

Quantitative detection of microplastics in water through fluorescence signal analysis

by Roberto Pizzoferrato^{1,*}, Yuliu Li¹, and Eleonora Nicolai^{2,*}

¹Department of Industrial Engineering, University of Rome Tor Vergata, Via del Politecnico 1, 00133 Rome, Italy; yuliuli1022@gmail.com

²Department of Experimental Medicine, University of Rome Tor Vergata, Via Montpellier 1, 00133 Rome, Italy

*Correspondence: pizzoferrato@uniroma2.it (R.P.); nicolai@med.uniroma2.it (E.N.)

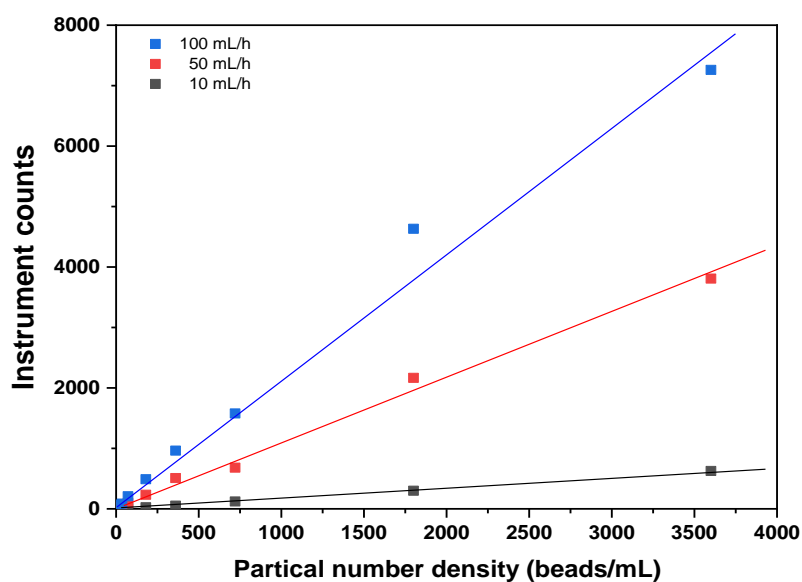


Figure S1. Experimental counts as a function of particle number density for different values of flow rate.

Model for the pulse duration distribution

We assume the parabolic velocity profile with circular symmetry inside a circular tube/capillary given by:

$$v(r) = v_M \left[1 - \left(\frac{r}{R} \right)^2 \right] \quad (1)$$

where:

$$v_M = 2 \langle v \rangle = 2 \frac{Q}{\pi R^2}$$

with Q as the liquid flow rate.

Eq. 1 gives:

$$\frac{dv}{dr} = -2v_M \frac{r}{R^2}$$

Let us consider the particle concentration n uniform inside the liquid and, thus, a uniform particle number density per unit volume. Therefore, the number of particles within a certain cylindrical annulus of ray r , thickness dr , and length L in the tube section will be:

$$N(r)dr = 2\pi n r dr L \quad (2)$$

From Eq. 1, we have:

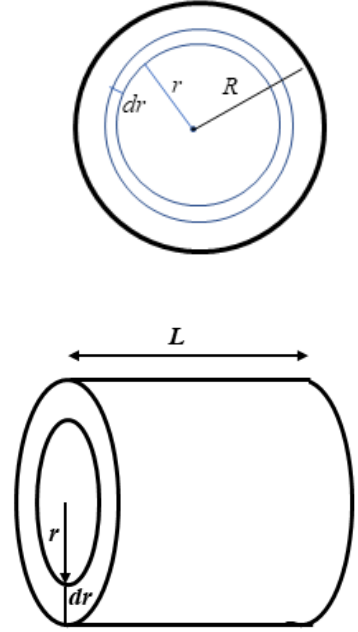
$$r = R \sqrt{1 - \frac{v}{v_M}} \quad , \quad dr = -\frac{R^2}{2v_M r} dv$$

which can be substituted in Eq. 2 (neglecting the sign) to obtain the velocity distribution:

$$N(v)dv = 2\pi n L R \sqrt{1 - \frac{v}{v_M}} \times \frac{R^2}{2Rv_M \sqrt{1 - \frac{v}{v_M}}} dv = \frac{\pi n R^2}{v_M} L dv \quad (3)$$

Therefore, the velocity statistical distribution is uniform up to v_M .

However, in a certain measurement time interval ΔT , only the particles within a distance from the laser excitation volume $L = v \Delta T$ will be counted. Therefore, the distribution of the number of counted particles over the time period T will be:

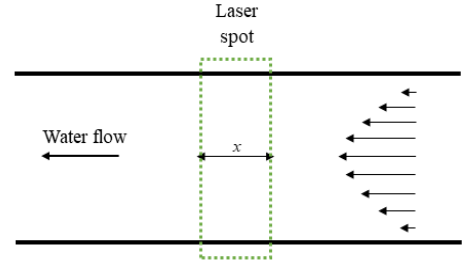


$$N(v)dv = \frac{\pi n R^2}{v_M} v \Delta T dv \quad (4)$$

The relation between the particle speed v and the transit time t across the illuminated path of length x (fluorescence pulse duration) is:

$$v = \frac{x}{t} \Rightarrow |dv| = \frac{x}{t^2} |dt|$$

Therefore, Eq. 4 gives the pulse duration distribution:



$$N(t)dt = \frac{\pi n R^2}{v_M} \frac{x}{t} \Delta T \frac{x}{t^2} dt = \frac{\pi n R^2}{v_M} \Delta T \frac{x^2}{t^3} dt = K \frac{\Delta T}{t^3} dt \quad (5)$$

$$\text{Up to } t_{min} = \frac{x}{v_M} = \frac{57 \times 10^{-6}}{0.177} = 322 \mu\text{s} \quad \text{according to a laminar flow at 10 mL/h}$$

This model does not have free parameters and gives a reasonable agreement with experimental distribution, as reported in Figure S2.

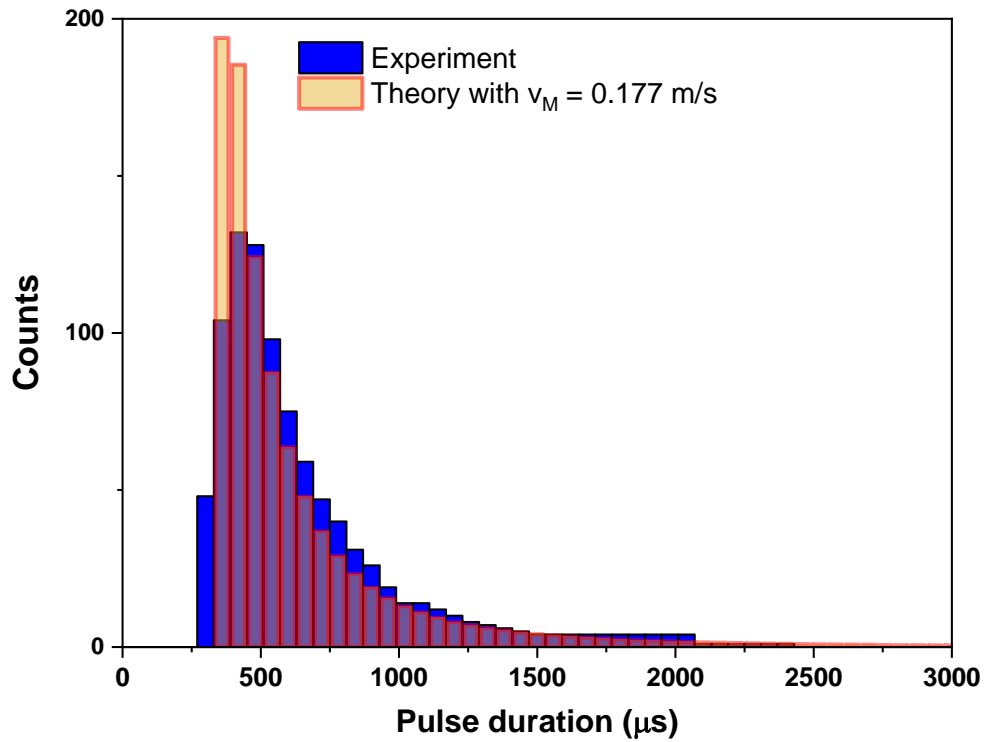


Figure S2. Experimental results for 10 μm OFPS microspheres at a flow rate of 10 mL/h. Theoretical distribution calculated according to Eq. 5 with no free parameters.

However, experimental verifications of Poiseuille's law often give values of v_M lower than the predicted one [45], and thus this can be considered as an adjustable parameter. A better fit was found for $v_M = 0.158$ m/s which gives:

$$t_{min} = \frac{x}{v_M} = \frac{57 \times 10^{-6}}{0.158} = 361 \mu s$$

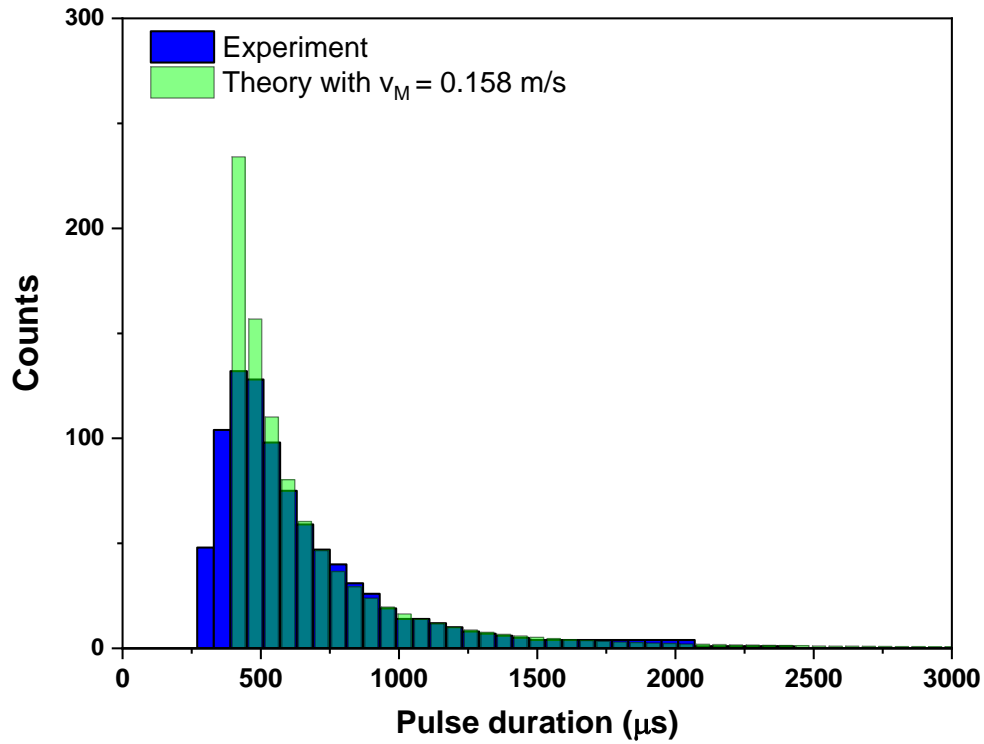


Figure S3. Experimental results for 10 μ m OFPS microspheres at a flow rate of 10 mL/h. Theoretical distribution calculated according to Eq. 5 with v_M as a free adjustable parameter.

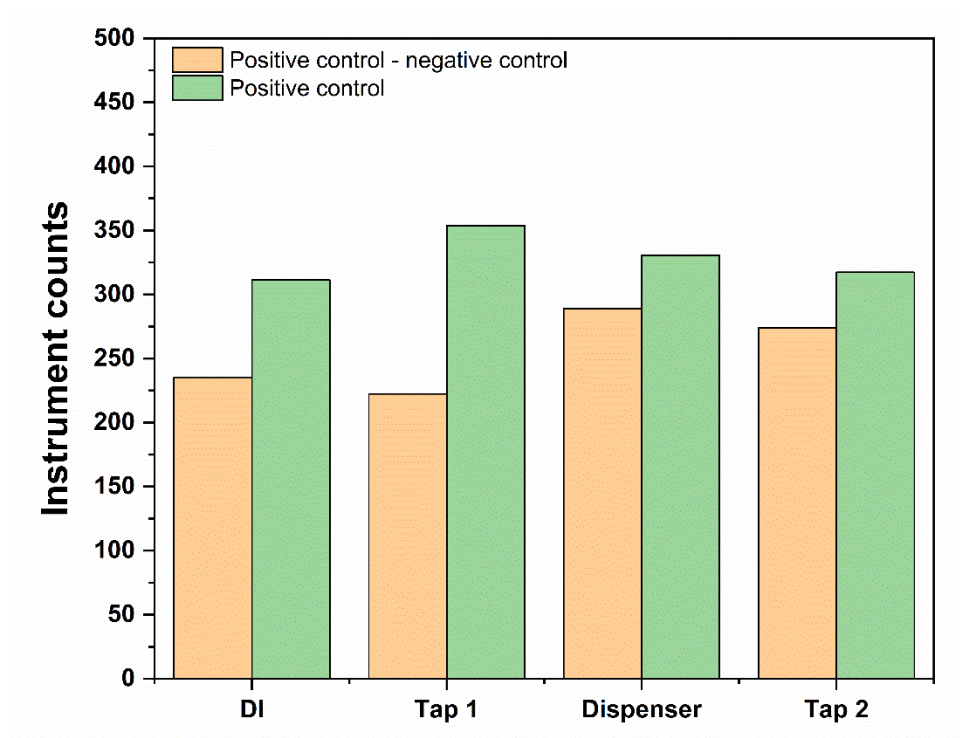


Figure S4. Observed counts in spiking experiment of stained PS MPs in different types of real water and in DI water. Water was filtered through a 0.7 μm filter and NR concentration was 0.1 mg/L.

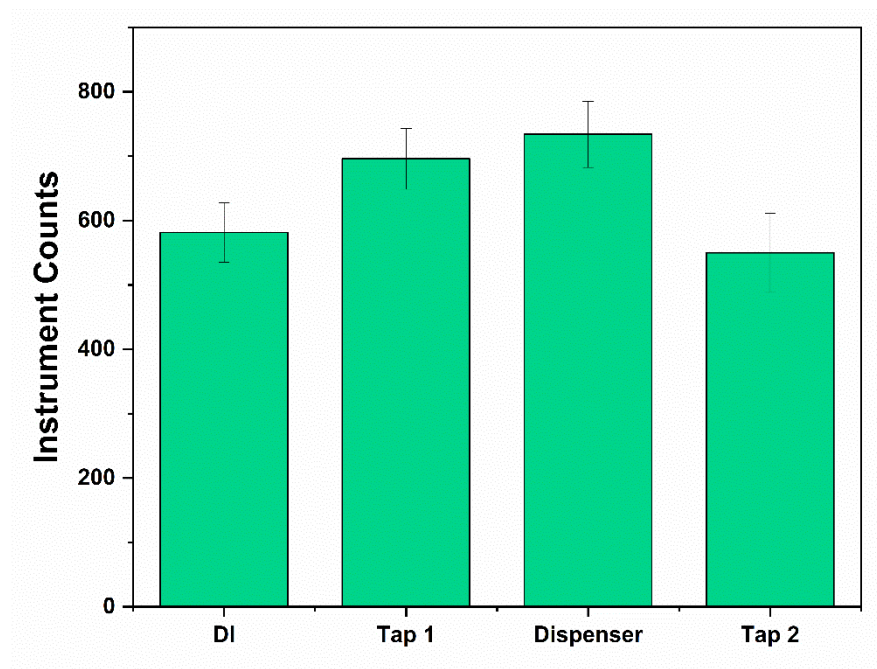


Figure S5. Observed counts in spiking experiment of stained PS MPs in different types of real water and in DI water. Water was filtered through a 40 μm filter and NR concentration was 5 mg/L.

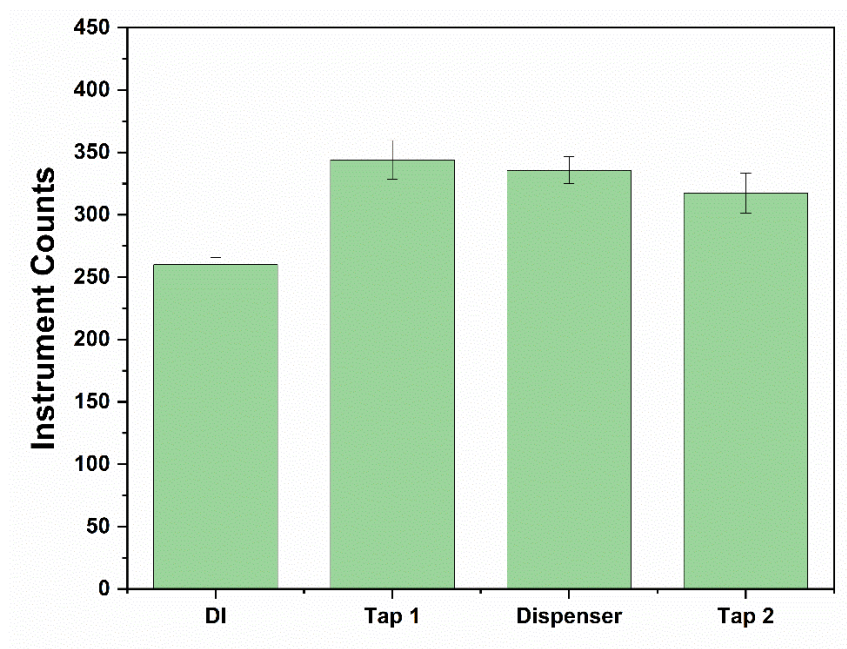


Figure S6. Observed counts in spiking experiment of stained PS MPs in different types of real water and in DI water. Water was filtered through a 40 μm filter and NR concentration was 0.1 mg/L.

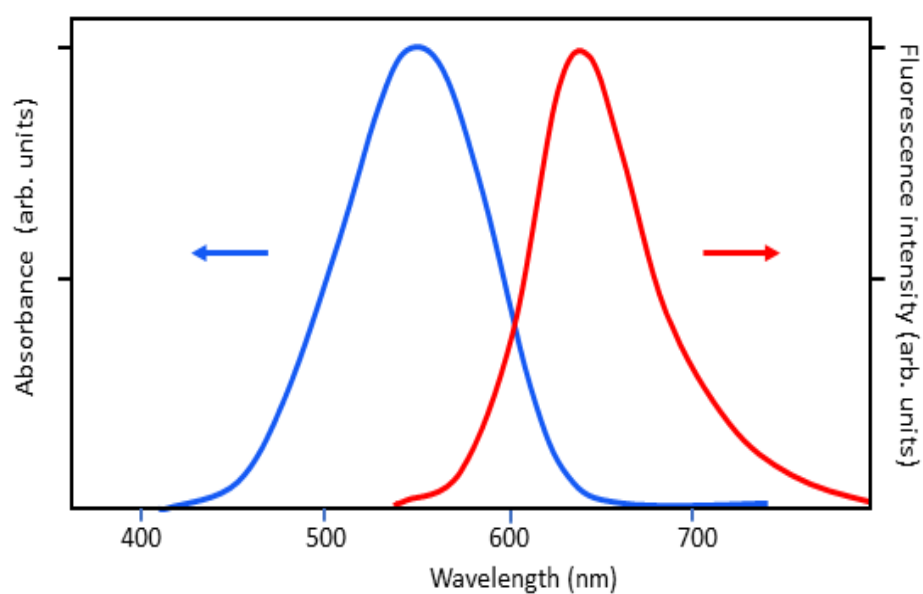


Figure S7. The absorption (blue curve) and emission spectrum (red curve) of Nile Red used in the present study.

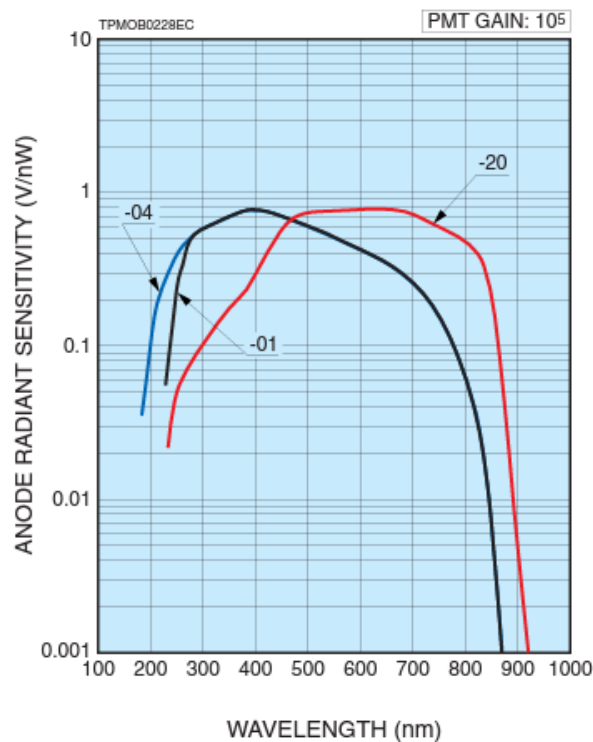


Figure S8. The spectral response (red curve) of the H10721-110 Hamamatsu photomultiplier used in the present study.

From:

<https://www.hamamatsu.com/eu/en/product/optical-sensors/pmt/pmt-module/voltage-output-type/H10723-20.html>

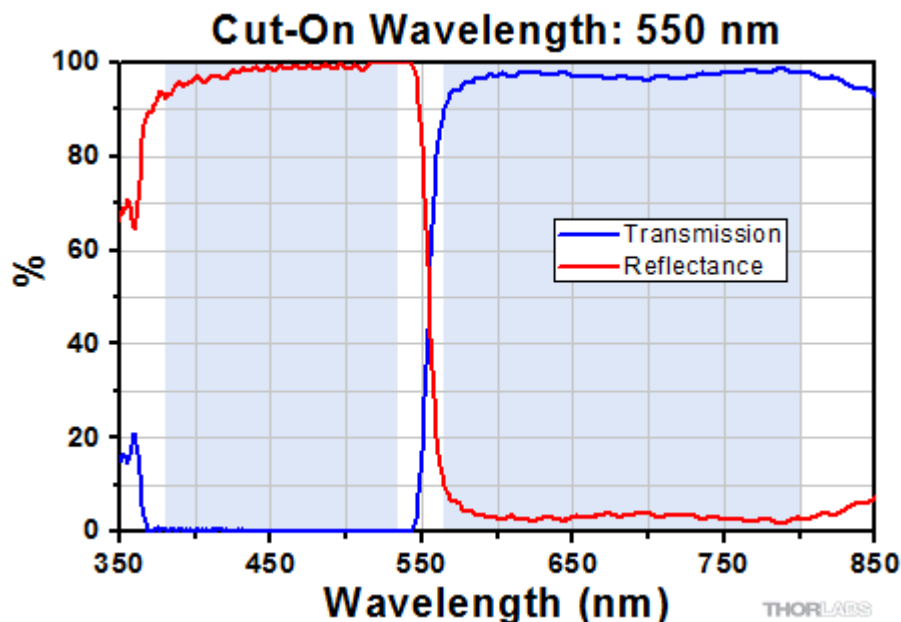


Figure S9. The transmission/reflection curves of the dichroic mirror Thorlabs DMLP550L used in the present study.

From:

https://www.thorlabs.com/newgrouppage9.cfm?objectgroup_id=3313&pn=DM LP550

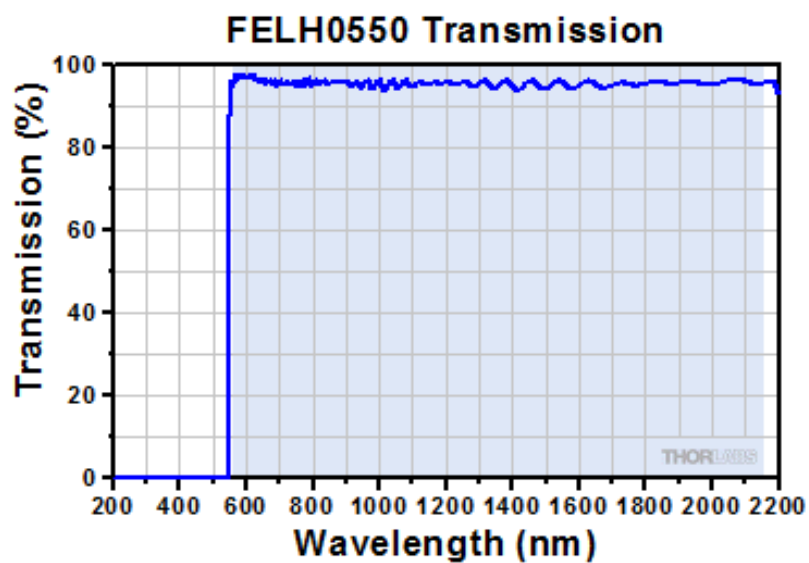


Figure S10. The transmission curve of the long-pass filter Thorlabs FEL0550 used in the present study.

From:

https://www.thorlabs.com/newgrouppage9.cfm?objectgroup_id=6082&pn=FELH0550