

Review

The Mechanisms of Plastic Food-Packaging Monomers' Migration into Food Matrix and the Implications on Human Health

Celia Muzeza ^{1,2}, Veronica Ngole-Jeme ² and Titus Alfred Makudali Msagati ^{1,*} 

- ¹ Institute for Nanotechnology and Water Sustainability (iNanoWS), College of Science, Engineering and Technology, University of South Africa, Science Campus, Roodepoort, Johannesburg 1709, South Africa
- ² Department of Environmental Science, College of Agriculture and Environmental Sciences, University of South Africa, Science Campus, Roodepoort, Johannesburg 1709, South Africa; ngolevm@unisa.ac.za
- * Correspondence: msagatam@unisa.ac.za; Tel.: +27-11-670-9482

Abstract: The development of packaging technology has become a crucial part of the food industry in today's modern societies, which are characterized by technological advancements, industrialization, densely populated cities, and scientific advancements that have increased food production over the past 50 years despite the lack of agricultural land. Various types of food-packaging materials are utilized, with plastic being the most versatile. However, there are certain concerns with regards to the usage of plastic packaging because of unreacted monomers' potential migration from the polymer packaging to the food. The magnitude of monomer migration depends on numerous aspects, including the monomer chemistry, type of plastic packaging, physical-chemical parameters such as the temperature and pH, and food chemistry. The major concern for the presence of packaging monomers in food is that some monomers are endocrine-disrupting compounds (EDCs) with a capability to interfere with the functioning of vital hormonal systems in the human body. For this reason, different countries have resolved to enforce guidelines and regulations for packaging monomers in food. Additionally, many countries have introduced migration testing procedures and safe limits for packaging monomer migration into food. However, to date, several research studies have reported levels of monomer migration above the set migration limits due to leaching from the food-packaging materials into the food. This raises concerns regarding possible health effects on consumers. This paper provides a critical review on plastic food-contact materials' monomer migration, including that from biodegradable plastic packaging, the monomer migration mechanisms, the monomer migration chemistry, the key factors that affect the migration process, and the associated potential EDC human health risks linked to monomers' presence in food. The aim is to contribute to the existing knowledge and understanding of plastic food-packaging monomer migration.

Keywords: food-packaging material (FPM); monomer migration; endocrine-disrupting compounds (EDCs)



Citation: Muzeza, C.; Ngole-Jeme, V.; Msagati, T.A.M. The Mechanisms of Plastic Food-Packaging Monomers' Migration into Food Matrix and the Implications on Human Health. *Foods* **2023**, *12*, 3364. <https://doi.org/10.3390/foods12183364>

Academic Editors: Jong-Whan Rhim and Wanli Zhang

Received: 28 July 2023

Revised: 19 August 2023

Accepted: 23 August 2023

Published: 7 September 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

In the past century, humans have mainly been sustained by locally grown seasonal foods that could meet their food demand within the food shelf life [1]. However, with the current technological advancements coupled with high levels of industrialization and better standards of living, which have catalysed the formation of large cities that are highly populated, there is virtually no space to grow food crop [1]. The scientific advancements in the last fifty years have nevertheless contributed to an increase in and variety in world food production [2], but demand remains high! Globally, these and other factors have resulted in the need to transport and store a variety of foods over long distances to consumers and for long times, respectively, taking a longer time beyond the storage life of the foods as a result. Due to the fact that the chemicals within food may be subjected to various environmental conditions, including oxygen (O₂), water vapour

(H₂O), and light, during transportation and storage, which possibly leads to microbial contamination or a loss of valuable properties, such as nutrition, colour, texture, and edibility [3], modern society households mainly depend on food refrigeration for various plastic food-packaging types as a method of food preservation. Industries also rely on numerous modern preservation techniques (high-pressure technology, irradiation, and hurdle technology) [3,4] and traditional preservation techniques, including packaging to meet the current demands of economic preservation and keeping food stable and safe to maintain the food quality [5] for consumer satisfaction. Therefore, in the food industry, food packaging serves as an indispensable multifunctional element and a sector currently representing a dynamic part of most economies, which currently contributes significantly to the Gross National Domestic Product (GDP) [6–9]. Research reports further speculate that, due to population expansion, the world's food supply will need to expand by 50% by 2050, which will consequently trigger a significant demand for food-contact materials [10,11]. In the developed world, the key driving factors, such as the increased plastic recycling infrastructure, the global population growth, a rise in feeding with processed and take-away foods due to consumers' busy lifestyles, and numerous other factors, increase the demand for convenience foods, which contributes positively towards the growth of the food industry [9]. In countries with emerging economies, South Africa, for instance, is a giant polymer producer of different food-packaging types. For the projected period of 2017–2027, the South African packaging market is anticipated to convert to an extraordinary growth region with a compound annual growth rate (CAGR) of 7.8% [12]. Plastic consumption per capita is particularly being projected to increase due to urbanization, urban–rural migration, and an increase in middle-income households [13]. As in the rest of the world, their modern food packaging makes use of ceramics, glass, metal, paper, paperboard, wood, and plastic material types to package a variety of retail and domestic food products [14–17]. Plastic dominates the packaging industry because it has numerous food-packaging advantages compared to its disadvantages, which have enormously contributed to its preference as a food-contact material to package various food items (Table 1). As such, it accounts for about 52% of most local markets [18]. Polypropylene (PP), polyethylene (PE), polyvinyl chloride (PVC), polystyrene (PS), polyethylene terephthalate (PET), and polyurethane (PU) are the polymer types globally dominating the plastic food-packaging industry [15,19,20]. The word 'plastic' is normally utilized to describe materials synthesized through the addition, condensation, or cross-linking polymerization of monomer units [21,22]. Plastics are present in various rigid and flexible forms to which, in addition to monomers, additives, for instance, plasticizers, adhesives, coatings and solvents, antioxidants, thermal and light protectants, and graphic information, are added [23] to ensure the characteristics needed for their function [16,24,25]. Heat and pressure is then applied to mould the polymers and to obtain the required final products and shapes, such as films, trays, and bottles [26].

Table 1. Applications, pros, and cons of various food-packaging materials.

Type of Packaging	Applications (Types of Foods)	Advantages	Disadvantages	References
Plastics	Fast foods Solid products, such as pasta, rice, biscuits, bread, and sugar Liquid products, such as concentrate juices, oils, and methylated spirits	<ul style="list-style-type: none"> - Inexpensive materials - Thermostability and microwaveability functional advantages - Optical properties - Unlimited sizes and shapes - Recyclable - Light weight - Strong - Oil and chemical resistance - Excellent barrier properties against gas and water vapour - Thermal stability and electrical insulation properties - Easily reused 	<ul style="list-style-type: none"> - Residual monomer and chemical migration - Variable permeability to gases and light 	[27]

However, despite the significant different roles that plastic food-packaging materials fulfil both domestically and within the food industry sector, in their natural state, plastic food-contact materials are limited in their ability to provide the required mechanical and barrier properties. This is because, without additives, for instance, plastic food packaging is limited in its food-packaging role. Additionally, most importantly, the scientific research evidence reveals that the chemical substances utilized during polymer synthesis, including the main building blocks (monomers), leach throughout the plastic product's life cycle [28–30]. More concerning is that, compared to other packaging material types, research confirms that chemical migration is more likely to occur in plastic packaging [31]. Migration describes the mass transfer of chemical substances from a higher-concentration region (the food-contact side) to a lower-concentration region (usually the food) until equilibrium is reached [32]. Monomers have been implicated as endocrine-disrupting compounds (EDCs) linked to serious human health problems that compromise consumer health, especially the safety of pregnant mothers and children, who make up the vulnerable group in the population [11,33–44]. Up to 906 chemicals have been linked with plastic food packaging of which 63 are classified as human health hazards; 68 are classified as environmental hazards; 7 are classified as persistent, bioaccumulative, and toxic; 15 are classified as endocrine-disrupting compounds (EDCs); and 34 are classified as potential EDCs [23]. However, these known values of the different chemical categories in plastic food packaging are but an insignificant proportion given that about 10,000 chemicals show potential capabilities of migration from plastics into food when subjected to various physicochemical conditions [23,32,45,46] during processing, transport, storage, and food preparation. However, to date, more than 2000 substances lack toxicological and detailed descriptions of their scope of use [23] due to numerous reasons, including the prevailing limitations in structure elucidation as a result of the lengthy modern analytical procedures used for the detection of monomers [47]. Additionally, the research contributions, which are mostly from the developed world, are biased on a few commonly known hazardous substances occurring in plastic food packaging, such as Bisphenol A and styrene (monomers) and phthalates (plasticizer additives) [48]. This is primarily due to the fact that it is difficult to find comprehensive information on these chemicals, including plastic monomers, in the public [20,28]. It is important that all the chemicals, including monomers, in food packaging are well accounted for so that their potential harm to humans is well understood and reduced. This is especially true because consumers interact with plastic packaging daily. The low amount of accounted data hence suggests that humans are susceptible to unknown harmful food-packaging chemicals daily. Additionally, there is scanty information on the treatability and level of the human health and environmental toxicity of the numerous known and unknown plastic food-contact chemicals, including monomers. Due to the fact that most of these substances have hardly been studied, about 1327 substances are, as a result, insufficiently governed across the world. Consequently, 901 substances are accepted for utilization in plastic food-packaging materials [49], but they have unknown impacts on human health. There is, therefore, an outcry for widespread research to ensure that food packaging, especially plastic packaging, maintains its main role of protecting food. A sustainable circular plastic economy which reduces and, even better, prevents the use of hazardous chemicals as well as increases information accessibility is therefore essential. Although research studies on plastic food-packaging chemical compounds' migration into food products are widely reported [24,26,50–52], previous review works mostly focus on the migration of additives, challenges in additive analyses in the food and biological matrices [53], and impacts on human health [54,55]. To add to the literature, this work provides a critical review on monomer migration, including the monomer migration mechanisms and chemistry in versatile plastic food packaging. Furthermore, monomer endocrine-disrupting effects are currently speculated to be one of the major reasons for most of the current global chronic illnesses. The aim is to shift the focus of the relevant authorities, especially in emerging-economy countries, from only addressing the environmental pollution of plastic packaging to also urgently addressing and regulating its health impacts due to migrating

monomers. This is because existing plastic packaging legislative authority regulations, for instance, those in South Africa, are biased towards environmental pollution. This is evidenced by the numerous legislative authority regulations, such as the National Environmental Management Waste Act, Act 59 of 2018, and the National Water Amendment Act, Act 27 of 2014 [56,57], and yet policies, education, and awareness that address the human health plastic toxicity effects are lacking!

2. Food-Contact Chemicals in Plastic Food-Packaging Types

Plastic food-packaging materials (FPMs) comprise different contact chemicals and a variety of synthetic materials made from different chemical compounds and their combinations thereof (Table 2) and are used to keep food safe during the transportation of diverse food products [58]. Table 2, for instance, illustrates the composition of a plastic yoghurt container.

Table 2. Composition of a plastic food-packaging material utilized to package yoghurt.

Packaging Material	Synthetic Materials Present	Food-Contact Chemicals		References
		Intentionally Added Substances (IASs)	Nonintentionally Added Substances (NIASs)	
Plastic packaging material	Aluminium Coatings Adhesives Printing inks	Monomers Oligomers Additives Pigments Metals	Impurities By-products of reactions Breakdown products Recycling-product contaminants Starting-material impurities Unwanted side products	[59–62]

A detailed analysis of the food-contact chemicals (FCCs) worldwide reveals that there are about 12,285 intentionally added substances (IASs) [63], some of which are the building blocks (monomers) of plastic packaging materials. Much more, although difficult to predict, there are nonintentionally added substances (NIASs) from numerous other possible reactions and transformations [23]. Additionally, there are various contaminants from the recycling processes in the synthesis of food-contact materials [64]. However, the global challenges related to food safety suggest that the current scientific knowledge demonstrates a limited detailed understanding of all the possible materials in a packaging type. This is more than important especially because, during the last few decades, plastic food-packaging materials have transformed significantly, with new materials, designs, and technologies such as microwaveability, evolving to enable packaging to respond to the increased demands of the modern consumer lifestyles [35]. As such, the current standards of manufacturing compliance may not sufficiently account for the possible migration implications of the packaging material. Tables 3 and 4 show the monomers and some additives added to common plastic food-packaging types. Interesting to note is that some additives, Bisphenol A, for instance, are also monomers in some plastic food-contact material types.

Table 3. Chemical structures of monomers commonly used in common plastic food-packaging types.

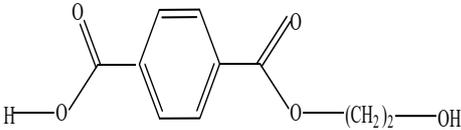
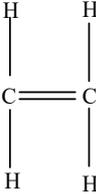
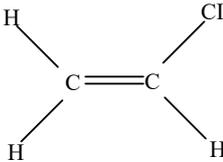
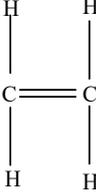
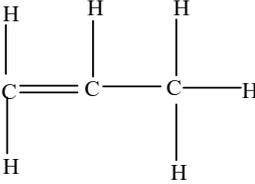
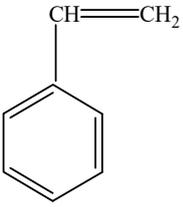
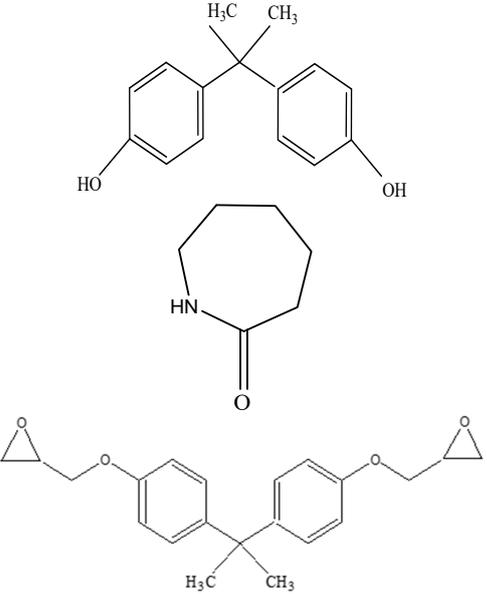
Plastic Type	Recycling Code and Symbol	Monomer Name	Monomer Structure	References
Polyethylene terephthalate (PET)		Ethylene terephthalate		[65,66]

Table 3. Cont.

Plastic Type	Recycling Code and Symbol	Monomer Name	Monomer Structure	References
High-density polyethylene (HDPE)		Ethylene		[65]
Polyvinyl chloride (PVC)		Vinyl chloride		[65]
Low-density polyethylene (LDPE)		Ethylene		[67]
Polypropylene (PP)		Propylene		[65,67]
Polystyrene (PS)		Styrene		[65]
Other		Bisphenol A for PC Caprolactam for Nylon-6 Bisphenol A diglycidyl ether for epoxy resins		[68,69]

Additives are added to a polymer for various functions (Table 4), such as to improve the overall characteristics of the polymer in accordance with its suitability for its end use [20]. However, they bind reversibly to the polymer system, and, as a result, monomers also easily leach into the food [20,70].

Table 4. Some additives present in plastic food-packaging materials.

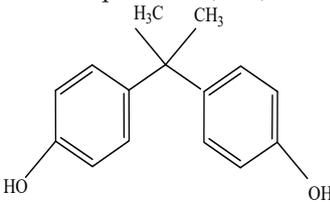
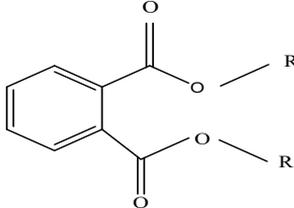
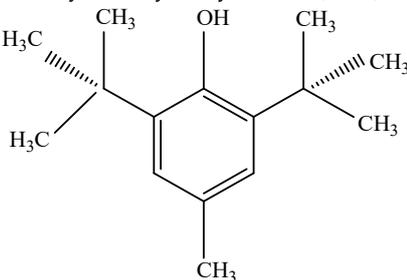
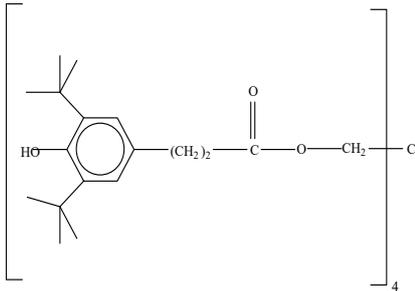
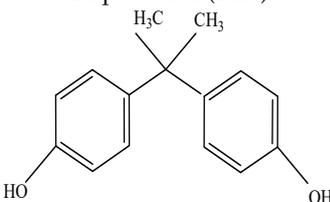
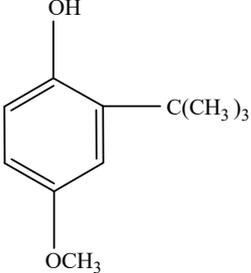
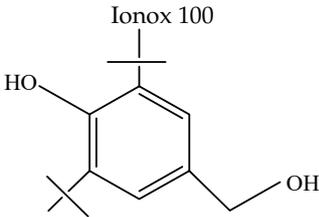
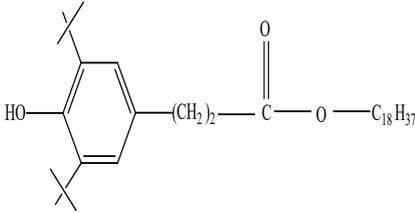
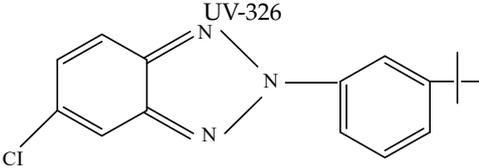
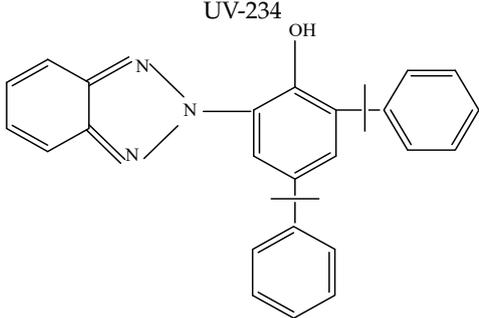
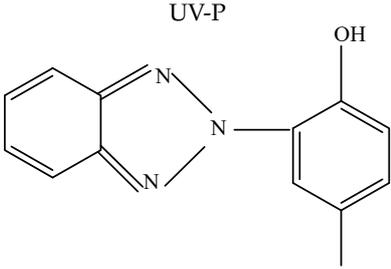
Additive Name	Function	Structure	Reference
Plasticizers	Increase the workability and flexibility of final product	<p>Bisphenol A (BPA)</p> 	[58,71]
		<p>Phthalates</p> 	
Antioxidants	Scavenge free radicals, reducing the oxidation process that exposure to light causes in polymers	<p>Butylated hydroxytoluene (BHT)</p> 	[72]
		<p>Irganox 1010</p> 	
		<p>Bisphenol A (BPA)</p> 	

Table 4. Cont.

Additive Name	Function	Structure	Reference
Antioxidants	Scavenge free radicals, reducing the oxidation process that exposure to light causes in polymers	Butylated hydroxyanisole (BHA)	[72]
			
			
Irganox 1076			
UV protectants	Stabilize polymers and prevent degradation		[73]
			
			

Recently, due to environmental concerns as a result of the non-biodegradability of plastics [74,75], there has been a drive towards the development of nontoxic eco-friendly biodegradable plastics [74]. Generally, three types currently exist based on their source of origin and method of production amongst which are various biopolymers produced through the chemical synthesis of renewable bio based monomers (Table 5).

Table 5. Monomers for some common biodegradable packaging.

Biopolymer	Monomers	References
Poly(lactic acid) (PLA)	Lactic acid	[74,76]
Poly(lactide aliphatic copolymer) (CPLA)	Lactide + aliphatic polyesters	[74,76]
Polyglycolide (PGA)	Glycolic acid	[74]
Polybutylene succinate (PBS)	Glycols + aliphatic polyesters	[74]
PBAT	1,4 butanediol + terephthalic acid + adipic acid	[74]

However, despite the seemingly acceptable organoleptic, mechanical, and chemical properties of biodegradable food packaging [77], its commercial application to date has been limited for numerous reasons, including the non-systematic knowledge on the migration of chemicals, including monomers [78]. This is because the utilized monomers have a low average molecular weight with the potential to diffuse through the polymeric matrix when utilized in food packaging. Strategies to improve biodegradable packaging performance involve the addition of a variety of substances, such as nano fillers and plasticizers, which also adds to the concerns of their migration into the food [79–82]. The increased addition of additives leads to undesirable interactions and the consequent migration of substances that may be more or less relevant for one than for the other [79]. Furthermore, the current research is centred on food simulants rather than real food products [78] and is concentrated on a few biopolymers, such as PLA [78]. The toxicological effects on animals are also lacking. Moreover, most countries of emerging economies do not have nanoparticle (NP)-specific regulations for aquatic systems, including wastewater treatment plants, where egested food with NP contaminants is finally deposited, which presents accurate, scientifically proven, and confirmed detection difficulties for the safe migration limit in food-packaging films in the respective countries [83]. Conclusively, more research on the migration of both components is essential to draw results on the safe utilization of biodegradable packaging with regards to chemical migration.

3. Packaging Monomers as Sources of Endocrine-Disrupting Compounds (EDCs) in Foods

The human body comprises tissues that interact with each other by means of hormones that control, for instance, reproduction, the early developmental processes, and the tissue and organ functions throughout adulthood [84]. Exogenous (non-natural) substances known as endocrine-disrupting compounds (EDCs) imitate the effects of natural hormones, preventing their production, release, transport, metabolism, binding, and elimination, which are essential for maintaining homeostasis, reproduction, and the developmental and behavioural processes in the human body, consequently causing adverse health effects [84]. Endocrine-disrupting compounds, such as monomers, in plastic food packaging thus disrupt the coordination and, consequently, the efficient functioning of the endocrine system, which is responsible for the regulation of various body processes [85–88]. Monomers are amongst the hundreds of EDCs globally utilized within the plastic food-packaging industry [89], such as additives (plasticizers, oxidants, and preservatives). Their EDC effect presents adverse human health problems across all consumers, with more health impacts on the younger generation, which poses a great challenge for future generations. Information on the types of EDCs present in the various plastic packaging materials is presented in Table 6.

Table 6. Uses of plastic food-packaging materials and EDCs contained in them.

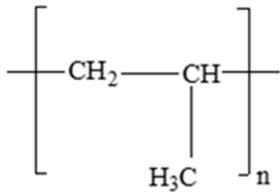
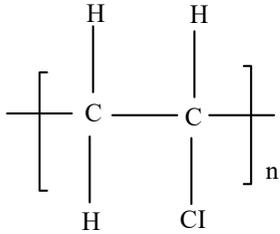
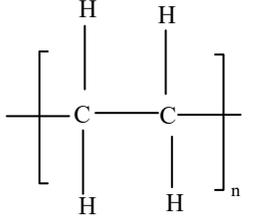
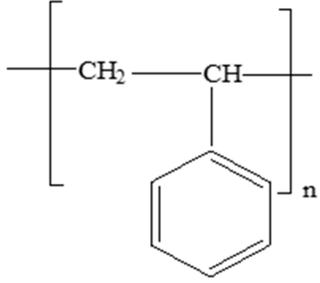
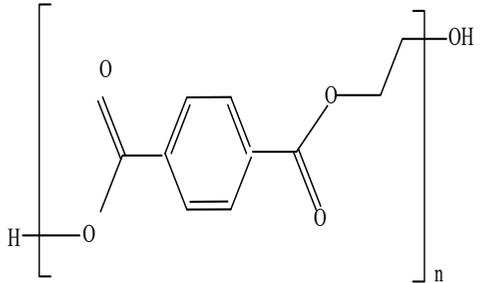
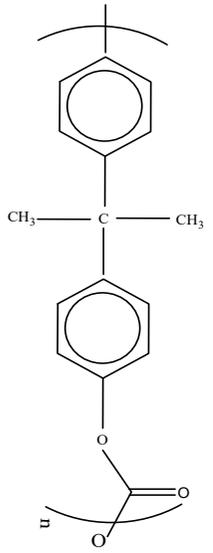
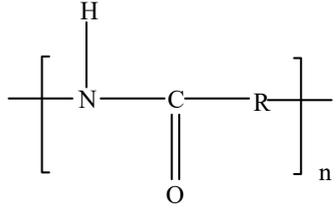
Packaging Material	Structure	EDCs Identified in Such Compounds	Uses of Packaging	Ref.
Polypropylene (PP)	 $\left[\text{CH}_2 - \underset{\text{H}_3\text{C}}{\text{CH}} \right]_n$	Antioxidants (vinyl and polymer with a methyl group) Plasticizers (phthalates)	Margarine tubs, microwaveable meal trays, lunch boxes, plastic bottle caps, and sweets and snack wrappers	[90–92]
Polyvinyl chloride (PVC)	 $\left[\begin{array}{c} \text{H} \quad \text{H} \\ \quad \\ -\text{C} - \text{C}- \\ \quad \\ \text{H} \quad \text{Cl} \end{array} \right]_n$	Heat stabilizers (Pb, Zn, and Sn compounds) Dioxins Plasticizers (phthalates) Bisphenol A (BPA)	Meat trays, bottles containing liquid foods (oils, vinegars, and beverage foods), flexible films for wrapping solid foods (fresh fruits, cheese, meat, and vegetables), coatings in metal cans, and lunch boxes	[93,94]
Polyethylene (HDPE and LDPE)	 $\left[\begin{array}{c} \text{H} \quad \text{H} \\ \quad \\ -\text{C} - \text{C}- \\ \quad \\ \text{H} \quad \text{H} \end{array} \right]_n$	Plasticizers (phthalates) Antioxidants Ethylene and olefins (butene, hexene, and octene)	Freezer bags; milk cartons; yoghurt, fruit juice, and soup pots; caps for plastic bottles; Tupperware; plastic grocery bags; and shrink wrap	[25]
Polystyrene (PS)	 $\left[\text{CH}_2 - \underset{\text{C}_6\text{H}_5}{\text{CH}} \right]_n$	Plasticizers (phthalates) Styrene	Disposable coffee cups; plastic food boxes; containers for yoghurt, ice cream, fruit juice, and cheese; egg cartons; and biscuit trays	[24,25,51]

Table 6. Cont.

Packaging Material	Structure	EDCs Identified in Such Compounds	Uses of Packaging	Ref.
Polyethylene terephthalate (PET)	 <p>The structure shows a repeating unit of polyethylene terephthalate (PET). It consists of a central benzene ring with two carbonyl groups (C=O) attached at the 1 and 4 positions. Each carbonyl group is linked to an oxygen atom, which is part of an ester linkage to a methylene group (-CH2-). The entire unit is enclosed in brackets with a subscript 'n', and a hydrogen atom (H) is attached to the oxygen on the left, and a hydroxyl group (-OH) is attached to the oxygen on the right.</p>	BPA Phthalates Dioxins Colourants Fillers Plasticizers	Water, soft drink, and alcohol beverage bottles as well as edible oil and fruit/vegetable punnets	[95,96]
Polycarbonate (PC)	 <p>The structure shows a repeating unit of polycarbonate (PC). It features a central carbon atom bonded to two methyl groups (CH3) and two phenyl rings. One phenyl ring is attached to a carbon atom that is part of a carbonate group (-O-C(=O)-O-), which is linked to another carbon atom. The entire unit is enclosed in brackets with a subscript 'n'.</p>	BPA Phenol Volatile aromatic and aliphatic hydrocarbons Chlorinated hydrocarbons	Recyclable beverage containers, ovenable frozen-food trays, and convenience meals	[97]
Polyamides (PAs)	 <p>The structure shows a repeating unit of polyamides (PAs). It consists of a nitrogen atom bonded to a hydrogen atom (H) and a carbonyl group (C=O). The nitrogen atom is also bonded to a carbon atom that is part of a chain (R). The entire unit is enclosed in brackets with a subscript 'n'.</p>	BPA 17α ethinyl estradiol Triclosan	Vacuum packaging of frozen foods, bacon, cheese, and fresh and processed meats	[98]

4. Monomer Migration into Food in Food Packaging

Since plastic packaging is produced through a polymerization process where monomers or building blocks are linked together, monomer residues are always present in plastics, although they are present at generally low concentrations of 0–2%. This is because not all added reactants will complete the reaction [30]. Furthermore, the majority of low-molecular-weight substances, like LDPE, are not covalently attached to the polymer chain. Instead, they take the form of branching chain structures, which prohibit the monomer units in the polymer chain structure from being packed closely together. As a result, these and residual monomers are able to diffuse all over the polymer matrix [32]. However, although monomers are generally stable and nontoxic when bound by the polymer matrix, interactions with food make them harmful and cause them to affect human health when they are later consumed with the food, and their concentrations increase in the body [39,99]. The migration process, which is influenced by numerous parameters, is divided into the following four primary steps. These are chemical compound diffusion through the polymers, diffused molecule desorption from the polymer surface, compound sorption at the plastic–food interface, and compound desorption throughout the food [20] (Figure 1).

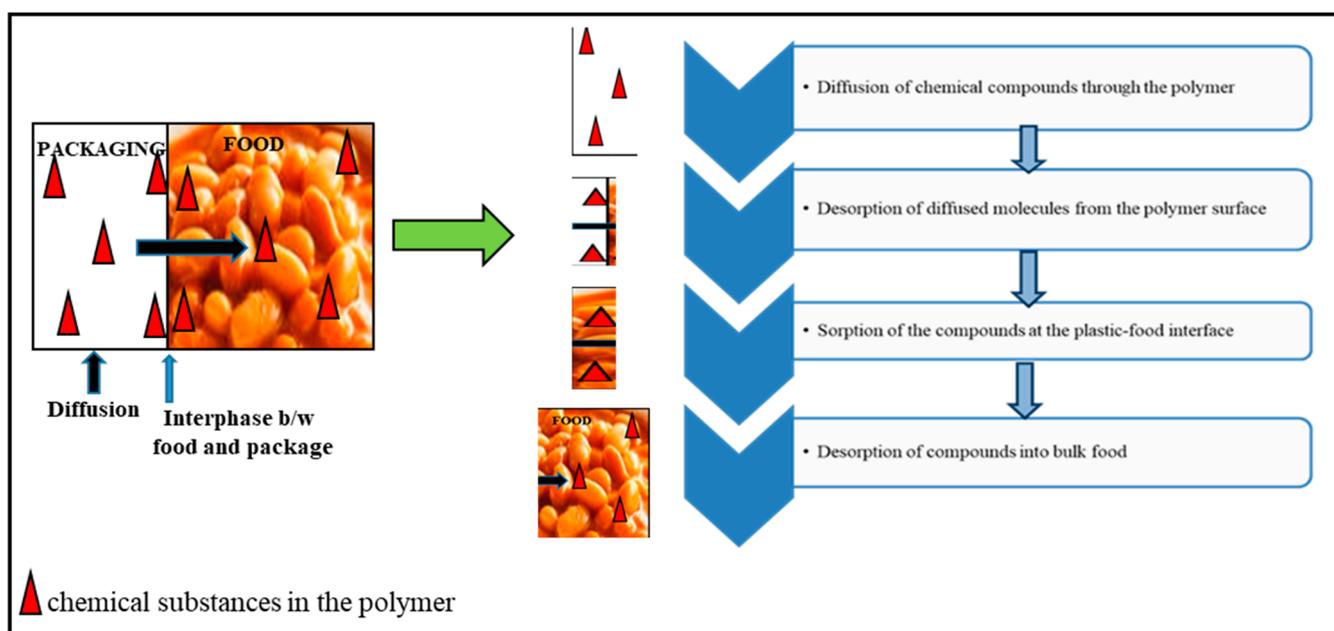


Figure 1. Monomer migration process.

Due to the fact that migration may introduce unwanted and dangerous chemical substances, which deteriorate the nutritional value, safety, and organoleptic qualities of packaged foods, the process is thus undesirable. Nonetheless, some transfer is unavoidable because food must be packaged before being purchased by the consumer [100]. To date, evidence from numerous researchers reveals monomer migration results from polystyrene, polyamides, polycarbonates, polyvinyl chloride, and polyurethanes plastic types into different foods and food simulants [101]. However, food simulants are commonly utilized in laboratory investigations to counteract the complexity of the physical structure of food, to understand migration more clearly, and for regulatory compliance reasons [102]. Commonly utilized food simulants include water for aqueous environments, 3% acetic acid for acidic food simulants, 10% ethanol for alcoholic food simulants, and refined olive oil for fatty food simulants as guided by the European Union [103] for migration tests on plastic food-contact materials. To verify the overall safety of the plastic contact material, the European Food Safety Authority (EFSA), for instance, indicates that the maximum limit of overall migration (the total of all the substances together) into a packaged food sample should be 60 mg kg^{-1} of the food or 10 mg dm^{-2} of the packaging material [20].

However, there are various complications with migration tests, including that the tests have long experimental workflows. Additionally, based on different cultures, which translate to how food is prepared in different nations, perhaps accurate and reliable research on monomer migration should focus on using real foods rather than food simulants to incorporate the different food preparation methods and spices that could otherwise affect chemical migration. The resultant OM and SM is, therefore, likely to be complicated and not comprehensively explained by food simulants. As such, simulants in comparison to real food samples risk omitting other possible interactions between food and the food's packaging. Furthermore, some materials, for instance, absorbent packaging materials, pose problems for accurate OM and SM testing. Migration tests also need to determine the temperature and humidity conditions to imitate the stress generation conditions that facilitate investigating the behaviour of the packaging material [32], which poses a huge challenge in the evaluation of OM for different food-packaging materials and, therefore, in that of the possible health and environmental effects.

4.1. Migration Mechanism Processes in the Migration of Monomers into Food

Migration occurs in a number of different ways, including contact, penetration, gas-phase, condensation, and set-off migration [104]

Contact migration

A direct substance transfer from the packing material's food-contact surface into the packed food is referred to as contact migration. Examples of contact transfers include the migration of materials from a plastic tub tray or wrapping into food and from a cardboard pizza box to the underside of the pizza (Figure 2) [104].

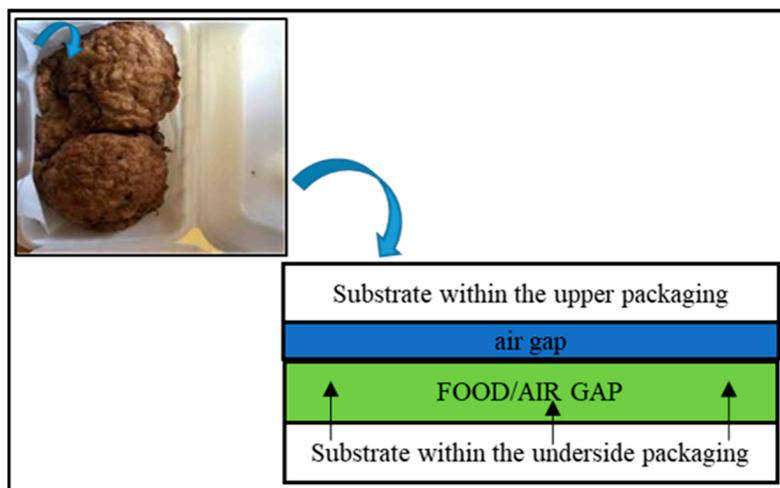


Figure 2. Contact migration mechanism in a pizza box.

Condensation/distillation migration

Condensation migration involves the leaching of chemical substances, particularly volatile components from food-packaging materials to food during heating stages, such as sterilization or boiling [104]. However, several migration studies' findings reveal distillation migration even before the above heating stages. As such, condensation leaching examples include microwave heating to cooking in cartons, trays, or plastic food containers (Figure 3) [104].

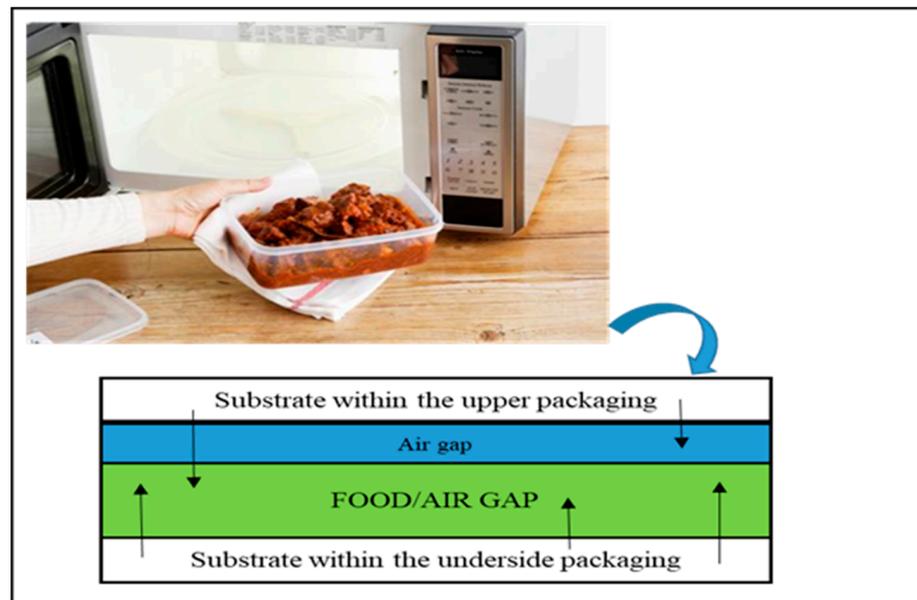


Figure 3. Condensation/distillation mechanism in microwave heating.

Gas-phase migration

Gas-phase migration relates to the permeation of volatile chemicals from the packaging coating on the outer layer through the airspaces within the plastic packaging and between the packaging material and food into the food through diffusion (Figure 4). Examples include the diffusion of mineral oil into meals through a plastic inner pouch (depending on the material's barrier qualities), an airspace within paper packaging, or a second airspace between the packaging and food [105].

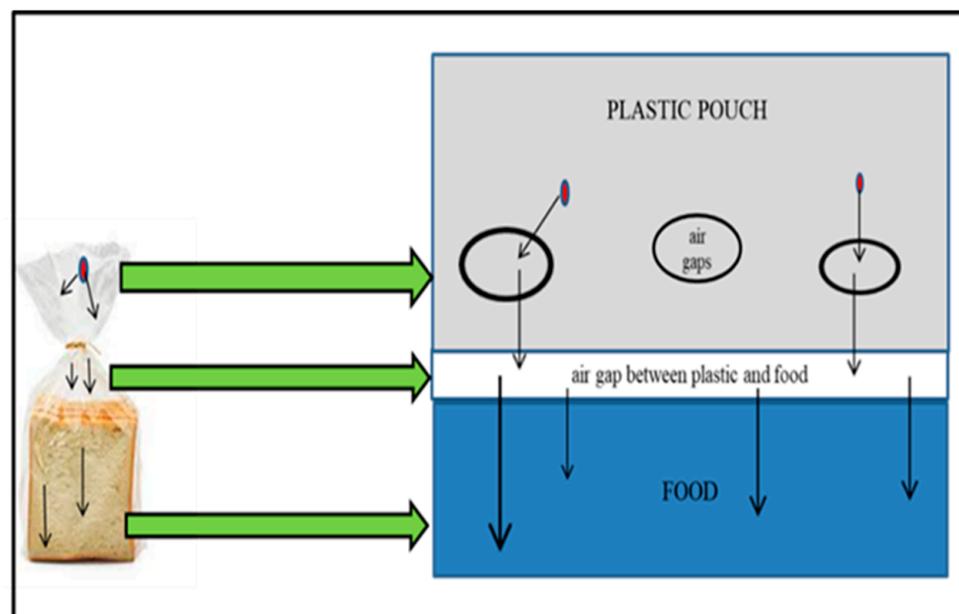


Figure 4. Gas-phase migration mechanism in plastic bread packaging.

Penetration migration

The penetration type of migration is the diffusion of chemical substances from the packaging non-food-contact surface (often a coated or printed surface) through the substrate and onto the packaging's food-contact surface (the inner layer) (Figure 5). Once the

migrating chemicals are on the food-contact surface, they then leach into the food through gas-phase or contact migration, contaminating the food [104].

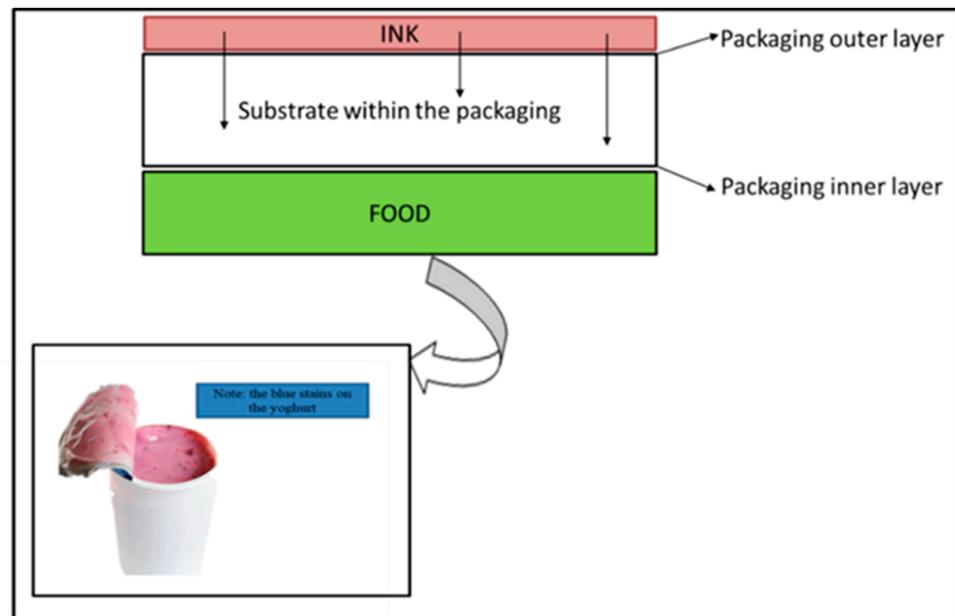


Figure 5. Penetration migration mechanism in a yoghurt container.

Set-off migration

Set-off migration describes chemical substance diffusion from the coatings, varnishes, or printed ink present on the outer printed non-food-contact side of the package material through the substrate towards the inner food-contact side due to the stacking of the printed items (Figure 6).

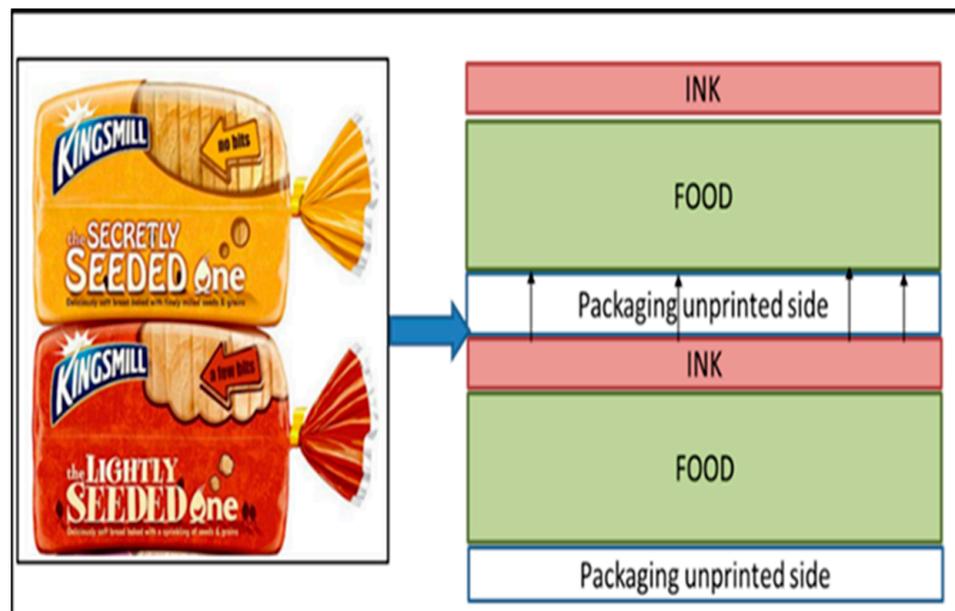


Figure 6. Set-off migration mechanism in stacked bread.

The set-off kind of migration can be either obvious or invisible. Once chemical compounds are on the food-contact surface, they are subsequently transferred throughout the food through gas-phase or contact migration, contaminating the food [104].

Due to the various chemicals present in plastic food packaging, small molecules, including monomer residues, oligomers, additives, reaction by-products, and adhesive components (a) as well as printing inks (b) from the outer layer of the packaging or from others in a stacked pile, diffuse and leach from the plastic material into the food (Figure 7) [62].

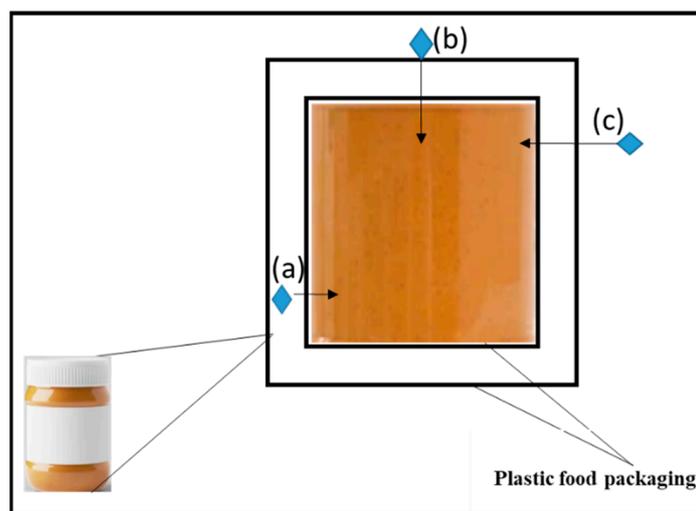


Figure 7. Illustration of the migration mechanism in plastic food packaging.

Several studies agree that, through the influence of several factors, migration either follows a set-off, contact, gas-phase, or penetration migration mechanism process depending on the present situation [105], and they further illustrate that in recycled plastic food packaging, environmental toxins, like pesticides, detergents, and persistent organic pollutants (c), are absorbed into the plastic packaging and are subsequently released again.

Some of the identified leaching monomers from plastic food-packaging materials that are particularly labelled as problematic include Bisphenol A, styrene, Bisphenol A diglyceride ether (BADGE), and caprolactam.

Bisphenol A (BPA) added as an antioxidant to polymers, for instance, can potentially migrate from PC or plastic resins commonly used in cans [106]. Its migration into different foods, including water and 10% and 50% ethanol, in PC and various plastic containers, such as PC baby bottles, baby bottle liners, non-PC baby bottles, and recyclable PC drinking bottles, has been recorded from the environment and from the can linings and PC bottles through investigations of several factors [107], and BPA's migration in evaporated milk, carrots in brine, minced beef in gravy, spring vegetable soup, and a food simulant (10% ethanol) has also been studied. The amount of migrated BPA was significantly higher in 10% ethanol ($68.3 \pm 9.0 \mu\text{g kg}^{-1}$) compared to the following foods: minced beef ($53.8 \pm 7.6 \mu\text{g kg}^{-1}$), milk ($49.8 \pm 10.9 \mu\text{g kg}^{-1}$), carrots ($47.2 \pm 5.1 \mu\text{g kg}^{-1}$), and soup ($45.7 \pm 5.0 \mu\text{g kg}^{-1}$). Bisphenol A diglyceride ether (BADGE) is also an epoxy resin polymer monomer utilized in internal food can linings. In a separate study [108], the amount of BADGE and BPA that leached into distilled water from two different can types that packaged tuna fish and jalapeño peppers was examined. The conclusions based on the study results are that both monomers migrate, although there are different factors influencing their overall migration.

Styrene monomers are always present in PS, acrylonitrile-butadiene-styrene, and polyamide packaging materials [109], which are widely utilized to package a range of dairy products, such as ice cream and yoghurt; bakery products; juices; meat; and fresh produce [110]. However, research reveals that residual styrene monomer levels vary in

similar packaging materials utilized to package similar products within different countries (Table 7) [111]. Furthermore, migration studies reveal that styrene monomer migration is dependent on several factors. In [112], studies on styrene migration from various PS food-contact packaging materials, including egg cartons, meat trays, plates, and cups, into oil showed that migration increased within days. With the exception of drink cups, migration was also proportional to the square root of the time increase. In a separate study with hot drinks, the migration of styrene strongly depended on the temperature and amount of fat in the hot drinks [113]. The styrene monomer migration level results in $\mu\text{g}/\text{L}$ varied from 0.61 to 8.15 for hot tea, 0.65 to 8.30 for hot milk, and 0.71 to 8.65 for hot cocoa milk in GPPS (general-purpose polystyrene) cups and from 0.48 to 6.85 for hot tea, 0.61 to 7.65 for hot milk, and 0.72 to 7.78 for hot cocoa milk in HIPS (high-performance polystyrene) cups at different temperatures and times [114]. The findings showed that hot cocoa milk had the highest degree of styrene leaching [114]. Further studies on styrene migration in aqueous and oily foods also revealed less styrene migration because the monomer is hydrophobic [115]. However, a recent study reveals that the effect of the fat content on the migration of styrene is insignificant in relation to the variability of other parameters [111].

Table 7. Residual styrene levels in PS packaging with similar products.

Country	Food Description	Residual Styrene Monomer Levels ($\mu\text{g}/\text{g}$)	Reference
Italy	Stirred yogurt, 3.2% fat	266 ± 1	[111]
Germany	Stirred yogurt, 3.5% fat	275 ± 2 – 351 ± 23	
Germany	Set yogurt, 3.5% fat	278 ± 12 – 308 ± 6	
Germany	Stirred sour cream with 10% fat	260 ± 8 – 292 ± 20	

Ref. [116] studies on caprolactam monomer migration from nylon 6 and nylon 6/66 polymers to oil when cooked in an oven showed that the nylon 6/66 oligomers that migrated due to the above made up nearly 43% of the existing oligomers in the utilized packaging material. In addition, Ref. [117] investigated how caprolactam moved from nylon 6 packaging to 95% ethanol. The samples analysed also included poultry breasts, ham, pate, turkey blanquettes, and bologna sausages. The findings showed that the migration of caprolactam was above the set EU standard of 15 mg kg^{-1} in 35% of the packaging for bologna sausage, 33% of the turkey blanquette packaging, 100% of the pate packaging, and 100% of the packaging for poultry breast [118]. Based on the continuous evidence of monomer leaching from plastic food packaging into the food, it is therefore important that, globally, industrial policies speak and implement enforcement measures that will ensure the compulsory synthesis of plastic packaging materials with efficient polymerization processes.

The concept of pyrolysis, which involves the thermal breakdown of organic molecules at a moderate temperature and in the absence of oxygen [119], can be used to inform on the bond dissociation energies (BDEs) of the monomers of different plastic polymers. The bond dissociation energy is a crucial thermodynamic quantity that represents the minimum energy required to break chemical bonds, in this instance, the monomer bonds from the polymer structure, so that they leach into the food. In addition, it also exemplifies the chemical activities of the free-radical reactions [120] in the plastic polymers, which are important in the chemical migration phenomenon and, thereafter, in food safety and quality. The larger the BDE, the stronger the chemical bond is, and the less likely the bond is to break. The bond dissociation energies of four common plastic packaging polymers calculated using two-density functional theory methods (DFTs) (B3P86/6 with the -31 G (d,p) basis set and M062X/6 with the -31 G (d) and $-31++\text{G (d,p)}$ basis sets) are shown in Table 8.

Table 8. Bond dissociation energies for some plastic polymers utilized in food packaging.

Plastic type	Different Methods' Chemical Bond Average Values (kJ mol ⁻¹)								Ref.	
	Bond types									
	C-C bonds		C-CH ₃ bonds		C-C aromatic bonds		C-Cl bonds			
	M06-2X/6	B3P86/6-31 G (d,p)	M062X/6-31 G (d)	B3P86/6-31 G (d,p)	M062X/6-31 G (d)	B3P86/6-31 G (d,p)	M062X/6-31 G (d)	B3P86/6-31 G (d,p)		
PE	364.3	350.9	-	-	-	-	-	-	-	
	0.003	0.003								
PP	357.1	329.5	361.9	342.6	-	-	-	-	-	
	0.003	0.003	0.003	0.003						
PS	331.5	291.7	-	-	424.1	395.9	-	-	-	[121,122]
	0.003	0.003			0.003	0.003				
PVC	373.8	345.8	-	-	-	-	355.6	343.7		
	0.003	0.003					0.003	0.003		

The BDE findings show that the main chain C-C bonds for PP (329.5) and PS (291.7) are generally weak. A comparison of the four polymers therefore suggests that the thermal stabilities of the four polymers are in the order of PE > PP > PS > PVC. Based on the bond dissociation energy equation indicated in Equation (1), the bond association constants of the monomers of the respective polymers calculated using Equation (2) are shown in Table 9 (coloured):



where [P] is the protein concentration/polymer, [L] is the ligand concentration/monomer/any molecule that the polymer binds, K_a is the association constant, K_D is the dissociation constant, and [PL] is the concentration of the protein ligand complex.

$$K_a = \frac{1}{K_d} \quad (2)$$

Table 9. Styrene and Bisphenol monomer interactions with functional groups of main nutrients.

Nutrients	Monomer	Reaction/Interaction
Carbohydrates	Styrene	

Table 9. Cont.

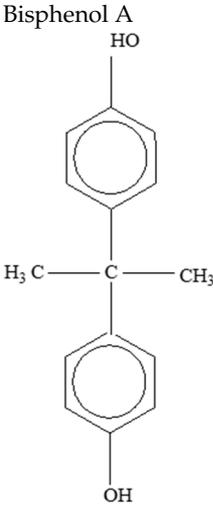
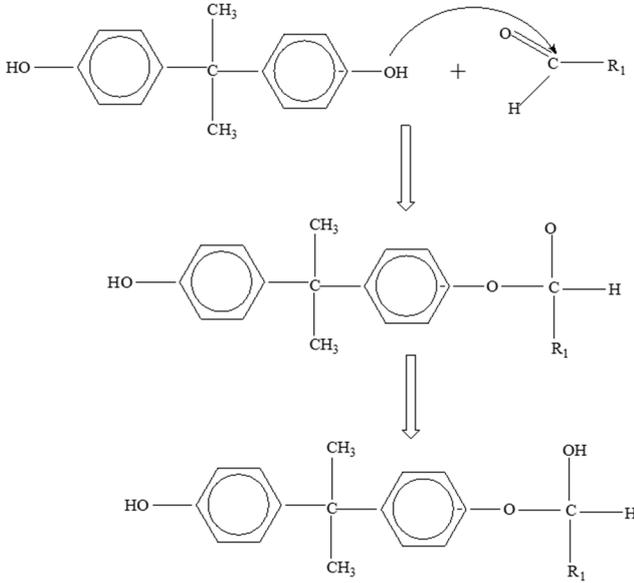
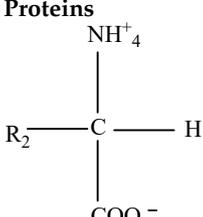
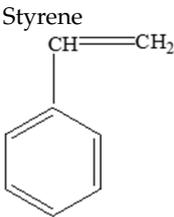
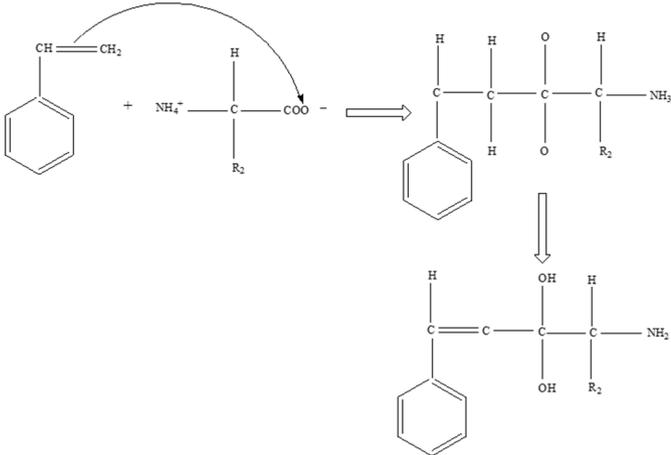
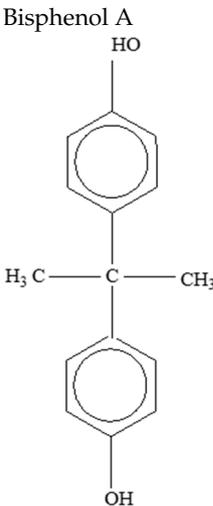
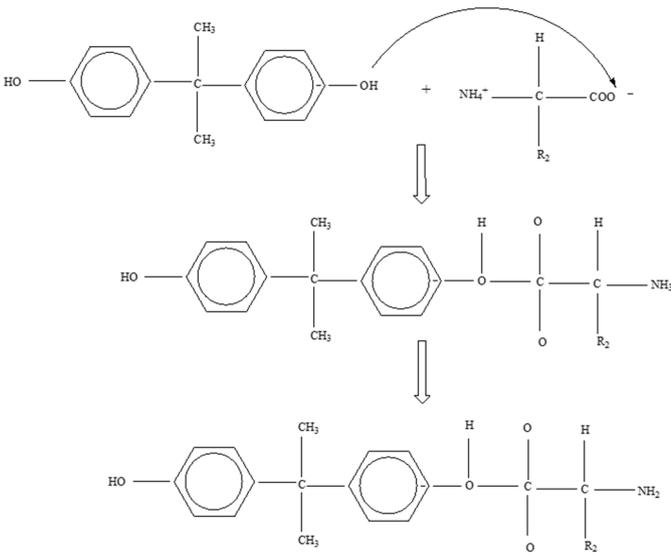
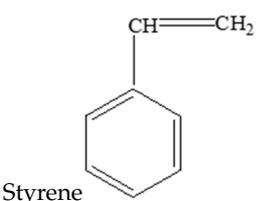
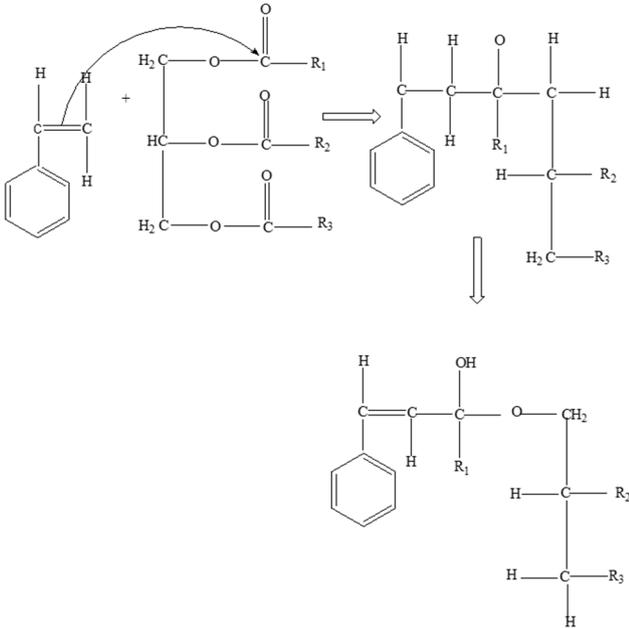
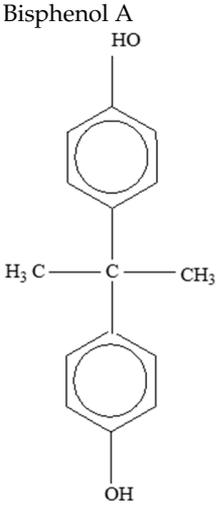
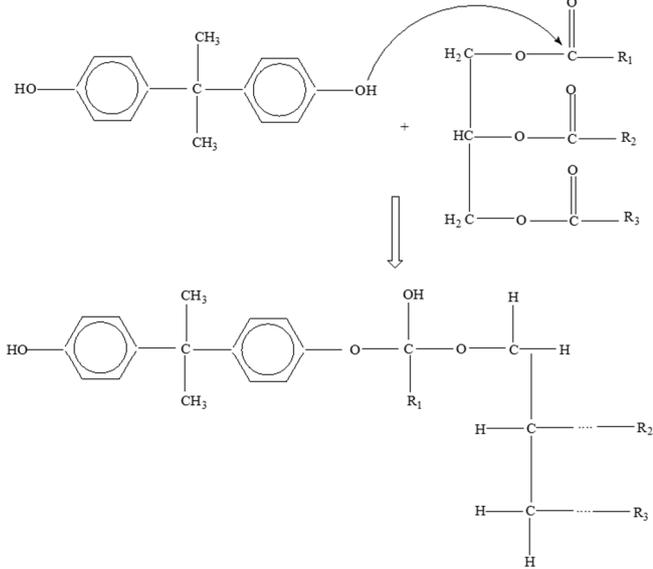
Nutrients	Monomer	Reaction/Interaction
	<p>Bisphenol A</p> 	
Proteins	 <p>Styrene</p> 	
	<p>Bisphenol A</p> 	

Table 9. Cont.

Nutrients	Monomer	Reaction/Interaction
Fats	 <p>Styrene</p>	
	 <p>Bisphenol A</p>	

4.2. Migration Mechanisms Involving Different Chemistries of Monomers

Throughout the previous twenty years, scientific research studies demonstrated that leaching from packaging materials into food simulants and food is a predictable diffusion process within the polymer network [123]. However, the migration chemistry mechanisms for different migrants, including monomers, are not the same. This is because, according to [124], simulating the migration process of each migrant from, for example, plastic packaging materials to food is difficult. The migration mechanism chemistries of two commonly researched monomers, for instance, are shown below:

Bisphenol A

Two different processes explain the leaching of Bisphenol A from polycarbonate polymeric materials. These processes are pH-dependent hydrolysis/decomposition, which occurs over time at the polymer surface (Figure 8), and the diffusion-controlled release of the leftover BPA monomers from the polymer [125].

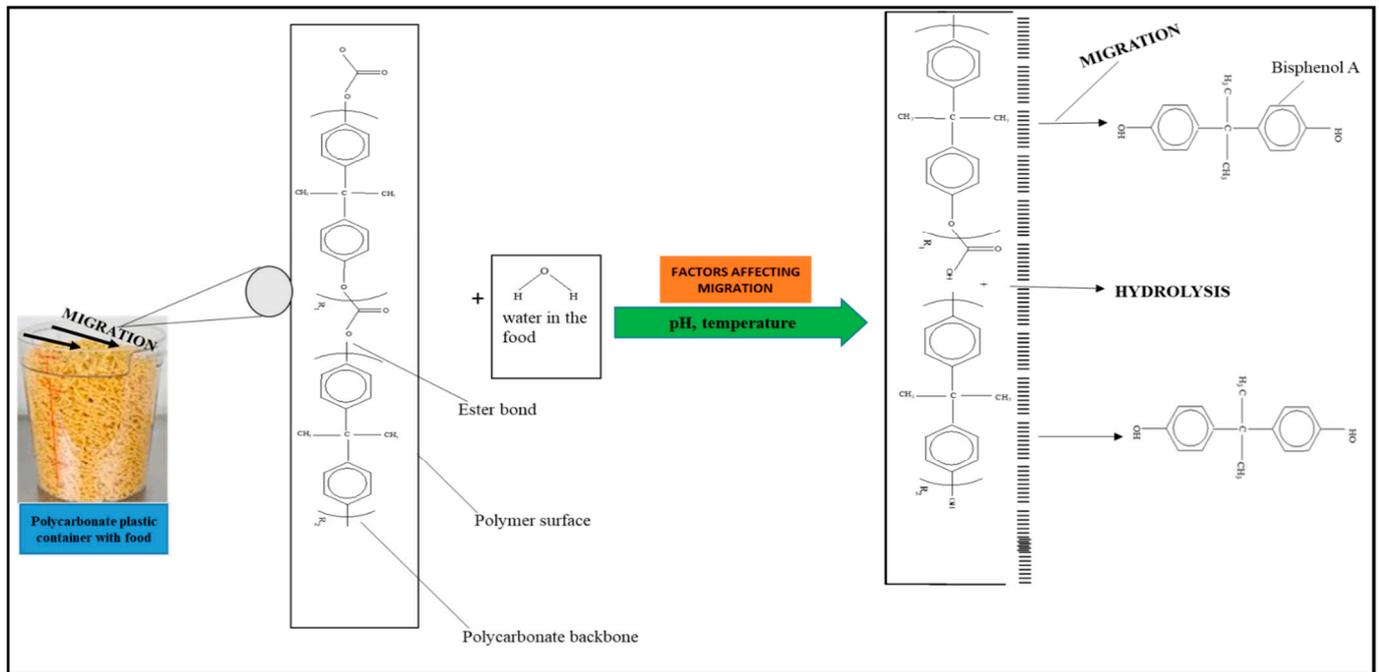


Figure 8. Hydrolysis migration mechanism of Bisphenol A from polycarbonate.

However, a comparison of the two migration methods shows that hydrolysis at the surface is the primary source of BPA migration into aqueous media and that it occurs at the carbonate–ester linkages of the PC backbone. In contrast, diffusion-controlled release only contributes to a relatively small part in overall migration [125].

Styrene

Styrene monomer migration (Figure 9) is primarily a diffusion-controlled process that follows Fick’s law [32].

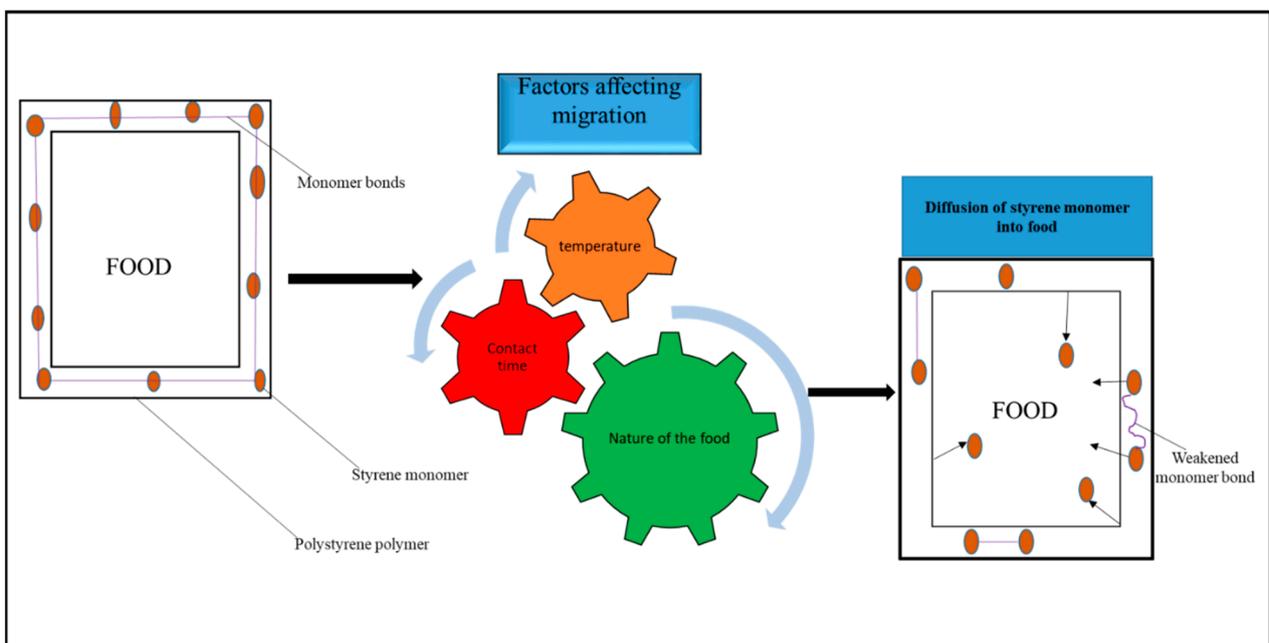


Figure 9. Migration mechanism of styrene from polystyrene.

In the presence of elements influencing migration, the styrene monomer diffuses from a higher-concentration zone (the polystyrene food packaging) to a lower-concentration zone,

which is the food, as result of weakened monomer/polymer interactions and increased solubility [126].

4.3. Factors Influencing the Migration of Food-Packaging Monomers into Food

Several factors govern the rate (kinetics) and general migration process from a food-packaging material into food [104,105]. The factors include aspects relating to the properties of the polymer material in interaction with the food (permeability, thickness, size, type, and format) and the properties of the migrant (polarity, molecular size, structure, and vapour pressure) as well as the state/properties and composition of the food materials, the starting migrant concentration in the packaging, the polymer matrix state, and the migrant components in contact with the food packaging [115,127]. The storage time, temperature, packaging size, period of contact, food surface area in relation to its volume such as with pasta, and packaging surface area ratio to the food product volume also affect chemical migration [105]. However, the primary factors affecting the migration process are as follows:

4.3.1. Nature of Foods

The food simulants and different foods utilized so far to show the impact of the nature of food on packaging substance leaching depict that foods with excess fat have significant migration rates [128]. The Bisphenol A migration studies, for instance, during the storage and can denting of PC containers with carrots in brine (0% fat), evaporated milk (8% fat), minced beef in gravy (20% fat), and spring vegetable soup (0.3% fat), displayed significantly higher BPA migration into 10% ethanol in fatty foods than in other foods. The detailed BPA migration results were $47.2 \pm 5.1 \mu\text{g kg}^{-1}$, $49.8 \pm 10.9 \mu\text{g kg}^{-1}$, $53.8 \pm 7.6 \mu\text{g kg}^{-1}$, and $45.7 \pm 5.0 \mu\text{g kg}^{-1}$, respectively [129]. The styrene migration studies also showed a migration increase with the fat content [110]. The above information is attributed to the lipophilic nature of the chemicals contained in the packaging materials. In a separate investigation, higher styrene migration levels were recorded in ethanol-containing solutions than those recorded in isooctane solutions. However, styrene did not migrate in aqueous food solutions [130]. A similar behaviour was also observed and noted for ϵ -caprolactam migration in the nylon 6 packaging. The samples analysed included poultry breasts, ham, turkey blanquettes, and bologna packages kept at 72–100 °C for 1–4 h. ϵ -caprolactam migration exceeded the EU set limit of 15 mg kg⁻¹ in 35% of the bologna sausage packaging, 33% of the turkey blanquette packaging, and 100% of both the pate and poultry breast packaging [117]. However, due to the numerous benefits of spices, food is usually cooked with a single spice or a mixture of spices. Seeds such as cumin, which are also utilized to produce spices, contain volatile oils. Perhaps it is important to further conduct experiments that show the contribution of spices in terms of influencing the leaching of monomers into food from packaging.

4.3.2. Nature of Contact

Research studies indicate that there is a relationship between migration rates and the nature of contact (direct or indirect) between food and the respective contact material. The mass transfer of the chemicals from the packaging to the food increases when there is direct contact between the food and the packaging material. Compared to direct contact, in an indirect medium, the gas medium between the packaging and food causes slower migration [131].

4.3.3. Period of Contact

Chemical migration largely depends on the duration of contact between the packaging and food [99]. Ref. [132], for instance, conducted research on the potential for PET oligomers to migrate from plastic packaging to different beverages and foods in ovens and microwaves at various temperatures with a focus on the temperature and exposure duration. Compared to oven heating, microwave heating showed less migration because of the shortened

exposure time (maximum of 15 min for MW and 80 min for oven heating). Ref. [104] also highlights that, depending on the nature of the food, for instance, solid or liquid, oily or aqueous, and a moisture or fat content, the food-packaging material compatible at the beginning of the shelf life may become incompatible at the end of the shelf life. With time, for instance, foods that contain water are likely to draw polar immigrants, while fatty foods attract nonpolar immigrants. The conclusions based on the research studies therefore indicate that the square root of the contact time of the food and packing material determines how much of the mass of the migrant substance is transferred [133].

4.3.4. Temperature during Contact

The temperature of the food directly influences the migration rate from the packaging into the food. In [128], it was discovered that migration rates rise as the temperature rises. Ref. [134] investigated styrene migration from various PS food-contact packaging materials, including egg cartons and meat trays, by exposing the materials to 8% ethanol and oil at 210 °C for 10 days, 490 °C for 4 days, and 65.5 °C for 1 day. The migration process exhibited a Fickian diffusion model. Migration increased from the first day to the tenth day and, for all materials with the exception of drink cups, was proportionate to the square root of the increase in time [112]. In a separate study on brand new PC baby bottles exposed to a temperature of 40–100 °C, the results showed a similar pattern, with the concentration of the BPA migrated into the food ranging from 0.03 ppb to 0.13 ppb at 40 °C to 95 °C, respectively [135]. Ref. [107] also used PC (baby bottles) and various other plastic containers (non-PC baby bottles) to study BPA migration into water and 10% and 50% ethanol. After 240 h at 40 °C, the average residual BPA content was higher in the 50% ethanol (2, 39 g L⁻¹) than in the water (1.88 g L⁻¹). The results showed that the higher the temperature and the longer the treatment periods are, the greater the BPA migration rate is [129]. Ref. [136] similarly came to the conclusion that the temperature has an inverse relationship with the log of the length of the equilibrium of a migratory material.

4.3.5. Packaging Material Characteristics

The composition of a food-packaging material significantly impacts substance migration. The migration of monomer additives, for instance, is dependent on the packaging material's thickness and plasticization. Thinner packaging allows for greater migration, and thicker packaging slows migration [21]. However, currently, research has not yet established any discernible relationship between the utilization of recycled components and the rate of migration [136].

4.3.6. Migrant Characteristics

The nature of the migratory substance affects the rate and amount of migration. For instance, highly volatile materials migrate at a faster rate, and lower migration rates are shown for substances with significantly higher molecular weights [24]. However, some monomers, such as vinyl chloride and ethylene, migrate quickly even at ambient temperatures [28]. Migration is also affected differently depending on whether the migratory substance is spherical or branched. For instance, experimental findings demonstrate that branched molecules display slower migration rates [128]. Ref. [108] also explored the possibility of BADGE and BPA leaching into distilled water from two different can types: one used for jalapeno peppers and the other used for tuna fish. The findings showed an increase in migration with time during storage for the jalapeno pepper cans. Bisphenol A migration from the tuna cans was, however, independent of the storage time, while BADGE migration during storage decreased over time due to its instability and ability to hydrolyse in an aqueous medium. Overall, the BPA and BADGE migration levels ranged from 0.25 to 4.3 g kg⁻¹ and from 0.6 to 83.4 g kg⁻¹ [108].

4.3.7. Migrant Concentration within the Packaging Material

Higher amounts of the migratory substances in the food matrix after a certain period of time in storage suggest that a mass transfer from the packaging into the food occurred at a higher rate as a result of a higher migratory compound concentration in the packaging material [137]. A study, for instance, conducted to investigate BPA migration under different factors, firstly involved processing the PC cans at 121 °C for 90 min and then storing them at 5 °C and 20 °C. Longer periods of storage were simulated by storing cans for up to 10 days–3 months. From the overall can coating BPA amount, the results showed 80% to 100% migration during processing. No BPA migration was observed in the simulants after processing. Therefore, the results suggested that there was a high migratory substance (BPA) concentration in the packaging material before processing [129].

4.3.8. State of Polymer Matrix

This phenomenon refers to whether the polymer matrix exists at the storage temperature in a rubbery or glassy form. Migration in glassy polymers, such as PE, is substantially slower compared to that in rubbery polymers [32]. Due to the fact that migration can be reduced through migration-informed manufacturing or the use of specially developed low-migration closures, toxicological risk assessments of migrants are therefore utilized to set the migration limits for food-packaging materials. The limitations are incorporated into Food-Contact Regulations with the intention of limiting exposure to safeguard human health. However, in most countries, ordinary consumers have no access, or they lack knowledge on such regulations and, therefore, remain vulnerable until there is a national crisis that leads to the discussion of the issue in the media. By such a time, it is likely that there could be fatalities too.

4.3.9. Migration Kinetics

Numerous factors affect the rate and speed of migration from food-packaging materials to food [99,104]. These include the features of the food-contact material, such as the thickness and permeability; the migrant chemical properties, including the molecular size, vapor pressure, polarity, structure, packaging material migrant's original concentration, temperature, and contact time; and, furthermore, the nature of the food interacting with the packaging material, that is, either real food or simulants [99,104]. Generally, for instance, small molecules, such as residual monomers, due to lower boiling points, migrate at a faster rate compared to larger ones [104]. Migration also increases significantly with increased temperatures that are accompanied by shorter contact times [99,104]. Migration also decreases with a decrease in the migrant starting concentration and the food-packaging material thickness. To the knowledge of the researcher, there is, however, no research to date that has compared the migration rates between real foods and simulants.

5. Interactions between Monomers and Food Nutrients

Once the monomer has leached, it combines covalently with the nutrients (Table 9) and/or the non-nutritive ingredients in the food.

The interactions are based on the functional groups of the main nutrients usually present in the food and the leached monomers. Additionally, the processing technologies, storage conditions, and duration also play a significant role [138]. Table 9 illustrates some possible interactions between the main nutrients present in the food and the monomers (bisphenol A and styrene).

6. Human Health Risks Due to Monomer Presence in Food

The initial food-packaging material regulations generally presupposed that, besides carcinogens, low-level chemical exposures, including EDCs, contained in food-packaging materials lower than the toxicologically determined no-effect levels had minimal health dangers to consumers [32]. However, to date, evidence from animal toxicological studies involving selected wildlife and human populations have raised more health questions

than have been answered [139] (Table 10), for instance, in plastic food packaging, due to numerous health effects.

Table 10. Human health effects of some monomers contained in plastic food packaging.

Monomer	Health Effects	References
Styrene	-Toxic effect on the liver, chromosomal abnormalities, carcinogen, mucous membrane irritation, eye irritation, gastrointestinal effects, CNS dysfunction (reaction time and memory), effects on some kidney enzyme functions and on the blood, stimulates cell replication, cell proliferation, and cytogenetic damage promotion.	[36,140]
Vinyl chloride	-Liver, kidney, and lung toxicity; effects on liver, kidney, lung, spleen, nervous system and blood; cancer; causes steatohepatitis; affects glucose homeostasis; and enhances alcoholic liver disease.	[141]
Bisphenol A	Breast, ovarian, uterine, prostate, and testicular cancer.	[142]
Caprolactam	Cause neurasthenia syndrome and damages the central nervous system.	[143]

However, the likelihood that consumers may experience negative health effects from any chemical contained in food mainly depends on the chemical toxicology and the exposure (dosage) as a result of the consumption of contaminated food. As such, currently, the utilization of ‘acceptable limits’ for different known chemicals is used to reduce the effects on humans. However, acceptable limits cannot exist for ‘unknown’ chemicals that, unfortunately, might be endocrine-disruptive and might have related or different adverse human health effects. This implies that, until a chemical is characterized and until their toxicological profile is determined, humans therefore remain vulnerable to their effects. There are several human EDC exposure routes, including the taking in of contaminated water and food, contaminated air inhalation, and chemical absorption through the skin, which are measurable using biological samples including breastmilk [89]. However, the consumption of contaminated food is continuously singled out as the major source of human exposure to EDCs across all age groups [144]. Once in the body, there are numerous independent toxicity actions that EDCs, including monomers, possibly interfere with, and they can block or imitate oestrogenic hormones, triggering diverse signalling pathways which yield diverse and divergent biological responses [145]. Alternatively, they could bioaccumulate in an organism’s lipid compartments and create mixed contaminated ‘body loads’ [89]. Currently, however, there are no studies that have focused on the impact matrices of EDC mixtures on the human body’s health. Table 11 presents the human health effects of some EDCs contained in food-packaging materials.

Table 11. Health effects of EDCs contained in food-packaging materials.

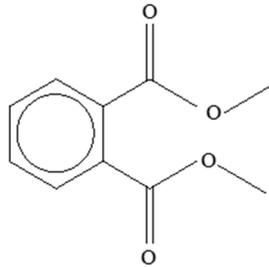
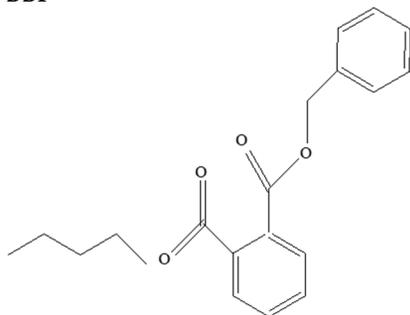
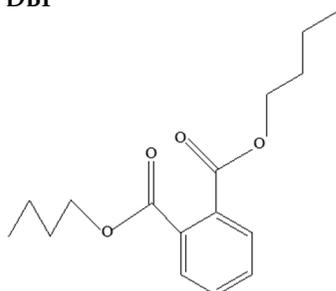
EDCs Present in Packaging Materials	Monomer Structures in the Food	EDC Health Effects	Sources
Plasticizers (phthalates)	<p>DMP</p> 	<p>Has antiandrogenic effects when it interacts with the androgen receptor. Interacts with the aryl hydrocarbon (AhR) and PPAR receptors. Affects thyroid signalling. Reproductive disorders, including low sperm count. Reduced anogenital distance in males. Increased risk of preterm birth. Elevated oestrogen levels in pregnant women. Birth defects. Thyroid axis dysfunction in men. Asthma. Hypospadias. Cryptorchidism. Neurobehaviour problems.</p>	[34,141,143,144]
	<p>BBP</p> 		
	<p>DBP</p> 		

Table 11. Cont.

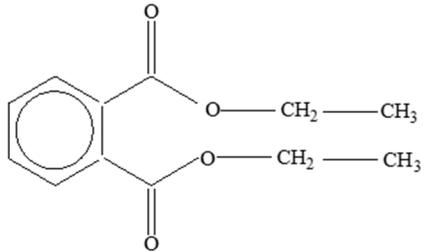
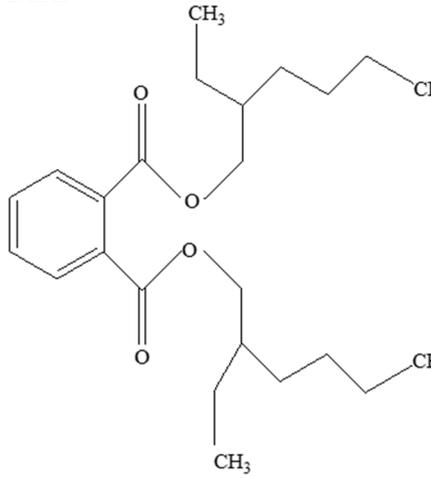
EDCs Present in Packaging Materials	Monomer Structures in the Food	EDC Health Effects	Sources
Plasticizers (phthalates)	<p>DEP</p> 	<p>Has antiandrogenic effects when it interacts with the androgen receptor. Interacts with the aryl hydrocarbon (AhR) and PPAR receptors. Affects thyroid signalling.</p>	[34,141,143,144]
	<p>DEHP</p> 		

Table 11. Cont.

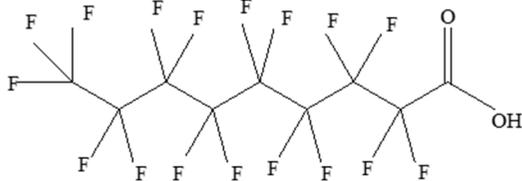
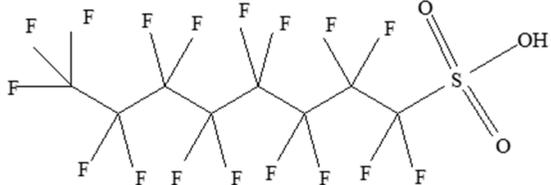
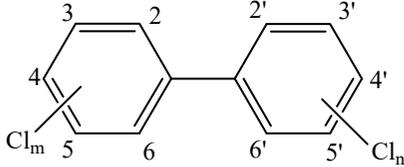
EDCs Present in Packaging Materials	Monomer Structures in the Food	EDC Health Effects	Sources
Perfluoroalkyl substances (PFASs)	<p data-bbox="465 405 904 432">Perfluoroalkyl carboxylic acids (PFCAs)</p>  <p data-bbox="465 632 882 659">Perfluoroalkyl sulfonic acids (PFSAs)</p> 	<p data-bbox="1055 472 1883 794">Decreased thyroid hormones. Has an effect on both pregnant women and children's thyroid hormone levels. Increases hyperactivity. Developmental and immune toxicity. Cancer. Weight gain. Kidney and testicular cancer. Liver degeneration. Changes in nervous system development. Suppressed immune response. Decreased foetal and birth weights.</p>	[145–148]
Dioxins	<p data-bbox="465 916 831 943">Polychlorinated biphenyls (PCB)</p> 	<p data-bbox="1055 874 1621 1166">Increased metabolism. Suppressed concentrations of thyroxine. Reduction in blood insulin and glucose levels. Increase in serum gastrin. Infertility and foetal loss. Decreased spermatogenesis. Decreased circulating androgens. Endometriosis. Inhibition of growth factor and vitamin A expression. Ovarian dysfunction.</p>	[149]

Table 11. Cont.

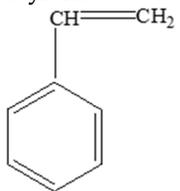
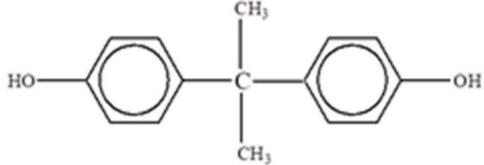
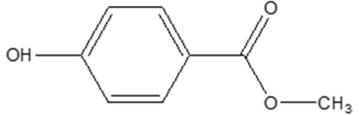
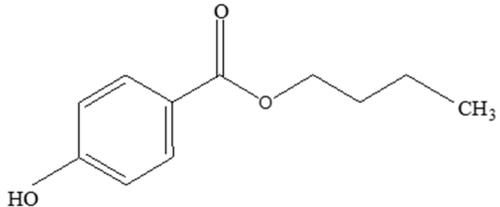
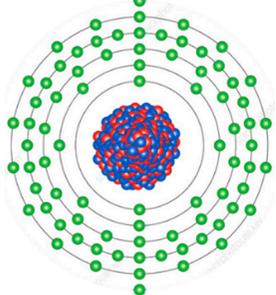
EDCs Present in Packaging Materials	Monomer Structures in the Food	EDC Health Effects	Sources
Styrene	<p>Styrene</p> 	<p>Reproductive toxicity. Developmental toxicity. Impaired immune response to concanavalin and reduced cell-mediated immunity. Neurotoxicity, which includes the suppression of the activity of the central nervous system, including slow reaction time and altered performance on neurobehavioural tests of memory and learning. Respiratory effects, including mucous membrane irritation. Gastrointestinal effects. Effects on the liver, kidney, and eye. Nasal irritation. Lung tumours.</p>	[36]
Bisphenol A	<p>BPA</p> 	<p>Oestrogenic properties. Interacts with a variety of nuclear receptors, including ERR, orphan receptor, oestrogen receptor, glucocorticoid receptor, human oestrogen-related receptor, PPARγ, androgen receptor, and gamma receptor. Disrupts the thyroid axis. Causes metabolic disorders, which result in hyperactivity, neurodevelopment disorders, and type 2 diabetes. Causes infertility. Gut permeability. Breast and prostate cancers. It directly impairs oxidative homeostasis and indirectly impairs redox homeostasis by increasing oxidative mediators and reducing antioxidant enzymes. Increases hydrogen peroxide and lipid peroxidation. Alters organogenesis of kidneys, brain, and testes in foetus. Anxiety in childhood. Cardiovascular function disorders. Increases hydrogen peroxide and lipid peroxidation. In menopausal women, it can bind to ER (oestrogen receptor), triggering noxious cellular responses, such as binding to and stimulating oestrogen receptors (ERs) as well as disrupting action of other steroid hormones and DNA methylation. Disrupts normal action of androgens and alters thyroid hormone synthesis.</p>	[150]

Table 11. Cont.

EDCs Present in Packaging Materials	Monomer Structures in the Food	EDC Health Effects	Sources
Parabens	<p data-bbox="465 408 640 435">Methylparaben</p> 	<p data-bbox="1055 459 1570 486">Exerts oestrogenic and antiandrogenic activities.</p> <p data-bbox="1055 491 1272 518">Results in fecundity.</p> <p data-bbox="1055 523 1406 550">Affects postnatal growth of boys.</p>	[35,151]
	<p data-bbox="465 560 808 587">Butylparaben, isobutylparaben</p> 		
Heavy metals	<p data-bbox="465 823 584 850">Cadmium</p> 	<p data-bbox="1055 887 1883 946">Cadmium, lead, mercury, and aluminium specifically linked to oestrogenic and breast-cancer-related effects.</p> <p data-bbox="1055 951 1883 1010">Mercury compounds also disrupt the thyroid gland function, the hypothalamic–pituitary–adrenal axis, and thyroid hormone function.</p> <p data-bbox="1055 1015 1883 1094">Lead inhibits cellular enzymes and binding of sulfhydryl groups. It also affects membrane stability of red blood cells, inducing functional disturbances in peripheral nerves and development of the skeleton.</p>	[152]

The extent at which humans are exposed to EDCs varies between countries due to variations in regulations [7]. The EU Member State and European Union (EU) regulations, for instance, list about 8030 chemical substances utilized in various food-packaging types [153]. However, in the United States (US) alone, about 10,787 chemical compounds are included in food both indirectly and directly as food additives [154], with many being used under the idea that they are generally recognized as safe (GRAS), even though they have not been reported to the US Food and Drug Administration (FDA) and have possibly not been tested to ascertain public safety. As a result, there is no published information on their use and possible exposure effects [45]. Presently, although in Africa, South Africa is amongst the few countries that control food-packaging materials mostly through general safety requirements due to its membership to the CODEX Alimentarius Commission. The Commission, created by the Food and Agriculture Organization (FAO) together with the World Health Organization (WHO) in 1963, creates international guidelines, food standards, and related texts like the codes of practice under the Joint FAO/WHO Food Standards Programme. In the above context, South Africans are protected by, for instance, the Foodstuffs, Disinfectants, and Cosmetics Act (FDCA) 54 of 1972, last reviewed as of 2009, which regulates food-related issues, such as importation into South Africa, including food packaging [155]. Specific laws that impact food-packaging materials are limited and lacking. They include R879/2011, which only forbids, among other things, the selling of Bisphenol-A-containing polycarbonate baby feeding bottles, and yet the BPA exposure of consumers occurs daily through everyday basic products, like food packaging. In addition, R962/2012 establishes the general hygienic standards for the transport of food and food premises.

According to Section 7(2) of R962/2012 [155]:

“A container shall be clean and free from any toxic substance, ingredient or any other substance liable to contaminate or spoil the food in the container”.

Given that South Africa has, as other upper-middle-income countries have, growing industries which attract an ever increasing population, food-packaging material use is therefore high, and standard consumer utilization practices vary and, in some instances, may possibly not be aligned with the material design. General regulations, such as those above, imply that producers continue to voluntarily comply with the expected standards and, hence, that substances with unacceptable hazard characteristics, such as reproductive toxicity, carcinogenicity, mutagenicity, bioaccumulation, and persistence or endocrine disruption, continue to be common in commerce, including their utilization in food-contact materials. Furthermore, the lack of consumer awareness concerning the chemical compounds in food packaging, such as plastics, and their effects on health continues, which greatly risks the health of consumers. Consumers in emerging countries, including South Africans, are therefore left to look after themselves by self-regulating legislation. There is therefore a significant likelihood that most people are unknowingly subjected to numerous individual and mixtures of FCC-associated chronic diseases [11,39,104].

7. Conclusions

Food packaging fulfils a significant duty in protecting food and enhancing people's standard of living, and, therefore, the utilization of plastic food packaging globally cannot be expected to decrease any time soon because plastic is a unique material with numerous benefits. However, the monomers contained within plastic food packaging are a significant source of food chemical contamination, with endocrine-disrupting effects that affect both current and future human generations and environmental health. Unfortunately, research trends indicate that most monomers' potential harm remains unaccounted for by science. Additionally, food migration studies utilize mostly food simulants rather than food products. Moreover, there is the broad consumer use and misuse of plastic packaging coupled with the nonawareness of the health concerns associated with its incorrect use. The European regulation on food packaging also continues to be criticized for its lack of revision to keep abreast with new scientific developments. Therefore, the research suggests that the entire human population is exposed to harmful substances with known and

unknown effects on health from plastic food packaging. To safeguard both the present and coming generations, the scientific community has more work to do. The identification and understanding of the chemistries of both known and unknown food-packaging chemicals are more than urgent. Furthermore, food migration research studies need to be developed and need to shift their focus to real foods rather than food simulants to avoid generalizing the migration of a group of foods that may seem similar because they belong to the same category of foods and yet differ by one or two chemicals, which has implications in the migration process. Especially in countries with emerging economies, chemical migration awareness and knowledge is crucial and urgent for the relevant legislative authorities for the formulation, development, implementation, enforcement, and review of policies that advocate for sustainable packaging. Awareness and knowledge amongst the general population promotes reflection, which encourages behavioural changes towards the healthy utilization of plastic food packaging and towards checking on the manufacturing compliance with the legislation and regulations on the type of polymer used for food-packaging materials.

Author Contributions: Conceptualization, V.N.-J., T.A.M.M. and C.M.; validation, T.A.M.M.; investigation, C.M.; writing—original draft preparation, C.M.; writing—review and editing, T.A.M.M. and V.N.-J.; funding acquisition, T.A.M.M. All authors have read and agreed to the published version of the manuscript.

Funding: Financial support from the internal grants of the Institute for Nanotechnology and Water Sustainability (iNanoWS) at the University of South Africa, towards the realization of this work is gratefully acknowledged.

Data Availability Statement: No new data were created.

Acknowledgments: The authors wish to thank the Institute for Nanotechnology and Water Sustainability (iNanoWS) in the College of Science and Engineering Technology (CSET) and the Department of Environmental Science in the College of Agriculture and Environmental Sciences (CAES) at the University of South Africa for their support and encouragement.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Ebnasajjad, S. (Ed.) *Plastic Films in Food Packaging Materials, Technology and Applications Plastics Design Library (PDL)*; William Andrew: Amsterdam, The Netherlands, 2013.
2. Coles, R.; McDowell, D.; Kirwan, M.J. *Food Packaging Technology*; Blackwell: Oxford, UK, 2003; Volume 3, pp. 9–15.
3. Leistner, L. Basic aspects of food preservation by hurdle technology. *Int. J. Food Microbiol.* **2000**, *55*, 181–186. [[CrossRef](#)] [[PubMed](#)]
4. Khan, H.; Flint, S.; Yu, P. Enterocins in food preservation. *Int. J. Food Microbiol.* **2010**, *141*, 1–10. [[CrossRef](#)] [[PubMed](#)]
5. Amit, S.K.; Uddin, M.M.; Rahman, R.; Islam, S.M.R.; Khan, M.S. A review on mechanisms and commercial aspects of food preservation and processing. *Agric. Food Secur.* **2017**, *6*, 51. [[CrossRef](#)]
6. Robertson, G.L. *Food Packaging: Principles and Practice*, 3rd ed.; CRC Press: Boca Raton, FL, USA, 2013.
7. Hoseinnejad, M.; Jafari, S.M.; Katouzian, I. Inorganic and metal nanoparticles and their antimicrobial activity in food packaging applications. *Crit. Rev. Microbiol.* **2018**, *44*, 161–181. [[CrossRef](#)]
8. Fortune Business, I. *Food Packaging Market Size, Share and COVID-19 Impact Analysis, by Material (Glass, Metal, Paper and Paperboard, Wood and Plastics), by Product Type (Rigid, Semi-Rigid and Flexible) by Application (Fruits and Vegetables, Bakery and Confectionery, Dairy Pr*; Fortune Business Insights Pvt. Ltd.: Maharashtra, India, 2021.
9. Grandview, R. *Food Packaging Market Size, Share and Growth Report, 2030*; Grand View Research Inc.: San Francisco, CA, USA, 2023.
10. Guillard, V.; Gaucel, S.; Fornaciari, C.; Angellier-Coussy, H.; Buche, P.; Gontard, N. The Next Generation of Sustainable Food Packaging to Preserve Our Environment in a Circular Economy Context. *Front. Nutr.* **2018**, *5*, 1–13. [[CrossRef](#)] [[PubMed](#)]
11. Muncke, J.; Andersson, A.M.; Backhaus, T.; Boucher, J.M.; Carney Almroth, B.; Castillo Castillo, A.; Chevrier, J.; Demeneix, B.A.; Emmanuel, J.A.; Fini, J.B.; et al. Impacts of food contact chemicals on human health: A consensus statement. *Environ. Health* **2020**, *19*, 25. [[CrossRef](#)]
12. Mordor, I. *Packaging Industry in South Africa Market-Growth, Trends, COVID 19 Impact, and Forecasts (2023–2028)*; Mordor Intelligence Pvt. Ltd.: Hyderabad, India, 2023; pp. 1–7.
13. Sadan, Z.; De Kock, L. *Plastics: Facts and Futures: Moving beyond Pollution Management towards a Circular Plastics Economy in South Africa*. 2020. Available online: https://wwfafrica.awsassets.panda.org/downloads/wwf_plastics_report_final_2nov2020.pdf (accessed on 30 September 2022).

14. Geueke, B.; Groh, K.; Muncke, J. Food packaging in the circular economy: Overview of chemical safety aspects for commonly used materials. *J. Clean. Prod.* **2018**, *193*, 491–505. [CrossRef]
15. Ncube, L.K.; Ude, A.U.; Ogunmuyiwa, E.N.; Zulkifli, R.; Beas, I.N. Environmental impact of food packaging materials: A review of contemporary development from conventional plastics to polylactic acid based materials. *Materials* **2020**, *13*, 4994. [CrossRef]
16. Piergiovanni, S.; Limbo, L. *Food Packaging Materials*; Springer: London, UK; New York, NY, USA, 2016; ISBN 9783319247304.
17. Rasul, S.F.; Noori, R.J.; Ali, K.M.; Khdir, R.B.; Ahmed, S.R.; Qadir, A.M. Roles of different packaging materials on the quality and shelf life of yogurt. *Food Sci. Technol.* **2022**, *42*, 1–6. [CrossRef]
18. The Department of Trade, Industry and Competition (DTIC). South African Plastics Industry. 2023. Available online: www.thedtic.gov.za/sectors-and-services-2/industrial-development/plastics/ (accessed on 4 March 2023).
19. Plastics Europe, Plastics—The Facts 2016—An Analysis of European Plastics Production, Demand and Waste Data; Brussels. 2016. Available online: www.plasticseurope.de/informations (accessed on 12 July 2022).
20. Hahladakis, J.N.; Velis, C.A.; Weber, R.; Iacovidou, E.; Purnell, P. An overview of chemical additives present in plastics: Migration, release, fate and environmental impact during their use, disposal and recycling. *J. Hazard. Mater.* **2018**, *344*, 179–199. [CrossRef]
21. Eyerer, P. (Ed.) Synthesis (Manufacture, Production) of Plastics. In *Polymers—Opportunities and Risks 1: General and Environmental Aspects*; Springer: Berlin/Heidelberg, Germany, 2010; Volume 11.
22. Ai, X.; Wang, D.; Li, X.; Pan, H.; Kong, J.; Yang, H.; Zhang, H.; Dong, L. The properties of chemical cross-linked poly(lactic acid) by bis(tert-butyl dioxy isopropyl) benzene. *Polym. Bull.* **2019**, *76*, 575–594. [CrossRef]
23. Groh, K.J.; Backhaus, T.; Carney-Almroth, B.; Geueke, B.; Inostroza, P.A.; Lennquist, A.; Leslie, H.A.; Maffini, M.; Slunge, D.; Trasande, L.; et al. Overview of known plastic packaging-associated chemicals and their hazards. *Sci. Total Environ.* **2019**, *651*, 3253–3268. [CrossRef]
24. Gelbke, H.P.; Banton, M.; Block, C.; Dawkins, G.; Eisert, R.; Leibold, E.; Pemberton, M.; Puijk, I.M.; Sakoda, A.; Yasukawa, A. Risk assessment for migration of styrene oligomers into food from polystyrene food containers. *Food Chem. Toxicol.* **2019**, *124*, 151–167. [CrossRef] [PubMed]
25. Baldev, R. Plastics and their role in food packaging. In *Plastics in Food Packaging*; Central Food Technological Research Institute: Mysore, India, 2005; pp. 1–32.
26. Sothornvit, R.; Krochta, J.M. Plasticizers in edible films and coatings. In *Innovation in Food Packaging. A Volume in Food Science and Technology*; Academic Press: Cambridge, MA, USA, 2005; pp. 403–428.
27. Marsh, K.; Bugusu, B. Food packaging—Roles, materials, and environmental issues. *J. Food Sci.* **2007**, *72*, 39–55. [CrossRef] [PubMed]
28. Hansen, E.; Denmark, C.; Nilsson, N.H.; Lithner, D.; Lassen, C.; Institute, D.T.; Sweden, C.; Lassen, C.-D.C. Hazardous Substances in Plastic Materials. 2013. Available online: http://www.miljodirektoratet.no/no/Publikasjoner/Publikasjoner/2013/Februar/Hazardous_substances_in_plastic_materials/ (accessed on 12 July 2022).
29. Gani, K.M.; Tyagi, V.K.; Kazmi, A.A. Occurrence of phthalates in aquatic environment and their removal during wastewater treatment processes: A review. *Environ. Sci. Pollut. Res. Int.* **2017**, *24*, 17267–17284. [CrossRef]
30. Environmental Protection Agency. *Survey and Investigation of Migration of Monomers in Toy Materials and Investigation of Migration of Monomers in Toy Materials*; Environmental Protection Agency: Washington, DC, USA, 2019.
31. Van Deventer, D.; Mallikarjunan, P. Optimizing an electronic nose for analysis of volatiles from printing inks on assorted plastic films. *Innov. Food Sci. Emerg. Technol.* **2002**, *3*, 93–99. [CrossRef]
32. Bhunia, K.; Sablani, S.S.; Tang, J.; Rasco, B. Migration of Chemical Compounds from Packaging Polymers during Microwave, Conventional Heat Treatment, and Storage. *Compr. Rev. Food Sci. Food Saf.* **2013**, *12*, 523–545. [CrossRef] [PubMed]
33. Kawa, I.A.; Akbar, M.; Fatima, Q.; Mir, S.A.; Jeelani, H.; Manzoor, S.; Rashid, F. Endocrine disrupting chemical Bisphenol A and its potential effects on female health. *Diabetes Metab. Syndr. Clin. Res. Rev.* **2021**, *15*, 803–811. [CrossRef]
34. Wójtowicz, A.K.; Sitarz-Głównia, A.M.; Szczesna, M.; Szychowski, K.A. The Action of Di-(2-Ethylhexyl) Phthalate (DEHP) in Mouse Cerebral Cells Involves an Impairment in Aryl Hydrocarbon Receptor (AhR) Signaling. *Neurotox. Res.* **2019**, *35*, 183–195. [CrossRef]
35. Jurewicz, J.; Radwan, M.; Wielgomas, B.; Karwacka, A.; Klimowska, A.; Kałużny, P.; Radwan, P.; Hanke, W. Parameters of ovarian reserve in relation to urinary concentrations of parabens. *Environ. Health* **2020**, *19*, 1–8. [CrossRef]
36. Ajaj, A.; J'bari, S.; Ononogbo, A.; Buonocore, F.; Bear, J.C.; Mayes, A.G.; Morgan, H. An insight into the growing concerns of styrene monomer and poly(styrene) fragment migration into food and drink simulants from poly(styrene) packaging. *Foods* **2021**, *10*, 1136. [CrossRef]
37. Sax, L. Polyethylene terephthalate May yield endocrine disruptors. *Environ. Health Perspect.* **2010**, *118*, 445–448. [CrossRef] [PubMed]
38. Moon, M.K.; Kim, M.J.; Jung, I.K.; Koo, Y.D.; Ann, H.Y.; Lee, K.J.; Kim, S.H.; Yoon, Y.C.; Cho, B.J.; Park, K.S.; et al. Bisphenol a impairs mitochondrial function in the liver at doses below the no observed adverse effect level. *J. Korean Med. Sci.* **2012**, *27*, 644–652. [CrossRef] [PubMed]
39. Pouech, C.; Kiss, A.; Lafay, F.; Léonard, D.; Wiest, L.; Cren-Olivé, C.; Vulliet, E. Human exposure assessment to a large set of polymer additives through the analysis of urine by solid phase extraction followed by ultra high performance liquid chromatography coupled to tandem mass spectrometry. *J. Chromatogr. A* **2015**, *1423*, 111–123. [CrossRef] [PubMed]
40. HEAL. *Food Contact Materials and Chemical Contamination*; HEAL: Brussels, Belgium, 2016.

41. Caner, C. *Fundamentals of the Sorption (Scalping) Phenomena in Packaged Foods*; Reference Module in Food Science; Elsevier: Amsterdam, The Netherlands, 2018; pp. 1–12.
42. Petrović, E.K.; Hamer, L.K. Improving the healthiness of sustainable construction: Example of polyvinyl chloride (PVC). *Buildings* **2018**, *8*, 28. [[CrossRef](#)]
43. Wang, K.; Zhao, Z.; Ji, W. Bisphenol A induces apoptosis, oxidative stress and inflammatory response in colon and liver of mice in a mitochondria-dependent manner. *Biomed. Pharmacother.* **2019**, *117*, 109182. [[CrossRef](#)] [[PubMed](#)]
44. Martínez-Ibarra, A.; Martínez-Razo, L.D.; MacDonald-Ramos, K.; Morales-Pacheco, M.; Vázquez-Martínez, E.R.; López-López, M.; Rodríguez Dorantes, M.; Cerbón, M. Multisystemic alterations in humans induced by bisphenol A and phthalates: Experimental, epidemiological and clinical studies reveal the need to change health policies. *Environ. Pollut.* **2021**, *271*, 116380. [[CrossRef](#)]
45. Neltner, T.G.; Alger, H.M.; O'Reilly, J.T.; Krinsky, S.; Bero, L.A.; Maffini, M.V. Conflicts of interest in approvals of additives to food: Determined to be generally recognized as safe: Out of balance. *JAMA Intern. Med.* **2013**, *173*, 2032–2036. [[CrossRef](#)]
46. Trier, X.; Taxvig, C.; Rosenmai, A.K.; Pedersen, G.A. *PFAS in Paper and Board for Food Contact, Options for Risk Management of Poly- and Perfluorinated Substances*; Noedic Council of Ministers: Copenhagen, Denmark, 2018.
47. Rosenmai, A.K.; Bengtström, L.; Taxvig, C.; Trier, X.; Petersen, J.H.; Svingen, T.; Binderup, M.; Barbara Medea Alice, v.V.-L.B.M.; Dybdahl, M.; Granby, K. An effect-directed strategy for characterizing emerging chemicals in food contact materials made from paper and board. *Food Chem. Toxicol.* **2017**, *106*, 250–259. [[CrossRef](#)]
48. Wagner, M. Know thy unknowns: Why we need to widen our view on endocrine disruptors. *J. Epidemiol Community Health* **2017**, *71*, 209–212. [[CrossRef](#)]
49. Wiesinger, H.; Wang, Z.; Hellweg, S. Deep Dive into Plastic Monomers, Additives, and Processing Aids. *Environ. Sci. Technol.* **2021**, *55*, 9339–9351. [[CrossRef](#)]
50. Grob, K.; Biedermann, M.; Scherbaum, E.; Roth, M.; Rieger, K. Food contamination with organic materials in perspective: Packaging materials as the largest and least controlled source? A view focusing on the European situation. *Crit. Rev. Food Sci. Nutr.* **2006**, *46*, 529–535. [[CrossRef](#)] [[PubMed](#)]
51. Azizi, D.; Arif, A.; Blair, D.; Dionne, J.; Fillion, Y.; Ouarda, Y.; Pazmino, A.G.; Pulicharla, R.; Rilstone, V.; Tiwari, B.; et al. A comprehensive review on current technologies for removal of endocrine disrupting chemicals from wastewaters. *Environ. Res.* **2022**, *207*, 112196. [[CrossRef](#)]
52. Pilevar, Z.; Bahrami, A.; Beikzadeh, S.; Hosseini, H.; Jafari, S.M. Migration of styrene monomer from polystyrene packaging materials into foods: Characterization and safety evaluation. *Trends Food Sci. Technol.* **2019**, *91*, 248–261. [[CrossRef](#)]
53. González-Sálamo, J.; Socas-Rodríguez, B.; Hernández-Borges, J. Analytical methods for the determination of phthalates in food. *Curr. Opin. Food Sci.* **2018**, *22*, 122–136. [[CrossRef](#)]
54. Giuliani, A.; Zuccarini, M.; Cichelli, A.; Khan, H.; Reale, M. Critical Review on the Presence of Phthalates in Food and Evidence of Their Biological Impact. *Int. J. Environ. Res. Public Health* **2020**, *17*, 5655. [[CrossRef](#)] [[PubMed](#)]
55. Wang, Y.; Qian, H. Phthalates and their impacts on human health. *Healthcare* **2021**, *9*, 603. [[CrossRef](#)] [[PubMed](#)]
56. National Environmental Management: Waste Act 59 of 2008. 2009; pp. 4–6. Available online: <https://www.gov.za/documents/national-environmental-management-waste-act> (accessed on 19 August 2023).
57. National Water Amendment Act 27 of 2014. South African Government. 2014; pp. 4–6. Available online: <https://www.gov.za/documents/national-water-amendment-act-0> (accessed on 19 August 2023).
58. Schrenk, D. Literature Report on Food Packaging Materials and Their Potential Impact on Human Health. 2014. Available online: http://presspage-production-content.s3.amazonaws.com/uploads/1081/profschrenk-foodpackagingmaterials_final.pdf (accessed on 12 July 2022).
59. Geueke, B. Dossier—Non-intentionally added substances (NIAS). *Food Packag. Forum* **2018**, *7*, 1–10. [[CrossRef](#)]
60. Yin, S.; Rajarao, R.; Gong, B.; Wang, Y.; Kong, C.; Sahajwalla, V. Thermo-delamination of metallised composite plastic: An innovative approach to generate Aluminium from packaging plastic waste. *J. Clean. Prod.* **2019**, *211*, 321–329. [[CrossRef](#)]
61. Seiko Kato, L.; Conte-junior, C.A. Safety of Plastic Food Packaging: The Challenges about Identification and Risk Assessment. *Polymers* **2021**, *13*, 33–43.
62. Muncke, J. Tackling the toxics in plastics packaging. *PLoS Biol.* **2021**, *19*, e3000961. [[CrossRef](#)] [[PubMed](#)]
63. Groh, K.J.; Geueke, B.; Martin, O.; Maffini, M.; Muncke, J. Overview of intentionally used food contact chemicals and their hazards. *Environ. Int.* **2021**, *150*, 106225. [[CrossRef](#)]
64. Birgit, G. *Dossier—Non-Intentionally Added Substances (NIAS)*; Food Packaging Forum: Zurich, Switzerland, 2013.
65. Mohanan, N.; Montazer, Z.; Sharma, P.K.; Levin, D.B. Microbial and Enzymatic Degradation of Synthetic Plastics. *Front. Microbiol.* **2020**, *11*, 580709. [[CrossRef](#)] [[PubMed](#)]
66. Walker, T.W.; Frelka, N.; Shen, Z.; Chew, A.K.; Banick, J.; Grey, S.; Kim, M.S.; Dumesic, J.A.; Van Lehn, R.C.; Huber, G.W. Recycling of multilayer plastic packaging materials by solvent-targeted recovery and precipitation. *Sci. Adv.* **2020**, *6*, eaba7599. [[CrossRef](#)] [[PubMed](#)]
67. Fazli, A.; Rodrigue, D. Waste rubber recycling: A review on the evolution and properties of thermoplastic elastomers. *Materials* **2020**, *13*, 782. [[CrossRef](#)]
68. Lohith, K.S.; Mahesh, S.V. Influence of cryogenic treatment on the friction co-efficient of nylon 6 and caprolactam—Graphite composite. *Ipsaj Int. J. Mech. Eng.* **2014**, *1*, 10–15.

69. Alizadeh Sahraei, A.; Mokarizadeh, A.H.; George, D.; Rodrigue, D.; Baniassadi, M.; Foroutan, M. Insights into interphase thickness characterization for graphene/epoxy nanocomposites: A molecular dynamics simulation. *Phys. Chem. Chem. Phys.* **2019**, *21*, 19890–19903. [CrossRef] [PubMed]
70. Liu, X.; Shi, J.; Bo, T.; Li, H.; Crittenden, J.C. Occurrence and risk assessment of selected phthalates in drinking water from waterworks in China. *Environ. Sci. Pollut. Res.* **2015**, *22*, 10690–10698. [CrossRef]
71. Freire, M.T.D.A.; Santana, I.A.; Reyes, F.G.R. Plasticizers in Brazilian food-packaging materials acquired on the retail market. *Food Addit. Contam.* **2006**, *23*, 93–99. [CrossRef]
72. Moore, M.; Han, I.; Acton, J.; Ogale, A.; Barmore, C.; Dawson, P. Effects of Antioxidants in Polyethylene Film on Fresh Beef Color. *J. Food Sci.* **2003**, *68*, 99–104. [CrossRef]
73. Lin, Q.; Li, B.; Song, H.; Li, X. Determination of 7 Antioxidants, 8 Ultraviolet Absorbents, and 2 Fire Retardants in Plastic food Package by Ultrasonic Extraction and Ultra Performance Liquid Chromatography. *J. Liq. Chromatogr. Relat. Technol.* **2011**, *34*, 730–743. [CrossRef]
74. Shaikh, S.; Yaqoob, M.; Aggarwal, P. An overview of biodegradable packaging in food industry. *Curr. Res. Food Sci.* **2021**, *4*, 503–520. [CrossRef] [PubMed]
75. Agarwal, A.; Shaida, B.; Rastogi, M.; Singh, N.B. Food Packaging Materials with Special Reference to Biopolymers-Properties and Applications. *Chem. Africa* **2023**, *6*, 117–144. [CrossRef]
76. Popa, M.; Mitelut, A.; Niculita, P.; Geicu, M.; Ghidurus, M.; Turtoi, M. Biodegradable materials for food packaging applications. *J. Environ. Prot. Ecol.* **2011**, *12*, 1825–1834.
77. Shershneva, E.G. Biodegradable Food Packaging: Benefits and Adverse Effects. *IOP Conf. Ser. Earth Environ. Sci.* **2022**, *988*, 022006. [CrossRef]
78. Bucci, D.Z.; Tavares, L.B.B.; Sell, I. Biodegradation and physical evaluation of PHB packaging. *Polym. Test.* **2007**, *26*, 908–915. [CrossRef]
79. Scarfato, L.; Di Maio, P.; Incarnato, L. Manufacturing of advanced biodegradable polymeric components. *J. Appl. Polym. Sci.* **2015**, *132*, 1–13. [CrossRef]
80. Zhang, W.; Rhim, J.W. Titanium dioxide (TiO₂) for the manufacture of multifunctional active food packaging films. *Food Packag. Shelf Life.* **2022**, *31*, 100806. [CrossRef]
81. Zhang, W.; Sani, M.A.; Zhang, Z.; McClements, D.J.; Jafari, S.M. High performance biopolymeric packaging films containing zinc oxide nanoparticles for fresh food preservation: A review. *Int. J. Biol. Macromol.* **2023**, *230*, 123188. [CrossRef]
82. Zhang, W.; Roy, S.; Rhim, J.W. Copper-based nanoparticles for biopolymer-based functional films in food packaging applications. *Compr. Rev. Food Sci. Food Saf.* **2023**, *22*, 1933–1952. [CrossRef]
83. Gvozdenko, A.A.; Siddiqui, S.A.; Blinov, A.V.; Golik, A.B.; Nagdalian, A.A.; Maglakelidze, D.G.; Statsenko, E.N.; Pirogov, M.A.; Blinova, A.A.; Sizonenko, M.N.; et al. Synthesis of CuO nanoparticles stabilized with gelatin for potential use in food packaging applications. *Sci. Rep.* **2022**, *12*, 12843. [CrossRef] [PubMed]
84. International Panel on Chemical Pollution. Overview Report III: Existing National, Regional, and Global Regulatory Frameworks Addressing Endocrine Disrupting Chemicals (EDCs). United Nations Environment Programme. 2017, pp. 1–59. Available online: <https://wedocs.unep.org/handle/20.500.11822/25636> (accessed on 12 July 2022).
85. Ferreira, A.P. Endocrine disruptors in sludge wastewater treatment plants: Environmental complications. *Acta Sci.* **2013**, *35*, 307–316. [CrossRef]
86. Tapia-Orozco, N.; Ibarra-Cabrera, R.; Tecante, A.; Gimeno, M.; Parra, R.; Garcia-Arrazola, R. Removal strategies for endocrine disrupting chemicals using cellulose-based materials as adsorbents: A review. *J. Environ. Chem. Eng.* **2016**, *4*, 3122–3142. [CrossRef]
87. Arambula, S.E.; Patisaul, H.B. *Endocrine Disrupting Chemicals and Behavior*, 2nd ed.; Elsevier Inc.: Amsterdam, The Netherlands, 2018.
88. Gadupudi, C.K.; Rice, L.; Xiao, L.; Kantamaneni, K. Endocrine Disrupting Compounds Removal Methods from Wastewater in the United Kingdom: A Review. *Science* **2019**, *1*, 11. [CrossRef]
89. Ribeiro, E.; Ladeira, C.; Viegas, S. EDCs mixtures: A stealthy hazard for human health? *Toxics* **2017**, *5*, 5. [CrossRef]
90. Gassman, N.R. Induction of oxidative stress by bisphenol A and its pleiotropic effects. *Environ. Mol. Mutagen.* **2017**, *58*, 60–71. [CrossRef]
91. Meli, R.; Monnolo, A.; Annunziata, C.; Pirozzi, C.; Ferrante, M.C. Oxidative stress and BPA toxicity: An antioxidant approach for male and female reproductive dysfunction. *Antioxidants* **2020**, *9*, 405. [CrossRef]
92. Tavakkoli, A.; Abnous, K.; Vahdati Hassani, F.; Hosseinzadeh, H.; Birner-Gruenberger, R.; Mehri, S. Alteration of protein profile in cerebral cortex of rats exposed to bisphenol a: A proteomics study. *Neurotoxicology* **2020**, *78*, 1–10. [CrossRef]
93. Patisaul, H.B.; Roberts, S.C.; Mabrey, N.; Mccaffrey, K.A.; Gear, R.B.; Braun, J.; Belcher, S.M.; Stapleton, H.M. Accumulation and Endocrine Disrupting Effects of the Flame Retardant Mixture Firemaster 550 in Rats: An Exploratory Assessment. *J. Biochem. Mol. Toxicol.* **2013**, *27*, 124–136. [CrossRef]
94. Ciacci, L.; Passarini, F.; Vassura, I. The European PVC cycle: In-use stock and flows. *Resour. Conserv. Recycl.* **2017**, *123*, 108–116. [CrossRef]
95. Biro, F.M.; Greenspan, L.C.; Galvez, M.P.; Pinney, S.M.; Teitelbaum, S.; Windham, G.C.; Dearthoff, J.; Herrick, R.L.; Succop, P.A.; Hiatt, R.A.; et al. Onset of breast development in a longitudinal cohort. *Pediatrics* **2013**, *132*, 1019–1027. [CrossRef]

96. Talpade, J.; Shrman, K.; Sharma, R.K.; Gutham, V.; Singh, I.R.; Meena, N.S. Bisphenol a: An endocrine disruptor. *J. Entomol. Zool. Stud.* **2018**, *6*, 394–397. Available online: <http://www.entomoljournal.com/archives/2018/vol6issue3/PartF/6-2-262-216.pdf> (accessed on 12 July 2022).
97. Alin, J.; Hakkarainen, M. Migration from polycarbonate packaging to food simulants during microwave heating. *Polym. Degrad. Stab.* **2012**, *97*, 1387–1395. [[CrossRef](#)]
98. Maryskova, A.; Rysova, M.; Novotny, M.; Sevcu, V. Polyamide-Laccase Nanofiber Membrane for Degradation of Endocrine-Disrupting Bisphenol A, 17 α -ethinylestradiol, and Triclosan. *Polymers* **2019**, *11*, 1560. [[CrossRef](#)]
99. Alamri, M.S.; Qasem, A.A.A.; Mohamed, A.A.; Hussain, S.; Ibraheem, M.A.; Shamlan, G.; Alqah, H.A.; Qasha, A.S. Food packaging's materials: A food safety perspective. *Saudi J. Biol. Sci.* **2021**, *28*, 4490–4499. [[CrossRef](#)]
100. Eker, B.; Icoz, A. *Packaging Materials and Effects on Quality of Life*; Center for Quality, Faculty of Engineering, University of Kragujevac: Kragujevac, Serbia, 2016; pp. 271–278.
101. Kradri, Z.; Mechnou, I.; Zyade, S. Migration of bisphenol A from epoxy coating to foodstuffs. *Mater. Today Proc.* **2021**, *45*, 7584–7587. [[CrossRef](#)]
102. *BS EN 1186-1:2002*; Materials and Articles in Contact with Foodstuffs—Plastics—Part 1: Guide to the Selection of Conditions and Test Methods for Overall Migration. British Standards Institution: London, UK, 2002; pp. 1–49.
103. European Commission. Commission Regulation (EU) No 10/2011 of 14 January 2011. 2011. Available online: <https://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2011:012:0001:0089:EN:PDF> (accessed on 12 July 2022).
104. İçöz, A.; Eker, B. Selection of Food Packaging Material, Migration and Its Effects on Food Quality. In Proceedings of the 1st International Conference on Quality of Life, Solo, Indonesia, 14–15 September 2016; pp. 201–210. Available online: http://cq.m.rs/2016/cd1/pdf/papers/focus_1/28.pdf (accessed on 18 July 2022).
105. Castle, L.; Defra, C.S.L. Chemical migration into food: An overview. In *Chemical Migration and Food Contact Materials*; Barnes, K., Sinclair, C., Watson, D., Eds.; Woodhead Publishing: Sawston, UK; CRC Press: Boca Raton, FL, USA, 2007; pp. 1–481, ISBN 9772081415.
106. Makowska, K.; Staniszewska, M.; Bodziach, K.; Calka, J.; Gonkowski, S. Concentrations of bisphenol a (BPA) in fresh pork loin meat under standard stock-farming conditions and after oral exposure—A preliminary study. *Chemosphere* **2022**, *295*, 133816. [[CrossRef](#)] [[PubMed](#)]
107. Kubwabo, C.; Kosarac, I.; Stewart, B.; Gauthier, B.R.; Lalonde, K.; Lalonde, P.J. Migration of bisphenol A from plastic baby bottles, baby bottle liners and reusable polycarbonate drinking bottles. *Food Addit. Contam.* **2009**, *26*, 928–937. [[CrossRef](#)]
108. Munguia-Lopez, E.M.; Soto-Valdez, H. Effect of heat processing and storage time on migration of bisphenol A (BPA) and bisphenol A-diglycidyl ether (BADGE) to aqueous food simulant from Mexican can coatings. *J. Agric. Food Chem.* **2001**, *49*, 3666–3671. [[CrossRef](#)]
109. Marć, M.; Formela, K.; Klein, M.; Namieśnik, J.; Zabiegała, B. The emissions of monoaromatic hydrocarbons from small polymeric toys placed in chocolate food products. *Sci. Total Environ.* **2015**, *530*, 290–296. [[CrossRef](#)] [[PubMed](#)]
110. Tawfik, M.S.; Huyghebaert, A. Polystyrene cups and containers: Styrene migration. *Food Addit. Contam.* **1998**, *15*, 592–599. [[CrossRef](#)] [[PubMed](#)]
111. Guazzotti, V.; Hendrich, V.; Gruner, A.; Fiedler, D.; Störmer, A.; Welle, F. Migration of Styrene in Yogurt and Dairy Products Packaged in Polystyrene: Results from Market Samples. *Foods* **2022**, *11*, 2120. [[CrossRef](#)] [[PubMed](#)]
112. Lickly, T.D.; Lehr, K.M.; Welsh, G.C. Migration of styrene from polystyrene foam food-contact articles. *Food Chem. Toxicol.* **1995**, *33*, 475–481. [[CrossRef](#)]
113. El-Ziney, M.G.; Tawfik, M.S. Migration Levels of Monostyrene from Polystyrene Containers to Dairy Products. *MOJ Food Process. Technol.* **2016**, *3*, 267–271. [[CrossRef](#)]
114. Khaksar, M.; Ghazi-Khansari, M. Determination of migration monomer styrene from GPPS (general purpose polystyrene) and HIPS (high impact polystyrene) cups to hot drinks. *Toxicol. Mech. Methods* **2009**, *19*, 257–261. [[CrossRef](#)]
115. Bradley, E.; Coulier, L. Report FD 07/01: An Investigation into the Reaction and Breakdown Products from Starting Substances Used to Produce Food Contact Plastics. 2007. Available online: <http://www.foodpackagingforum.org/wp-content/uploads/2014/06/Bradley-and-Coulier-2007.pdf> (accessed on 12 July 2022).
116. Begley, T.H.; Gay, M.L.; Hollifield, H.C. Determination of migrants in and migration from nylon food packaging. *Food Addit. Contam.* **1995**, *12*, 671–676. [[CrossRef](#)]
117. Bomfim, M.V.J.; Zamith, H.P.S.; Abrantes, S.M.P. Migration of ϵ -caprolactam residues in packaging intended for contact with fatty foods. *Food Control* **2011**, *22*, 681–684. [[CrossRef](#)]
118. European Commission. Commission Regulation (EC) No 1282/2011 of 28 November 2011 Amending and Correcting Commission Regulation (EU) No 10/2011 on Plastic Materials and Articles Intended to Come into Contact with Food. 2011. Available online: <https://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2011:328:0022:0029:En:PDF> (accessed on 12 July 2022).
119. Huang, H.B.; Pan, W.H.; Chang, J.W.; Chiang, H.C.; Guo, Y.L.; Jaakkola, J.J.K.; Huang, P.C. Does exposure to phthalates influence thyroid function and growth hormone homeostasis? The Taiwan Environmental Survey for Toxicants (TEST) 2013. *Environ. Res.* **2017**, *153*, 63–72. [[CrossRef](#)]
120. Younker, J.M.; Beste, A.; Iii, A.C.B. Computational study of bond dissociation enthalpies for lignin model compounds: B-5 Arylcoumaran. *Chem. Phys. Lett.* **2012**, *545*, 100–106. [[CrossRef](#)]

121. Huang, J.B.; Zeng, G.S.; Li, X.S.; Cheng, X.C.; Tong, H. Theoretical studies on bond dissociation enthalpies for model compounds of typical plastic polymers. *IOP Conf. Ser. Earth Environ. Sci.* **2018**, *167*, 012029. [[CrossRef](#)]
122. Majder-Łopatka, M.; Węsierski, T.; Ankowski, A.; Ratajczak, K.; Duraliski, D.; Piechota-Polanczyk, A.; Polanczyk, A. Thermal analysis of plastics used in the food industry. *Materials* **2022**, *15*, 248. [[CrossRef](#)] [[PubMed](#)]
123. Begley, T.H.; Biles, J.E.; Cunningham, C.; Piringer, O. Migration of a UV stabilizer from polyethylene terephthalate (PET) into food simulants. *Food Addit. Contam.* **2004**, *21*, 1007–1014. [[CrossRef](#)] [[PubMed](#)]
124. Naziruddin, M.A.; Nurulhuda, K.; Sulaiman, R.; Sanny, M. Assessment of residual styrene monomer migration into yoghurt packed in High Impact Polystyrene pots using a modelling approach. *Food Control* **2023**, *148*, 109612. [[CrossRef](#)]
125. Mercea, P. Physicochemical Processes Involved in Migration of Bisphenol A from Polycarbonate. *J. Appl. Polym. Sci.* **2009**, *112*, 579–593. [[CrossRef](#)]
126. Oduneye, T. *Migration of Styrene from Polystyrene Food Containers into Food Simulants: A Meta-Analysis*; Master of Applied Science; Dalhousie University: Halifax, NS, Canada, 2020; Volume 91, pp. 1–108.
127. Muncke, J. Endocrine disrupting chemicals and other substances of concern in food contact materials: An updated review of exposure, effect and risk assessment. *J. Steroid Biochem. Mol. Biol.* **2011**, *127*, 118–127. [[CrossRef](#)] [[PubMed](#)]
128. Triantafyllou, V.I.; Akrida-Demertzi, K.; Demertzis, P.G. A study on the migration of organic pollutants from recycled paperboard packaging materials to solid food matrices. *Food Chem.* **2007**, *101*, 1759–1768. [[CrossRef](#)]
129. Goodson, A.; Robin, H.; Summerfield, W.; Cooper, I. Migration of bisphenol A from can coatings—Effects of damage, storage conditions and heating. *Food Addit. Contam.* **2004**, *21*, 1015–1026. [[CrossRef](#)]
130. Paraskevopoulou, D.; Achilias, D.S.; Paraskevopoulou, A. Migration of styrene from plastic packaging based on polystyrene into food simulants. *Polym. Int.* **2012**, *61*, 141–148. [[CrossRef](#)]
131. Anderson, W.A.C.; Castle, L. Benzophenone in cartonboard packaging materials and the factors that influence its migration into food. *Food Addit. Contam.* **2003**, *20*, 607–618. [[CrossRef](#)]
132. Castle, L.; Mayo, A.; Crews, C.; Gilbert, J. Migration of poly(ethylene terephthalate) (PET) oligomers from PET plastics into food during microwave and conventional cooking and into bottled beverages. *J. Food Prot.* **1989**, *52*, 337–342. [[CrossRef](#)]
133. Arvanitoyannis, I.S.; Bosnea, L. Migration of Substances from Food Packaging Materials to Foods. *Crit. Rev. Food Sci. Nutr.* **2004**, *44*, 63–76. [[CrossRef](#)] [[PubMed](#)]
134. Lickly, T.D.; Breder, C.V.; Rainey, M.L. A Model for Estimating the Daily Dietary Intake of a Substance from Food-Contact Articles: Styrene from Polystyrene Food Contact Polymers. *Regul. Toxicol. Pharmacol.* **1995**, *21*, 406–417. [[CrossRef](#)] [[PubMed](#)]
135. Nam, S.H.; Seo, Y.M.; Kim, M.G. Bisphenol A migration from polycarbonate baby bottle with repeated use. *Chemosphere* **2010**, *79*, 949–952. [[CrossRef](#)]
136. Poças, M.; Oliveira, J.C.; Pereira, J.R.; Brandsch, R.; Hogg, T. Modelling migration from paper into a food simulant. *Food Control* **2011**, *22*, 303–312. [[CrossRef](#)]
137. Boccaci Mariani, M.; Chiacchierini, E.; Gesumundo, C. Potential migration of Diisopropyl naphthalenes from recycled paperboard packaging into dry foods. *Food Addit. Contam.* **1999**, *16*, 207–213. [[CrossRef](#)] [[PubMed](#)]
138. Garban, Z.; Garban, G.; Ghibu, G.D.; Baltă, C.; Avacovici, A.E.; Mitroi, E.M.; Miclău, L. Xenobiochemistry at the interface of packaging materials-food. Note I. Packaging materials and specific interactions in vivo. *J. Agroalim. Process. Technol.* **2010**, *16*, 265–275.
139. Acerini, C.L.; Hughes, I.A. Endocrine disrupting chemicals: A new and emerging public health problem? *Arch. Dis. Child.* **2006**, *91*, 633–638. [[CrossRef](#)]
140. Groh, K.J.; Muncke, J. In Vitro Toxicity Testing of Food Contact Materials: State-of-the-Art and Future Challenges. *Compr. Rev. Food Sci. Food Saf.* **2017**, *16*, 1123–1150. [[CrossRef](#)]
141. Zelko, I.N.; Taylor, B.S.; Das, T.P.; Watson, W.H.; Sithu, I.D.; Wahlang, B.; Malovichko, M.V.; Cave, M.C.; Srivastava, S. Effect of vinyl chloride exposure on cardiometabolic toxicity. *Environ. Toxicol.* **2022**, *37*, 245–255. [[CrossRef](#)]
142. Jones, B.A.; Watson, N.V. Perinatal BPA exposure demasculinizes males in measures of affect but has no effect on water maze learning in adulthood. *Horm. Behav.* **2012**, *61*, 605–610. [[CrossRef](#)]
143. Zhu, J.; Wang, Q.; Han, L.; Zhang, C.; Wang, Y.; Tu, K.; Peng, J.; Wang, J.; Pan, L. Effects of caprolactam content on curdlan-based food packaging film and detection by infrared spectroscopy. *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* **2021**, *245*, 118942. [[CrossRef](#)] [[PubMed](#)]
144. Rudel, R.A.; Gray, J.M.; Engel, C.L.; Rawsthorne, T.W.; Dodson, R.E.; Ackerman, J.M.; Rizzo, J.; Nudelman, J.L.; Brody, J.G. Food packaging and bisphenol A and bis(2-ethylhexyl) phthalate exposure: Findings from a dietary intervention. *Environ. Health Perspect.* **2011**, *119*, 914–920. [[CrossRef](#)] [[PubMed](#)]
145. Genthe, B.; Steyn, M. *Health Risk Assessment Protocol for Endocrine Disrupting Chemicals*; CSIR, Natural Resources and the Environment: Stellenbosch, South Africa, 2008.
146. Stroheker, T.; Cabaton, N.; Nourdin, G.; Régnier, J.F.; Lhuguenot, J.C.; Chagnon, M.C. Evaluation of anti-androgenic activity of di-(2-ethylhexyl)phthalate. *Toxicology* **2005**, *208*, 115–121. [[CrossRef](#)] [[PubMed](#)]
147. Boas, M.; Frederiksen, H.; Feldt-Rasmussen, U.; Skakkebaek, N.E.; Hegedüs, L.; Hilsted, L.; Juul, A.; Main, K.M. Childhood exposure to phthalates: Associations with thyroid function, insulin-like growth factor I, and growth. *Environ. Health Perspect.* **2010**, *118*, 1458–1464. [[CrossRef](#)] [[PubMed](#)]

148. Wolff, M.S.; Teitelbaum, S.L.; McGovern, K.; Windham, G.C.; Pinney, S.M.; Galvez, M.; Calafat, A.M.; Kushi, L.H.; Biro, F.M. Phthalate exposure and pubertal development in a longitudinal study of US girls. *Hum. Reprod.* **2014**, *29*, 1558–1566. [[CrossRef](#)] [[PubMed](#)]
149. Dufour, P.; Pirard, C.; Seghaye, M.; Charlier, C. Association between organohalogenated pollutants in cord blood and thyroid function in newborns and mothers from Belgian population. *Environ. Pollut.* **2018**, *238*, 389–396. [[CrossRef](#)] [[PubMed](#)]
150. Peretz, J.; Vrooman, L.; Ricke, W.A.; Hunt, P.A.; Ehrlich, S.; Hauser, R.; Padmanabhan, V.; Taylor, H.S.; Swan, S.H.; Vandervoort, C.A.; et al. Bisphenol A and reproductive health: Update of Experimental and Human Evidence, 2007–2013. *Environ. Health Perspect.* **2014**, *122*, 775–786. [[CrossRef](#)]
151. Kim, J.; Chevrier, J. Exposure to parabens and prevalence of obesity and metabolic syndrome: An analysis of the Canadian Health Measures Survey. *Sci. Total Environ.* **2020**, *713*, 135116. [[CrossRef](#)]
152. Shimizu, Y.; Kambayashi, Y.; Tsujiguchi, H.; Hara, A.; Hori, D.; Thi Thu Nguyen, T.; Suzuki, F.; Hamagishi, T.; Yamada, Y.; Nakamura, H.; et al. Relationship between the Use of Parabens and Allergic Diseases in Japanese Adults—A Cross-Sectional Study. *J. Multidiscip. Sci. J.* **2018**, *1*, 148–158. [[CrossRef](#)]
153. Simoneau, C.; Raffael, B.; Garbin, S.; Hoekstra, E.; Mieth, A.; Lopes, J.A.; Reina, V. JRC Science for Policy Report: Non-Harmonised Food Contact Materials in the EU: Regulatory and Market Situation Baseline Study Final Report. 2016. Available online: <https://doi.org/10.2788/234276> (accessed on 12 July 2022).
154. Neltner, T.G.; Kulkarni, N.R.; Alger, H.M.; Maffini, M.V.; Bongard, E.D.; Fortin, N.D.; Olson, E.D. Navigating the U.S. Food Additive Regulatory Program. *Compr. Rev. Food Sci. Food Saf.* **2011**, *10*, 342–368. [[CrossRef](#)]
155. South Africa. Department of Health, Foodstuffs, Cosmetics and Disinfectants Act, 1972 (Act No. 54 of 1972). 2018. Available online: www.gpwonline.co.za (accessed on 12 July 2022).

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.