

**Table S1.** *In vitro* and *in vivo* antifungal results of pristine CEO compared to BCNC/GelA-CEO emulsions against *P. expansum* and *B. cinerea*. For any investigated test/fungus pair, different lowercase letters within the same row denote significant differences ( $p < 0.05$ ) between mean values due to the different modes of administration of CEO (pristine or encapsulated).

	Concentrations [μL/L]	In vitro test (AI after 2 weeks, %)		In vivo test (Lesion diameter after 21 days, mm)	
		Pristine CEO	BCNC/GelA- CEO emulsions	Pristine CEO	BCNC/GelA- CEO emulsions
<i>P. expansum</i>	75	29.3 ± 1.8 <sup>b</sup>	19.4 ± 4.4 <sup>a</sup>	39.1 ± 1.0 <sup>b</sup>	19.0 ± 2.6 <sup>a</sup>
	150	55.4 ± 1.1 <sup>b</sup>	23.7 ± 3.5 <sup>a</sup>	27.5 ± 2.1 <sup>b</sup>	14.0 ± 1.0 <sup>a</sup>
	300	100.0 ± 0.0 <sup>b</sup>	45.7 ± 10.1 <sup>a</sup>	14.2 ± 0.5 <sup>b</sup>	9.5 ± 0.5 <sup>a</sup>
	600	100.0 ± 0.0 <sup>a</sup>	100.0 ± 0.0 <sup>a</sup>	9.2 ± 1.3 <sup>a</sup>	7.0 ± 1.3 <sup>a</sup>
	1200	100.0 ± 0.0 <sup>a</sup>	100.0 ± 0.0 <sup>a</sup>	6.2 ± 2.2 <sup>a</sup>	6.3 ± 1.0 <sup>a</sup>
<i>B. cinerea</i>		In vitro test (AI after 2 weeks, %)		In vivo test (Lesion diameter after 21 days, mm)	
	75	3.4 ± 0.5 <sup>a</sup>	44.6 ± 8.1 <sup>b</sup>	96.7 ± 3.2 <sup>b</sup>	80.3 ± 3.1 <sup>a</sup>
	150	89.6 ± 1.3 <sup>a</sup>	90.7 ± 16.0 <sup>a</sup>	65.3 ± 2.4 <sup>b</sup>	59.6 ± 2.6 <sup>a</sup>
	300	100.0 ± 0.0 <sup>a</sup>	100.0 ± 0.0 <sup>a</sup>	42.7 ± 6.4 <sup>a</sup>	50.3 ± 3.2 <sup>a</sup>
	600	100.0 ± 0.0 <sup>a</sup>	100.0 ± 0.0 <sup>a</sup>	27.2 ± 4.3 <sup>a</sup>	25.3 ± 4.1 <sup>a</sup>
	1200	100.0 ± 0.0 <sup>a</sup>	100.0 ± 0.0 <sup>a</sup>	11.9 ± 5.1 <sup>a</sup>	15.3 ± 1.5 <sup>a</sup>

**Table S2.** Chemical composition of CEO, ZEO, and SEO by GC-MS analysis (adapted from Tahmasebi et al., 2020).

components	CEO (%)	ZEO (%)	SEO (%)	Retention index
<i>α-Thujene</i>	1.53	3.63	2.32	933
<i>α-pinene</i>	-	0.43	2.64	938
<i>Benzaldehyde</i>	0.37	-	-	944
<i>Camphene</i>	-	0.15	-	946
<i>3-Octanone</i>	-	0.14	-	966
<i>β-pinene</i>	-	0.39	1.83	973
<i>β-Myrcene</i>	-	0.71	2.47	981
<i>α-Phellandrene</i>	-	0.13	-	990
<i>3-Carene</i>	0.65	-	-	998
<i>α-Terpinene</i>	-	0.52	5.76	1008
<i>p-Cymene</i>	1.54	8.94	16.55	1009
<i>Limonene</i>	-	0.56	1.45	1018
<i>β-Phellandrene</i>	0.37	-	-	1024

<i>Eucalyptol</i>	1.15	0.43	-	1034
<b><i>γ-Terpinene</i></b>	0.08	<b>5.96</b>	<b>21.89</b>	1048
<i>Linalool</i>	-	1.13	-	1098
<i>β-Terpineol, cis-</i>	-	0.1	-	1135
<i>α-Terpineol</i>	0.08	1.54	-	1189
<i>Thymol methyl ether</i>	-	0.67	-	1216
<i>Carvacrol methyl ether</i>	-	0.98	-	1225
<i>Carvone</i>	-		1.21	1243
<i>Linalool acetate</i>	2.65	-	-	1248
<b><i>Cinnamaldehyde</i></b>	<b>80.82</b>	-	-	1263
<b><i>Thymol</i></b>	-	<b>32.68</b>	1.94	1281
<b><i>Carvacrol</i></b>	-	<b>30.57</b>	<b>38.43</b>	1294

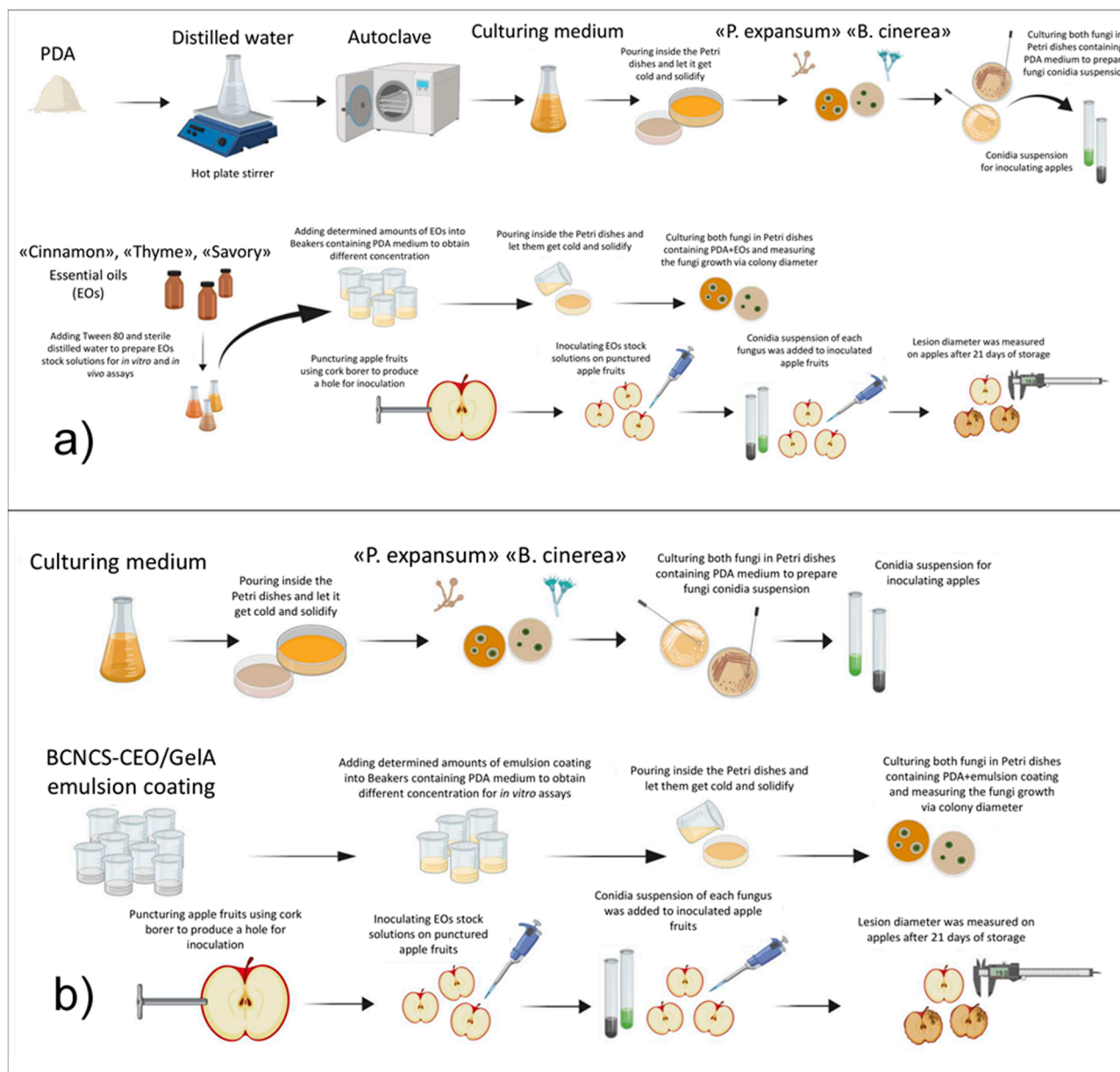
components	CEO (%)	ZEO (%)	SEO (%)	Retention index
<i>Thymol acetate</i>	-	1.45	-	1332
<i>Carvacrol acetate</i>	-	1.78	-	1348
<i>Caryophyllene, (Z)-</i>	-	2.38	-	1404
<i>Aromadendrene</i>	-	1.21	-	1441
<i>Caryophyllene</i>	3.57	-	-	1456
<i>(Z)-Cinnamic acid</i>	0.12	-	-	1471
<i>γ-Bisabolene, (E)-</i>	-	0.8	2.32	1523
<i>Spathulenol</i>	-	0.83	-	1563
<i>2H-1- Benzopyran-2-one</i>	0.1	-	-	1581
<i>Cinnamyl acetate</i>	0.14	-	-	1635
<i>Caryophyllene oxide</i>	0.03	0.78	-	1961
<b>Total</b>	97.66	98.46	98.81	

**Table S3.** Schematic description of the protocol used for cell counting (adapted from Selvakumaran & Jell, 2005).

1. Make a cell suspension according to the sub-culture protocol.
2. Prepare a hemocytometer and coverslip by spraying with 70% alcohol and drying with a clean tissue.
3. Fix a coverslip on the hemocytometer by moistening the coverslip with water or exhaled air, slide the coverslip over the chamber, and move back and forth exerting slight pressure until Newton's rings (rainbow-colored rings) appear.
4. Place 0.2 ml of a suitable cell suspension (in complete medium) in a sealed container.
5. Add 0.2 ml of 0.4% trypan blue stain and mix thoroughly (dilution factor of 2).
6. Allow to stand for 2–3 minutes at 15 to 30 °C (room temperature). Prolonged exposure to trypan blue kills the cells.
7. With a pipette, fill both chambers of the hemocytometer. Do not over- or under-fill the chambers. Make sure that there are no air bubbles.
8. Under a microscope count the number of viable (unstained) and non-viable (stained blue) cells in eight-ten  $4 \times 4$  squares or  $0.1 \text{ cm}^2$  area. Count the cells in the squares and touch the left and top middle line.
9. Calculate the number of viable and non-viable cells/ml using the formula given below.

Number of viable cells/ml =	The average number of viable cells per $0.1 \text{ cm}^2$ area $\times 10^4$ (correction factor for volume of shaded area) $\times 2$ (dilution factor, 0.2 ml of cell suspension in 0.4 ml)
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10. Then, by having the total volume of fungi suspension and multiplying by the number of cells in 1 ml obtained from the above formula, the final concentrations will be obtained.



**Figure S1.** Schematic representation of the procedure used to assess the *in vitro* (a) and *in vivo* (b) antifungal activity of both pristine EOs (CEO, ZEO, and SEO) and BCNC/GelA-CEO emulsions.

## References

1. Tahmasebi, M.; Golmohammadi, A.; Nematollahzadeh, A.; Davari, M.; Chamani, E. Control of nectarine fruits postharvest fungal rots caused by *Botrytis cinerea* and *Rhizopus stolonifer* via some essential oils. *J. Food Sci. Technol.* **2020**, *57*, 1647–1655. <https://doi.org/10.1007/s13197-019-04197-4>
2. Selvakumaran, J.; Jell, G. A guide to basic cell culture and applications in biomaterials and tissue engineering. In *Biomaterials, Artificial Organs and Tissue Engineering*, 1st ed.; Hench, L.L., Jones, J.R., Eds.; Woodhead Publishing Series in Biomaterials, 2005; pp. 215–226. <https://doi.org/10.1533/9781845690861.4.215>