

Article

# Development and Validation of Rapid RP-HPLC and Green Second-Derivative UV Spectroscopic Methods for Simultaneous Quantification of Metformin and Remogliflozin in Formulation Using Experimental Design

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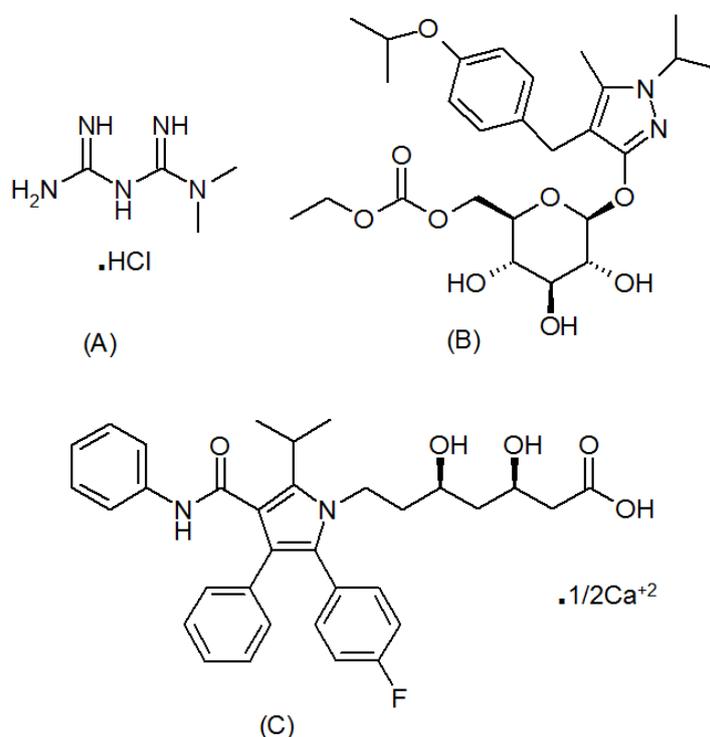


**Abstract:** Recently, a new formulation containing metformin HCl (MFH) and remogliflozin etabonate (RGE) has been approved for the management of diabetes mellitus. However, only one analytical method has been reported for the simultaneous determination of both the analytes. Therefore, the current study was designed to develop simple UV derivative spectroscopic and rapid RP-HPLC methods for simultaneous determination of MFH and RGE. The chromatographic separation of MFH and RGE was performed using a monolithic C18 column with an optimized chromatographic conditions carried out by full factorial Box–Behnken design model. The spectroscopic technique was based on the determination of peak amplitude of second-order derivative UV spectra at zero crossings. Further, both the methods were validated and compared statistically using Student's-t-test and F-test, and employed for the concurrent estimation of MFH and RGE in laboratory mixed solutions and formulations. Perturbation plots and response surface models showed the effect of chromatographic parameters and the final chromatographic condition was selected from 47 solutions suggested by the desirability function. Further, UV spectroscopic and HPLC procedures showed good linearity in the range of 1–24 µg/mL and 2–150 µg/mL for RGE and 2–30 µg/mL and 5–200 µg/mL for MFH, respectively. The average percent assay was found to be 99.51% and 99.80% for MFH and 99.60% and 100.07% for RGE by spectroscopic and HPLC methods, respectively. The proposed methods were simple, accurate, precise, and rapid. Therefore, they can be used for regular quality control of MFH and RGE formulations and dissolution studies as well.

**Keywords:** metformin; remogliflozin; HPLC; box–behnken design; derivative spectroscopy; optimization; formulation

## 1. Introduction

Diabetes mellitus is a leading cause of death all over the world [1–3]. Hence, better glycemic control is essential to reduce diabetic complications such as kidney failure, retinopathy, neuropathy, cardiovascular complications, etc. [4]. Combination therapy using oral hypoglycemic agents having a diverse mechanism of action is generally suggested over monotherapy to achieve the desired therapeutic target and to reduce side effects [5]. Recently, a new formulation containing metformin HCl and remogliflozin etabonate has been approved for the management of diabetes mellitus. Metformin HCl (MFH, Figure 1A) is extensively used as an oral hypoglycemic drug, which has multiple modes of action, namely, reducing the intestine absorption of glucose by increasing the anaerobic metabolism of glucose in enterocytes, decreasing the rate of hepatic gluconeogenesis, and insulin sensitivity, thereby increasing the utilization of glucose by peripheral cells [6–8].



**Figure 1.** Chemical structure of metformin HCl (A), remogliflozin etabonate (B), and atorvastatin (C).

Remogliflozin etabonate (RGE, Figure 1B) is a recently developed insulin-independent oral hypoglycemic agent. [9,10]. It acts by inhibiting sodium-glucose cotransporter-2 (SGLT2), an enzyme accountable for reabsorption of sugar in the kidneys, thereby; increase the elimination of sugar in the urine. Apart from glycemic control, SGLT-2 inhibitors possess many beneficial effects that include lowering of body weight, reduction of systolic blood pressure, and lowering hemoglobin A1c levels. Hence, RGE is more helpful when co-administered with metformin, particularly in patients with cardiac and renal diseases, who require further reduction of hemoglobin A1c level [11–16].

Literature depicts many analytical methods for quantification of metformin alone using spectrophotometric [17], HPLC [18], LCMS [19], capillary electrophoresis [20] in both formulations and biological samples. Metformin was also quantified in combination with other antidiabetic drugs using spectrophotometric [21], HPLC [22–26], LCMS [27,28], capillary electrophoresis [29,30]. Metformin was also determined by the HPLC method along with canagliflozin [31] and with empagliflozin [32]. Quantification of RGE from bulk and in house tablets has been reported by Ankita et al. [33] using a spectroscopic method and high-performance thin-layer chromatography, whereas Sigafos et al [34] utilized LC/MS method for quantification of RGE from plasma. Recently, analytical method using

UPLC has been reported [35] for the concurrent determination of MFH and RGE from formulation. However, this study was aimed to develop simple, eco-friendly spectroscopic method and rapid RP-HPLC technique using atorvastatin as internal standard for concurrent quantification of MFH and RGE from solutions prepared in the laboratory and formulation.

The use of a direct UV spectroscopic method for the determination of multicomponent formulations is difficult if the spectra are overlapping. Hence, derivative spectroscopic methods are extensively used to resolve overlapping spectra for the concurrent determination of a combination of compounds [36–38]. In addition, the derivative techniques improve sensitivity and avoid interference from formulation excipients [36–40]. Hence, the second-derivative spectroscopic method has been performed to determine MFH and RGE simultaneously without extraction or separation of components from the formulation. As per guidelines of USFDA, the pharmaceutical industries are using quality by design methodology for developing rapid and economical RP-HPLC method [41]. The design of the experiment with multivariate optimization is generally used to optimize the analytical methods because it utilizes statistical concepts in comparing the simultaneous effects of different variables [41–47]. Further, a validated analytical method will be more efficient, rapid, and economical, as fewer experiments need to be carried out when compared with univariate optimization. Hence, the rapid RP-HPLC method has been developed using a monolithic column adopting multivariate optimization of HPLC parameters.

## 2. Materials and Methods

### 2.1. Instruments

The spectroscopic method was developed using the UV–VIS spectrophotometer (Shimadzu UV-1700, Shinagawa city, Tokyo, Japan). The instrument was attached to a personal computer installed with UV-probe software (Shimadzu Ver. 2.0, Shinagawa city, Tokyo, Japan). Analyte solutions were scanned using 10 mm quartz cuvettes with scan speed adjusted to fast mode, sampling intermission of 0.1 nm, and slit width 2 nm. An Agilent technologies HPLC instrument (Agilent 1200 series, Waldbronn, Germany) used was connected to mobile phase degasser, quaternary pump, autosampler, and diode array detector. Chem station (Ver. B 4.0.2) Agilent software was used to monitor the signals.

### 2.2. Analytes and Reagents

Active pharmaceutical ingredients of metformin HCl and remogliflozin etabonate were purchased from Bioteck Private Limited, Hyderabad, India. The internal standard atorvastatin calcium (AVC) was procured from Sigma Aldrich (Buchs, Switzerland). Chromosolv acetonitrile purchased from Honeywell (Morris Plains, NJ, USA) was HPLC grade and ethanol used for dissolving RGE was of analytical grade purchased from Sigma Aldrich (St. Louis, MO, USA). Sodium dodecyl sulfate (SDS), potassium dihydrogen phosphate, and orthophosphoric acid were obtained from Sigma (St. Louis, MO, USA) were analytical grade. Ultrapure water arranged in our laboratory utilizing Milli Q (Millipore, Burlington, MA, USA) water purification system was utilized during the investigations.

### 2.3. Preparation of Standard Solutions

All three standards MFH, RGE, and AVC were weighed accurately and dissolved separately into water, ethanol, and methanol respectively to get 1000 µg/mL concentration. Further, working standard and laboratory mixed solutions of MFH and RGE for UV spectroscopic methods were prepared by diluting with ultra-pure water. For the HPLC method, a series of working standard and laboratory mixed solutions of analytes along with internal standards were prepared by mixing all three solutions in the required quantity using the mobile phase.

#### 2.4. Preparation of Sample Solutions

A recently approved new combination of MFH and RGE was not available in the Saudi market, therefore, simulated tablets consisting of RGE 100 mg along with MFH 500 mg/1000 mg were prepared by mixing the required quantity of RGE, MFH, and tablet excipients mannitol, talc, magnesium stearate, microcrystalline cellulose, colloidal silicon dioxide, and polyvinyl pyrrolidone. Tablet powder equivalent to 100 mg of RGE along with MFH 500 mg and 1000 mg were measured separately and transferred to a 50 mL graduated flask containing 25 mL ethanol. The flasks were mixed properly by sonication to dissolve the drugs. Then the solutions were filtered into other graduated flasks and the finishing volume was attained with ethanol. Further, to achieve the amount of the drugs in the linearity range the sample solutions were diluted with water and the mobile phase for the UV spectroscopic and chromatographic methods, respectively.

#### 2.5. Chromatographic Conditions

The chromatographic separation of MFH, RGE, and IS was achieved on a Chromolith (50 mm × 4.6 mm i.d. 5 µm particle size) (Merck, Darmstadt, Germany) C18 column with isocratic elution. The mobile phase utilized for the separation of analytes and IS was comprising of acetonitrile and mixture of 25 mM sodium dodecyl sulfate, 10 mM potassium dihydrogen phosphate (pH adjusted to 3.5 with orthophosphoric acid) in a ratio of 42%:58% (v/v), respectively. The mobile phase was pumped at a rate of 2 mL/min at room temperature. The chromatogram was recorded by measuring the UV absorbance of the eluted analytes by a diode array detector at 230 nm. Different concentrations of working standard solutions, laboratory mixed solutions, and sample solutions were transferred into vials and placed into the auto-sampler. The injection volume of auto-sampler was set at 20 microliters for investigation.

#### 2.6. Optimization of Chromatographic Conditions by Experimental Design

One of the multivariate optimization techniques, Box–Behnken design model was adopted to elevate the effect of chromatographic conditions [48–50]. The Second-order three-level full factorial design experiment was performed by 17 runs including 5 central points at low, intermediate, and high levels of variables coded as −1, 0, and +1 respectively. The three independent variables, at three levels considered for optimization, were the concentration of sodium dodecyl sulfate (15 mM, 20 mM, and 25 mM), percent of acetonitrile (35%, 40%, and 45%) and pH of the mobile phase (2.5, 3.5 and 4.5). The 17 runs suggested by the software were run randomly to avoid any residual effects and the resolution between MFH peak to RGE peak and resolution between RGE peak to IS peak were considered as dependent response factors. Polynomial equations were generated by Design Expert software (Version 12, Stat-Ease Inc. Minneapolis, MN, USA) and its validation was performed by analysis of variance (ANOVA). A second-order quadratic model polynomial equation with coefficients of the influence of chromatographic variable parameters on the response was expressed as below:

$$R = \delta_0 + \delta_1 A + \delta_2 B + \delta_3 C + \delta_4 AC + \delta_5 AB + \delta_6 BC + \delta_7 A^2 + \delta_8 B^2 + \delta_9 C^2$$

where R represents the response (resolution between peaks); A, B, and C are three independent variables % of SDS, % of acetonitrile, and pH of the mobile phase respectively;  $\delta_0$  represents the intercept  $\delta_1$ – $\delta_9$  represents the regressions coefficients computed from the responses of 17 runs, showing the effects of each factors individual, square, and combined effects of factors. Both perturbation and 3D surface response models were generated using Design Expert software (Version 12, Stat-Ease Inc. Minneapolis, MN, USA) for easy representation for presenting the effect of chromatographic variable parameters on the selected response and optimize the chromatographic method.

## 2.7. Method Validation

The developed analytical methods have to be validated as per the ICH recommendations to prove the methods proposed are suitable for quality control. Validation of the proposed techniques [51] was performed for linearity, the limit of detection, the limit of quantification, accuracy, and precision. Additionally, the HPLC technique has been validated for system suitability and robustness.

### 2.7.1. System Suitability for HPLC Method

For the HPLC method, the system suitability test was carried out by analyzing 20 µg/mL and 100 µg/mL solutions of RGE and MFH respectively by maintaining IS concentration at 50 µg/mL. The investigation was carried out in triplicate and the various parameters determined were peak area ratios, theoretical plate, tailing factor, resolution, and retention time.

### 2.7.2. Linearity

#### Second-derivative Spectroscopic Method

Aliquots of MFH and RGE were transferred separately into two series of 10 mL graduated flask to obtain seven solutions of 2.0–30 µg/mL (2.0, 5, 10, 15, 20, 25, and 30 µg/mL) and 1 to 24 µg/mL (1, 4, 8, 12, 16, 20, and 24 µg/mL), respectively. The UV absorption spectra were recorded in the wavelength 200–300 nm using ethanol-water as blank and stored spectra were converted into second-order derivative spectra by 8 nm as  $\Delta\lambda$  with amplification feature 100. The absorption of MFH spectra was measured at wavelength 235.5 nm, a zero crossing for RGE. Similarly, the peak amplitude of RGE spectra was measured at wavelength 243.0 nm, a zero crossing for MFH. The calibration curves and regression equations were constructed from these peak amplitudes and corresponding concentrations of MFH and RGE.

#### HPLC Method

The required quantity of MEF and RGE was transferred to series of vials to get the eight concentrations levels in between 5 to 200 µg/mL (5, 50, 75, 100, 125, 150 and 200 µg/mL) and 2 to 150 µg/mL (2, 25, 50, 75, 100, 125 and 150 µg/mL) respectively. Further, an aliquot of the IS solution was transferred to all the vials to get a concentration of 50 µg/mL. Chromatographs were documented by injecting 20 µL of each sample in triplicate. Peak area ratios of MEF to IS and RGE to IS, calculated from the chromatogram that were used to generate calibration curves using the corresponding concentration of MEF and RGE.

### 2.7.3. Limit of Detection and Quantification

Limit of detection and quantification explains the sensitivity of the analytical techniques. Hence, LOD and LOQ were determined for both the methods using 3.3 d/s and 10 d/s equations, respectively, where “d” is the standard deviation of the response intercept and “s” is the slope of the standardization curve.

### 2.7.4. Precision and Accuracy

The precision of both analytical methods was evaluated in terms of inter-day and intraday precision. For determination of inter-day precision, four diverse concentrations of MEF and RGE solutions were evaluated in triplicate on the same day covering the entire calibration range. Similarly, for the determination of intraday precision the above-prepared solutions were analyzed on three consecutive days. The precision of the method was presented as relative standard deviation (RSD). Accuracy of both the methods was evaluated using the above solutions and expressed as percent relative error (%RE).

### 2.7.5. Recovery Studies

The recovery of both analytes from the formulation by both analytical methods was evaluated by the standard addition method. In this, formerly evaluated sample solution consisting of a known amount of MFH and RGE were spiked with three different concentration levels 50%, 100%, and 150%. Briefly, for spectroscopic method recovery study was executed by adding standard solutions of MFH (5, 10, and 15  $\mu\text{g/mL}$ ) and RGE (1, 2, and 3  $\mu\text{g/mL}$ ) to the formerly studied solution containing 10  $\mu\text{g/mL}$  of MFH and 2  $\mu\text{g/mL}$  of RGE. Similarly, recovery study of HPLC method was performed by analyzing the solution after adding three different concentration of standard MFH (25, 50, and 75  $\mu\text{g/mL}$ ) and RGE (5, 10, and 15  $\mu\text{g/mL}$ ) to the formerly examined sample comprising of 50  $\mu\text{g/mL}$  MFH and 10  $\mu\text{g/mL}$  RGE. Further, 50  $\mu\text{g/mL}$  of IS was maintained in these solutions. The recovery study result was expressed in terms of percent recovery and percent relative error.

### 2.7.6. Robustness Using Experimental Design

The robustness of the liquid chromatographic procedure for the concurrent determination of MFH and RGE was evaluated by fractional factorial design. The parameters were slightly but deliberately changed from the optimized conditions for both analytes to study their effect on the considered response. The three parameters designated on the importance of parameters witnessed during experimental runs were a percentage of acetonitrile (40%, 42%, and 44%), pH (3.3, 3.5, and 3.7) of the mobile phase and flow rate (1.8 mL/min, 2 mL/min and 2.2 mL/min). The responses examined were peak area ratios of MFH to IS and RGE to IS. The responses recorded were explored with design expert software to know the effect of simultaneous changes in the parameters. The experiments were accomplished in random order to avoid the influence of unrestricted features. The Pareto chart was generated from the peak area ratios obtained for both analytes, to know the effect of slight changes in the experimental conditions.

### 2.8. Analysis of Laboratory Mixed Solutions and Formulations

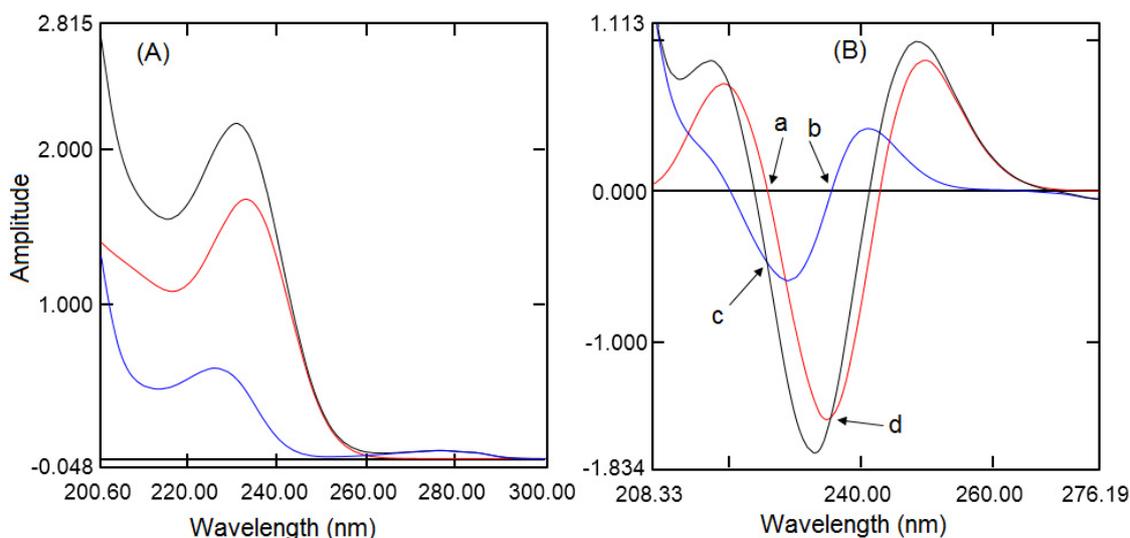
For spectrophotometric method, six solutions consisting of both analytes in different ratios of 2:5, 10:5, 10:20, 20:20, 5:30, and 20:30  $\mu\text{g/mL}$  of RGE and MFH were mixed. Then the UV absorption was recorded in between the wavelength 200 to 300 nm and the recorded spectra were converted to second-order derivative spectra. Then the peak amplitude was recorded at a wavelength of 243 nm for RGE and 235.5 nm for MFH. The concentrations were computed from the respective regression equations. Similarly, formulation samples were analyzed by diluting the formulation solutions to get an amount of analytes in the range of calibration curve. The formulation is available in two different ratios of RGE and MFH; hence, samples for each strength were prepared and analyzed.

For HPLC analysis various aliquots of standard solutions of RGE and MFH were transferred to vials to get the concentration in different ratios of 5:25, 5:100, 75:200, 75:100, 150:25, 150:200  $\mu\text{g/mL}$ , respectively by maintaining the IS concentration at 50  $\mu\text{g/mL}$ . The experiments were conducted as per the optimized procedure explained in the chromatographic condition and the quantity of both analytes, was computed using the respective regression equations. Further, the optimized HPLC method was utilized for the determination of RGE and MFH from the dosage form without prior separation. Assay of in house prepared tablets consisting of 100 mg of RGE and 500 mg/1000 mg of MFH was performed for each tablet. The percent drug content was determined by employing the respective linearity equations.

## 3. Results and Discussion

The second-order derivative UV spectrophotometric technique was developed for the simultaneous quantification of RGE and MFH because zero-order UV spectra of RGE and MFH showed complete overlap (Figure 2A). Further, derivatization eliminates the absorption effects of co analytes and excipients. Different order derivative spectra were studied, the first derivative spectra of MFH and RGE showed only one zero crossings and second-order derivatization showed better precision with  $\Delta\lambda$

8 nm with a scaling factor of 100, hence, the 2nd derivative has been selected for analysis of the binary mixture. Further, the second-order derivatization of RGE displayed two zero crossings at wavelength 220.1 nm and 235.5 nm where MFH had some absorption, whereas MFH showed two zero crossings at wavelength 225.9 nm and 243.0 nm where RGE had good peak amplitude. However, RGE showed better amplitude at 225.9 nm but the sensitivity was good at 243.0 nm, hence, 243.0 nm was selected for further analysis of RGE. MFH showed better amplitude and good sensitivity at 235.5 nm compared to a wavelength of 220.1 nm, hence, 235.5 was selected for further analysis. Furthermore, Figure 2B showed the same peak amplitude for pure standard solution spectra of RGE and MFH as of mixture of RGE and MFH spectra containing the same amount of analytes.



**Figure 2.** Normal UV absorption spectra of RGE (blue), MFH (red), and mixture (black) (A); second-derivative spectra of RGE (blue), MFH (red), and mixture (black) (B). Points a and b are zero crossings of MFH and RGE, respectively. Points c and d show the same peak amplitude for RGE and MFH in the pure sample and mixture, respectively.

### 3.1. Optimization of HPLC Method

To develop a rapid chromatographic method monolithic column (50 mm) has been selected. Monolithic column, also known as silica rod, because the column is packed with one piece of porous substance with flow pores and mesopores. Large flow pores allow for use of the mobile phase at a high flow rate and small mesopores increase the total surface area for interaction with the analyte for better separation. Monolithic columns can be operated up to 10 mL/min due to low back pressure; however, for separation of three analytes, 2 mL/min will be appropriate. The preliminary chromatographic study demonstrated that metformin was eluted at dead volume with more than 30% organic solvents along with phosphate buffer, whereas with less than 30% of organic solvent RGE was eluted after 6 min. Hence, from our previous report [52] to increase the retention time of MFH sodium dodecyl sulfate was employed in the mobile phase and 10 mM potassium dihydrogen phosphate was sufficient to give sharp peaks. Further, to optimize the chromatographic condition Box–Behnken's experimental design was adopted. From the preliminary studies, the important factors considered for optimization were the amount of sodium dodecyl sulfate (A), the percentage of organic modifier (B), and mobile phase pH(C). Two responses considered were resolution between MFH and RGE and resolution between RGE and IS. To develop the Box–Behnken experimental model [47–49], the number of experiments required can be determined by the equation  $N = 2\kappa(\kappa - 1) + \theta$ , where  $\kappa$  is independent parameters and  $\theta$  is the number of center point experiments. Hence, two response surface models have been created from the 17 runs and applied for computing the relations between the independent variables and

responses (Table 1). The best-fit models were suggested by the software based on the results, such as *p*-value, determination coefficients, adjusted and predicted R<sup>2</sup> value, and model F value.

**Table 1.** Box–Behnken model for optimization of chromatographic condition.

Experiment Number	Percent SDS	Percent Acetonitrile	pH	Resolution of MFH to RGE	Resolution of RGE to IS
1	20	40	3.5	14.93	5.93
2	25	40	2.5	6.11	5.57
3	25	40	4.5	15.94	3.00
4	20	35	4.5	26.72	1.54
5	20	35	2.5	11.68	6.85
6	25	45	3.5	8.36	4.02
7	20	40	3.5	15.48	6.12
8	20	45	4.5	10.84	1.38
9	20	40	3.5	14.91	5.85
10	15	40	2.5	11.15	6.2
11	20	40	3.5	15.11	6.04
12	25	35	3.5	18.25	5.11
13	20	45	2.5	7.75	4.74
14	15	35	3.5	21.59	5.39
15	20	40	3.5	15.29	6.17
16	15	40	4.5	19.32	0.87
17	20	40	3.5	14.93	5.93

The second-order polynomial equation representing the quadratic relationship was suggested by the software for resolution between MFH and RGE. This was computed using the least square method and found to be:

$$\text{Resolution of MFH to RGE} = -28.08831 + 0.3262\% \text{SDS} - 0.3811\% \text{ACN} + 36.1765 \text{pH} + 0.002\% \text{SDS} * \text{ACN} + 0.083\% \text{SDS} * \text{pH} - 0.5975\% \text{ACN} * \text{pH} - 0.02673\% \text{SDS}^2 + 0.01797\% \text{ACN}^2 - 1.34575 \text{pH}^2$$

The first order influence of percent of acetonitrile and mobile phase pH, separately, and their simultaneous effect was found to be most significant (*p* < 0.0001) on the resolution of MFH to RGE. Further, the interaction of percent of SDS and pH of the mobile phase with percent of acetonitrile was also most significant. The pure quadratic effect of acetonitrile and pH were highly significant. However, percent of acetonitrile had a negative effect whereas mobile phase pH ensured a positive influence on the resolution of MFH to RGE. The predicted R<sup>2</sup> value (0.9817) was very close to the adjusted R<sup>2</sup> value (0.9963) indicating satisfactory fitting of the model (since the difference is less than 0.2). The suggested model gave an F-value of 475.83 denotes the significant model, further the *p*-value was less than 0.05 for all parameters indicating model expressions were significant. Therefore, a quadratic model was exhibited to be significant for the resolution of MFH to RGE. Adequate precision indicates the signal to noise ratio and it was found to be 83.348 and that indicate adequate signal (A precision value more than four is desirable).

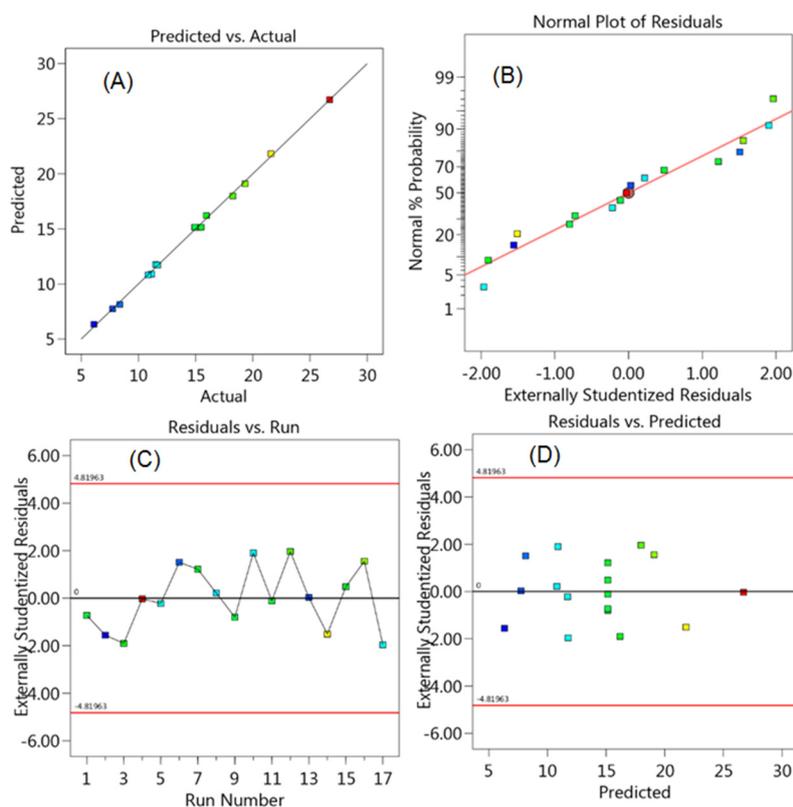
The quadratic relationship model suggested for resolution between RGE and IS by the software was found to be:

$$\text{Resolution of RGE to IS} = -38.44587 + 0.432850\% \text{SDS} + 2.04195 \text{ACN}\% + 2.75575 \text{pH} - 0.0037\% \text{SDS} * \text{ACN} \% + 0.138\% \text{SDS} * \text{pH} + 0.0975 \text{ACN}\% * \text{pH} - 0.01884\% \text{SDS}^2 - 0.03014 \text{ACN}\%^2 - 1.641 \text{pH}^2$$

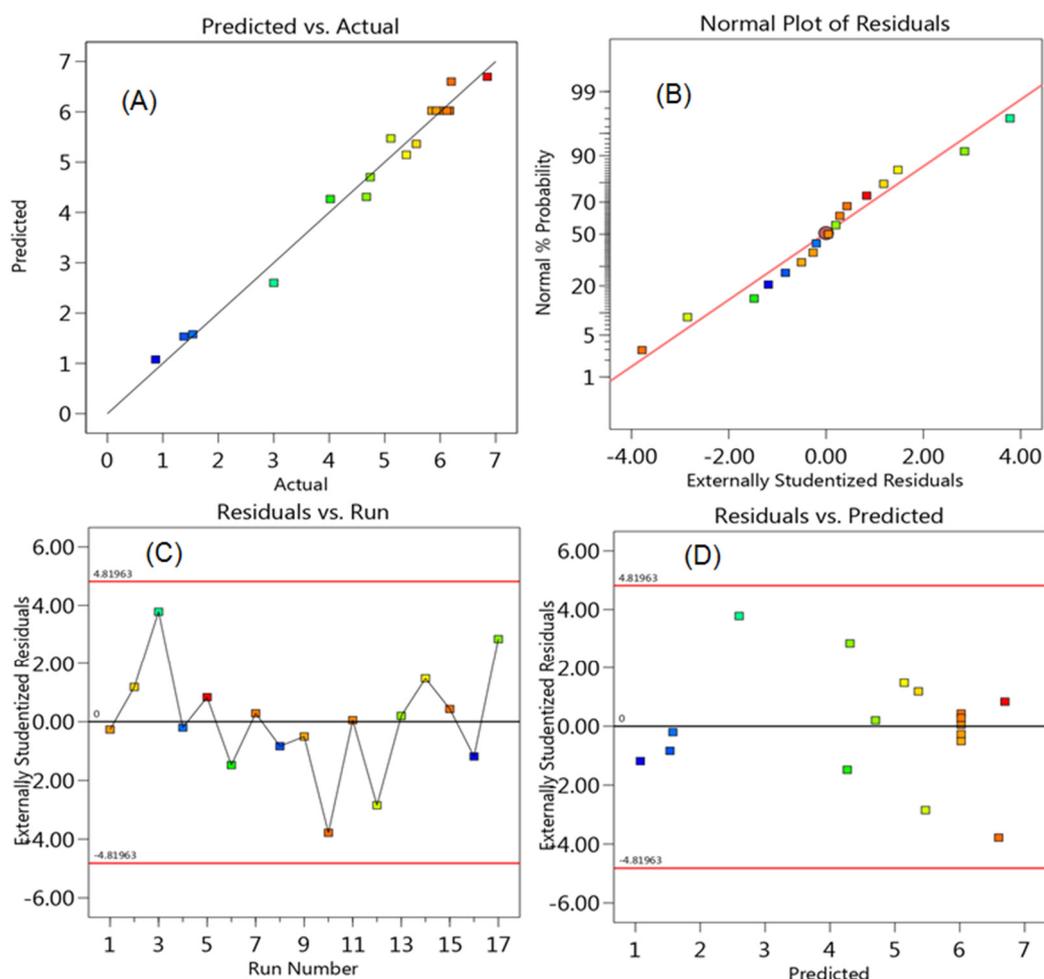
The first order influence and pure quadratic influence of percent of acetonitrile and mobile phase pH were most significant (*p* < 0.001), similarly, the interaction of percent of SDS and percent of acetonitrile with pH of the mobile phase was significant (*p* < 0.001). Further percent of acetonitrile and

mobile phase pH had a positive influence on the resolution of RGE to IS. The quadratic relationship model showed a better fit of the model with reverence to its  $p$ -value. A narrow difference was observed between the predicted  $R^2$  values (0.859) and the adjusted  $R^2$  value (0.963) indicating acceptable fitting of the model (since the difference is less than 0.2). The model F-value of 47.25 suggests the significant model because the  $p$ -value was 0.0001, which is  $\leq 0.05$ , showing that the model terms were significant. Hence, the quadratic relationship model was found to be significant for the resolution of RGE to IS. Adequate precision indicates the signal to noise ratio and it was found to be 20.3347, indicating a sufficient signal (a precision value more than four is desirable).

Design Expert software also provides different types of diagnostic plots, a comparison of predicted residual plots against actual plots, which help in identifying the thorough fit of the model. A good-correlated, predicted and the actual value is demonstrated by the proper scattering of both values along the average line. The model is considered statistically acceptable if the predicted  $R^2$  and adjusted  $R^2$  values are within 20%. The present model suggested by the software showed that the predicted and actual values (Figures 3A and 4A) were close to each other. Similarly, normal probability residuals (Figures 3B and 4B) showed that almost all residual points are adjacent to the diagonal line. Further residual plots (Figure 3C,D and Figure 4C,D) against the run number and predicted values showed a random nonlinear scattering style without any outliers. In addition, the residual points were close to the projected regression line at zero indicated the perfectness of the prediction. The residual points above the zero lines (positive values) indicates a low prediction, whereas below the zero lines (negative values) indicate over prediction.

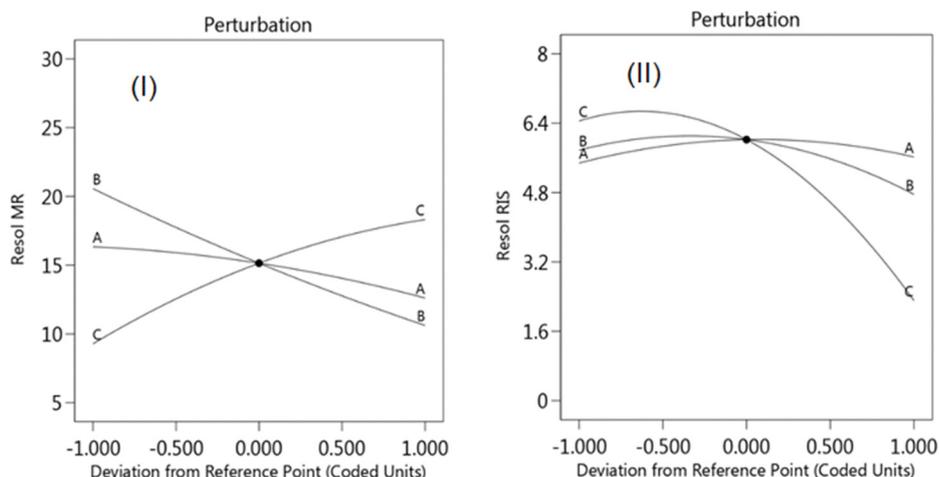


**Figure 3.** Diagnostic plots for resolution of MFH to RGE showing the predicted vs. actual (A) normal plot of residuals (B), residuals against run (C) and predicted values (D).

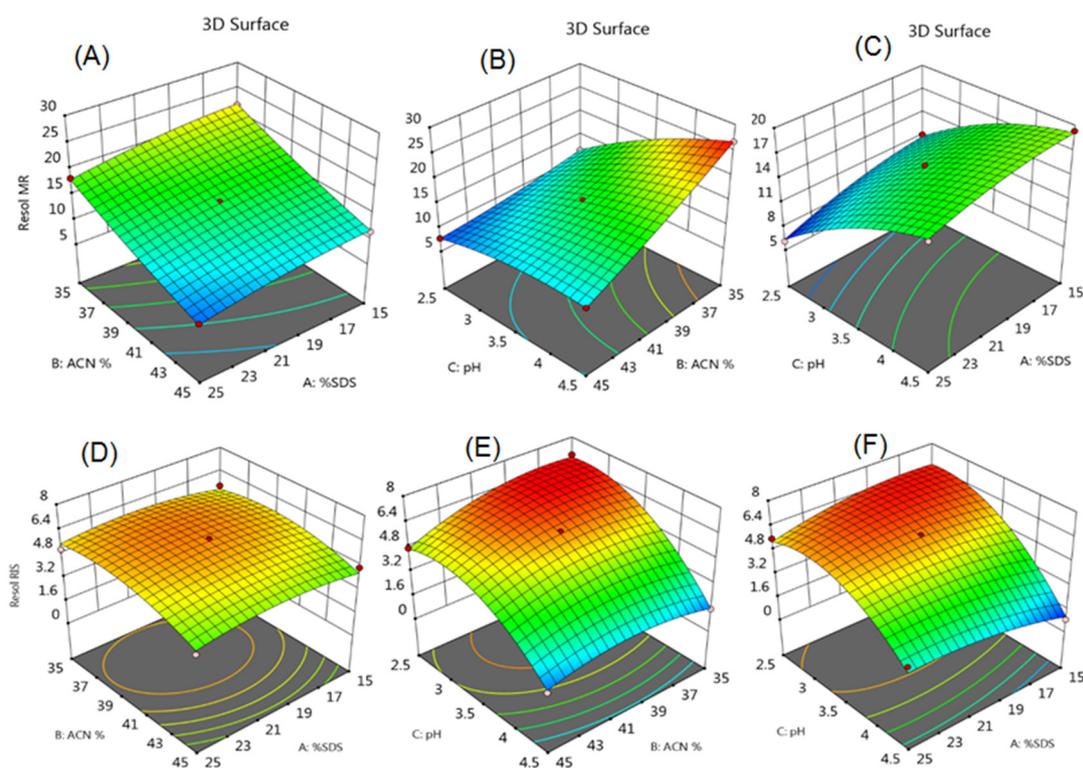


**Figure 4.** Diagnostic plots for resolution of RGE to IS showing the predicted vs. actual (A) normal plot of residuals (B), residuals against run (C) and predicted values (D).

The perturbation plots (Figure 5) and response surface models (RSM) clearly showed the effect of the concentration of SDS, percent of acetonitrile, and mobile phase pH on the resolution of drugs. The perturbation plot, Figure 5I, showed that percent of acetonitrile and mobile phase pH had an incredible influence on the resolution between MFH and RGE. With the decrease in the mobile phase pH, the resolution was decreased, whereas the percentage of acetonitrile showed the opposite effect. Further percent of SDS showed a slight effect on the resolution of MFH to RGE. Figure 5II showed that with an increase in the percent of acetonitrile and mobile phase pH, decrease in the resolution between RGE and IS. The influence of the combined effect of different factors was observed in response surface models (Figure 6). The RSM of the resolution of MFH to RGE, Figure 6A–C, showed that the decrease in the amount of acetonitrile and SDS increased the resolution, whereas an increase in the mobile phase pH with the decrease in the amount of acetonitrile drastically increased the resolution. Further, the increase in the pH of mobile phase increased the resolution without much effect of the percent of SDS. The RSM of resolution between RGE and IS (Figure 6D–F) showed that a decrease in the amount of acetonitrile and SDS showed a slight increase in the resolution. Whereas, the decrease in the pH along with the amount of acetonitrile increased the resolution. Similarly, the decrease in the mobile phase pH and amount of SDS increased the resolution.



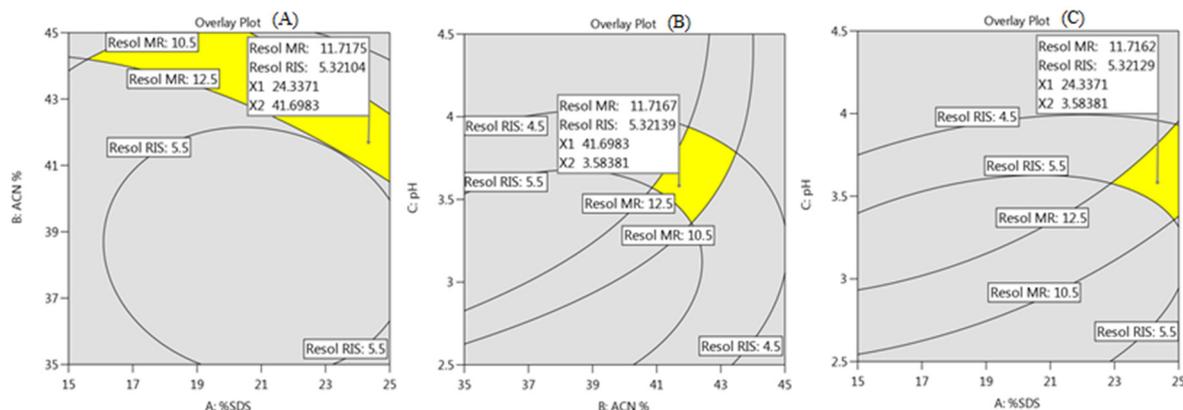
**Figure 5.** Perturbation plots showing the effect of percent of SDS (A), percent of acetonitrile (B) and pH of the mobile phase (C) on the resolution of MFH to RGE (I) and RGE to IS (II).



**Figure 6.** Response surface models showing the effect of percent of SDS, percent of acetonitrile and pH of the mobile phase on the resolution of MFH to RGE (A–C) and RGE to IS (D–F).

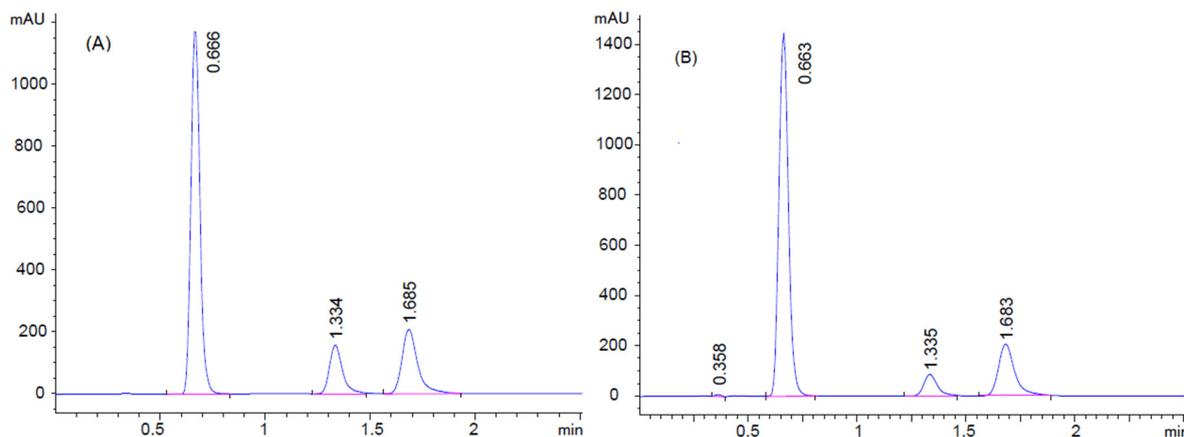
Multiple response process study indicated the effect of different parameters, further for final chromatographic condition, desirability function approach [53] was performed. In this, the conditions of the independent variable were assessed to get a targeted output of responses. RSM demonstrated that a decrease in the acetonitrile concentration and increase in the pH exhibited better resolution, however, the run time was more than 4 min. Further, the low concentration of SDS eluted the MFH just after the dead volume; hence, there is a chance of interference with formulation excipients. Hence, to increase the retention time of MFH and to develop the rapid HPLC method the concentration of SDS was set to 20 mM to 25 mM and percent of acetonitrile was set to 40–45%. Figure 7 showed the chromatographic condition for good resolution of the analytes and the desirability of independent

variables and response. The desirability value ( $0 \leq d \leq 1$ ) near to 1 indicates it is inside the acceptable range, hence, the quality of the response is acceptable. Software suggested 47 solutions; the optimum chromatographic conditions for developing rapid HPLC method with the good resolution were 25 mM SDS, 42% acetonitrile concentration, and mobile phase pH 3.5 with a desirability of 1.



**Figure 7.** Overlay plot for optimum chromatographic conditions suggested by Design Expert software. % SDS against % of acetonitrile (A); % of acetonitrile against mobile phase pH (B); %SDS against mobile phase pH (C).

To validate the suggested conditions, five experiments were conducted using the final mobile phase consisting of 10 mM phosphate buffer with 25 mM SDS, (pH 3.5) and acetonitrile in a ratio of 58:42 *v/v*, respectively. The resolution of MFH to RGE and RGE to IS was found to be 11.77 and 5.14, respectively, as compared to the predicted 11.71 and 5.32 by the model. The chromatogram showed good resolution, with good symmetric peak shape in the short run time. (Figure 8). An optimized chromatographic condition was used for running standard and sample solutions (Figure S1).



**Figure 8.** Representative chromatograms of standard solution (A) and formulation solution (B). The retention time of RFH:0.66 min, RGE:1.33 min, and IS:1.68 min.

### 3.2. Validation

#### 3.2.1. System Suitability Tests for the HPLC Method

As per the standard guidelines, the determination of system suitability parameters is a part of chromatographic method developments. Retention time, resolution, theoretical plate, and tailing factor were calculated and tabulated in Table 2. The theoretical plate was found to be low because of the short monolithic column.

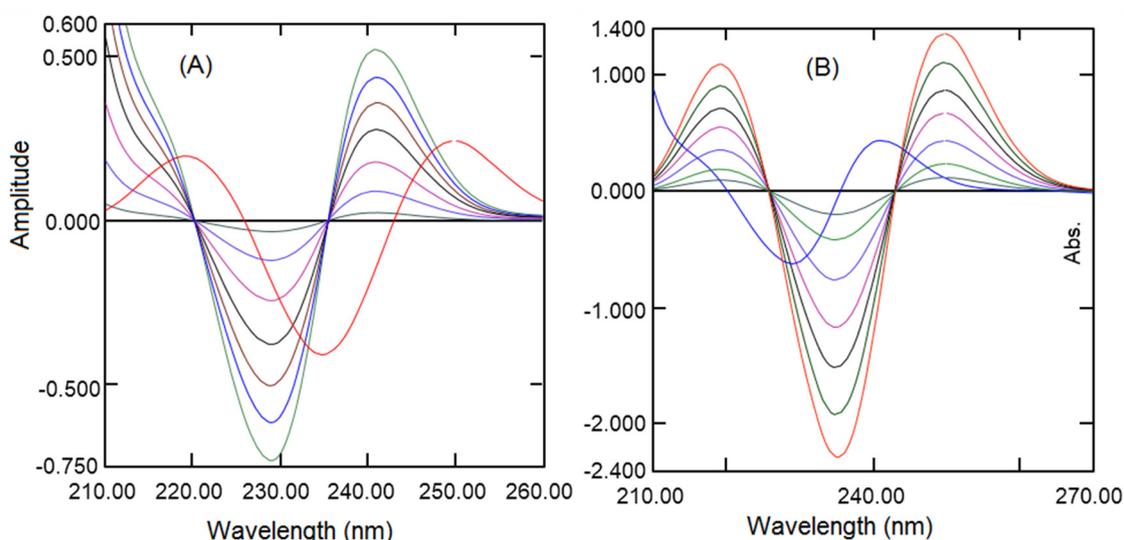
**Table 2.** System suitability results for analytes by RP HPLC method.

Parameters	MFH	RGE	IS
Retention Time ± SD	0.66 ± 0.019	1.31 ± 0.022	1.69 ± 0.012
Peak area ± SD	4294.15 ± 41.48 <sup>a</sup>	363.12 ± 6.95 <sup>b</sup>	1070.01 ± 25.57 <sup>c</sup>
Resolution ± SD	-	11.77 ± 0.65 <sup>d</sup>	5.14 ± 0.25 <sup>e</sup>
Theoretical plate ± SD	748.54 ± 15.83	1003.62 ± 25.71	1151.91 ± 47.28
Tailing factor ± SD	0.97 ± 0.034	0.89 ± 0.013	0.81 ± 0.024

<sup>a</sup> 100 µg/mL; <sup>b</sup> 20 µg/mL; <sup>c</sup> 50 µg/mL solution; <sup>d</sup> between MFH and RGE; <sup>e</sup> between RGE and IS; SD: Standard Deviation of six replicates.

### 3.2.2. Linearly

A calibration curve (Figure S2) was constructed for both analytes by computing the absorption of second-derivative spectra at 243 nm for RGE (Figure 9A) and 235.5 nm for MFH (Figure 9B). The RGE exhibited good linearity in the concentration of 1 to 24 µg/mL and MFH in the concentration of 2 to 30 µg/mL with a good regression coefficient ( $R^2 \geq 0.999$ ) Table 3. The spectra were measured in triplicate for each concentration of both analytes.



**Figure 9.** Second-derivative spectra of metformin (2.0, 5, 10, 15, 20, 25, and 30 µg/mL) with RGE (red) (A) and second derivative spectra of RGE (1, 2, 4, 8, 12, 16 20 and 24 µg/mL) with MFH (blue) (B).

**Table 3.** Calibration parameters results for both the proposed methods.

Parameters	HPLC Method		UV Spectroscopic Method	
	MFH	RGE	MFH	RGE
Linearity				
Wavelength (nm)	230.0	230.0	235.5	243.0
Linearity range (µg/mL)	5–200	2–150	2–30	1–24
Slope	0.0353	0.0163	0.0753	0.0248
Intercept	+0.355	+0.019	+0.015	+0.0035
Regression coefficient (r2)	0.9989	0.9993	0.9997	0.9991
Sensitivity				
LOD (µg/mL)	1.49	0.48	0.54	0.24
LOQ (µg/mL)	4.53	1.56	1.68	0.76

HPLC calibration curves (Figure S3) were developed by drawing a graph between the peak area ratios for both the analytes to IS against the respective concentration of RGE and MEH. The linear

relationship for RGE and MEH was accomplished in the concentration range of 2–150 µg/mL, and 5–200 µg/mL respectively (Table 3).

### 3.2.3. Limit of Detection and Quantification

The LOD and LOQ were calculated utilizing the standard deviation of the response and slopes of the three calibration curves. The results are presented in Table 3, low LOD and LOQ for both analytes revealed good sensitivity of both the proposed analytical procedures.

### 3.2.4. Precision and Accuracy

Inter day and intraday precision and accuracy of both the methods were determined by evaluating four different concentrations of both analytes on the same day and three following days respectively. (Figure S4) The %RSD was found to be in the range of 0.61–1.78 for RGE and 0.68–1.91 For MFH (Table 4) indicating the good precision of the optimized spectroscopic and HPLC methods. The %RE was found to be –1.75 to 1.81 for RGE and –1.96 to 1.20 for MFH, and exhibited good accuracy of the methods.

**Table 4.** Precision and accuracy results of MFH and RGE by the proposed methods.

Drug	Amount of Drug [µg/mL]	Inter Day			Intra Day		
		Amount Found Mean <sup>a</sup> ± SD	%RSD	%RE	Amount Found Mean <sup>b</sup> ± SD	%RSD	%RE
UV spectroscopic method							
RGE	2	2.01 ± 0.02	1.00	0.50	1.99 ± 0.03	1.51	–0.50
	8	7.86 ± 0.14	1.78	–1.75	7.93 ± 0.11	1.39	–0.88
	15	14.9 ± 0.25	1.68	–0.67	15.07 ± 0.25	1.66	0.47
	20	19.76 ± 0.27	1.37	–1.20	19.8 ± 0.12	0.61	–1.00
MFH	5	4.93 ± 0.07	1.42	–1.40	5.06 ± 0.09	1.78	1.20
	15	15.09 ± 0.13	0.86	0.60	14.79 ± 0.17	1.15	–1.40
	20	19.72 ± 0.31	1.57	–1.40	19.75 ± 0.16	0.81	–1.25
	30	29.87 ± 0.36	1.21	–0.43	29.68 ± 0.34	1.15	–1.07
HPLC Method							
RGE	5	5.01 ± 0.07	1.40	0.20	4.87 ± 0.08	1.64	–2.60
	50	49.69 ± 0.43	0.87	–0.62	50.13 ± 0.32	0.64	0.26
	100	101.81 ± 1.45	1.42	1.81	98.69 ± 0.93	0.94	–1.31
	150	148.89 ± 2.08	1.40	–0.74	148.82 ± 1.99	1.34	–0.79
MFH	25	24.67 ± 0.39	1.58	–1.32	25.15 ± 0.17	0.68	0.60
	50	49.11 ± 0.94	1.91	–1.78	49.02 ± 0.69	1.41	–1.96
	100	99.03 ± 1.34	1.35	–0.97	98.73 ± 1.52	1.54	–1.27
	200	201.59 ± 1.83	0.91	0.80	197.78 ± 1.59	0.80	–1.11

<sup>a</sup> n = 3; <sup>b</sup> n = 9; SD: Standard deviation; %RSD: Percent relative standard deviation; %RE: Percent relative error.

### 3.2.5. Recovery Study

A recovery study of both the analytical approaches was assessed by analyzing both analytes at three dissimilar concentration levels (50%, 100%, and 150%) and calculating the percentage recovery and %RE. Results (Table 5) showed a mean percent recovery of 99.61% for RGE and 99.19% for MEH by the spectroscopic method and 99.57% for RGE and 98.96% for MEH by the HPLC method. The %RE was also found to be below ±2% for both analytes by both the methods demonstrating the high recovery of analytes by both the analytical approaches.

Table 5. Recovery studies.

Drug	UV Spectroscopic Method			HPLC Method		
	Amount Added in [µg/mL]	%Recovery	%RE	Amount Added in [µg/mL]	%Recovery	%RE
RGE	1	101.00	1.00	5	100.40	0.40
	2	98.50	-1.50	10	99.50	-0.50
	3	99.33	-0.67	15	98.80	-1.20
	Across Mean	99.61			99.57	
	%RSD	1.27			0.80	
MFH	5	98.40	-1.60	25	99.24	-0.76
	10	100.10	0.10	50	98.06	-1.94
	15	99.07	-0.93	75	99.59	-0.41
	Across Mean	99.19			98.96	
	%RSD	0.85			0.81	

### 3.2.6. Robustness

According to ICH, a robustness test should be performed to know the effect of slight changes in the experimental condition on the analysis results. The three important factors considered are the percentage of acetonitrile, mobile phase pH, and flow rate. Further, the final concentration of the analytes was calculated by determining the peak area ratios of the analyte to the IS. Hence, the peak area ratio of MFH to IS and RGE to IS was selected as a response for studying the effect of changes in the experimental conditions. The experiments were conducted by altering three factors at three levels according to Box–Behnken experimental design (Table S1). The Pareto charts, generated using Design Expert software, displays the influence of the coincident discrepancy of variables on the peak area ratios. If the effects are beyond the Bonferroni Limit, those factors are highly significant, whereas the effects beyond the t-value limit are significant and effects less than the t-value limit are not significant. The Pareto diagram (Figure 10I,II) showed that a slight change in all three factors did not show any impact on the chromatogram peak area, indicating the robustness of the proposed HPLC method.

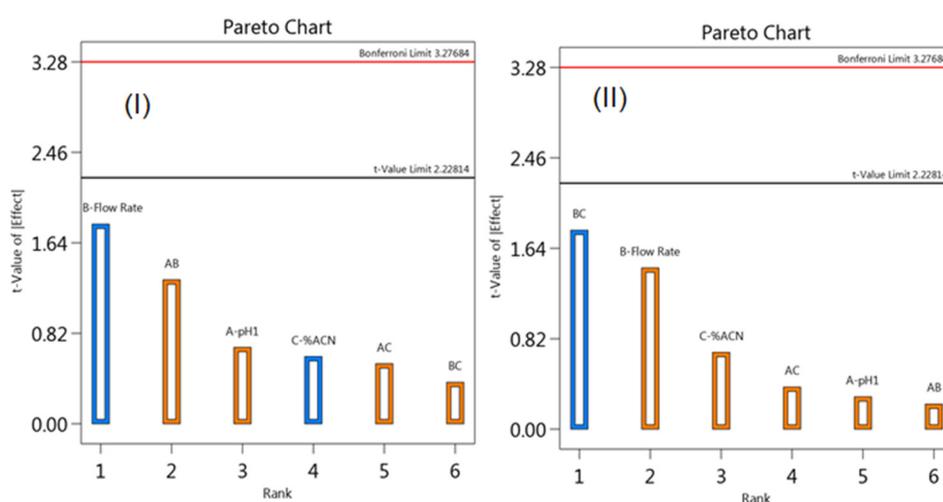


Figure 10. Pareto Chart showing the effects of percent of SDS (A), percent of acetonitrile (B) and pH of the mobile phase (C) on the resolution of MFH to RGE (I) and RGE to IS (II).

### 3.3. Analysis of Laboratory Mixed Solutions and Formulations

Second-derivative spectroscopic and optimized HPLC methods were applied for the concurrent quantification of MFH and RGE from laboratory prepared solutions (Figure S4) and formulation (Figure S4). The percent assay was found to be 98.29–101.35% for MFG and 98.13–101.39% for RGE

from laboratory mixed solutions (Table 6). The amount of MFH and RGE determined from formulation agree with the actual amount (Table 7), indicating the absence of effect of formulation excipient on the analysis. Further, both the proposed procedures were statistically compared using the student-t-test and F-test. The experimental t and F values (Table 7) were less than the critical values, indicated that the results obtained by both the methods were non-significant.

**Table 6.** Analysis results of laboratory mixed solutions by the proposed methods.

Ratios <sup>a</sup> RGE: MFH	UV Method (%Recovery ± SD)		Ratiosa RGE: MFH	HPLC Method (%Recovery ± SD)	
	Remogliflozin	Metformin		Remogliflozin	Metformin
2:05	101.23 ± 0.01	99.50 ± 0.02	5:25	100.90 ± 0.07	98.29 ± 0.09
5:30	99.68 ± 0.04	98.58 ± 0.03	5:100	99.29 ± 0.19	99.04 ± 0.17
10:05	100.98 ± 0.15	99.71 ± 0.08	75:200	101.06 ± 0.21	101.35 ± 0.06
10:20	98.82 ± 0.09	101.32 ± 0.12	75:100	99.18 ± 0.11	99.77 ± 0.15
20:20	99.16 ± 0.32	100.94 ± 0.14	150:200	101.35 ± 0.24	98.51 ± 0.23
20:30	101.39 ± 0.21	99.30 ± 0.18	150:25	98.13 ± 0.35	98.39 ± 0.31

<sup>a</sup> ratio amount in µg/mL.

**Table 7.** Assay of formulation and statistical calculation results.

	UV Spectroscopic Method		HPLC Method	
	RGE	MFH	RGE	MFH
Label Claim (mg)	100	500	100	500
Amount taken [µg/mL]	4	20	20	100
Amount found [µg/mL]	3.95	20.04	19.83	98.48
Label Claim (mg)	100	1000	100	1000
Amount taken [µg/mL]	3	30	15	150
Amount found [µg/mL]	2.97	29.78	15.12	148.24
Mean %estimation <sup>a</sup>	99.51	99.60	99.80	100.07
%RSD	1.34	0.89	0.78	1.08
n	6	6		
Student t-test (2.306) <sup>b</sup>	0.711	0.450		
F (6.388) <sup>c</sup>	3.521	1.436		

<sup>a</sup> Mean of three determination at each level; <sup>b</sup> critical value of t at  $p = 0.05$ ; <sup>c</sup> critical value of F at  $p = 0.05$ .

#### 4. Conclusions

Two simple analytical procedures were established for the concurrent determination of RGE and MFH from laboratory mixed solutions and tablets without prior separation. The UV spectroscopic method was economical and eco-friendly as water has been used as a solvent. The second-order derivative technique was used which eliminated the absorption effect of another analyte as well as excipients. HPLC method was optimized using the design of the experiment and data were generated employing Design-Expert software. The independent variables percent of SDS, percent of acetonitrile, and mobile phase pH exhibited a substantial influence on the resolution of analytes, which was confirmed by ANOVA. The actual  $R^2$  and adjusted  $R^2$  values were close and adequacy precision ratio was high indicating the good fit of the model and can navigate through the design space. The analysis results were also presented as diagnostic plots, perturbation plots, and response surface models. The optimized chromatographic technique was simple, rapid, precise, and robust for the concurrent quantification of RGE and MFH from solid dosage forms.

**Supplementary Materials:** The following are available online at <http://www.mdpi.com/2297-8739/7/4/59/s1>, Figure S1. Chromatograms of standard solutions of metformin and remogliflozin (MFH:RGE ( $\mu\text{g/mL}$ ) = 25:5 (A); 50:25 (B), 75:50 (C); 100:75 (D); 125:100 (E); 150:125 (F); to 200:150 (G) and samples 100:10 (H) and 200:20 (I). Figure S2. Calibration curves for RGE and MFH by second-derivative UV spectroscopic method. Figure S3: Calibration curves for RGE and MFH by RP-HPLC method. Figure S4. Normal and second-derivative UV spectra of MFH and RGE for accuracy and precision, laboratory mixed solutions, and formulations. Table S1. Results of the Box–Behnken model for robustness study.

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