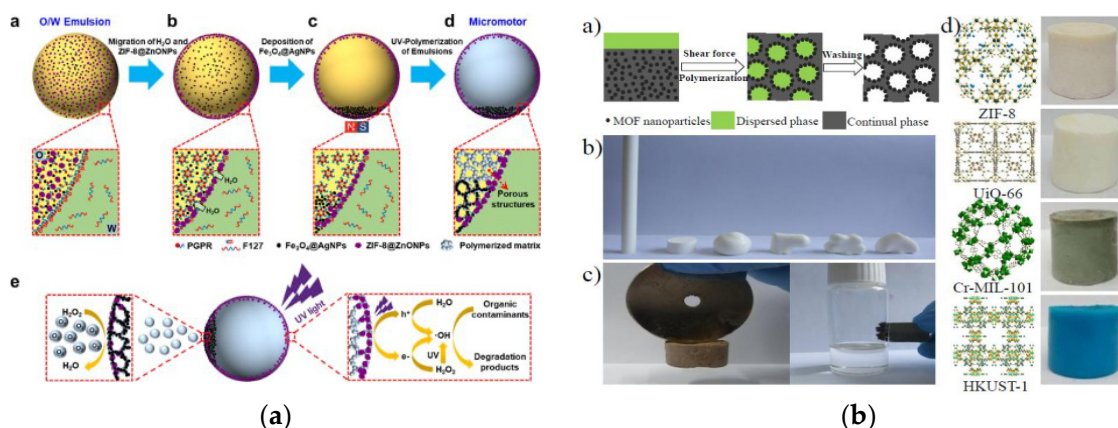
**Figure 4.** (a) Schematic diagram of the preparation of magnetic  $\text{Fe}_3\text{O}_4@\text{SiO}_2@\text{HKUST-1}$  composites [95]. Copyright Journal of Materials Chemistry A, 2015. (b) Schematic of the synthesis of  $\text{Rh-Ni@MOF-74(Ni)}$  composite [96]. Copyright ACS Applied Materials & Interfaces, 2016.

Bimetallic alloys NPs composed of noble-metals (Pt, Pd, and Rh) and transition metals ( $\text{M} = \text{Fe}$ ,  $\text{Co}$ ,  $\text{Ni}$ ,  $\text{Cu}$ , and  $\text{Zn}$ ) are low-cost and have superior performance in catalysis, which attracted increasing attention in recent years [97–99]. It is worth noting that the transition metal atoms in bimetallic NP tend to be oxidized and dissolved by dissolved oxygen, resulting in chemical dealloying in solution. Therefore, bimetallic NPs can be used as the self-sacrificial template to synthesis the required MNPs@MOF composite material with definite structure [100,101]. Chen and his co-workers reported a MOF-74(Ni)-encapsulated Rh-Ni hierarchical hetero structures ( $\text{Rh-Ni@MOF-74(Ni)}$ ) using magnetic Rh-Ni-alloyed nanoflowers (NFs) as a self-sacrificial template [96] (Figure 4b). The encapsulation state and thickness of the formed MOFs shell were well-tuned via template dealloying by changing the Ni

content in the Rh-Ni NFs template. More interestingly, when using Rh-Ni@MOF-74 (Ni) as the catalyst, due to the confinement effect of the MOF shell, enhanced catalytic performance was observed for the selective hydrogenation of alkynes to cis products.

### 2.3.2. Emulsion Template Method

Emulsion template method is a new method of preparing porous materials in recent years [102,103]. This method generally uses emulsion microdroplets from microfluidics as template, and the target material is deposited on the surface of template by physical or chemical methods. After formation, the template was removed, leaving the neat pore structure [104,105]. Figure 5a showed a simple and general strategy for continuous fabrication of MMOFs [106]. Using  $\text{Fe}_3\text{O}_4$ @AgNPs integrated into controllable emulsion micro-droplets as template, uniform porous MMOFs decorated with  $\text{Fe}_3\text{O}_4$ @AgNPs at the bottom and ZIF-8@ZnONPs on the surface was successfully synthesized. Through the directional migration of emulsion droplets in the limited microspace driven by interface energy and magnetic field, the spatial position of ZIF-8@ZnONPs and  $\text{Fe}_3\text{O}_4$ @AgNPs in MMOFs can be precisely controlled. Different kinds of metal-based composite MOFs materials can grow in the polymer macropores to generate MMOF materials. Jin and his co-workers used emulsion droplet as template to prepare MMOF composite materials through interfacial nanoassembly/emulsion polymerization [107]. The fluid properties of the emulsion droplets can process the MOFs composite material into a specific task shape. The embedded  $\text{Fe}_3\text{O}_4$  NPs offered the MOF composite magnetic properties and excellent mechanical stability (Figure 5b). Chen et al. prepared a magnetic porous polyacrylamide polymer using  $\text{Fe}_3\text{O}_4$  and polyvinyl alcohol under water bath conditions, and successfully induced UiO-66 and Fe-MIL-101(-NH<sub>2</sub>) crystals of MOFs in the high-temperature emulsion template [108].



**Figure 5.** (a) Schematic illustration of the fabrication process of MOF-integrated photocatalytic micromotors [106]. Copyright ACS Applied Materials Interfaces, 2020. (b) Illustration of the interfacial assembly/polymerization fabrication procedure for the preparation of MMOFs [107]. Copyright Communication, 2013.

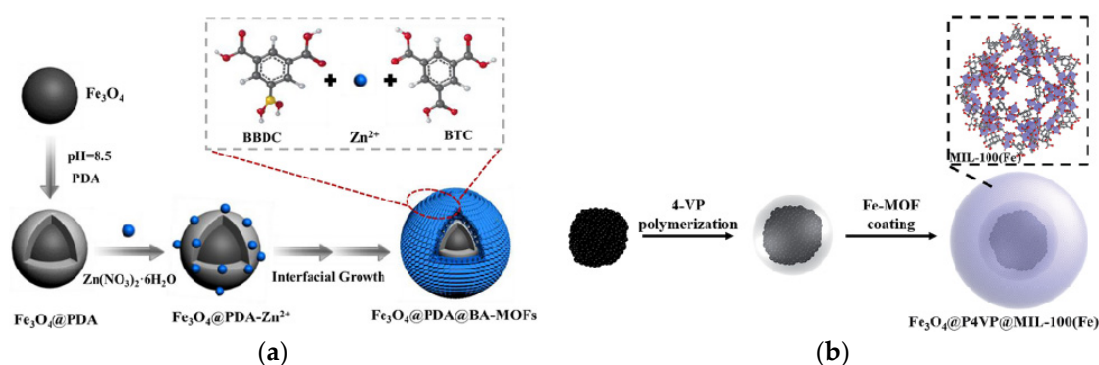
In emulsion template method, it can effectively avoid the decrease of specific surface area caused by the addition of MNPs, and can also construct macroporous polymers, thus accelerating the effective transport of matrix in the pore tunnel. However, this method requires a large amount of inorganic solvent and the preparation process is cumbersome [109].

### 2.4. Layer by Layer Self-Assembly Method

As a simple and efficient surface modification method, layer by layer (LbL) self-assembly technology has attracted increasing attention in the preparation of nanomaterials and promoted the development of material preparation research [110,111]. In the process of MMOFs preparation, before mixing with MOFs ligand solution, appropriate functional groups can be modified on the surface

of magnetic nanomaterials, so that the MOFs ligands can gradually extend on the surface of magnetic nanomaterials and spontaneously assemble into MMOFs [112,113]. Generally, the chemical groups modified on the surface of magnetic nanomaterials can promote the growth of MOFs crystal nucleus, which is beneficial to the formation of core-shell structure, improving the stability of MMOFs [114,115].

Figure 6a illustrated a LbL self-assembly strategy to fabricate  $\text{Fe}_3\text{O}_4$ @polydopamine (PDA)@boric-acid (BA)-functional MOFs ( $\text{Fe}_3\text{O}_4$ @PDA@BA-MOFs) for highly selective capture and separation of natural flavone from complex extraction samples [116]. The magnetic core  $\text{Fe}_3\text{O}_4$  was prepared using a solvothermal reaction and self-polymerized of PDA onto the surface. Due to the hydroxyl and amino groups of PDA,  $\text{Zn}^{2+}$  was easily adhered to the core surface, which can promote the growth of MOFs crystal nucleus. The  $\text{Fe}_3\text{O}_4$ @PDA@BA-MOFs with ultrahigh surface area, uniform morphology, excellent hydrophilicity and strong magnetic responsiveness were demonstrated to have high selectivity, obvious binding kinetics, large adsorption capacity, and excellent reusability (above 91.23% at least six repeated adsorption-desorption cycles), which would be good candidates for the enrichment and purification of luteolin. Liu et al., prepared the magnetic composite of MOF- $\text{Fe}_3\text{O}_4$ @HKUST-1/MIL-100(Fe) by LbL self-assembly method and applied it for the detection of methylene blue in foods [117]. Great recoveries in the range of 70–110% signified the application prospect of MMOFs as SPE adsorbent in the pretreatment of food matrices. Miao and the co-workers have prepared the core of  $\text{Fe}_3\text{O}_4$ @poly(4-vinylpyridine) (P4VP) and introduced the MIL-100(Fe) shell through LbL self-assembly [118] (Figure 6b). This novel magnetic  $\text{Fe}_3\text{O}_4$ @P4VP@MIL-100(Fe) composite with core-shell material was applied as the catalyst for selective oxidation of alcohols with remarkable reusability (maintained after ten reuses).



**Figure 6.** (a) Illustration of LbL assembly strategy to fabricate the  $\text{Fe}_3\text{O}_4$ @PDA@BA-MOFs composite [116]. Copyright Chemical Engineering Journal, 2018. (b) Schematic illustration of the synthesis of  $\text{Fe}_3\text{O}_4$ @P4VP@MIL-100(Fe) composite [118]. Copyright RSC Advances, 2017.

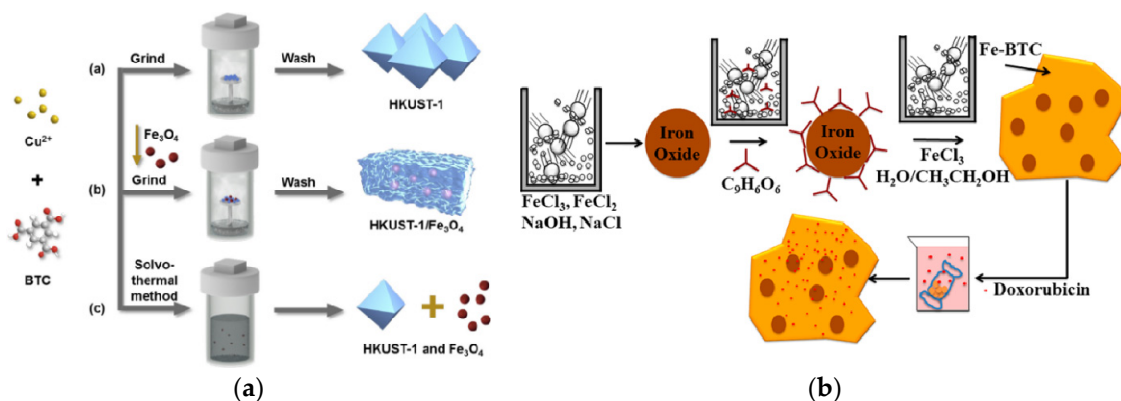
LbL self-assembly method can also precisely control the thickness or properties of MMOFs by adjusting the number of self-assembly cycles, and the reaction conditions are relatively mild. However, it usually takes a long time to obtain the desired thickness of MMOFs, and the types of self-assembled MOFs ligands are few, which limits the development of new MOFs with crystal structure.

### 2.5. Other Strategies for MMOFs Synthesis

Dry gel conversion method was also used for the synthesis of MMOFs. In dry gel conversion method, the reactants were first separated from the solvent and steamed at a high temperature to induce the MOFs material to grow around MNPs [119,120]. The preparation of MMOFs by dry gel conversion method was first proposed by Tan et al., in 2017 [121]. Figure 7a has shown a synthesis process of MMOFs by dry gel conversion method. The solid separated from the solvent was stewing with *N,N*-dimethylformamide at high temperature to induce the growth of MOFs around MNPs, and successfully prepared a MMOFs material  $\text{Fe}_3\text{O}_4$ /HKUST-1. The characteristic of growing around the MNPs avoids the blockage of material pores caused by the presence of unreacted magnetic



nanomaterials in the prepared MOFs material channel, reduces the loss of organic solvents to a certain extent, and needs less time for preparation. It's worth noting that the uniformity of the MMOFs prepared by this method needs to be improved [122,123].



**Figure 7.** (a) Schematic diagram for MMOFs synthesis by the dry gel conversion method [121]. Copyright Journal of Hazardous Materials, 2017. (b) Schematic diagram for MMOFs synthesis by mechanical grinding method [124]. Copyright Inorganic Chemistry, 2018.

At room temperature, MOFs can be connected to the surface of MNPs with organic linker, which can promote the growth of MOFs. Through liquid-assisted grinding in water or ethanol, the MMOFs composite can be obtained [125–127]. This procedure for MMOFs synthesis is named “mechanical grinding method”. Bellusci et al., first verified the feasibility of the grinding method to obtain a magnetic composite of MOFs (Figure 7b). A magnetic  $\text{Fe}_3\text{O}_4$  NPs were first prepared using ball grinding method by inducing the mechanical and chemical reaction, and further functionalized by neat grinding with benzene-1,3,5-tricarboxylic acid to obtain a composite consisting of  $\text{Fe}_3\text{O}_4$  NPs encapsulated in a MOF matrix. The prepared MMOFs exhibits magnetic characteristics and high porosity and can be used as the carrier for targeted drug delivery and release [124].

Although there are only few types of MOFs ligands available and the properties of the prepared MMOFs are not uniform, this mechanical grinding method also has great potential in future studies on the preparation of MMOFs due to its simple synthesis process and low cost.

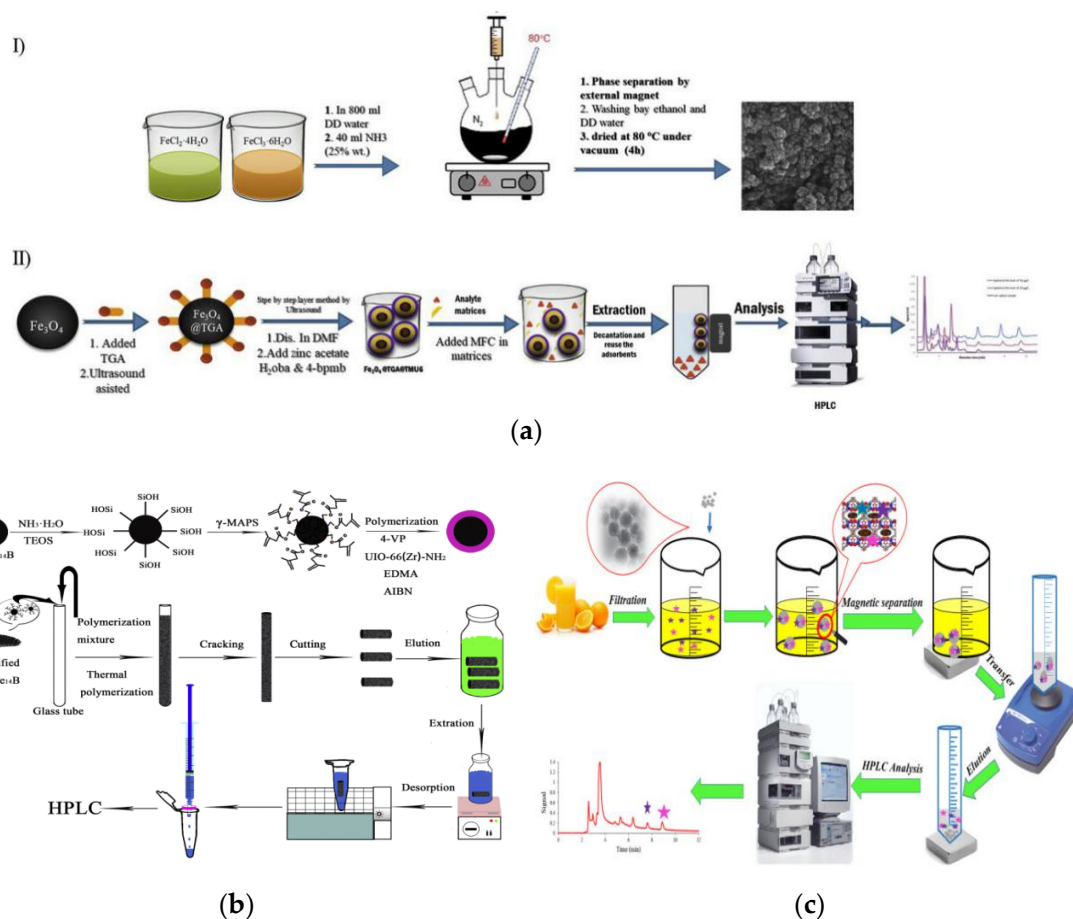
### 3. Applications of Magnetic MOFs for Food Contaminants Extraction

#### 3.1. Residues of Pesticide and Veterinary Drugs

The residue of pesticide and veterinary drugs in food are the main contaminants in food, which has aroused wide concern [128–130]. In the process of crop production, pesticides such as insecticides and herbicides are used in large quantities and enter human and animal bodies through food intake, resulting in accumulative toxicity. For the production of animal-derived foods (meat, eggs, milk, etc.), various antibiotics and other veterinary drugs are used to treat livestock diseases and promote animal growth. Improper use of veterinary drugs resulted in their residues in food samples [131,132]. After the foods containing the residue of pesticides or veterinary drugs are ingested by human body, it may induce various allergic reactions or diseases, which bring great threat to human health.

In the detection process of the residue of pesticide and veterinary drugs in food, MMOFs are characterized by easy preparation, simple operation and high adsorption efficiency, which can effectively reduce matrix interference and improve the detection sensitivity of the target [133–136]. Figure 8a has shown the synthesis of a MMOF ( $\text{Fe}_3\text{O}_4$ @ thioglycolic acid (TGA) @TMU-6) and the use for MSPE of some organophosphorus pesticides (OPPs) in rice and environmental water samples [137]. Due to the large surface area and unique porous structure, as well as the  $\pi$ - $\pi$ , hydrophobic interaction between the analytes and MOF ligands, the prepared MMOFs composites have a high affinity for

OPPs. The detection limit (LOD) of phosalone, chlorpyrifos, and profenofos achieved to 0.5, 1 and 0.5  $\mu\text{g/L}$ , respectively. The Zn/Co-MOFs-derived magnetic nanoporous carbons synthesized by Li et al., via one-step carbonization also exhibit high specific surface area and high extraction efficiency for OPPs. Coupled with gas chromatography-flame photometric detection (GC-FPD), a simple, rapid, selective and highly sensitive analysis for trace five OPPs in fruits was achieved [138]. A UIO-66(Zr)-NH<sub>2</sub> hybrid magnetic stir bar developed by Yang and the co-workers was successfully applied for adsorption and extraction of five sulfonylurea herbicides in food samples [139] (Figure 8b), which obtained higher extraction efficiency than the blank stir bar without UIO-66(Zr)-NH<sub>2</sub>. The LOD for five analytes was in the range of 0.04–0.84  $\mu\text{g/L}$  with the recoveries of 68.8–98.1% in real samples. The UIO-66(Zr)-NH<sub>2</sub> hybrid magnetic stir bar possessed excellent advantages of both supermagnetic performance and large specific surface area, which is an optimal adsorbent for food analysis. The MMOFs composites (Fe<sub>3</sub>O<sub>4</sub>@TMU-21) with a shell-core structure prepared by Yamini et al. through hybrid preparation method showed good selectivity and high recovery for pyrethrin pesticides in orange juice [140] (Figure 8c).



**Figure 8.** (a) Synthesis of Fe<sub>3</sub>O<sub>4</sub>@TGA@TMU-6 for MSPE of OPPs in water and rice [137]. Copyright Talanta, 2020. (b) Preparation and stir-bar sorptive extraction scheme of the hybrid UIO-66(Zr)-NH<sub>2</sub> magnetic stir bar [139]. Copyright Microchemical Journal, 2018. (c) MSPE using Fe<sub>3</sub>O<sub>4</sub>@TMU-21 and HPLC analysis procedure of trace pyrethroids [140]. Copyright Microchemical Journal, 2019.

Up to now, more and more studies on the development and application of MMOFs for analyzing pesticide and veterinary drug residues were reported. The detection targets also extended to fluoroquinolones [141,142], sulfonamides [143], ampicillin [144], nitroimidazoles [145], benzoylurea insecticides [146], tetracycline [147], triazine herbicides [148] et al., Duo et al., synthesized a novel, spinel-like magnetic metal-organic skeleton material (Fe<sub>3</sub>O<sub>4</sub>-NH<sub>2</sub>@MOF-235) by a two-step

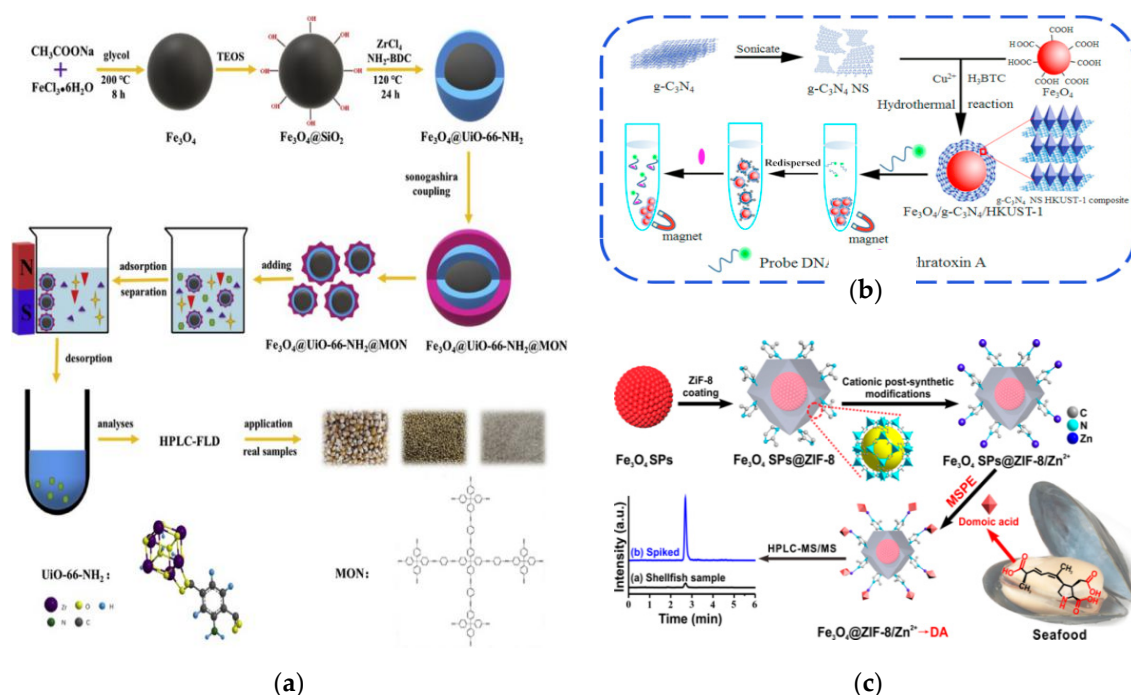
solvothermal method, and used as SPE adsorbent for simultaneous extraction and determination of five benzoyl urea pesticides in honey, fruit juice and tap water samples [149]. The deposition and hydrophobic interaction between  $\text{Fe}_3\text{O}_4\text{-NH}_2\text{@MOF-235}$  and the analyte plays an important role in the adsorption process, and the maximum adsorption capacity can be reached within 5 min. The magnetic  $\text{Fe}_3\text{O}_4\text{-NH}_2\text{@MOF-235}$  composite combined with SPE-HPLC is demonstrated to be a convenient method for the detection of benzourea insecticides in actual samples.

### 3.2. Toxins

#### 3.2.1. Mycotoxins

Mycotoxin is a kind of secondary metabolites produced by fungi, which is the common contaminants in grains, oils and other foods [150,151]. Each step in the food production including processing, storage and transportation is susceptible to fungal infection, which is why mycotoxin pollution is difficult to avoid. Ingesting foods with high levels of mycotoxin can cause serious harm to human body [152,153]. The food matrix composition is complex and the content of mycotoxins in food is very low, which brings difficulties to the detection of mycotoxins contamination. MSPE technique using MMOFs as the purification material has high enrichment efficiency, which has been widely used in the analysis of mycotoxins in foods.

Aflatoxins, produced by filamentous fungal species, are the most toxic type of mycotoxins and great effort has been made in developing of adsorptive materials for effective probing the target aflatoxins. Li et al., have constructed a core-shell structure  $\text{Fe}_3\text{O}_4\text{@UiO-66-NH}_2\text{@microporous organic network (MON)}$  composite and used it as sorbent to separate aflatoxins [154] (Figure 9a). The obtained sorbent possessed excellent selectivity and sensitivity with LODs in the range of 0.15–0.87  $\mu\text{g/L}$  by combination with HPLC analysis. And due to the presence of MON coating, the hydro-stability and adsorption efficiency of adsorbents were significantly improved. Another composite of  $\text{MIL53(Al)-SiO}_2\text{@Fe}_3\text{O}_4$ , prepared in a typical Stöber synthesis process and ultrasonic agitation co-precipitation method, was applied in multi-component adsorption for aflatoxin B1 in winter herbal tea, which obtained a wider linear range (0.5–150  $\text{ng/mL}$ ), a lower LOD (0.5  $\text{ng/mL}$ ), and an acceptable recovery rate (70.7–96.5%) [155]. A  $\text{MIL-101(Cr)/Fe}_3\text{O}_4\text{@SiO}_2\text{@propylthiouracil}$  composite developed by Sabeghi et al., was used to simultaneously isolate and determine four aflatoxins (B1, B2, G1, G2) in pistachio samples [156]. The developed strategy was shown to be facile, sensitive and accurate for aflatoxins detection, having great prospect for aflatoxins detection from other complex samples. Hu et al., prepared a MMOFs ( $\text{Fe}_3\text{O}_4\text{/graphitic-phase carbon nitride(g-C}_3\text{N}_4\text{)/HKUST-1}$ ) through the LbL self-assembly method, which was used as a MSPE adsorbent to detect ochratoxin A in corn [157] (Figure 9b). Under the optimal conditions, the  $\text{Fe}_3\text{O}_4\text{/g-C}_3\text{N}_4\text{/HKUST-1}$  exhibit better sensitivity (LOD: 2.57  $\text{ng/mL}$ ) and higher recovery (96.5–101.4%) for ochratoxin A, which confirmed the application prospect of magnetic MOFs as SPE adsorbent in food sample pretreatment.



**Figure 9.** (a) Schematic illustration of the synthesis and application of  $\text{Fe}_3\text{O}_4@\text{UiO-66-NH}_2@\text{MON}$  [154]. Copyright Journal of Hazardous Materials, 2020. (b) Biosensing detection of ochratoxin A based on  $\text{Fe}_3\text{O}_4/\text{g-C}_3\text{N}_4/\text{HKUST-1}$  composite [157]. Copyright Biosensors and Bioelectronics, 2016. (c) Scheme for preparing  $\text{Fe}_3\text{O}_4$  SPs@ZIF-8/ $\text{Zn}^{2+}$  particles and the use in MSPE for domoic acid detection [158]. Copyright Analytical Chemistry, 2019.

### 3.2.2. Algal Toxins

Due to climate change and large amounts of industrial wastewater discharge, environmental issues such as eutrophication of water and cyanobacteria blooms have also attracted widespread attention. Harmful algae can produce a large number of algal toxins to pollute water sources, which also can enter human body through the digestive tract, causing diarrhea, nerve paralysis, liver damage, and even death in serious cases [159,160].

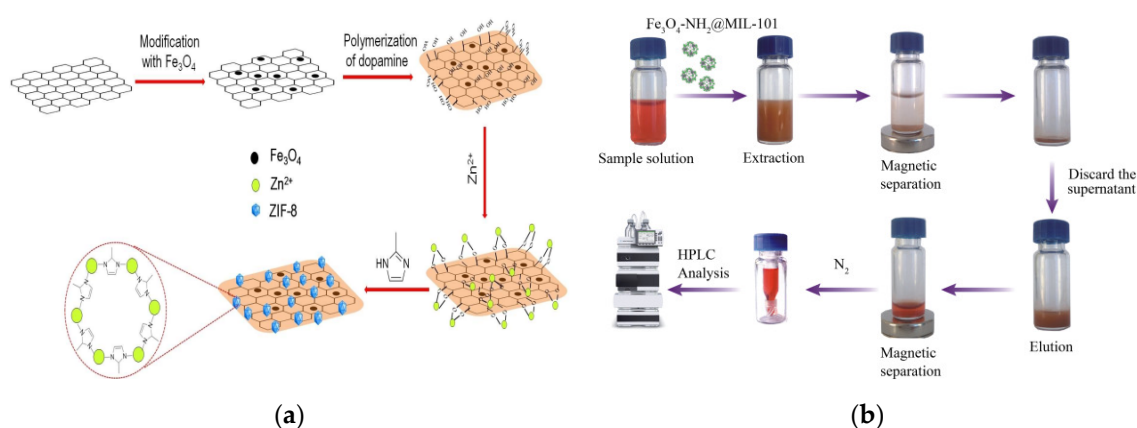
Microcystin-LR with high content and strong toxicity, belonging to class 2B carcinogen, was released by cyanobacteria in eutrophic water. Chen et al., successfully synthesized  $\text{Fe}_3\text{O}_4@\text{poly-dopamine(PDA)}@\text{Cu-MOFs}$  and further applied to analyze traces of microcystin-LR [161]. It is found that the  $\text{Fe}_3\text{O}_4@\text{PDA}@\text{Cu-MOFs}$  composite material exhibits ultra-high surface area, strong magnetic response and excellent hydrophilicity. Combined with matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF-MS), microcystin-LR in water was accurately and sensitively detected. Huang et al., fabricated the MMOFs ( $\text{Fe}_3\text{O}_4$  superparticles (SPs) @ZIF-8/ $\text{Zn}^{2+}$ ) for the adsorption of domoic acid from food samples [158]. Through the strong electrostatic interactions and chelation with coordinatively unsaturated  $\text{Zn}^{2+}$  sites on the surface of magnetic MOFs, the adsorption of domoic acid can be finished within 5 min. By employing  $\text{Fe}_3\text{O}_4$  SPs@ZIF-8/ $\text{Zn}^{2+}$  as sorbent for MSPE, followed by liquid chromatography-mass spectrometry (LC-MS), a facile, rapid, efficient, and sensitive detection of trace domoic acid in marine products was realized (Figure 9c).

### 3.3. Illegal Additives

As an important part of food industry, food additives play an irreplaceable role in food production, processing and storage. However, the emergence of illegal additives such as melamine, Sudan dyes and plasticizer compounds has brought new threats to food safety [162,163]. In the analysis of illegal additives in foods, MMOFs materials with simple preparation and high adsorption efficiency are also



used for food matrix purification and trace contaminants enrichment [164–166]. Lu et al. have proposed a location-controlled strategy for functionalization of hydrophilic magnetic graphene by ZIF-8 MOFs to construct uniform and robust nanosheets (magG@PDA@ZIF-8) [167] (Figure 10a). The prepared magG@PDA@ZIF-8 has regular porous structure and large surface area with unique  $\pi$ - $\pi$  stacking electron system, strong magnetism and excellent water dispersibility, which was further used to enrich nine phthalates in cola beverages. Yamini et al., synthesized the  $\text{Fe}_3\text{O}_4$ @TMU-24 materials were demonstrated an effective sorbent for preconcentration of the plasticizer compounds [168]. Shi et al., fabricated a MMOFs ( $\text{Fe}_3\text{O}_4$ - $\text{NH}_2$ @MIL-101) through in-situ synthesis method, which can be used for MSPE pretreatment to effectively isolate six Sudan dyes [169]. Sudan dyes were efficiently separated from the complex matrices using only 3 mg of  $\text{Fe}_3\text{O}_4$ - $\text{NH}_2$ @MIL-101 within 2 min, making pretreatment easier, faster and more effective. Figure 10b has shown the MSPE process for Sudan dyes detection using  $\text{Fe}_3\text{O}_4$ - $\text{NH}_2$ @MIL-101 as adsorbent. Zhou prepared a  $\text{Fe}_3\text{O}_4$ @PEI-MOF-5 material by a facile two-step solvothermal approach for effective enrichment of malachite green (MG) and crystal violet (CV) in fish samples [170]. Th new material was demonstrated to have high magnetization and chemical stability, a large surface area and good adsorption to MG and CV. By combining the  $\text{Fe}_3\text{O}_4$ @PEI-MOF-5 material with HPLC-MS, an effective enrichment and detection method for MG and CV was subsequently developed with linearity range of 1–500 ng/mL (MG) and 0.25–500 ng/mL (CV), LOD of 0.30 ng/mL (MG) and 0.08 ng/mL (CV), respectively.

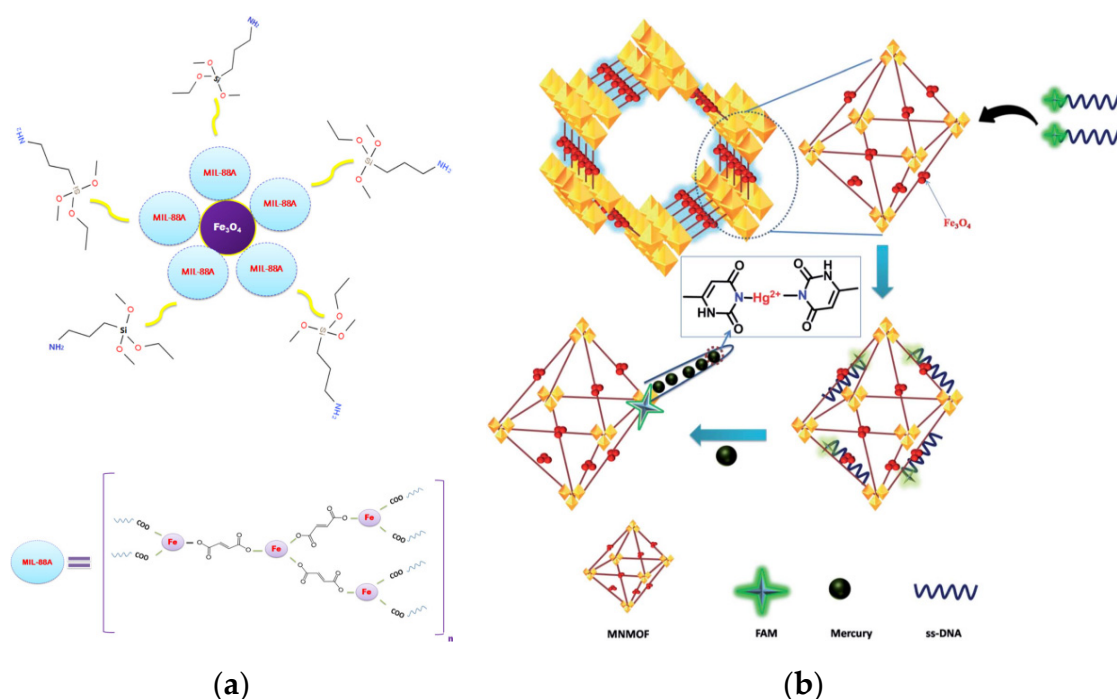


**Figure 10.** (a) Scheme of synthetic process of magG@PDA@ZIF-8 [167]. Copyright Chemistry Select, 2018. (b) MSPE for Sudan dyes using  $\text{Fe}_3\text{O}_4$ - $\text{NH}_2$ @MIL-101 as adsorbent [169]. Copyright Journal of Chromatography B, 2018.

### 3.4. Heavy Metal Ions

The analysis of heavy metal elements in food is another research hotspot of food contaminant detection [171,172]. The excessive discharge of industrial sewage containing heavy metals, the unreasonable use of pesticides containing heavy metals and other factors have resulted in the excessive heavy metals in the water and soil where the food raw materials are produced [173,174]. The pollution of heavy metals is difficult to be biodegraded and can enter human body through water or food chain, resulting in accumulative toxicity. Modern medicine research has proved that the accumulation of heavy metals is likely to induce cardiovascular, liver and kidney diseases and even cancers, which has posed a serious threat to human health [175]. The detection of trace heavy metals in food matrix also requires matrix pretreatment, target ion enrichment and other processes. The composite of MMOFs is simple to prepare, stable in structure and function, and can replace the traditional adsorbent to achieve more efficient and faster detection [176,177]. As a result, various MMOFs were synthesized and applied in the extraction of  $\text{Hg}^{2+}$  [178–180],  $\text{Sn}^{2+}$  [181],  $\text{Cr}^{3+}$  [182],  $\text{Pb}^{2+}$  [183] from food samples.

Figure 11a has illustrated the fabrication of novel MOFs meso-porous  $n\text{Fe}_3\text{O}_4@\text{MIL-88A}(\text{Fe})/3\text{-aminopropyltrimethoxysilane}$  (APTMS) nanocomposite, which exhibited excellent selective adsorption performance for heavy metal ions [184]. The maximum adsorption capacity for  $\text{Cd}^{2+}$ ,  $\text{Pb}^{2+}$  and  $\text{Cr}^{2+}$  achieved to 693.0, 536.22 and 1092.22 mg/g, respectively. It was worth noting that this  $n\text{Fe}_3\text{O}_4@\text{MIL-88A}(\text{Fe})/\text{APTMS}$  nanocomposite was easily regenerated and the adsorptive removal values were decreased by only 3% after five consecutive recycling processes. Safari et al., have developed a MMOFs composite consisting of phenylthiosemicarbazide MNPs ( $\text{Fe}_3\text{O}_4@\text{PTSC}$ ) and MIL-101(Cr), and further utilized in MSPE to effectively isolate and simultaneously determine the heavy metals including  $\text{Cd}^{2+}$ ,  $\text{Pb}^{2+}$  and  $\text{Ni}^{2+}$  in agricultural and seafood samples [185]. By combining with flame atomic absorption spectrometry (FAAS), the developed MSPE method obtained the lower LODs (0.07–0.5 mg/kg), lower quantification limits (0.2–2.0 mg/kg), wide linear ranges (0.25–250 mg/kg) and good extraction recoveries (97.5–99.0%). Figure 11b illustrated a DNA fluorescence assay comprising a MMOFs functionalized with fluorescein amidite (FAM)-labeled ssDNA for  $\text{Hg}^{2+}$  detection [186]. The fluorescence intensity showed a drastic decrease upon the presence of  $\text{Hg}^{2+}$ . The methods exhibited good sensitivity (LOD: 8 nM) and selectivity for  $\text{Hg}^{2+}$  over other co-existing metal ions. These merits made it an ideal platform for  $\text{Hg}^{2+}$  sensing applications. A novel MMOFs from dithizone-modified  $\text{Fe}_3\text{O}_4$  nanoparticles and a copper-(benzene-1,3,5-tricarboxylate) MOF were successfully prepared and applied to the rapid extraction of trace quantities of heavy metal ions in fish, sediment, soil, and water samples [187]. By combined with FAAS, the LOD for  $\text{Cd}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Ni}^{2+}$ , and  $\text{Pb}^{2+}$  achieved to 0.12, 0.39, 0.98, and 1.2 ng/mL, respectively. Li et al., prepared  $\text{Fe}_3\text{O}_4@\text{MOF}@\text{covalent organic framework (COF)}$  by solvothermal methods for selectively separation and preconcentration of  $\text{Cu}^{2+}$  for the first time [188]. With a high density of nitrogen- and oxygen-containing functional groups on the surface, the  $\text{Fe}_3\text{O}_4@\text{MOF}@\text{COF}$  composite could efficiently extract and preconcentrate of  $\text{Cu}^{2+}$  ions in aqueous solutions, with a maximum adsorption capacity of 37.29 mg/g and LOD of 37.6 nm ( $n = 11$ ) for  $\text{Cu}^{2+}$ .



**Figure 11.** (a) Proposed structure of  $n\text{Fe}_3\text{O}_4@\text{MIL-88A}(\text{Fe})/\text{APTMS}$  [184]. Copyright Journal of Hazardous Materials, 2019. (b) Schematic illustration of the DNA fluorescence assay with MMOFs as a sensing platform [186]. Copyright RSC Advances, 2020.

#### 4. Conclusions

MMOFs have a unique structure and good adsorption performance and are easy to separate, which have become high-efficiency adsorbents widely used in the pretreatment process of food matrices and play an important role in the detection of trace contaminants in foods. (Table 2) However, the existing MMOFs still have the disadvantages of few kinds and poor stability, which cannot meet the different application requirements. The difficulties in large-scale preparation caused by relatively complicated synthesis process and the large difference between batches are also the factors restricting its further development.

Future development direction: (1) Green and efficient synthesis of MMOFs for food matrices pretreatment is very necessary. The exploration on the synthesis and application strategies of new, green, environmentally friendly and efficient MMOFs has become a new research hotspot. (2) The development and application of broad-spectrum MMOFs materials that can enrich and separate a variety of contaminants can significantly improve the pretreatment efficiency of food matrices and increase the detection throughput, which has broad application prospects. (3) Improving the matching of MMOFs materials and analytical instruments, and developing integrated, automated, online sample pretreatment technologies and equipment are of great significance to the detection and control of contaminants in foods.

**Table 2.** Analysis of Various Contaminants in Foods Using MMOFs as SPE Sorbent Combined with Different Instruments.

Materials	Analyte	Instrument	Sample	Conditions (Sorbent Amount; Eluent)	LOD	Linear Range	RSD	Recovery	Ref
Fe <sub>3</sub> O <sub>4</sub> /ZIF-8/IL	Pyrethroids	GC-MS/MS	Tea infusions (2 g)	10 mg; 0.8 mL of acetonitrile	0.0065–0.1017 µg/L	0.5–50 µg/L	≤9.70%	81.5–98.1%	[51]
Fe <sub>3</sub> O <sub>4</sub> @TGA@TMU-6	Phosalone Chlorpyrifos Profenofos	HPLC-UV	Rice (20 g)	2 mg; 0.1 mL of 1-butanol	0.15–0.87 µg/L	7.5–75 µg/L 10–100 µg/L 10–150 µg/L	4.8–7.3%	88–107.2%	[137]
MNPCs-Zn/Co-MOF	OPPs	GC-FPD	Fruit (1 g)	10 mg; 0.2 mL of acetone/ethyl acetate (1:1, v/v)	0.018–0.045 µg/L	0.1–100 µg/L	3.5–9.7%	84–116%	[138]
Nd <sub>2</sub> Fe <sub>14</sub> B-UIO-66, (Zr)-NH <sub>2</sub>	Sulfonylurea	HPLC	Water (5 mL)	5 mg; 0.2 mL of methanol-acetic acid (9:1, v/v)	0.04–0.84 µg/L	10–700 µg/L	≤13.8%	68.8–98.1%	[139]
Fe <sub>3</sub> O <sub>4</sub> @UiO-66-NH <sub>2</sub> @MON	Aflatoxins	HPLC	Corn, rice, millet (5 g)	10 mg; 6 mL of acetonitrile	1 µg/L	0.15–0.87 µg/L	–	87.3–101.8%	[154]
MIL53(Al)-SiO <sub>2</sub> @Fe <sub>3</sub> O <sub>4</sub>	Aflatoxin B1	FTIR	Herbal teas (no indication)	100 mg; 2 mL of Me <sub>2</sub> CO/MeCN/CH <sub>2</sub> Cl <sub>2</sub> (1:1:2, v/v)	0.5 ng/mL	0.5–150 ng/mL	<4.3%	70.7–96.5%	[155]
Fe <sub>3</sub> O <sub>4</sub> /g-C <sub>3</sub> N <sub>4</sub> /HKUST-1	Ochratoxin A	Fluorescent biosensor	Corn (no indication)	–	2.57 ng/mL	5.0–160.0 ng/mL	<2.5%	96.5–101.4%	[157]
Fe <sub>3</sub> O <sub>4</sub> @PDA@Cu-MOFs	Microcystin	MALDI-TOF-MS	Water (80 µL)	20 µg; 10 µL of NH <sub>4</sub> HCO <sub>3</sub> (0.25 mol/L)	0.015 mg/L	0.05–4 mg/L	6.1–8.2%	98.67–106.15%	[160]
Fe <sub>3</sub> O <sub>4</sub> SPs@ZIF-8/Zn <sup>2+</sup>	Domoic acid	LC-MS	Shellfish (5 g)	1.0 mg; 0.4 mL of aqueous histidine solution (3 mmol/L)	0.2 pg/mL	1.0–1000 pg/mL	≤3.4%	93.1–102.3%	[161]
magC@PDA@ZIF-8	Phthalates	HPLC	Beverages (1 mL)	20 mg; 1 mL of acetone	0.003–0.09 ng/mL	50–8000 ng/mL	<4.5%	91.5–104.7%	[167]
Fe <sub>3</sub> O <sub>4</sub> -NH <sub>2</sub> @MIL-101	Sudan dyes	HPLC	Tomato (4 g)	3 mg; 1 mL × 2 ethyl acetate	0.5–2.5 µg/kg	0.01–25 µg/mL	≤9.2%	69.6–92.6%	[169]
Fe <sub>3</sub> O <sub>4</sub> @1-phenylthiosemicarbazide/MIL-101(Cr)	Cd <sup>2+</sup> ; Pb <sup>2+</sup> ; Ni <sup>2+</sup>	FAAS	Shrimp, Cucumber, Tomato, Parsley (1 g)	13 mg; 2.2 mL of 0.85 mol/L HCl	0.07–0.5 mg/kg	0.07–0.5 mg/kg 0.2–2.0 mg/kg 0.25–250 mg/kg	4.5–7.3%	97.5–99.0%	[185]
Fe <sub>3</sub> O <sub>4</sub> -NH <sub>2</sub> @MIL-101(Fe)	Hg <sup>2+</sup>	Fluorescent biosensor	Water (no indication)	–	8 nm	2–20 nm	–	About 70%	[186]
M-MOF/β-CD	Triazole Fungicides	HPLC-MS/MS	Lettuce, tomato (10 g)	10 mg; 4 mL of acetone-sodium citrate buffer	0.25–1.0 mg/L	0.25–1.0 µg/L	1.5–7.3%	86.44–119.83%	[189]
Fe <sub>3</sub> O <sub>4</sub> @JUC-48	Sulfonamides	HPLC	Chicken, pork, shrimp (5 g)	25 mg; 0.8 mL of methanol	3.97–1000 ng/g	3.97–1000 ng/g	<4.5%	76.1–102.6%	[190]



**Author Contributions:** J.Y. coordinated and organized the writing of the entire manuscript and complete the part of Sections 2 and 3; Y.W. completed the part of Section 2; M.P. provided the idea and completed the Section 1; X.X. and K.L. checked the language and format of the manuscript; L.H. completed the Section 4; S.W. provided the framework of the paper and checked the quality of the article. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by National Natural Science Foundation of China (No. 31972147), Tianjin Technical Expert Project (No. 19JCTPJ52700), Project of Tianjin Science and Technology Plan (No. 19PTSYJC00050), the Open Project Program of State Key Laboratory of Food Nutrition and Safety, Tianjin University of Science and Technology (No. SKLFNS-KF-202011), and the Open Project Program of Key Laboratory for Analytical Science of Food Safety and Biology, Ministry of Education (No. FS2001). The APC was funded by the Open Project Program of State Key Laboratory of Food Nutrition and Safety, Tianjin University of Science and Technology (SKLFNS-KF-202011).

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Kantiani, L.; Llorca, M.; Sanchis, J.; Farre, M.; Barcelo, D. Emerging food contaminants: A review. *Anal. Bioanal. Chem.* **2010**, *398*, 2413–2427. [PubMed]
2. Alexander, J.; Barlow, S.; Benford, D.; Bolger, M.; Cantrill, R.; Cressey, P.; De Nijs, M.; Edwards, S.; Feeley, M.; Mueller, U.; et al. Evaluation of certain food additives and contaminants. *WHO Tech. Rep. Ser.* **2017**, *1002*, 1–166.
3. Callen, C.; Bhatia, J.; Czerkies, L.; Klish, W.J.; Gray, G.M. Challenges and considerations when balancing the risks of contaminants with the benefits of fruits and vegetables for infants and toddlers. *Nutrients* **2018**, *10*, 1572.
4. Rotariu, L.; Lagarde, F.; Jaffrezic-Renault, N.; Bala, C. Electrochemical biosensors for fast detection of food contaminants trends and perspective. *Trends Anal. Chem.* **2016**, *79*, 80–87.
5. Raeisossadati, M.J.; Danesh, N.M.; Borna, F.; Gholamzad, M.; Ramezani, M.; Abnous, K.; Taghdisi, S.M. Lateral flow based immunobiosensors for detection of food contaminants. *Biosens. Bioelectron.* **2016**, *86*, 235–246.
6. Frenich, A.G.; Romero-Gonzalez, R.; Aguilera-Luiz, M.D. Comprehensive analysis of toxics (pesticides, veterinary drugs and mycotoxins) in food by UHPLC-MS. *Trends Anal. Chem.* **2014**, *63*, 158–169.
7. Justino, C.I.L.; Duarte, K.R.; Freitas, A.C.; Panteleitchouk, T.S.L.; Duarte, A.C.; Rocha-Santos, T.A.P. Contaminants in aquaculture: Overview of analytical techniques for their determination. *Trends Anal. Chem.* **2016**, *80*, 293–310.
8. Li, D.A.; Lv, D.Y.; Zhu, Q.X.; Li, H.; Chen, H.; Wu, M.M.; Chai, Y.F.; Lu, F. Chromatographic separation and detection of contaminants from whole milk powder using a chitosan-modified silver nanoparticles surface-enhanced Raman scattering device. *Food Chem.* **2017**, *224*, 382–389. [PubMed]
9. Chiou, J.C.; Leung, A.H.H.; Lee, H.W.; Wong, W.T. Rapid testing methods for food contaminants and toxicants. *J. Integr. Agric.* **2015**, *14*, 2243–2264.
10. Huang, X.L.; Aguilar, Z.P.; Xu, H.Y.; Lai, W.H.; Xiong, Y.H. Membrane-based lateral flow immunochromatographic strip with nanoparticles as reporters for detection: A review. *Biosens. Bioelectron.* **2016**, *75*, 166–180.
11. Xu, M.L.; Gao, Y.; Han, X.X.; Zhao, B. Detection of pesticide residues in food using surface-enhanced Raman spectroscopy: A review. *J. Agric. Food Chem.* **2017**, *65*, 6719–6726. [PubMed]
12. Naing, N.N.; Lee, H.K. Microextraction and analysis of contaminants adsorbed on atmospheric fine particulate matter: A review. *J. Chromatogr. A* **2020**, *1627*, 461433. [PubMed]
13. Zhang, Y.Y.; Li, G.L.; Wu, D.; Li, X.T.; Yu, Y.X.; Luo, P.J.; Chen, J.; Dai, C.J.; Wu, Y.N. Recent advances in emerging nanomaterials based food sample pretreatment methods for food safety screening. *Trends Anal. Chem.* **2019**, *121*, 115669.
14. Burato, J.S.D.; Medina, D.A.V.; de Toffoli, A.L.; Maciel, E.V.S.; Lancas, F.M. Recent advances and trends in miniaturized sample preparation techniques. *J. Sep. Sci.* **2020**, *43*, 202–225.
15. Xin, J.H.; Wang, X.; Li, N.; Liu, L.; Lian, Y.J.; Wang, M.L.; Zhao, R.S. Recent applications of covalent organic frameworks and their multifunctional composites for food contaminant analysis. *Food Chem.* **2020**, *330*, 127255.
16. Li, G.Q.; Ma, G.H. Recent applications of solid-phase extraction techniques for analysis of trace residues and contaminants in food. *Chin. J. Chromatogr.* **2011**, *29*, 606–612.

17. Li, Z.; Yu, B.; Cong, H.L.; Yuan, H.; Peng, Q.H. Recent development and application of solid phase extraction materials. *Rev. Adv. Mater. Sci.* **2017**, *49*, 87–111.
18. Mashile, G.P.; Nomngongo, P.N. Recent application of solid phase based techniques for extraction and preconcentration of cyanotoxins in environmental matrices. *Crit. Rev. Anal. Chem.* **2017**, *47*, 119–126.
19. Farajzadeh, M.A.; Dabbagh, M.S. Development of a dispersive solid phase extraction method based on in situ formation of adsorbent followed by dispersive liquid-liquid microextraction for extraction of some pesticide residues in fruit juice samples. *J. Chromatogr. A* **2020**, *1627*, 461398.
20. Yu, X.; Yu, L.; Ma, F.; Li, P.W. Quantification of phenolic compounds in vegetable oils by mixed-mode solid-phase extraction isotope chemical labeling coupled with UHPLC-MS/MS. *Food Chem.* **2021**, *334*, 127572.
21. Fumes, B.H.; Silva, M.R.; Andrade, F.N.; Nazario, C.E.D.; Lanças, F.M. Recent advances and future trends in new materials for sample preparation. *Trends Anal. Chem.* **2015**, *71*, 9–25.
22. Maciel, E.V.S.; Toffoli, A.L.; Neto, E.S.; Nazario, C.E.D.; Lancas, F.M. New materials in sample preparation: Recent advances and future trends. *Trends Anal. Chem.* **2019**, *119*, 115633.
23. Chen, Q.L.; Zhu, L.; Chen, J.X.; Jiang, T.; Ye, H.Z.; Ji, H.; Tsang, S.W.; Zhao, Z.Z.; Yi, T.; Chen, H.B. Recent progress in nanomaterial-based assay for the detection of phytotoxins in foods. *Food Chem.* **2019**, *277*, 162.
24. Herrero-Latorre, C.; Barciela-Garcia, J.; Garcia-Martin, S.; Pena-Creciente, R.M.; Otarola-Jimenez, J. Magnetic solid-phase extraction using carbon nanotubes as sorbents: A review. *Anal. Chim. Acta* **2015**, *892*, 10–26.
25. He, M.; Huang, L.J.; Zhao, B.S.; Chen, B.B.; Hu, B. Advanced functional materials in solid phase extraction for ICP-MS determination of trace elements and their species—A review. *Anal. Chim. Acta* **2017**, *973*, 1–24.
26. Er, E.O.; Bozyigit, G.D.; Buyukpinar, C.; Bakirdere, S. Magnetic nanoparticles based solid phase extraction methods for the determination of trace elements. *Crit. Rev. Anal. Chem.* **2020**, *18*, 1–18.
27. Capriotti, A.L.; Cavaliere, C.; Foglia, P.; La Barbera, G.; Samperi, R.; Ventura, S.; Lagana, A. Mycoestrogen determination in cow milk: Magnetic solid-phase extraction followed by liquid chromatography and tandem mass spectrometry analysis. *J. Sep. Sci.* **2016**, *39*, 4794–4804.
28. Patino-Ropero, M.J.; Diaz-Alvarez, M.; Martin-Esteban, A. Molecularly imprinted core-shell magnetic nanoparticles for selective extraction of triazines in soils. *J. Mol. Recognit.* **2017**, *30*, e2593.
29. Mohamed, F.A.; Khashaba, P.Y.; El-Wakil, M.M.; Shahin, R.Y. Fabrication of water compatible and biodegradable super-paramagnetic molecularly imprinted nanoparticles for selective separation of memantine from human serum prior to its quantification: An efficient and green pathway. *Int. J. Biol. Macromol.* **2019**, *140*, 140–148.
30. Wierucka, M.; Biziuk, M. Application of magnetic nanoparticles for magnetic solid-phase extraction in preparing biological, environmental and food samples. *Trends Anal. Chem.* **2014**, *59*, 50–58.
31. Rais, S.; Islam, A.; Ahmad, I.; Kumar, S.; Chauhan, A.; Javed, H. Preparation of a new magnetic ion-imprinted polymer and optimization using Box-Behnken design for selective removal and determination of Cu (II) in food and wastewater samples. *Food Chem.* **2021**, *334*, 127563.
32. Pan, L.; Liu, G.; Li, H.; Meng, S.; Han, L.; Shang, J.; Chen, B.; Platero-Prats, A.E.; Lu, W.; Zou, X.D.; et al. A resistance-switchable and ferroelectric metal-organic framework. *J. Am. Chem. Soc.* **2014**, *136*, 17477–17483.
33. Qu, B.T.; Lai, J.C.; Liu, S.; Liu, F.; Gao, Y.D.; You, X.Z. Cu- and Ag-based metal-organic frameworks with 4-Pyranone-2,6-dicarboxylic acid: Syntheses, crystal structures, and dielectric properties. *Cryst. Growth Des.* **2015**, *15*, 1707–1713.
34. Wang, C.H.; Liu, X.L.; Demir, N.K.; Chen, J.P.; Li, K. Applications of water stable metal-organic frameworks. *Chem. Soc. Rev.* **2016**, *45*, 5107–5134.
35. Zhang, M.W.; Chen, Y.P.; Bosch, M.; Gentle, T.; Wang, K.C.; Feng, D.W.; Wang, Z.Y.U.; Zhou, H.C. Symmetry-guided synthesis of highly porous metal-organic frameworks with fluorite topology. *Angew. Chem.* **2014**, *126*, 834–837.
36. Banerjee, D.; Cairns, A.J.; Liu, J.; Motkuri, R.K.; Nune, S.K.; Fernandez, C.A.; Krishna, R.; Strachan, D.M.; Thallapally, P.K. Potential of metal-organic frameworks for separation of Xenon and Krypton. *Acc. Chem. Res.* **2015**, *48*, 211–219.
37. Xu, Y.X.; Li, Q.; Xue, H.G.; Pang, H. Metal-organic frameworks for direct electrochemical applications. *Coordin. Chem. Rev.* **2018**, *376*, 292–318.

38. Liu, C.S.; Li, J.J.; Pang, H. Metal-organic framework-based materials as an emerging platform for advanced electrochemical sensing. *Coordin. Chem. Rev.* **2020**, *213*, 222. [[CrossRef](#)]
39. Seo, J.S.; Whang, D.; Lee, H.; Jun, S.I.; Oh, J.; Jeon, Y.J.; Kim, K. A homochiral metal-organic porous material for enantioselective separation and catalysis. *Nature* **2000**, *404*, 982–986.
40. Horcajada, P.; Chalati, T.; Serre, C.; Gillet, B.; Sebrie, C.; Baati, T.; Eubank, J.F.; Heurtaux, D.; Clayette, P.; Kreuz, C.; et al. Porous metal-organic-framework nanoscale carriers as a potential platform for drug delivery and imaging. *Nat. Mater.* **2010**, *2*, 172–178.
41. Ghaemi, F.; Amiri, A. Microcrystalline cellulose/metal-organic framework hybrid as a sorbent for dispersive micro-solid phase extraction of chlorophenols in water samples. *J. Chromatogr. A* **2020**, *1626*, 461386.
42. Giliopoulos, D.; Zamboulis, A.; Giannakoudakis, D.; Bikiaris, D.; Triantafyllidis, K. Polymer/metal organic framework (MOF) nanocomposites for biomedical applications. *Molecules* **2020**, *25*, 185. [[CrossRef](#)]
43. Li, J.R.; Sculley, J.; Zhou, H.C. Metal-organic frameworks for separations. *Chem. Rev.* **2012**, *112*, 869–932.
44. Gu, Z.Y.; Yang, C.X.; Chang, N.; Yan, X.P. Metal-organic frameworks for analytical chemistry: From sample collection to chromatographic separation. *Acc. Chem. Res.* **2012**, *45*, 734–745.
45. Furukawa, H.; Cordova, K.E.; O’Keeffe, M.; Yaghi, O.M. The chemistry and applications of metal-organic frameworks. *Science* **2013**, *341*, 1230444.
46. Zhu, Q.L.; Xu, Q. Metal-organic framework composites. *Chem. Soc. Rev.* **2014**, *43*, 5468–5512.
47. Wang, R.Y.; Zhang, C.Y.; Wang, S.P.; Zhou, Y.Y. Synthesis and application of magnetic metal-organic frameworks. *Prog. Chem.* **2015**, *27*, 945–952.
48. Hashemi, B.; Zohrabi, P.; Raza, N.; Kim, K.H. Metal organic frameworks as advanced sorbents for the extraction and determination of pollutants from environmental, biological, and food media. *Trends Anal. Chem.* **2017**, *97*, 65–82.
49. Wang, D.; Zeng, F.; Hu, X.; Li, C.; Su, Z. Synthesis of a magnetic 2D Co@NC-600 material by designing a MOF precursor for efficient catalytic reduction of water pollutants. *Inorg. Chem.* **2020**, *59*, 12672–12680.
50. Mohamed, A.K.; Mahmoud, M.E. Encapsulation of starch hydrogel and doping nanomagnetite onto metal-organic frameworks for efficient removal of fluvastatin antibiotic from water. *Carbohydr. Polym.* **2020**, *245*, 116438.
51. Huang, X.D.; Liu, Y.N.; Liu, H.F.; Liu, G.Y.; Xu, X.M.; Li, L.Y.; Lv, J.; Gao, H.X.; Xu, D.H. Magnetic solid-phase extraction of pyrethroid insecticides from tea infusions using ionic liquid-modified magnetic zeolitic imidazolate framework-8 as an adsorbent. *RES Adv.* **2019**, *9*, 39272–39281.
52. Han, Q.; Sun, Y.; Ding, K.; Chen, X.; Han, T. Preparation of multitarget immunomagnetic beads based on metal-organic frameworks and their application in food samples. *J. Chromatogr. B* **2020**, *1158*, 122341.
53. Li, S.Q.; Zhang, X.D.; Huang, Y.M. Zeolitic imidazolate framework-8 derived nanoporous carbon as an effective and recyclable adsorbent for removal of ciprofloxacin antibiotics from water. *J. Hazard. Mater.* **2017**, *321*, 711–719. [[PubMed](#)]
54. Liang, C.H.; Zhang, X.D.; Feng, P.; Chai, H.X.; Huang, Y.M. ZIF-67 derived hollow cobalt sulfide as superior adsorbent for effective adsorption removal of ciprofloxacin antibiotics. *Chem. Eng. J.* **2018**, *344*, 95–104.
55. Maya, F.; Cabello, C.P.; Frizzarin, R.M.; Estela, J.M.; Palomino, G.T.; Cerda, V. Magnetic solid-phase extraction using metal-organic frameworks (MOFs) and their derived carbons. *Trends Anal. Chem.* **2017**, *90*, 142–152.
56. Jiang, H.L.; Li, N.; Cui, L.; Wang, X.; Zhao, R.S. Recent application of magnetic solid phase extraction for food safety analysis. *Trends Anal. Chem.* **2019**, *120*, 115632.
57. Gao, Y.H.; Liu, G.; Gao, M.K.; Huang, X.D.; Xu, D.H. Recent advances and applications of magnetic metal-organic frameworks in adsorption and enrichment removal of food and environmental pollutants. *Crit. Rev. Anal. Chem.* **2020**, *50*, 472–484.
58. Pan, S.D.; Chen, X.H.; Li, X.H.; Jin, M.C. Nonderivatization method for determination of glyphosate, glufosinate, bialaphos, and their main metabolites in environmental waters based on magnetic metal-organic framework pretreatment. *J. Sep. Sci.* **2019**, *42*, 1045–1050.
59. Aghayi-Anaraki, M.; Safarifard, V. Fe<sub>3</sub>O<sub>4</sub>@MOF magnetic nanocomposites: Synthesis and applications. *Eur. J. Inorg. Chem.* **2020**, *20*, 1916–1937.
60. He, K.; Tsui, C.; Yeung, K. Flow synthesis of magnetic metal-organic frameworks. *Abstr. Pap. Am. Chem. Soc.* **2015**, *250*, 174.
61. Li, H.Q.; Sadiq, M.M.; Suzuki, K.; Falcaro, P.; Hill, A.J.; Hill, M.R. Magnetic induction framework synthesis: A general route to the controlled growth of metal-organic frameworks. *Chem. Mater.* **2017**, *29*, 6186–6190.

62. Stock, N.; Biswas, S. Synthesis of metal-organic frameworks (MOFs): Routes to various MOF topologies, morphologies, and composites. *Chem. Rev.* **2012**, *43*, 933–969.
63. Ma, Y.J.; Jiang, X.X.; Lv, Y.K. Recent advances in preparation and applications of magnetic framework composites. *Chem. Asian J.* **2019**, *14*, 3515–3530. [PubMed]
64. Abazari, R.; Mahjoub, A.R.; Molaie, S.; Ghaffarifar, F.; Ghasemi, E.; Slawin, A.M.Z.; Carpenter-Warren, C.L. The effect of different parameters under ultrasound irradiation for synthesis of new nanostructured  $\text{Fe}_3\text{O}_4$ @bio-MOF as an efficient anti-leishmanial in vitro and in vivo conditions. *Ultrason. Sonochem.* **2018**, *43*, 248–261.
65. Du, F.Y.; Qin, Q.; Deng, J.C.; Ruan, G.H.; Yang, X.Q.; Li, L.H.; Li, J.P. Magnetic metal-organic framework MIL-100(Fe) microspheres for the magnetic solid-phase extraction of trace polycyclic aromatic hydrocarbons from water samples. *J. Sep. Sci.* **2016**, *39*, 2356–2364.
66. Zhang, S.L.; Yao, W.X.; Ying, J.B.; Zhao, H.T. Polydopamine-reinforced magnetization of zeolitic imidazolate framework ZIF-7 for magnetic solid-phase extraction of polycyclic aromatic hydrocarbons from the air-water environment. *J. Chromatogr. A* **2016**, *1452*, 18–26. [PubMed]
67. Jiang, Z.W.; Dai, F.Q.; Huang, C.Z.; Li, Y.F. Facile synthesis of a  $\text{Fe}_3\text{O}_4$ /MIL-101(Fe) composite with enhanced catalytic performance. *RSC Adv.* **2016**, *6*, 86443–86446.
68. Hu, Y.L.; Huang, Z.L.; Liao, J.; Li, G.K. Chemical bonding approach for fabrication of hybrid magnetic metal-organic framework-5: High efficient adsorbents for magnetic enrichment of trace analytes. *Anal. Chem.* **2013**, *85*, 6885–6893.
69. Pena-Mendez, E.M.; Mawale, R.M.; Conde-Gonzalez, J.E.; Socas-Rodriguez, B.; Havel, J.; Ruiz-Perez, C. Metal organic framework composite, nano- $\text{Fe}_3\text{O}_4$ @Fe-(benzene-1,3,5-tricarboxylic acid), for solid phase extraction of blood lipid regulators from water. *Talanta* **2020**, *207*, 120275.
70. Li, L.N.; Chen, Y.L.; Yang, L.; Wang, Z.T.; Liu, H.W. Recent advances in applications of metal-organic frameworks for sample preparation in pharmaceutical analysis. *Coordin. Chem. Rev.* **2020**, *411*, 213235.
71. Xiang, W.L.; Gebhardt, S.; Glaser, R.; Liu, C.J. Millimeter-scale magnetic spherical metal-organic framework core-shell structured composites for recyclable catalytic applications. *Micropor. Mesopor. Mat.* **2020**, *300*, 110152.
72. Cao, C.H.; Xiao, L.; Chen, C.H.; Shi, X.W.; Cao, Q.H.; Gao, L. In situ preparation of magnetic  $\text{Fe}_3\text{O}_4$ /chitosan nanoparticles via a novel reduction-precipitation method and their application in adsorption of reactive azo dye. *Powder Technol.* **2014**, *260*, 90–97.
73. Shi, Z.N.; Xu, C.; Guan, H.; Li, L.; Fan, L.; Wang, Y.X.; Liu, L.; Meng, Q.T.; Zhang, R. Magnetic metal organic frameworks (MOFs) composite for removal of lead and malachite green in wastewater. *Colloids Surf. A* **2018**, *539*, 382–390.
74. Bhattacharjee, A.; Gumma, S.; Purkait, M.K.  $\text{Fe}_3\text{O}_4$  promoted metal organic framework MIL-100(Fe) for the controlled release of doxorubicin hydrochloride. *Micropor. Mesopor. Mat.* **2018**, *259*, 203–210.
75. Liu, J.F.; Qiu, H.J.; Zhang, F.; Li, Y. Zeolitic imidazolate framework-8 coated  $\text{Fe}_3\text{O}_4$ @ $\text{SiO}_2$  composites for magnetic solid-phase extraction of bisphenols. *New J. Chem.* **2020**, *44*, 5324–5332.
76. Zhou, Z.X.; Gao, Z.J.; Shen, H.; Li, M.Q.; He, W.T.; Su, P.; Song, J.Y.; Yang, Y. Metal-organic framework in situ post-encapsulating DNA-enzyme composites on a magnetic carrier with high stability and reusability. *ACS Appl. Mater. Inter.* **2020**, *12*, 7510–7517.
77. Xu, J.Y.; Zhai, X.P.; Gao, L.F.; Chen, P.; Zhao, M.; Yang, H.B.; Cao, D.F.; Wang, Q.; Zhang, H.L. In situ preparation of a MOF-derived magnetic carbonaceous catalyst for visible-light-driven hydrogen evolution. *RSC Adv.* **2016**, *6*, 2011–2018.
78. Wu, Y.N.; Zhou, M.M.; Li, S.; Li, Z.H.; Li, J.; Wu, A.Z.; Li, G.T.; Li, F.T.; Guan, X.H. Magnetic metal-organic frameworks: Gamma- $\text{Fe}_2\text{O}_3$ @MOFs via confined in situ pyrolysis method for drug delivery. *Small* **2014**, *10*, 2927–2936.
79. Liu, Y.; Gong, C.S.; Lin, L.S.; Zhou, Z.J.; Liu, Y.J.; Yang, Z.; Shen, Z.Y.; Yu, G.C.; Wang, Z.T.; Wang, S.; et al. Core-shell metal-organic frameworks with fluorescence switch to trigger an enhanced photodynamic therapy. *Theranostics* **2019**, *9*, 2791–2799. [PubMed]
80. Yang, Q.X.; Zhao, Q.Q.; Ren, S.S.; Lu, Q.Q.; Guo, X.M.; Chen, Z.J. Fabrication of core-shell  $\text{Fe}_3\text{O}_4$ @MIL-100(Fe) magnetic microspheres for the removal of Cr(VI) in aqueous solution. *J. Solid State Chem.* **2016**, *244*, 25–30.
81. Jiang, Z.W.; Li, Y.F. Facile synthesis of magnetic hybrid  $\text{Fe}_3\text{O}_4$ /MIL-101 via heterogeneous coprecipitation assembly for efficient adsorption of anionic dyes. *J. Taiwan Inst. Chem. E.* **2016**, *59*, 373–379.



82. Imaz, I.; Hernando, J.; Ruiz-Molina, D.; Maspoch, D. Metal–organic spheres as functional systems for guest encapsulation. *Angew. Chem. Int. Ed.* **2009**, *48*, 2325–2329.
83. Lohe, M.R.; Gedrich, K.; Freudenberger, T.; Kockrick, E.; Dellmann, T.; Kaskel, S. Heating and separation using nanomagnet-functionalized metal-organic frameworks. *Chem. Commun.* **2011**, *47*, 3075–3077.
84. Fang, Q.L.; Shen, Y.; Chen, B.L. Synthesis, decoration and properties of three-dimensional graphene-based macrostructures: A review. *Chem. Eng. J.* **2015**, *264*, 753–771.
85. Liu, Y.D.; Goebel, J.; Yin, Y.D. Templated synthesis of nanostructured materials. *Chem. Soc. Rev.* **2013**, *42*, 2610–2653.
86. Cheng, J.J.; Mallet, N.; Baaziz, W.; Ersen, O.; Gombart, E.; Alard, V.; Majimel, J.; Delville, M.H.; Treguer-Delapierre, M.; Duguet, E. Template-directed synthesis of titania nanocages with four tetrahedrally arranged open windows. *Chem. Eur. J.* **2018**, *24*, 6917–6921.
87. Yue, Y.F.; Fulvio, P.F.; Dai, S. Hierarchical metal–organic framework hybrids: Perturbation-assisted nanofusion synthesis. *Acc. Chem. Res.* **2015**, *48*, 3044.
88. Chen, C.; Feng, N.J.; Guo, Q.R.; Li, Z.; Li, X.; Ding, J.; Wang, L.; Wan, H.; Guan, G.F. Template-directed fabrication of MIL-101(Cr)/mesoporous silica composite: Layer-packed structure and enhanced performance for CO<sub>2</sub> capture. *J. Colloid Interf. Sci.* **2018**, *513*, 891–902.
89. Yang, X.J.; Liang, T.; Sun, J.X.; Zaworotko, M.J.; Chen, Y.; Cheng, P.; Zhang, Z.J. Template-directed synthesis of photocatalyst-encapsulating metal-organic frameworks with boosted photocatalytic activity. *ACS Catal.* **2019**, *9*, 7486–7493.
90. Ma, K.K.; Wang, Y.F.; Chen, Z.J.; Islamoglu, T.; Lai, C.L.; Wang, X.W.; Fei, B.; Farha, O.K.; Xin, J.H. Facile and scalable coating of metal–organic frameworks on fibrous substrates by a coordination replication method at room temperature. *ACS Appl. Mater. Inter.* **2019**, *11*, 22714–22721.
91. Ding, B.; Wang, J.; Chang, Z.; Xu, G.Y.; Hao, X.D.; Shen, L.F.; Dou, H.; Zhang, X.G. Self-sacrificial template-directed synthesis of metal–organic framework-derived porous carbon for energy-storage devices. *ChemElectroChem* **2016**, *3*, 668–674.
92. Huang, Y.F.; Zhang, L.; Ma, L.; Li, Y.; Zhong, C.L. Fe<sub>3</sub>O<sub>4</sub>@Cu/C and Fe<sub>3</sub>O<sub>4</sub>@CuO composites derived from magnetic metal–organic frameworks Fe<sub>3</sub>O<sub>4</sub>@HKUST-1 with improved peroxidase-like catalytic activity. *Catal. Lett.* **2019**, *150*, 815–825.
93. Chen, L.N.; Li, H.Q.; Yan, M.W.; Yuan, C.F.; Zhan, W.W.; Jiang, Y.Q.; Xie, Z.X.; Kuang, Q.; Zheng, L.S. Ternary alloys encapsulated within different MOFs via a self-sacrificing template process: A potential platform for the investigation of size-selective catalytic performances. *Small* **2017**, *13*, 1700683.
94. Liu, T.; Li, P.; Yao, N.; Kong, T.G.; Cheng, G.Z.; Chen, S.L.; Luo, W. Self-sacrificial template-directed vapor-phase growth of MOF assemblies and surface vulcanization for efficient water splitting. *Adv. Mater.* **2019**, *31*, 1806672.
95. Huang, L.J.; He, M.; Chen, B.B.; Hu, B. A designable magnetic MOF composite and facile coordination-based post-synthetic strategy for the enhanced removal of Hg<sup>2+</sup> from water. *J. Mater. Chem. A* **2015**, *21*, 11587–11595.
96. Chen, L.N.; Li, H.Q.; Zhan, W.W.; Cao, Z.M.; Chen, J.Y.; Jiang, Q.R.; Jiang, Y.Q.; Xie, Z.X.; Kuang, Q.; Zheng, L.S. Controlled encapsulation of flower-like Rh–Ni alloys with MOFs via tunable template dealloying for enhanced selective hydrogenation of alkyne. *ACS Appl. Mater. Inter.* **2016**, *8*, 31059–31066.
97. El-Feky, S.A.; Al-Sherbini, E.A. Fabrication and optical absorption properties of gold-silver and gold-platinum alloy nanoparticles formed by laser ablation. *Curr. Nanosci.* **2013**, *9*, 192–196.
98. Kumar, M.K.; Do, J.Y.; Reddy, A.K.; Kang, M. Natural solar light-driven preparation of plasmonic resonance-based alloy and core-shell catalyst for sustainable enhanced hydrogen production: Green approach and characterization. *Appl. Catal. B Environ.* **2018**, *231*, 137–150.
99. Li, L.; Wang, Y.Z.; Wang, X.X.; Song, K.K.; Jian, X.D.; Qan, P.; Bai, Y.; Su, Y.J. Size and stoichiometry effect of FePt bimetal nanoparticle catalyst for CO oxidation: A DFT study. *J. Phys. Chem. C* **2020**, *124*, 8706–8715.
100. Zhou, L.; Mao, J.Y.; Ren, Y.; Yang, J.Q.; Zhang, S.R.; Zhou, Y.; Liao, Q.F.; Zeng, Y.J.; Shan, H.Q.; Xu, Z.X.; et al. Biological spiking synapse constructed from solution processed bimetal core-shell nanoparticle based composites. *Small* **2018**, *14*, 1800288.
101. Ma, R.Q.; Jiang, H.Q.; Wang, C.; Zhao, C.B.; Deng, H.X. Multivariate MOFs for laser writing of alloy nanoparticle patterns. *Chem. Commun.* **2020**, *56*, 2715–2718.
102. Yuan, L.; Li, X.Y.; Ge, L.M.; Jia, X.Q.; Lei, J.F.; Mu, C.D.; Li, D.F. Emulsion template method for the fabrication of gelatin-based scaffold with controllable pore structure. *ACS Appl. Mater. Inter.* **2019**, *11*, 269–277.

103. Chen, J.; Azhar, U.; Wang, Y.K.; Liang, J.H.; Geng, B. Preparation of fluoropolymer materials with different porous morphologies by an emulsion template method using supercritical carbon dioxide as a medium. *RSC Adv.* **2019**, *9*, 11331–11340.
104. Kim, H.; Lah, M.S. Templated and template-free fabrication strategies for zero-dimensional hollow MOF superstructures. *Dalton Trans.* **2017**, *46*, 6146–6158.
105. Zhu, H.; Zhang, Q.; Zhu, S.P. MOFs via transient pickering emulsion template. *Adv. Mater. Interfaces* **2016**, *3*. [[CrossRef](#)]
106. Chen, L.; Zhang, M.J.; Zhang, S.Y.; Shi, L.; Yang, Y.M.; Liu, Z.; Ju, X.J.; Xie, R.; Wang, W.; Chu, L.Y. Simple and continuous fabrication of self-propelled micromotors with photocatalytic metal–organic frameworks for enhanced synergistic environmental remediation. *ACS Appl. Mater. Inter.* **2020**, *12*, 35120–35131.
107. Jin, P.; Tan, W.L.; Huo, J.; Liu, T.T.; Liang, Y.; Wang, S.Y.; Bradshaw, D. Hierarchically porous MOF/polymer composites via interfacial nanoassembly and emulsion polymerization. *J. Mater. Chem. A* **2018**, *6*, 20473–20479.
108. Chen, L.J.; Ding, X.; Huo, J.; El Hankari, S.; Bradshaw, D. Facile synthesis of magnetic macroporous polymer/MOF composites as separable catalysts. *J. Mater. Sci.* **2019**, *54*, 370–382.
109. Zhu, H.; Zhang, Q.; Zhu, S.P. Assembly of a metal–organic framework into 3D hierarchical porous monoliths using a pickering high internal phase emulsion template. *Chem. Eur. J.* **2016**, *22*, 8751–8755.
110. Iost, R.M.; Crespiello, F.N. Layer-by-layer self-assembly and electrochemistry: Applications in biosensing and bioelectronics. *Biosens. Bioelectron.* **2012**, *31*, 1–10.
111. Ariga, K.; Yamauchi, Y.; Rydzek, G.; Ji, Q.M.; Yonamine, Y.; Wu, K.C.W.; Hill, J.P. Layer-by-layer nanoarchitectonics: Invention, innovation, and evolution. *Chem. Lett.* **2014**, *45*, 36–68.
112. Rawtani, D.; Agrawal, Y.K. Emerging strategies and applications of layer-by-layer self-assembly. *Nanobiomedicine* **2014**, *1*, 8.
113. Ohhashi, T.; Tsuruoka, T.; Nawafune, H.; Akamatsu, K. Controlled synthesis of metal-organic framework films on metal nanoparticles by the versatile layer-by-layer assembly approach. *Tetrahedron* **2014**, *39*, 153–156.
114. Wang, N.X.; Liu, T.J.; Shen, H.P.; Ji, S.L.; Li, J.R.; Zhang, R. Ceramic tubular mof hybrid membrane fabricated through in situ layer-by-layer self-assembly for nanofiltration. *AIChE J.* **2016**, *62*, 538–546.
115. So, M.C.; Jin, S.; Son, H.J.; Wiederrecht, G.P.; Farha, O.K.; Hupp, J.T. Layer-by-layer fabrication of oriented porous thin films based on porphyrin-containing metal-organic frameworks. *J. Am. Chem. Soc.* **2013**, *135*, 15698.
116. Liu, S.C.; Ma, Y.; Gao, L.; Pan, J.M. pH-responsive magnetic metal-organic framework nanocomposite: A smart porous adsorbent for highly specific enrichment of c is-diol containing luteolin. *Chem. Eng. J.* **2018**, *341*. [[CrossRef](#)]
117. Liu, J.; Yang, F.; Zhang, Q.L.; Chen, W.; Gu, Y.F.; Chen, Q. Construction of hierarchical Fe<sub>3</sub>O<sub>4</sub>@HKUST-1/MIL-100(Fe) microparticles with large surface area through layer-by-layer deposition and epitaxial growth methods inorganic chemistry. *Inorg. Chem.* **2019**, *58*, 3564–3568.
118. Miao, Z.; Shu, X.; Ramella, D. Synthesis of a Fe<sub>3</sub>O<sub>4</sub>@P<sub>4</sub>VP@metal–organic framework core–shell structure and studies of its aerobic oxidation reactivity. *RSC Adv.* **2017**, *7*, 2773–2779.
119. Rice, C.M.; Davis, Z.H.; McKay, D.; Bignami, G.P.M.; Chitac, R.G.; Dawson, D.M.; Morris, R.E.; Ashbrook, S.E. Following the unusual breathing behaviour of (17)O-enriched mixed-metal (Al,Ga)-MIL-53 using NMR crystallography. *Phys. Chem. Chem. Phys.* **2020**, *22*, 14514–14526. [[PubMed](#)]
120. Lee, Y.R.; Jang, M.S.; Cho, B.Y.; Kwon, H.J.; Kim, S.; Ahn, W.S. ZIF-8: A comparison of synthesis methods. *Chem. Eng. J.* **2015**, *271*, 276–280.
121. Tan, P.; Xie, X.Y.; Liu, X.Q.; Pan, T.; Gu, C.; Chen, P.F.; Zhou, J.Y.; Pan, Y.C.; Sun, L.B. Fabrication of magnetically responsive HKUST-1/Fe<sub>3</sub>O<sub>4</sub> composites by dry gel conversion for deep desulfurization and denitrogenation. *J. Hazard. Mater.* **2017**, *321*, 344–352.
122. Rocio-Bautista, P.; Pino, V.; Ayala, J.H.; Pasan, J.; Ruiz-Perez, C.; Afonso, A.M. A magnetic-based dispersive micro-solid-phase extraction method using the metal-organic framework HKUST-1 and ultra-high-performance liquid chromatography with fluorescence detection for determining polycyclic aromatic hydrocarbons in waters and fruit tea infusions. *J. Chromatogr. A* **2016**, *1436*, 42–50.
123. Gokpinar, S.; Diment, T.; Janiak, C. Environmentally benign dry-gel conversions of Zr-based UiO metal–organic frameworks with high yield and the possibility of solvent re-use. *Dalton Trans.* **2017**, *46*, 9895–9900. [[PubMed](#)]

124. Bellusci, M.; Guglielmi, P.; Masi, A.; Padella, F.; Singh, G.; Yaacoub, N.; Peddis, D.; Secci, D. Magnetic metal–organic framework composite by fast and facile mechanochemical process. *Inorg. Chem.* **2018**, *57*, 1806–1814.
125. Zhang, R.; Tao, C.A.; Chen, R.; Wu, L.F.; Zou, X.X.; Wang, J.F. Ultrafast synthesis of Ni-MOF in one minute by ball milling. *Nanomaterials* **2018**, *8*, 1067.
126. Sun, J.K.; Chen, C.; Cai, L.X.; Ren, C.X.; Tan, B.; Zhang, J. Mechanical grinding of a single-crystalline metal-organic framework triggered emission with tunable violet-to-orange luminescence. *Chem. Commun.* **2014**, *50*, 15956–15959.
127. Park, K.M.; Kim, H.; Murray, J.; Koo, J.; Kim, K. A facile preparation method for nanosized MOFs as a multifunctional material for cellular imaging and drug delivery. *Supramol. Chem.* **2016**, *29*, 441–445.
128. Zhou, J.W.; Zou, X.M.; Song, S.H.; Chen, G.H. Quantum dots applied to methodology on detection of pesticide and veterinary drug residues. *J. Agric. Food Chem.* **2018**, *66*, 1307–1319.
129. Kumar, A.; Bhattacharyya, A.; Shinde, R.; Dhanshetty, M.; Elliott, C.T.; Banerjee, K. Development and validation of a multiresidue method for pesticides and selected veterinary drugs in animal feed using liquid- and gas chromatography with tandem mass spectrometry. *J. Chromatogr. A* **2020**, *1627*, 461416.
130. Ninga, E.; Sapozhnikova, Y.; Lehotay, S.J.; Lightfield, A.R.; Monteiro, S.H. High-throughput mega-method for the analysis of pesticides, veterinary drugs, and environmental contaminants by ultra-high-performance liquid chromatography-tandem mass spectrometry and robotic mini-solid-phase extraction cleanup + low-pressure gas chromatography-tandem mass spectrometry, part 2: Catfish. *J. Agric. Food Chem.* **2020**. [[CrossRef](#)]
131. Jadhav, M.R.; Pudale, A.; Raut, P.; Utture, S.; Shabeer, T.P.A.; Banerjee, K. A unified approach for high-throughput quantitative analysis of the residues of multi-class veterinary drugs and pesticides in bovine milk using LC-MS/MS and GC-MS/MS. *Food Chem.* **2019**, *272*, 292–305.
132. Byzova, N.A.; Serchenya, T.S.; Vashkevich, I.I.; Zherdev, A.V.; Sviridov, O.V.; Dzantiev, B.B. Lateral flow immunoassay for rapid qualitative and quantitative control of the veterinary drug bacitracin in milk. *Microchem. J.* **2020**, *156*, 104884.
133. Li, B.; Zhou, X.Z.; Cheng, F.S.; Wei, X.J.; Wang, W.W.; Zhang, J.Y. Research progress on analysis and detection techniques of veterinary drug residues in animal foods. *Agric. Biotechnol.* **2019**, *8*, 60–69.
134. Wu, Z.L.; Wang, Y.P.; Xiong, Z.K.; Ao, Z.M.; Pu, S.Y.; Yao, G.; Lai, B. Core-shell magnetic Fe<sub>3</sub>O<sub>4</sub>@Zn/Co-ZIFs to activate peroxymonosulfate for highly efficient degradation of carbamazepine. *Appl. Catal. B Environ.* **2020**, *277*, 119136.
135. Yi, L.H.; Yan, Y.H.; Tang, K.Q.; Ding, C.F. Facile preparation of polymer grafted ZIF-8 modified magnetic nanosphere for effective identification and capture phosphorylated peptides and glycosylated peptides. *Anal. Methods UK* **2020**, *38*. [[CrossRef](#)]
136. Ghiasi, A.; Malekpour, A.; Mahpishanian, S. Metal-organic framework MIL101 (Cr)-NH<sub>2</sub> functionalized magnetic graphene oxide for ultrasonic-assisted magnetic solid phase extraction of neonicotinoid insecticides from fruit and water samples. *Talanta* **2020**, *217*, 121120.
137. Shakourian, M.; Yamini, Y.; Safari, M. Facile magnetization of metal–organic framework TMU-6 for magnetic solid-phase extraction of organophosphorus pesticides in water and rice samples. *Talanta* **2020**, *218*, 121139.
138. Li, D.D.; He, M.; Chen, B.B.; Hu, B. Metal organic frameworks-derived magnetic nanoporous carbon for preconcentration of organophosphorus pesticides from fruit samples followed by gas chromatography-flame photometric detection. *J. Chromatogr. A* **2019**, *1583*, 19–27.
139. Yang, J.H.; Cui, C.X.; Qu, L.B.; Chen, J.; Zhou, X.M.; Zhang, Y.P. Preparation of a monolithic magnetic stir bar for the determination of sulfonylurea herbicides coupled with HPLC. *Microchem. J.* **2018**, *141*, 369–376.
140. Yamini, Y.; Safari, M. Magnetic Zink-based metal organic framework as advance and recyclable adsorbent for the extraction of trace pyrethroids. *Microchem. J.* **2019**, *146*, 134–141.
141. Gordi, Z.; Ghorbani, M.; Khakhiyani, M.A. Adsorptive removal of enrofloxacin with magnetic functionalized graphene oxide@ metal-organic frameworks employing D-optimal mixture design. *Water Environ. Res.* **2020**, *92*, 1935–1947.
142. Lian, L.L.; Zhang, X.Y.; Hao, J.; Lv, J.Y.; Wang, X.Y.; Zhu, B.; Lou, D.W. Magnetic solid-phase extraction of fluoroquinolones from water samples using titanium-based metal-organic framework functionalized magnetic microspheres. *J. Chromatogr. A* **2018**, *1579*, 1–8.

143. Mendiola-Alvarez, S.Y.; Palomino, G.T.; Guzman-Mar, J.; Hernandez-Ramirez, A.; Hinojosa-Reyes, L.; Cabello, C.P. Magnetic porous carbons derived from cobalt(II)-based metal-organic frameworks for the solid-phase extraction of sulfonamides. *Dalton Trans.* **2020**, *49*, 8959–8966.
144. Bagheri, A.R.; Ghaedi, M. Magnetic metal organic framework for pre-concentration of ampicillin from cow milk samples. *J. Pharm. Anal.* **2020**, *10*, 365–375.
145. Wang, Q.Q.; Liu, W.H.; Hao, L.; Wang, C.; Wu, Q.H.; Wang, Z. Fabrication of magnetic porous organic framework for effective enrichment and assay of nitroimidazoles in chicken meat. *Food Chem.* **2020**, *332*, 127427.
146. Wang, X.M.; Ma, X.M.; Huang, P.F.; Wang, J.; Du, T.T.; Du, X.Z.; Lu, X.Q. Magnetic Cu-MOFs embedded within graphene oxide nanocomposites for enhanced preconcentration of benzenoid-containing insecticides. *Talanta* **2018**, *181*, 112–117.
147. Du, F.Y.; Sun, L.S.; Tan, W.; Wei, Z.Y.; Nie, H.G.; Huang, Z.J.; Ruan, G.H.; Li, J.P. Magnetic stir cake sorptive extraction of trace tetracycline antibiotics in food samples: Preparation of metal-organic framework-embedded polyHIPE monolithic composites, validation and application. *Anal. Bioanal. Chem.* **2019**, *411*, 2239–2248. [PubMed]
148. Liang, L.; Wang, X.H.; Sun, Y.; Ma, P.Y.; Li, X.P.; Piao, H.L.; Jiang, Y.X.; Song, D.Q. Magnetic solid-phase extraction of triazine herbicides from rice using metal-organic framework MIL-101(Cr) functionalized magnetic particles. *Talanta* **2018**, *179*, 512–519.
149. Duo, H.X.; Lu, X.F.; Wang, S.; Wang, L.C.; Guo, Y.; Liang, X.J. Synthesis of magnetic metal-organic framework composites, Fe<sub>3</sub>O<sub>4</sub>-NH<sub>2</sub>@MOF-235, for the magnetic solid-phase extraction of benzoylurea insecticides from honey, fruit juice and tap water samples. *New J. Chem.* **2019**, *43*, 12563–12569.
150. D’Mello, J.P.F.; Macdonald, A.M.C.; Postel, D.; Dijksma, W.T.P.; Dujardin, A.; Placinta, C.M. Pesticide use and mycotoxin production in fusarium and aspergillus phytopathogens. *Eur. J. Plant Pathol.* **1998**, *104*, 741–751.
151. Bata, A.; Lasztity, R. Detoxification of mycotoxin-contaminated food and feed by microorganisms. *Trends Food Sci. Technol.* **1999**, *10*, 223–228.
152. Kabak, B.; Dobson, A.D.W.; Var, I. Strategies to prevent mycotoxin contamination of food and animal feed: A review. *Crit. Rev. Food Sci.* **2006**, *46*, 593–619.
153. Xu, B.J.; Jia, X.Q.; Gu, L.J.; Sung, C.K. Review on the qualitative and quantitative analysis of the mycotoxin citrinin. *Food Control* **2006**, *17*, 271–285.
154. Li, C.Y.; Liu, J.M.; Wang, Z.H.; Lv, S.W.; Zhao, N.; Wang, S. Integration of Fe<sub>3</sub>O<sub>4</sub>@UiO-66-NH<sub>2</sub>@MON core-shell structured adsorbents for specific preconcentration and sensitive determination of aflatoxins against complex sample matrix. *J. Hazard. Mater.* **2020**, *384*, 121348.
155. Durmus, Z.; Kurt, B.Z.; Gazioglu, I.; Sevgi, E.; Hancer, C.K. Spectrofluorimetric determination of Aflatoxin B1 in winter herbal teas via magnetic solid phase extraction method by using metal-organic framework (MOF) hybrid structures anchored with magnetic nanoparticles. *Appl. Organomet. Chem.* **2020**, *34*, e5375.
156. Sabeghi, M.B.; Ghasempour, H.R.; Koohi, M.K.; Karimi, N. Synthesis and application of a novel functionalized magnetic MIL-101(Cr) nanocomposite for determination of aflatoxins in pistachio samples. *Res. Chem. Intermed.* **2020**, *46*, 4099–4111.
157. Hu, S.S.; Ouyang, W.J.; Guo, L.H.; Lin, Z.Y.; Jiang, X.H.; Qiu, B.; Chen, G.N. Facile synthesis of Fe<sub>3</sub>O<sub>4</sub>/g-C<sub>3</sub>N<sub>4</sub>/HKUST-1 composites as a novel biosensor platform for ochratoxin A. *Biosens. Bioelectron.* **2017**, *92*, 718–723.
158. Huang, C.H.; Qiao, X.Z.; Sun, W.M.; Chen, H.; Chen, X.Y.; Zhang, L.; Wang, T. Effective extraction of domoic acid from seafood based on postsynthetic-modified magnetic zeolite imidazolate framework-8 particles. *Anal. Chem.* **2019**, *91*, 2418–2424.
159. Hallegraeff, G.M. Ocean climate change, phytoplankton community responses, and harmful algal blooms: A formidable predictive challenge. *J. Phycol.* **2010**, *46*, 220–235.
160. Christensen, V.G.; Khan, E. Freshwater neurotoxins and concerns for human, animal, and ecosystem health: A review of anatoxin-a and saxitoxin. *Sci. Total Environ.* **2020**, *736*, 139515. [PubMed]
161. Li, Z.J.; Gong, C.C.; Huo, P.P.; Deng, C.H.; Pu, S.Z. Synthesis of magnetic core-shell Fe<sub>3</sub>O<sub>4</sub>@PDA@Cu-MOFs composites for enrichment of microcystin-LR by MALDI-TOF MS analysis. *RSC Adv.* **2020**, *10*, 29061–29067.
162. Piatkowska, M.; Jedziniak, P.; Zmudzki, J. Multiresidue method for the simultaneous determination of veterinary medicinal products, feed additives and illegal dyes in eggs using liquid chromatography-tandem mass spectrometry. *Food Chem.* **2016**, *197*, 571–580.



163. Zhang, T.; Li, Z.H.; Li, Y.; Liu, N.; Liu, K. A combined raman and DFT study of illegal food additives methenamine. *J. Insp. Quar.* **2013**, *23*, 36–47.
164. Zhang, N.; Huang, C.H.; Tong, P.; Feng, Z.M.; Wu, X.P.; Zhang, L. Moisture stable Ni-Zn MOF/g-C<sub>3</sub>N<sub>4</sub> nanoflowers: A highly efficient adsorbent for solid-phase microextraction of PAHs. *J. Chromatogr. A* **2015**, *1556*, 37–46.
165. Ventura, K.; Arrieta, R.A.; Marcos-Hernandez, M.; Jabbari, V.; Powell, C.D.; Turley, R.; Lounsbury, A.W.; Zimmerman, J.B.; Gardea-Torresdey, J.; Wong, M.S.; et al. Superparamagnetic MOF@GO Ni and Co based hybrid nanocomposites as efficient water pollutant adsorbents. *Sci. Total Environ.* **2020**, *738*, 139213.
166. Yuan, Y.; Zheng, X.W.; Lin, H.Z.; Li, Y.Y.; Yang, M.; Liu, X.L.; Deng, C.H.; Fan, Z.Q. Development of a hydrophilic magnetic amino-functionalized metal-organic framework for the highly efficient enrichment of trace bisphenols in river water samples. *Talanta* **2020**, *221*, 120713.
167. Lu, Y.J.; Wang, B.C.; Yan, Y.H.; Liang, H.Z. Location-controlled synthesis of hydrophilic magnetic metal-organic frameworks for highly efficient recognition of phthalates in beverages. *Chem. Select* **2018**, *3*, 12440–12445.
168. Yamini, Y.; Safari, M.; Morsali, A.; Safarifard, V. Magnetic frame work composite as an efficient sorbent for magnetic solid-phase extraction of plasticizer compounds. *J. Chromatogr. A* **2018**, *1570*, 38–46.
169. Shi, X.R.; Chen, X.L.; Hao, Y.L.; Li, L.; Xu, H.J.; Wang, M.M. Magnetic metal-organic frameworks for fast and efficient solid-phase extraction of six Sudan dyes in tomato sauce. *J. Chromatogr. B* **2018**, *1086*, 146–152.
170. Zhou, Z.H.; Fu, Y.Q.; Qin, Q.; Lu, X.; Shi, X.Z.; Zhao, C.X.; Xu, G.W. Synthesis of magnetic mesoporous metal-organic framework-5 for the effective enrichment of malachite green and crystal violet in fish samples. *J Chromatogr. B* **2018**, *1560*, 19–25.
171. Zhong, X.; Chen, Z.W.; Li, Y.Y.; Ding, K.B.; Liu, W.S.; Liu, Y.; Yuan, Y.Q.; Zhang, M.Y.; Baker, A.J.M.; Yang, W.J.; et al. Factors influencing heavy metal availability and risk assessment of soils at typical metal mines in Eastern China. *J. Hazard. Mater.* **2020**, *400*, 123289.
172. Leng, J.S.; Gao, X.M.; Wang, L.X. Hazards of heavy metal pollution in food and progress in study on analysis technologies. *Farm Prod. Process.* **2015**, *23*, 50–53.
173. Ma, J.F.; Chen, Y.P.; Antoniadis, V.; Wang, K.B.; Huang, Y.Z.; Tian, H.W. Assessment of heavy metal(loid)s contamination risk and grain nutritional quality in organic waste-amended soil. *J. Hazard. Mater.* **2020**, *399*, 123095.
174. Zhang, X.Y.; Zhong, T.Y.; Liu, L.; Ouyang, X.Y. Impact of soil heavy metal pollution on food safety in China. *PLoS ONE* **2015**, *10*, e0135182.
175. Li, J.; Yue, X.L.; Cheng, Y.H.; Bao, L.Y.; Yu, Y.Y.; Ren, X.M. Research progress on heavy metal pollution in soil and the effect on heavy metal residues in vegetables. *J. Food Saf. Food Qual.* **2019**, *10*, 5299–5305.
176. Zhang, H.K.; Wang, Z.Y.; Cai, S.S. Research progress of ICP-MS in determination of heavy metal elements in food and relative products. *Food Res. Dev.* **2016**, *37*, 195–200.
177. Manousi, N.; Giannakoudakis, D.A.; Rosenberg, E.; Zachariadis, G.A. Extraction of metal ions with metal-organic frameworks. *Molecules* **2019**, *24*, 4605.
178. Abolhasani, J.; Khanmiri, R.H.; Babazadeh, M.; Ghorbani-Kalhor, E.; Edjlali, L.; Hassanpour, A. Determination of Hg(II) ions in sea food samples after extraction and preconcentration by novel Fe<sub>3</sub>O<sub>4</sub>@SiO<sub>2</sub>@polythiophene magnetic nanocomposite. *Environ. Monit. Assess.* **2015**, *187*, 554.
179. Ghorbani-Kalhor, E.; Hosseinzadeh-Khanmiri, R.; Abolhasani, J.; Babazadeh, M.; Hassanpour, A. Determination of mercury(II) ions in seafood samples after extraction and preconcentration by a novel functionalized magnetic metal-organic framework nanocomposite. *J. Sep. Sci.* **2015**, *38*, 1179–1186.
180. Yang, J.; Dong, X.; Zhen, X.T.; Chen, Y.; Wang, Y.; Wang, Q.Y.; Hu, Y.H.; Xie, T.; Wang, S.L.; Cao, J. Metal organic framework assisted in situ complexation for miniaturized solid phase extraction of organic mercury in fish and *Dendrobium officinale*. *Talanta* **2020**, *209*, 120598. [PubMed]
181. Esmaeilzadeh, M. Ultrasound-assisted dispersive magnetic solid phase extraction based on metal-organic framework/1-(2-pyridylazo)-2-naphthol modified magnetite nanoparticle composites for speciation analysis of inorganic tin. *New J. Chem.* **2019**, *43*, 4929–4936.
182. Hassanpour, A.; Hosseinzadeh-Khanmiri, R.; Babazadeh, M.; Abolhasani, J.; Ghorbani-Kalhor, E. Determination of heavy metal ions in vegetable samples using a magnetic metal-organic framework nanocomposite sorbent. *Food Addit. Contam. A* **2015**, *32*, 725–736.

183. Ghorbani-Kalhor, E.; Hosseinzadeh-Khanmiri, R.; Babazadeh, M.; Abolhasani, J.; Hassanpour, A. Synthesis and application of a novel magnetic metal-organic framework nanocomposite for determination of Cd, Pb, and Zn in baby food samples. *Can. J. Chem.* **2015**, *93*, 518–525.
184. Mahmoud, M.E.; Amira, M.F.; Selim, S.M.; Mohamed, A.K. Amino-decorated magnetic metal-organic framework as a potential novel platform for selective removal of chromium (VI), cadmium (II) and lead (II). *J. Hazard. Mater.* **2020**, *381*, 120979.
185. Mehraban, M.; Manoochehri, M.; Taromi, F.A. Trace amount determination of Cd(II), Pb(II) and Ni(II) ions in agricultural and seafood samples after magnetic solid phase extraction by MIL-101(Cr)/phenylthiosemicarbazide-functionalized magnetite nanoparticle composite. *New J. Chem.* **2018**, *42*, 17636–17643.
186. Marieeswaran, M.; Panneerselvam, P. A magnetic nanoscale metal-organic framework (MNMOF) as a viable fluorescence quencher material for ssDNA and for the detection of mercury ions via a novel quenching-quenching mechanism. *RSC Adv.* **2020**, *10*, 3705–3714.
187. Taghizadeh, M.; Asgharinezhad, A.A.; Pooladi, M.; Barzin, M.; Abbaszadeh, A.; Tadjarodi, A. A novel magnetic metal organic framework nanocomposite for extraction and preconcentration of heavy metal ions, and its optimization via experimental design methodology. *Microchim. Acta* **2013**, *180*, 1073–1084.
188. Li, W.T.; Shi, W.; Hu, Z.J.; Yang, T.; Chen, M.L.; Zhao, B.; Wang, J.H. Fabrication of magnetic Fe<sub>3</sub>O<sub>4</sub>@metal organic framework@covalent organic framework composite and its selective separation of trace copper. *Appl. Surf. Sci.* **2020**, *530*, 147254.
189. Liu, G.Y.; Li, L.Y.; Gao, Y.H.; Gao, M.K.; Huang, X.D.; Lv, J.; Xu, D.H. A beta-cyclodextrin- functionalized magnetic metal organic framework for efficient extraction and determination of prochloraz and triazole fungicides in vegetables samples. *Ecotox. Environ. Saf.* **2019**, *183*, 109546.
190. Xia, L.; Liu, L.J.; Lv, X.X.; Qu, F.; Li, G.L.; You, J.M. Towards the determination of sulfonamides in meat samples: A magnetic and mesoporous metal-organic framework as an efficient sorbent for magnetic solid phase extraction combined with high-performance liquid chromatography. *J. Chromatogr. A* **2017**, *1500*, 24–31.

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