

**Supplementary Table S1. Primers used for PCR**

Primer	Target	Sequence 5'-3'	Reference
<b>1- <math>\beta</math>-Lactamases</b>			
TEM-F	<i>bla<sub>TEM</sub></i>	ATAAAATTCTTGAAGACGAAA	[1]
TEM-R		GACAGTTACCAATGCTTAATC	
SHV-F	<i>bla<sub>SHV</sub></i>	TTATCTCCCTGTTAGCCACC	[1]
SHV-R		GATTGCTGATTTCGCTCGG	
OXA- F	<i>bla<sub>OXA</sub></i>	TCAACTTCAAGATCGCA	[2]
OXA-R		GTGTGTTAGAATGGTGA	
CTX-M-F	<i>bla<sub>CTX-M</sub></i>	CGCTTGCGATGTGCAG	[3]
CTX-M-R		ACCGCGATATCGTTGGT	
CMY-F	<i>bla<sub>CMY</sub></i>	GACAGCCTTTCTCCACA	[4]
CMY-R		TGGAACGAAGGCTACGTA	
OXY-F	<i>bla<sub>OXY</sub></i>	GTTTTGGTAACTGTGACGGG	[5]
OXY-R		AGAGTCAGAGTGTGTCAG	
<b>2- Plasmid-mediated quinolone resistance genes</b>			
qnrA-F	<i>qnrA</i>	ATTCTCACGCCAGGATTG	[6]
qnrA-R		GATCGGAAAGGTTAGGTCA	
qnrB-F	<i>qnrrB</i>	GATCGTGAAGCCAGAAAGG	[6]
qnrB-R		ACGATGCCCTGGTAGTTGTCC	
qnrS-F	<i>qnrS</i>	ACGACATTCTGCAACTGCAA	[6]
qnrS-R		TAAATTGGCACCTGTAGGC	
aac(6')-Ib-F	<i>aac(6')-Ib</i>	TTGCGATGCTCATGAGTGGCTA	[7]
aac(6')-Ib-R		CTCGAATGCCCTGGCGTCTT	
<b>3- Tetracyclines</b>			
<i>tet(A)-F</i>	<i>tet(A)</i>	GCTACATCCTGCTTGCCTTC	[8]
<i>tet(A)-R</i>		CATAGATGCCGTGAAGAGG	
<i>tet(B)-F</i>	<i>tet(B)</i>	TTGGTTAGGGCAAGTTTG	[8]
<i>tet(B)-R</i>		GTAATGGCCAATAACACCG	
<i>tet(C)-F</i>	<i>tet(C)</i>	CTTGAGAGCCTCAACCCAG	[8]
<i>tet(C)-R</i>		ATGGTCGTATCTACCTGCC	
<i>tet(D)-F</i>	<i>tet(D)</i>	AAACCATTACGGCATTCTGC	[8]
<i>tet(D)-R</i>		GACCGGATAACACCATCCATC	
<i>tet(E)-F</i>	<i>tet(E)</i>	AAACACATCCTCCATACGC	[8]
<i>tet(E)-R</i>		AAATAGGCCACAACCGTCAG	
<b>4- Phylogenetic group</b>			
ChuA.1b	<i>chuA</i>	ATGGTACCGGACGAACCAAC	[9]
ChuA.2.2		TGCCGCCAGTACCAAAGACA	
YjaA.1b	<i>yjaA</i>	CAAACGTGAAGTGTCAAGGAG	[9]
YjaA.2b		AATGCGTTCTCAACCTGTG	
TspE4C2.1b	<i>tspE4C</i>	CACTATTGTAAGGTATC	[9]
TspE4C2b		AGTTTATCGCTGCCAGGTG	
AceK.f	<i>arpA</i>	AACGCTATTGCCAGCTTGC	[9]
ArpA1.r		TCTCCCCATACCGTACGCTA	
<b>5- Virulence genes</b>			
ETEC	<i>lt</i>	GGCGACAGATTACCGTGC	[10]
	<i>st</i>	CGGTCTATATTCCCTGTT	[10]
EPEC	<i>bfpA</i>	ATTTTCTTCTGTATTGTCTT	[10]
	<i>eaeA</i>	CACCCGGTACAAGCAGGATT	[10]
STEC	<i>stx1</i>	AATGGTGCTTGCCTGCTGC	[10]
	<i>stx2</i>	GCCGTTATCCAACCTGGTA	[10]
EIEC	<i>ial</i>	GACCCGGCACAAGCATAAGC	[10]
		CCACCTGCAGCAACAAAGAGG	
		CTGGATTAAATGCGCATAGTG	[10]
		AGAACGCCACTGAGATCATC	
		GGCACTGCTGAAACTGCTCC	[10]
		TCGCCAGTTATCTGACATTCTG	
		GGTATGATGATGATGAGTCCA	[10]
		GGAGGCCACAATTATTCC	
<b>6- ERIC-PCR</b>			
ERIC2	ERIC	AAGTAAGTGACTGGGTGAGCG	[11]

## References

1. Weill, F.X.; Demartin, M.; Tande, D.; Espie, E.; Rakotoarivony, I.; Grimont, P.A. SHV-12-like extended-spectrum-beta-lactamase-producing strains of *Salmonella enterica* serotypes Babelsberg and Enteritidis isolated in France among infants adopted from Mali. *J. Clin. Microbiol.* **2004**, *42*, 2432–2437, <https://doi.org/10.1128/JCM.42.6.2432-2437.2004>.
2. Ahmed, A.M.; Motoi, Y.; Sato, M.; Maruyama, A.; Watanabe, H.; Fukumoto, Y.; Shimamoto, T. Zoo animals as reservoirs of gram-negative bacteria harboring integrons and antimicrobial resistance genes. *Appl. Environ. Microbiol.* **2007**, *73*, 6686–6690, <https://doi.org/10.1128/AEM.01054-07>.
3. Bonnet, R.; Sampaio, J.L.; Labia, R.; De Champs, C.; Sirot, D.; Chanal, C.; Sirot, J. A novel CTX-M β-lactamase (CTX-M-8) in cefotaxime-resistant *Enterobacteriaceae* isolated in Brazil. *Antimicrob. Agents Chemother.* **2000**, *44*, 1936–1942, <https://doi.org/10.1128/aac.44.7.1936-1942.2000>.
4. Briñas, L.; Zarazaga, M.; Saenz, Y.; Ruiz-Larrea, F.; Torres, C. β-lactamases in ampicillin-resistant *Escherichia coli* isolates from foods, humans, and healthy animals. *Antimicrob. Agents Chemother.* **2002**, *46*, 3156–3163, <https://doi.org/10.1128/aac.46.10.3156-3163.2002>.
5. Fevre, C.; Jbel, M.; Passet, V.; Weill, F.X.; Grimont, P.A.; Brisson, S. Six groups of the OXY β-lactamase evolved over millions of years in *Klebsiella oxytoca*. *Antimicrob. Agents Chemother.* **2005**, *49*, 3453–3462, <https://doi.org/10.1128/AAC.49.8.3453-3462.2005>.
6. Cattoir, V.; Poirel, L.; Rotimi, V.; Soussy, C.J.; Nordmann, P. Multiplex PCR for detection of plasmid-mediated quinolone resistance *qnr* genes in ESBL-producing enterobacterial isolates. *J. Antimicrob. Chemother.* **2007**, *60*, 394–397, <https://doi.org/10.1093/jac/dkm204>.
7. Robicsek, A.; Jacoby, G.A.; Hooper, D.C. The worldwide emergence of plasmid-mediated quinolone resistance. *Lancet Infect. Dis.* **2006**, *6*, 629–640, [https://doi.org/10.1016/S1473-3099\(06\)70599-0](https://doi.org/10.1016/S1473-3099(06)70599-0).
8. Ng, L.K.; Martin, I.; Alfa, M.; Mulvey, M. Multiplex PCR for the detection of tetracycline resistant genes. *Mol. Cell Probes* **2001**, *15*, 209–215, <https://doi.org/10.1006/mcpr.2001.0363>.
9. Clermont, O.; Christenson, J.K.; Denamur, E.; Gordon, D.M. The Clermont *Escherichia coli* phylo-typing method revisited: Improvement of specificity and detection of new phylo-groups. *Environ. Microbiol. Rep.* **2013**, *5*, 58–65, <https://doi.org/10.1111/1758-2229.12019>.
10. López-Saucedo, C.; Cerna, J.F.; Villegas-Sepulveda, N.; Thompson, R.; Velazquez, F.R.; Torres, J.; Tarr, P.I.; Estrada-Garcia, T. Single multiplex polymerase chain reaction to detect diverse loci associated with diarrheagenic *Escherichia coli*. *Emerg. Infect. Dis.* **2003**, *9*, 127–131, <https://doi.org/10.3201/eid0901.010507>.
11. Versalovic, J.; Koeuth, T.; Lupski, J.R. Distribution of repetitive DNA sequences in eubacteria and application to fingerprinting of bacterial genomes. *Nucleic Acids Res.* **1991**, *19*, 6823–6831, <https://doi.org/10.1093/nar/19.24.6823>.