


Review

Oxidative Potential Induced by Ambient Particulate Matters with Acellular Assays: A Review

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Abstract: Acellular assays of oxidative potential (OP) induced by ambient particulate matters (PMs) are of great significance in screening for toxicity in PMs. In this review, several typical OP measurement techniques, including the respiratory tract lining fluid assay (RTLFL), ascorbate depletion assay (AA), dithiothreitol assay (DTT), chemiluminescent reductive acridinium triggering (CRAT), dichlorofluorescein assay (DCFH) and electron paramagnetic/spin resonance assay (EPR/ESR) are discussed and their sensitivity to different PMs species composition, PMs size distribution and seasonality is compared. By comparison, the DTT assay tends to be the preferred method providing a more comprehensive measurement with transition metals and quinones accumulated in the fine PMs fraction. Specific transition metals (i.e., Mn, Cu, Fe) and quinones are found to contribute OP^{DTT} directly whereas the redox properties of PMs species may be changed by the interactions between themselves. The selection of the appropriate OP measurement methods and the accurate analysis of the relationship between the methods and PM components is conducive to epidemiological researches which are related with oxidative stress induced by PMs exposure.

Keywords: ambient particles; oxidative potential; acellular assay; transition metals; quinones

1. Introduction

Numerous epidemiological and experimental studies between exposure to ambient particulate matters (PMs) and lots of adverse health effects such as respiratory and cardiovascular diseases and diabetes have been carried out [1–5]. Compared to other air pollutants, PMs are complex in chemical composition and source origination, and their capability to absorb large amounts of toxic chemicals [6,7]. Adverse health effects which are most potentially caused by PMs are thought to be PMs-induced oxidative activity. Reactive oxygen species (ROS) are produced when PMs interact with epithelial cells and macrophages.

There are two kinds of ways for PMs to induce ROS in the human body: (1) oxidants existing on and/or within the particle itself are deposited in respiratory systems; (2) cells are stimulated by certain chemicals in PMs to produce excess ROS or specific biochemicals interact with components in PMs to

produce ROS [8,9]. ROS are highly reactive due to their unpaired electrons, and they include hydroxyl radical ($\bullet\text{OH}$), hydrogen peroxide (H_2O_2), organic peroxy radicals (RO_2), superoxide radical ($\text{O}_2^{\bullet-}$) and hypochlorite ion (OCl^-) [9].

Previous studies mostly defined “oxidative potential” (OP) as the generation of ROS and depletion of antioxidants catalyzed by inhaled PMs [9,10]. Oxidative potential, representing the capacity of PMs to oxidize molecules with the generation of ROS, can be used as a plausible metric of PMs toxicity [11]. Compared to the cellular assays, acellular assays have the advantage of faster reading speed, lower price, less control environments and being suitable for automation. Each acellular OP assay has certain specificity for the ROS or the precise type of ROS-inducers, leading to the fact that none of the methods has been used as a standard method to assess toxicity of ambient particles [12].

Considering that different assays focus on different chemical fractions of the oxidative activity caused by the PMs, there are no clear criteria for which OP assay and results are likely to be the best [13]. This work briefly describes some commonly adopted acellular OP assays, including electron paramagnetic/spin resonance (ESR assay, OP^{ESR}), dithiothreitol assay (DTT assay, OP^{DTT}), dichlorofluorescein assay (DCFH assay, OP^{DCFH}), ascorbic acid assay (AA assay, $\text{OP}^{\text{AA-only}}$) and the respiratory tract lining fluid assay (RTLFL assay, OP^{RTLFL}) firstly. Their sensitivity to different PMs species composition, PMs size distribution and seasonality are discussed in the following part with the aim of providing a preferred method to measure OP. Because of the comprehensiveness of the DTT assay, its driving factors are introduced as the focus in this review. Understanding the differences between acellular OP techniques, as well as the varying relationship to PMs composition and size distribution, is conducive to future research on investigating the relevance between epidemiologic disease and OP assay.

2. Oxidative Potential Measurement Methods

Oxidative potential is considered as a reasonable indicator of PMs toxicity. Some acellular methodologies are used to quantify the OP and demonstrate the complex mechanisms of the generation of ROS. Some assays measure the OP of PMs by the loss of a proxy of cellular reductants (i.e., OP^{DTT}), or endogenous antioxidant species (i.e., RTLFL assay, AA assay). ESR assay measures the generation of hydroxyl radical in the presence of H_2O_2 . Fluorescence intensity in DCFH assay and the intensity of the emitting light in CRAT assay are both converted into equivalent H_2O_2 concentrations. In order to ensure the standardization of interlaboratory measurement, residual oil fly ash (ROFA) always is selected as a positive control and inert carbon black as a negative control. Blank filters are also routinely extracted and run through assays system [14–17].

2.1. Respiratory Tract Lining Fluid Assay

The respiratory tract lining fluid (RTLFL) represents the first detoxifying environment encountered by inspired particulate matter [18] and has been shown to contain both small molecular weight and high concentrations of the antioxidants uric acid (UA) and ascorbic acid (AA), and reduced glutathione (GSH) [14]. The structure of urate and its initial degradation products can be seen in Figure 1. GSH is a tripeptide and its reaction with reactive species in PMs can often generate thiyl (GS^\bullet) radicals. Any available GS^- can react quickly with GS^\bullet to form glutathione radical disulfide anion ($\text{GSSG}^{\bullet-}$) and in turn $\text{O}_2^{\bullet-}$ (Equation (3)). This assay detects concentration variation of antioxidant molecules in the simplified synthetic human respiratory tract lining liquid after mixing with particle suspension [17]. PMs sample and composite antioxidant solution (200 μM) pH were adjusted to pH 7.0. The mixture was then transferred into an incubator maintained at 37 °C for 4 h [14]. Antioxidant concentrations were determined after incubations by high-pressure liquid chromatography (HPLC) and glutathione-reductase enzyme recycling method respectively [19]. The extent to which PMs

depleted antioxidants provided not only a quantitative PMs oxidative potential, but also reflected reactions that may occur at the air-lung interface in the body.

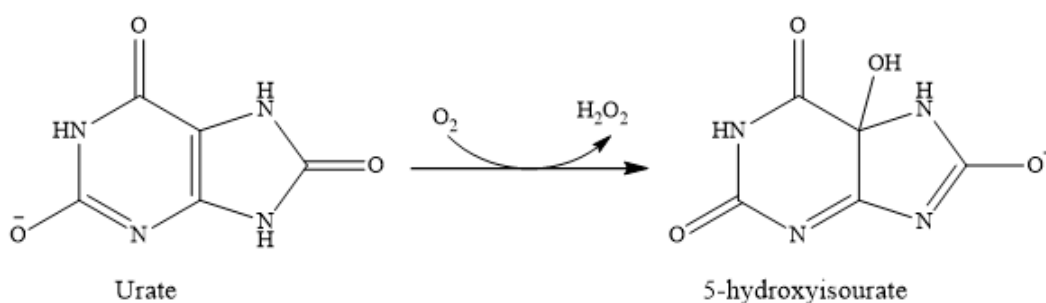


Figure 1. Urate can be oxidized by urate oxidase and forms 5-hydroxyisourate (HIU) initially [20].

The Ascorbate Depletion Assay

In the ascorbate (AA) depletion assay, only ascorbate acid is used which is thought to be a simplified version of the RTLF assay [21]. AA is oxidized to dehydroascorbic acid with the decrease in redox active substances in PMs. Figure 2 shows the structure of ascorbate and its oxidation and degradation products. In the AA assay, PMs extracts were incubated at 37 °C, after adding ascorbate acid. The reaction occurred directly, and was then measured in a UV/VIS-spectrophotometer with spectral scans (190–350 nm) performed at intervals of 10 min [13,22]. The depletion rate of AA was applied to represent the PM oxidative potential.

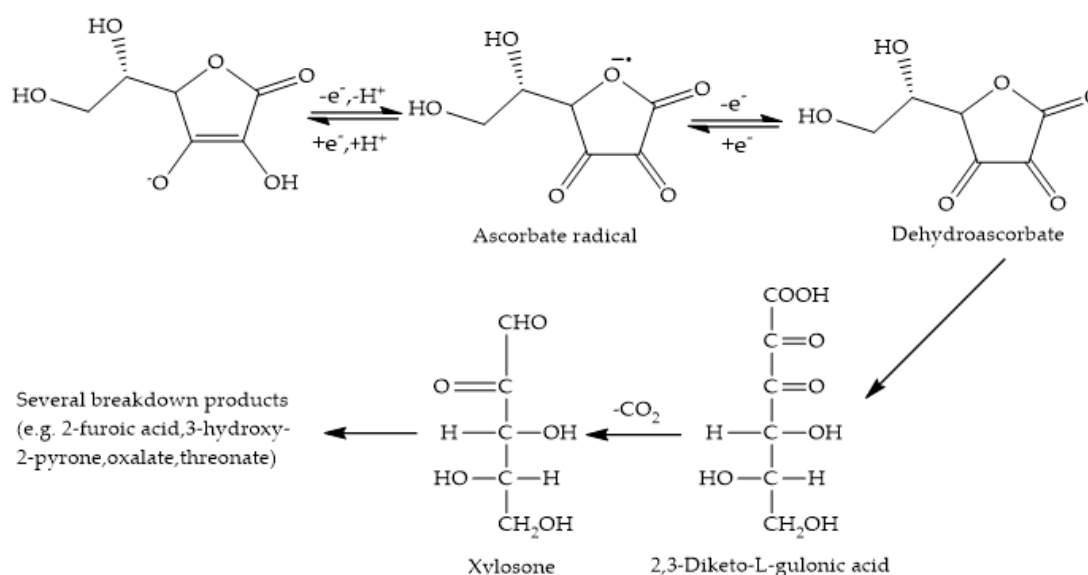


Figure 2. Structure of ascorbate and its oxidation and degradation products [23].

2.2. Dithiothreitol Assay

The dithiothreitol (DTT) could reduce oxygen to the superoxide anion ($\text{O}_2^{\bullet-}$) that could then form other ROS (i.e., $\bullet\text{OH}$). Redox active compounds oxidized DTT to its disulfide form and transferred

an electron to O_2 , generating $O_2^{\bullet-}$ (Figure 3) [8,24]. In the DTT assay, particles were first extracted using Milli-Q or methanol. The PMs extraction solution was then incubated for a period with DTT in a phosphate buffer. At preset times, an aliquot removed from the mixture was added with trichloroacetic acid (TCA) (quench reaction), Ethylene Diamine Tetraacetic Acid (EDTA) and 5,5'-dithiobis-(2-nitrobenzoic acid) (DTNB) solution. The reaction of residual DTT and DTNB formed 2-nitro-5-thiobenzoic acid (TNB), which could be quantified by a UV/VIS spectrophotometer at 412 nm [21,25,26]. When DTT was in excess, the consumption rate of DTT which was dependent on the linear slope of DTT depletion was proportional to the concentration of redox-active species in the PMs sample. The OP responses could be normalized by the volume of sampled air as an exposure metrics or by the PMs mass to represent the intrinsic ability of the PMs to deplete relevant antioxidants.

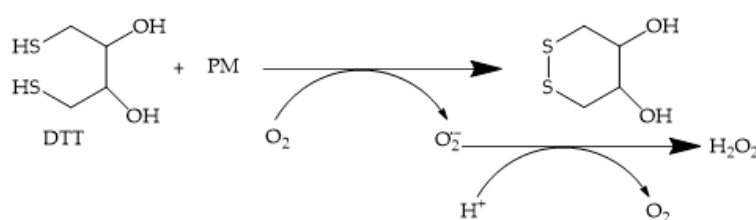


Figure 3. Reactions between dithiothreitol (DTT) and O_2 with particulate matters (PMs) as a catalyst [27].

2.3. Chemiluminescent Reductive Acridinium Triggering Assay

Chemiluminescence of acridine esters under alkaline conditions is the foundation of the chemiluminescent reductive acridinium triggering (CRAT) assay [28]. The mechanism of this reaction can be seen in Figure 4. DTT or GSH were adopted as reductants in the CRAT assay to produce H_2O_2 , which reacted with acridinium ester after adding a slightly alkaline buffer into the solution. The emitting light during this reaction was determined by about 1 s with a luminescence meter. The intensity of the emitting light could be applied to quantify rates of H_2O_2 production [21]. CRAT, a relatively new methodology, has not yet been widely used in large air pollution studies.

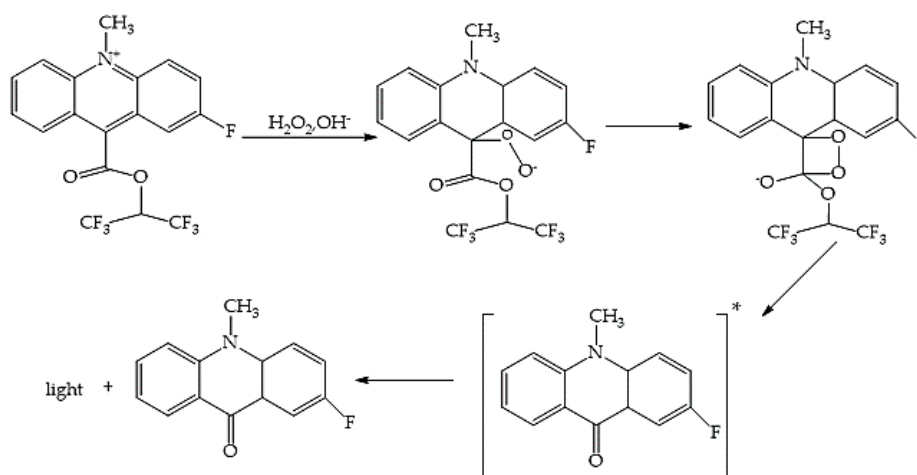


Figure 4. Mechanism for the chemiluminescence reactions of chemiluminescent reductive acridinium triggering (CRAT) assay [28].

2.4. Dichlorofluorescein Assay

Dichlorofluorescein is a non-fluorescent reagent that fluoresces when oxidized [29]. In the presence of horseradish peroxidase (HRP), DCFH could be rapidly oxidized to a fluorescent compound (DCF) (Figure 5). Prior to analysis, the DCFH in sodium phosphate buffer (pH = 7.2) was mixed with horseradish peroxidase (HRP) in a ratio [30]. This experiment was performed under dark conditions

to prevent DCFH photooxidation and reduce the variability of background H_2O_2 concentration [31]. Sample filters were suspended in a beaker containing DCFH–HRP, and sonicated to extract ROS from the particles. The formed DCF was determined by fluorescence at the excitation wavelengths of 485 nm and emission wavelengths of 530 nm. The measured fluorescence intensities were converted into equivalent H_2O_2 concentrations using least-squares analysis with a H_2O_2 calibration curve [29,32,33].

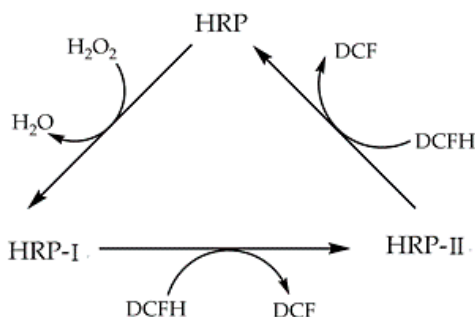


Figure 5. H_2O_2 reacts with horseradish peroxidase (HRP), following the reaction with dichlorofluorescein (DCFH) to produce fluorescence product fluorescent compound (DCF) [31].

2.5. Electron Paramagnetic/Spin Resonance Assay

The electron paramagnetic/spin resonance (EPR/ESR) assay was developed for the direct detection and quantification of materials containing unpaired electrons, such as free radicals or transition metal ions [13,34,35]. The physical assay can be measured by free radicals from the particles collected on filters. Stronger free radical signals generated from coal combustion fine particles ($\text{PM} < 1.1 \mu\text{m}$) could be measured (Figure 6).

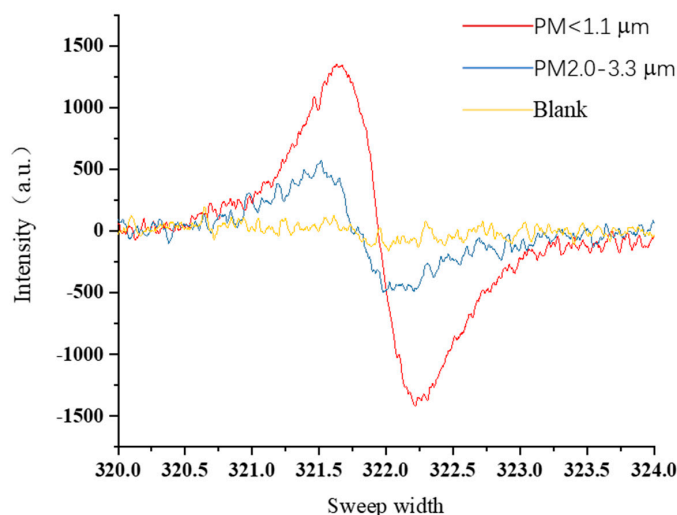


Figure 6. Free radicals generated from coal combustion particles. Free radical intensity varied with different particle sizes. Fine particles ($\text{PM} < 1.1 \mu\text{m}$) could generate stronger free radicals (present study, not published).

Additionally, the ESR assay could be used to test different free radicals with different spin traps, and the free radical spectrum pattern was different with different spin traps. Four peaks could be found in the pattern with 5,5-dimethyl-1-pyrroline-N-oxide (DMPO) spin trap, while only three peaks showed with 1-hydroxy-2,2,6,6-tetramethyl-4-oxo-piperidine (TEMPONE-H) (Figure 7). Oxidative potential of particles on filters based on the capacity of PMs to generate $\bullet\text{OH}$ via Fenton-type reaction in the presence of H_2O_2 with 5,5-dimethyl-1-pyrroline-N-oxide (DMPO) as a specific spin-trap [36,37].

DMPO–OH adducts could be formed from direct trapping of $\bullet\text{OH}$ (Equation (4)) or the decomposition of DMPO–OOH (Equation (5)) [36]. The OP was calculated from the sum of total amplitudes of the DMPO–OH quartet signal and expressed as the total amplitude arbitrary units divided by the volume of sampled air or weight of PMs [38,39].

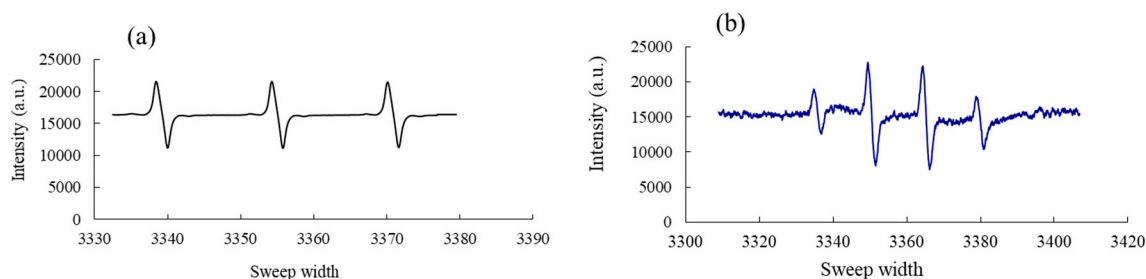
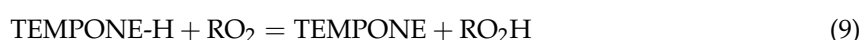


Figure 7. ESR spectra showed different patterns with different spin traps: (a) 1-hydroxy-2,2,6,6-tetramethyl-4-oxo-piperidine (TEMPONE-H) and (b) 5,5-dimethyl-1-pyrroline -N-oxide (DMPO) (present study, not published).

In addition to DMPO, another spin-trap 1-hydroxy-2,2,6,6-tetramethyl-4-oxo-piperidine (TEMPONE-H) was adapted to quantify total concentrations of ROS (Figure 7). TEMPONE-H is a closed shell molecule that could be transformed to radicals by deprotonation in a reaction with radicals. TEMPONE-H reduces peroxynitrite to NO_2 (Equation (6)) and reduces NO_2 to nitrite (Equation (7)). Moreover, TEMPONE-H can react with superoxide radicals (Equation (8)) and peroxy radicals (Equation (9)) with a formation of 1-hydroxy-2,2,6,6-tetramethyl-4-oxo-piperidinoxyl (TEMPONE) [40,41].



3. Comparison of Acellular Assays

Chemical constituents of PMs drive different responses by different assay types, and particle sizes also have significant influences on OP outcomes [9]. In order to provide a preferred method to measure OP, the sensitivity to different PMs species composition, size distribution and seasonality are discussed.

3.1. Sensitivity of Different Acellular Assays

Different OP assays capture different ROS species due to different redox reactions [9]. The DCFH assay measures particle-bound ROS that are inherently existing on and/or within the particle itself [29]. OP^{ESR} measures the certain production of ROS over time whereas OP^{RTL} and OP^{AA} measure the depletion of target molecules [28].

Certain OP assays respond differently by panels of chemicals and may also vary due to interactions between different components in PMs. For example, the DTT assay has different responses to metals and organic compounds. The OP^{DTT} was also sensitive to organic compounds (i.e., water-soluble organic carbon (WSOC), elemental carbon (EC), organic carbon (OC), black carbon, quinones and hydroxyquinones). OP^{CRAT} could react to trace metals Fe or Cu and quinones too [42,43]. $\text{OP}^{\text{AA-only}}$ was almost exclusively related with Cu(II) [44]. In the RTL assay, OP^{AA} and OP^{GSH} responded to different components of PMs as they were not significantly correlated with one another [45]. OP^{GSH}

was uniquely correlated with water-soluble Cu [19] and OP^{AA} had a strong positive correlation with Fe and Cu, the main elements that track non-exhaust traffic emissions [46]. Interestingly, OP^{AA} -only had a much stronger correlation with Cu(II) than OP^{AA} in the RTLF assay [44]. The OP^{ESR} was highly correlated with the traffic-related PMs components (i.e., Fe, Cu). OP^{ESR} and OP^{AA} showed the most similar results due to their large metal dependences [42].

3.2. PMs Size Distribution

Generally, redox active species with PMs size indicate that the OP of PMs is size determined [24]. OP^{DTT} in $PM_{2.5}$ was significantly higher than that in PM_{10} on a per mass basis [42] and per volume unit [47]. But sometimes, it was the reverse case for OP^{ESR} [42]. The OP^{ESR} in PM_{10} was 4.6 times greater than that in $PM_{2.5}$ when expressed per volume unit, and 3.1 times greater when expressed per mass unit showing higher OP in larger PMs size fractions. OP^{AA} was also more affected by components in coarse PMs as its responses were significantly higher for PM_{10} than that for $PM_{2.5}$ [48].

However, for a more refined division of particle size, OP^{DTT} was unimodal (Figure 8). Volume-normalized OP^{DTT} and mass-normalized OP^{DTT} peaked at submicron 0.56–1 μm and 0.1–0.32 μm respectively [24]. OP^{DTT} and OP^{DCFH} were similar in distribution, peaking at 0.32–1.8 μm , but AA assay was completely different. OP^{AA} was almost exclusively sensitive to PMs in the coarse mode (3.2–5.6 μm) [49]. The particles from brake/tire wear containing very high concentrations of Cu, Fe and Mn also belonged to the coarse fraction in the range of 3.2–5.6 μm . The similar distribution sizes indicated that OP^{AA} was significantly sensitive to PMs in the coarse mode produced by vehicular traffic, such as brake wear and re-suspended road dust. OP^{DTT} and OP^{DCFH} were more sensitive to fine PMs generated by combustion processes [49]. In the study of Fang et al., (2017), water-soluble OP^{DTT} peaked near 1–2.5 μm whereas water-insoluble OP^{DTT} was bimodal with both fine and coarse fractions due to the absorbed reactive species on the surfaces of PMs [50]. Overall, it seems that ultrafine and fine PMs are more important to OP^{DTT} value, but more work is required to reach precise conclusions.

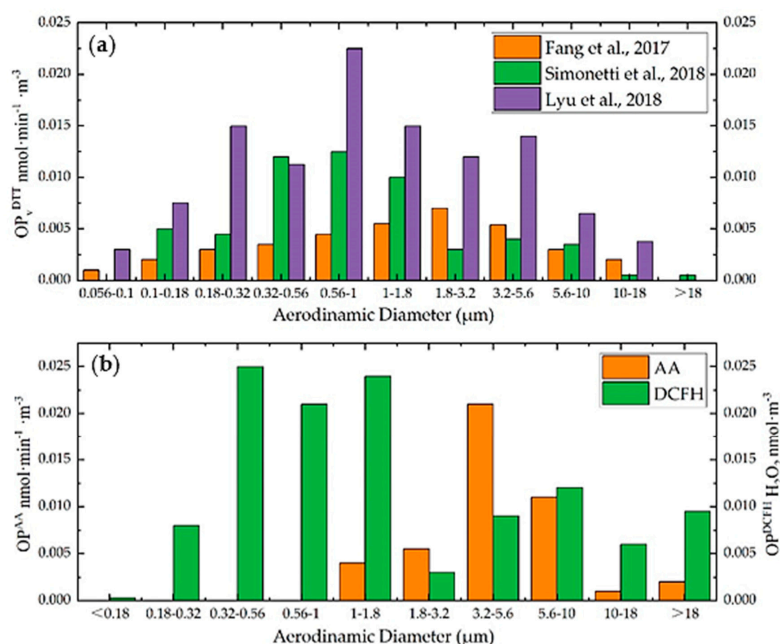


Figure 8. Oxidative potential of water-soluble fraction extracted from different size particles on a per volume unit. (a) the highest level of the oxidative potential (OP) was found in the size range from 1–2.5 μm , 0.32–1.8 μm and 0.56–1 μm by using DTT assay respectively [24,49,50]; (b) OP^{AA} was almost sensitive to particles in the coarse mode (3.2–5.6 μm) and OP^{DCFH} peaked at 0.32–1.8 μm [49].

3.3. OP Related with the Chemical Composition of PMs Collected in Different Seasons

Generally, significantly higher OP response could be measured in winter than that in summer for DTT and AA assays. The volume- and mass-based DTT (DTTv and DTTm) values of PM_{2.5} were significantly higher in the cold season possibly driven by biomass burning emission seasonality [51–53]. Average OP^{DTT} was 1.5 times and OP^{AA} was 2.3 times higher in the cold season than in the warm season [48]. However, OP^{AA} reached highest values near heavily trafficked highways and OP^{DTT} was more spatially homogeneous [44]. Calas et al., (2018) reported seasonal variations of the OP value by using four acellular assays (DTT, AA, RTLf and ESR) over a 1-year period, and showed only OP^{ESR} did not show seasonality and showed weaker relationships with other assays and chemical species [54]. But sometimes the OP data are different. In an urban background area of Athens, Greece, the OP value that used the DCFH assay was higher in summer, mainly as a result of higher concentrations of EC and WSOC during the warm season [55]. In Beijing, the seasonal averages of OP^{DTT} exhibited peak values in summer. This was correlated with the higher concentration of water-soluble organic components in summer produced by the photochemical formation of secondary organic aerosols [56].

3.4. Correlation with Health Impacts

Among the studies about the relationship between epidemiology and OP of PMs, the OP^{DTT} assay was a main method. OP^{DTT} has been found to be linked with several biological end points, such as, fraction of nitric oxide (FeNO) in exhaled breath [57], and mammalian cell cytotoxicity of PM_{2.5} [58], increased relative risk for asthma [5] and congestive heart failure [59]. A study in the Netherlands found that asthma and rhinitis were positively related with OP^{DTT}, but not significantly associated with OP^{ESR} [60]. Maikawa et al., (2016) investigated the relationship between airway inflammation in asthmatic children (FeNO) and oxidative potential of PM_{2.5} using RTLf assay. A positive correlation was found between FeNO and OP^{GSH} but not found between PM_{mass} or OP^{AA} [61]. OP^{DTT} and OP^{GSH} are the most relevant to health among several OP assays [9,13].

From the above, the DTT assay tends to be the preferred method to evaluate the oxidative potential, providing a more comprehensive measurement due to its high sensitivity to transition metals and organic compounds, which are mainly accumulated in fine PMs fractions (Table 1) [10]. DTT assay is highly correlated with health effects to better understand the link between PMs and human health. The DTT approach is the major point of discussion in the next section in this review in terms of driving forces.

Table 1. Comparison of acellular assays to assess the oxidative potential.

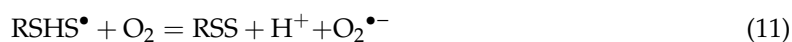
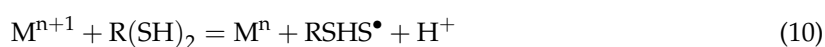
Assay	Requirements	Estimated Way	Sensitivity	Sources	Characteristic	Reference
DTT	fast, inexpensive, easy to perform and suitable for automation	the depletion rate of chemical proxies for cellular reductants	organic compounds; traffic-related metals; inorganic ions	biomass burning; brake/tire wear; traffic/fossil fuel combustion; photochemical aging	associated with fine fraction	[24,42,47,50]
AA		the antioxidant loss rate	metals	non-exhaust traffic emissions	associated with coarse particles	[48,49]
GSH		the antioxidant loss rate	Cu	non-exhaust traffic emissions	not a strong marker for traffic	[43]
DCFH		the increase in fluorescence intensity over time	organic compounds; inorganic ions	anthropic combustion; secondary aerosol	associated with fine fraction	[49]
CRAT	has not been widely used	the chemiluminescence reaction	transition metals, quinones	ambient particles	highly correlated with PM mass concentration	[21]
ESR	relatively little material, inexpensive	the ability of PMs to generate $\bullet\text{OH}$	transition metals; organic components	ambient particles	associated with coarse/fine particles	[42]

4. Drivers of Oxidative Potential

Considering that the PMs is consisted of organic and inorganic matters, these components could exacerbate free radical reaction and contribute to the measured OP response, and therefore the relationship between the OP and the main chemical components would play a key role in understanding the health risk (Table 2). In general, these following inorganic and organic species in the PMs showed the closest relationships with the OP.

4.1. Trace Metals

Metals, constituted 6–13% of the PMs mass [62], have been characterized in detail such as Ca, Mg, Ba, Al which originate from resuspension of soil dust emissions [49,63,64], while Fe, Cu, Zn, Cr, Cd are the transition metals associated with non-exhaust traffic emissions [62]. K could be considered as tracers of the biomass burning [33,48]



Transition metals (M, above) which have a certain amount of oxidative potential can catalyze the oxidation of DTT easily in Equation (10) [65]. The superoxide anion is formatted by the reaction between electron donors (DTT) and molecular oxygen (Equation (11)) [66]. Taking transition metals Cu and Fe for example, Cu(I) and Fe(II) can generate the highly reactive oxidant hydroxyl radical via the Fenton reaction (Equation (13)). And Cu(II) can be reduced to Cu(I), and Fe(III) to Fe(II) to complete a redox cycle (Equation (14)).

Table 2. Summary of driving species of oxidative potential (OP) mainly measured by DTT assay reported in literatures.

Location	Particles	Seasons	Sampling Period	Assay	Driving Species	Reference
Atlanta	PM _{2.5}	One year	Jan–Dec, 2017	DTT	BrC, EC, K, Fe, Cu	[51]
Central Mediterranean Sea	PM ₁₀ , PM _{2.5}	One year	2014–2015	DTT, AA	K ⁺ , NO ₃ [−] , Ba, Cd, Cu, Fe, Mn, P, V, OC, EC	[48]
Atlanta	PM _{2.5}	One year	2017	DTT, RTLF	WSOC, OC, EC, Fe, Cu, Mn	[19]
the University of Illinois, Urbana–Champaign	Ambient PM _{2.5}	Spring	Feb–Apr, 2017	DTT	HULIS, Fe, Cu, Mn	[67]
the Central Mediterranean basin	PM ₁₀	One year	Dec 2014–Oct 2015	DTT AA	Ba, Cd, Ce, Cr, Cu, Fe, EC, OC	[68]
Indo-Gangetic Plain	PM _{2.5}	Winter	2014	DTT	OC, EC, WSOC,	[64]
Beijing	PM _{2.5}	One year	2012	DTT	HULIS	[69]
Italy	PM ₁₀	One year	Feb–Nov 2015 Apr–May 2016	DTT, AA	SO ₄ ^{2−} , NH ₄ ⁺ , K ⁺ , Mg ²⁺ , Ca ²⁺ , Ca, Mg, K, Mn, Cu, Rb, Zn, WSOC	[70]
the littoral zone of the Bohai Sea	PM _{2.5}	One year	2016	DTT	WSOC, EC, Mn, Co, Fe, Cr, Cd, SO ₄ ^{2−} , NH ₄ ⁺ , NO ₃ [−]	[52]
Italy	Size-segregated PMs	spring	Feb–Mar, 2017	AA, DTT, DCFH	Cu, Fe, Mn, As, B, Cd, Cr, Mo, Se, Ni, Pb, K, Rb	[49]
the Los Angeles Basin	PM _{2.5}	winter	Oct 2014–Jan 2015; Nov 2015–Jan 2016	DTT	Ba, Cr, Cu, Mn, Ni, Pb, Sb and Zn, EC, OC	[71]

Abbreviations: BrC: brown carbon; EC: elemental carbon; OC: organic carbon; HULIS: Humic-like Substances, WSOC: water-soluble organic carbon.

Numerous studies have assessed the intrinsic OP induced by metals [24,27,72,73]. A summary of studies is presented in Figure 9. In this review, we followed the procedure of Lu et al., (2019) for the DTT assay [74]. Interestingly, the units of this review are different from other literatures, but the underlying trends of the intrinsic OP of quinones and metals are the same. As shown in Figure 9, Cu(II) and Mn(II) were the most active metals in the DTT assay and Co was the third most reactive metal. The reactivity of other transition metals followed the pattern $V(V) \approx Ni(II) > Pb(II) \approx Fe(II) > Fe(III)$. In Figure 10, Cu(II) and Mn(II) were well fitted by power functions, but the concentration responses for Fe(II) and Fe(III) were linear [24]. Even though the intrinsic OP of metals have been quantified by using inorganic salt solutions, the importance for OP assessment of ambient PMs also depends upon the mass concentration of each metal. For example, Fe which made a modest contribution on OP^{DTT} values accounted for the majority of DTT loss from typical ambient $PM_{2.5}$ due to its abundant concentrations [24,73]. Therefore, Fe was responsible for the DTT assumption. However, the solubility of metal ions can be enhanced by the presence of organic ligands in the PMs. For example, the solubility of Fe was strongly correlated with the concentration of oxalate [75]. Sulfate was an important proxy for Fe solubility by affecting aerosol pH [62].

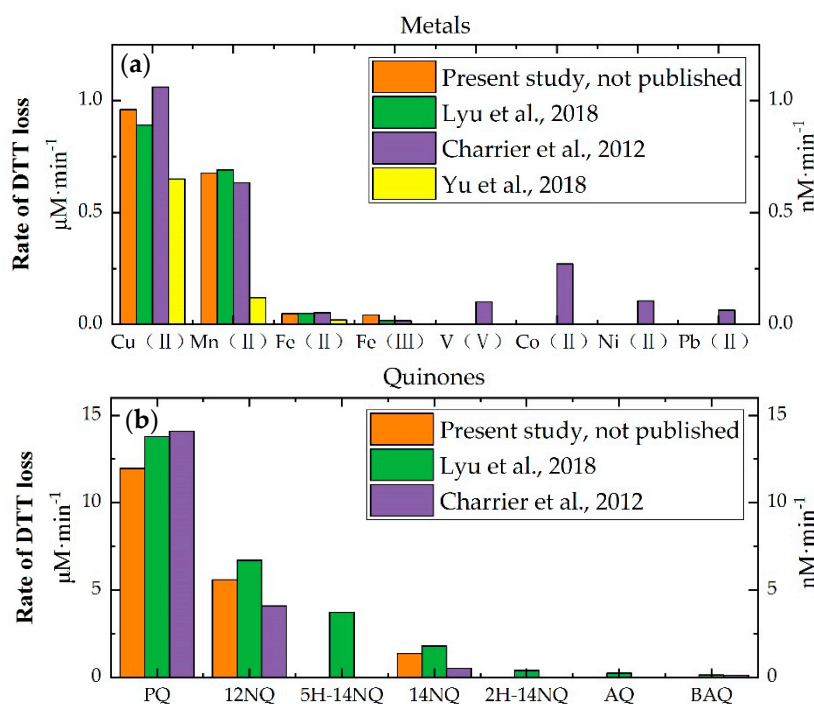


Figure 9. Intrinsic OP of individual (a) metals and (b) quinones [24,67,73]. The concentration of each species was 1 μM . The data in this paper are in $nM \cdot min^{-1}$ while in other literatures they are in $\mu M \cdot min^{-1}$.

The concentration of H^+ could easily destroy the mineral structures to release the structural or interlayer Fe [76]. The dissolved Fe from minerals (e.g., pyrite, illite, chlorite or kaolinite) had considerable oxidative potential [76]. Moreover, minerals could induce ROS formation as a “carrier” of more toxic species such as metals and surface-absorbed polycyclic aromatic hydrocarbons (PAHs). Besides, Fe and Cu were very efficient in forming complexes with Suwanee river fulvic acid (SRFA), making them unlikely to exist in free ionic forms and significantly altering their OP [72].

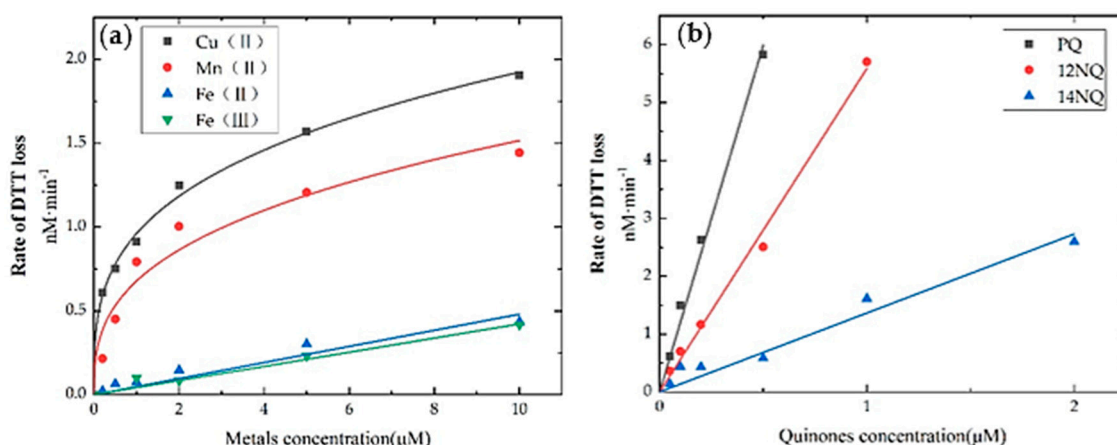


Figure 10. Rate of DTT loss as a function of concentration of redox-active species: (a) results for the four metals that oxidized DTT; (b) results for the three DTT-reactive quinones (present study, not published).

The correlations between OP^{DTT} and metal concentration in ambient PMs do not indicate causation, since particle species tend to be highly covariate. In winter, the strong correlation with non-redox active metal K should be analyzed carefully because this association may be due to a similar increase in K and quinones concentrations, rather than establishing an inevitable causal relationship [77]. The redox inert ions (i.e., Zn, Cd, Pb) also contribute to the oxidative stress of PMs exposure by interacting with cysteinyl thiols on glutathione and key regulatory proteins [66,78]. And Ca^{2+} , Mg^{2+} , potentially presenting in large quantities in $PM_{2.5}$, can strongly modulate PMs acidity [79].

4.2. Carbonaceous Species

Carbonaceous species include elemental carbon (EC), organic carbon (OC) and water soluble organic carbon (WSOC). OC may be widely used as a tracer of biomass burning [80]; polycyclic aromatic hydrocarbons (PAHs) are emitted during incomplete combustion processes including traffic [55]; and quinones are the markers of photochemical formation of secondary organic aerosols (SOA) [33].

Quinones are capable of destroying the proximal thiol of DTT [65,81]. Taking the reaction between DTT and 14NQ for example in the Figure 11, DTT has two thiol groups in close proximity that readily form a stable six membered ring with 14NQ reduced (Reaction 1). And then 14NQ rearranges to form the 1,4-naphthoquinol (Reaction 2). Quinol forms a free radical-semiquinone through two ways (Reaction 3_a, 3_b). Then the semiquinone is transformed to the original quinone by reacting with O_2 to complete a redox cycle (Reaction 4). Similarly, many researchers started with the standard solutions of known quinones (9,10-phenanthraquinone, 1,2-naphthoquinone, 1,4-naphthoquinone, 2-hydroxy-1,4-naphthoquinone, 9,10-anthracenequinone, benz(a)anthracene-7,12-quinone and 5-hydroxy-1,4-naphthoquinone) measuring the oxidation potential of quinones (Figure 9) [67]. As shown in Figure 10, the rates of DTT consumption from quinones were proportional to their concentration [24]. PQ was the most reactive species, followed by 12NQ, 5H-14NQ, 14NQ, 2H-14NQ, AQ, and BAQ. On a concentration normalized basis, quinones, especially PQ, could be much more reactive than other species, but their concentrations were generally low [73]. Quinones in traffic-related PMs, especially NQ and AQ, had considerable pro-inflammatory and genotoxic potential which could cause lung impairment. As the secondary derivatives of PAHs in the air, quinones made a great contribution to the toxicity of airborne PMs [82]. There exists correlation between mass level of PAHs, quinones and DTT consumption [73]. Quinones in the atmosphere are formed by tropospheric conversions of precursors PAHs via photochemical reactions and reactions with $\bullet OH$, nitrate radicals and ozone (O_3) [24,83,84]. Many studies showed a correlation between OC and/or WSOC and OP^{DTT} of PMs collected during different seasons [63,64,74,85]. Besides, Chen et al.,

(2019) found that the light absorbing substance BrC3 and the fluorescent substance C7 were important contributors to the DTT activity of water-soluble PMs [8].

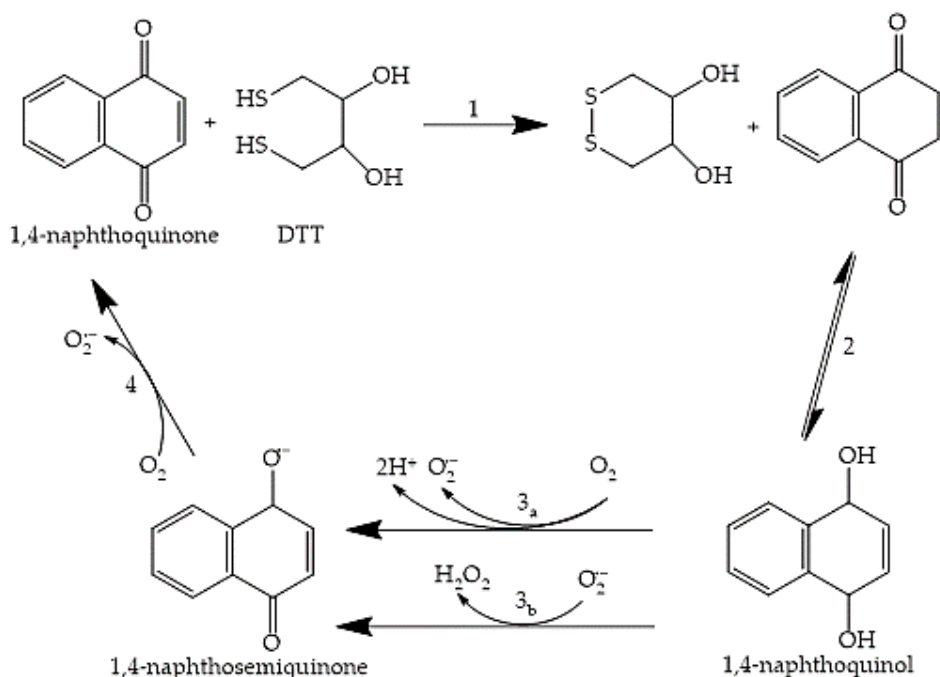
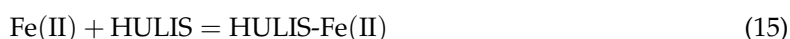
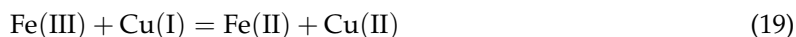
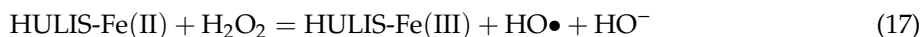
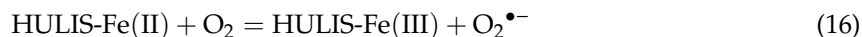


Figure 11. The reaction mechanism between DTT and 1,4-naphthoquinone (14NQ), including 1, 2, 3a, 3b and 4. Reaction 1 shows DTT readily forms a stable six membered ring with 14NQ reduced; Reaction 2 shows 14NQ rearranges to form the 1,4-naphthoquinol; Quinols form a free radical-semiquinone through Reaction 3a or 3b and the semiquinone is transformed to the original quinone by Reaction 4 [65].

Biomass burning which could release high levels of particulate matter with PAHs and volatile organic compounds (VOCs) played a primary role in PMs capability to generate ROS [49,86]. And there was also a large amount of quinones in fresh wood smoke organic aerosols, which could well explain the DTT consumption [87]. In the meantime, OP^{DTT} caused by vehicle emission particles was high because of the semivolatile organic species [85].

There could be both synergistic and antagonistic interactions among PMs components, which could cause different OP responses [88]. Humic-like substances (HULIS) are water soluble compounds [89]. HULIS, consisting of powerful chelating functional groups (i.e., carboxyl (-COOH), hydroxyl (-CH₂OH) and carbonyl (-COCH₃), can chelate the transition metals in aerosols and are capable of participating in redox cycling. Higher OP^{DTT} value was been found in HULIS-Fe(II) complexes than Fe(II) alone [74]. Fe(II) itself can produce ROS species through Fenton reactions (Equation (13)), and Fenton-based HULIS-Fe(II) complex system can be expressed through Equations (15)–(17). Quinones could catalyze the generation of H₂O₂ and •OH [27]. And the interactions between quinones and Fe were additive in OP^{DTT} but synergistically in forming •OH. Magnesium could have synergistic effects with quinones, but Cu were different in the OP^{DTT} assay [67]. When mixing with Cu, the OP^{DTT} value of AQ decreased significantly [88]. And there could be synergistic effects on •OH generation for the mixtures of Cu(II) + Fe(II), PQ + Fe(II), and Cu(II) + Fe(II) + 1,2-NQ, Cu(II) + Fe(II) + PQ [90]. Fe(II) should be rapidly oxidized to Fe(III) (Equation (13) (18)), and Cu(I) can also readily reduce Fe(III) (Equation (19)). Cu(II) is continuously reduced by O₂•⁻, providing a steady source of reduced iron (Equation (14)) [91]. This may explain the synergistic effect of Fe(II) and Cu(II).





4.3. Ionic Species

K^+ , Cl^- , Na^+ , SO_4^{2-} , NO_3^- , NH_4^+ are the main ions in the PMs [63,92]. These ions also played roles in assessment of the OP. According to Patel et al., (2018), there was a significant negative correlation ($p < 0.05$) with SO_4^{2-} , NO_3^- , NH_4^+ which indicated that they do not affect DTT activity [64]. But SO_2 and NO_2 which are the main precursors of secondary inorganic aerosols showed strong correlations with DTT [63]. Secondary acids, such as ammonium sulfate, can have an indirect effect on OP through reducing pH in the assay of OP^{DTT} [62,76].

4.4. Water-Insoluble Components

It was noted that insoluble fraction of PMs had a relationship with the OP. The OP^{DTT} values of ambient particles without filtration were higher than that of water-soluble fraction [93]. In the studies of Gao et al., (2020) and Fang et al., (2017), water-insoluble OP ($\text{OP}^{\text{WI-DTT}}$) was determined by the difference between the water-soluble ($\text{OP}^{\text{WS-DTT}}$) and total OP ($\text{OP}^{\text{Total-DTT}}$). The $\text{OP}^{\text{WI-DTT}}$ comprised 20% of total PMs OP on average [50,51]. However, the main drives of $\text{OP}^{\text{WS-DTT}}$ and $\text{OP}^{\text{WI-DTT}}$ appeared to be the same, implying that there is a connection between $\text{OP}^{\text{WI-DTT}}$ and dust surface property.

5. Conclusions

Acellular assays including the RTLF, AA, GSH, DTT, CRAT, DCFH and ESR for measurement of OP were reviewed. We compared sensitivity to different PMs species composition, PMs size distribution and seasonality of different assays and concluded that the DTT assay could be the preferred method due to its sensitivity to tracers of combustion derived transition metals and aromatic organic compounds. Specific transition metals (i.e., Mn, Cu, Fe) and quinones made a great contribution to OP^{DTT} value. However, the interactions among PMs components which may change the redox properties of PMs species cannot be ignored, and also insoluble fraction of PMs could not be ignored in assessment of the oxidative potential of PMs.

Additionally, considering that the OP has several dimensions, a combination of two or more OP measures may be needed to create an accurate predictor for health effects.

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References

1. Hamanaka, R.B.; Mutlu, G.M. Particulate Matter Air Pollution: Effects on the Cardiovascular System. *Front. Endocrinol.* **2018**, *9*, 680. [CrossRef] [PubMed]
2. Lim, C.C.; Thurston, G.D. Air Pollution, Oxidative Stress, and Diabetes: A Life Course Epidemiologic Perspective. *Curr. Diabetes Rep.* **2019**, *19*, 1–14. [CrossRef] [PubMed]

3. Rao, X.; Zhong, J.; Brook, R.D.; Rajagopalan, S. Effect of Particulate Matter Air Pollution on Cardiovascular Oxidative Stress Pathways. *Antioxid. Redox Signal.* **2018**, *28*, 797–818. [\[CrossRef\]](#) [\[PubMed\]](#)
4. Rajper, S.A.; Ullah, S.; Wang, J.-C. Exposure to air pollution and self-reported effects on Chinese students: A case study of 13 megacities. *PLoS ONE* **2018**, *13*, e0194364. [\[CrossRef\]](#)
5. Delfino, R.J.; Staimer, N.; Tjoa, T.; Gillen, D.L.; Schauer, J.J.; Shafer, M.M. Airway inflammation and oxidative potential of air pollutant particles in a pediatric asthma panel. *J. Expo. Sci. Environ. Epidemiol.* **2013**, *23*, 466–473. [\[CrossRef\]](#) [\[PubMed\]](#)
6. Romano, S.; Perrone, M.; Becagli, S.; Pietrogrande, M.; Russo, M.; Caricato, R.; Lionetto, M. Ecotoxicity, genotoxicity, and oxidative potential tests of atmospheric PM10 particles. *Atmos. Environ.* **2020**, *221*, 117085. [\[CrossRef\]](#)
7. Song, Y.; Zhang, Y.; Li, R.; Chen, W.; Chung, C.K.A.; Cai, Z. The cellular effects of PM2.5 collected in Chinese Taiyuan and Guangzhou and their associations with polycyclic aromatic hydrocarbons (PAHs), nitro-PAHs and hydroxy-PAHs. *Ecotoxicol. Environ. Saf.* **2020**, *191*, 110225. [\[CrossRef\]](#)
8. Chen, Q.; Wang, M.; Wang, Y.; Zhang, L.; Li, Y.; Han, Y. Oxidative Potential of Water-Soluble Matter Associated with Chromophoric Substances in PM2.5 over Xi'an, China. *Environ. Sci. Technol.* **2019**, *53*, 8574–8584. [\[CrossRef\]](#) [\[PubMed\]](#)
9. Bates, J.T.; Fang, T.; Verma, V.; Zeng, L.; Weber, R.J.; Tolbert, P.E.; Abrams, J.Y.; Sarnat, S.E.; Klein, M.; Mulholland, J.A.; et al. Review of Acellular Assays of Ambient Particulate Matter Oxidative Potential: Methods and Relationships with Composition, Sources, and Health Effects. *Environ. Sci. Technol.* **2019**, *53*, 4003–4019. [\[CrossRef\]](#)
10. Leni, Z.; Künzi, L.; Geiser, M. Air pollution causing oxidative stress. *Curr. Opin. Toxicol.* **2020**, 1–8. [\[CrossRef\]](#)
11. Øvrevik, J. Oxidative Potential Versus Biological Effects: A Review on the Relevance of Cell-Free/Abiotic Assays as Predictors of Toxicity from Airborne Particulate Matter. *Int. J. Mol. Sci.* **2019**, *20*, 4772. [\[CrossRef\]](#)
12. Weber, S.; Uzu, G.; Calas, A.; Chevrier, F.; Besombes, J.-L.; Charron, A.; Salameh, D.; Ježek, I.; Močnik, G.; Jaffrezo, J.-L. An apportionment method for the oxidative potential of atmospheric particulate matter sources: Application to a one-year study in Chamonix, France. *Atmos. Chem. Phys. Discuss.* **2018**, *18*, 9617–9629. [\[CrossRef\]](#)
13. Gao, D.; Ripley, S.; Weichenthal, S.; Pollitt, K.J.G. Ambient particulate matter oxidative potential: Chemical determinants, associated health effects, and strategies for risk management. *Free. Radic. Biol. Med.* **2020**, *151*, 7–25. [\[CrossRef\]](#)
14. Mudway, I.S.; Stenfors, N.; Duggan, S.T.; Roxborough, H.; Zielinski, H.; Marklund, S.L.; Blomberg, A.; Frew, A.J.; Sandström, T.; Kelly, F.J. An in vitro and in vivo investigation of the effects of diesel exhaust on human airway lining fluid antioxidants. *Arch. Biochem. Biophys.* **2004**, *423*, 200–212. [\[CrossRef\]](#)
15. Ayres, J.G.; Borm, P.; Cassee, F.R.; Castranova, V.; Donaldson, K.; Ghio, A.; Harrison, R.M.; Hider, R.; Kelly, F.; Kooter, I.M.; et al. Evaluating the Toxicity of Airborne Particulate Matter and Nanoparticles by Measuring Oxidative Stress Potential—A Workshop Report and Consensus Statement. *Inhal. Toxicol.* **2008**, *20*, 75–99. [\[CrossRef\]](#)
16. Godri, K.J.; Duggan, S.T.; Fuller, G.W.; Baker, T.; Green, D.; Kelly, F.J.; Mudway, I.S. Particulate Matter Oxidative Potential from Waste Transfer Station Activity. *Environ. Health Perspect.* **2009**, *118*, 493–498. [\[CrossRef\]](#)
17. Crobeddu, B.; Aragao-Santiago, L.; Bui, L.-C.; Boland, S.; Squiban, A.B. Oxidative potential of particulate matter 2.5 as predictive indicator of cellular stress. *Environ. Pollut.* **2017**, *230*, 125–133. [\[CrossRef\]](#)
18. Zielinski, H.; Mudway, I.S.; Bérubé, K.; Murphy, S.; Richards, R.; Kelly, F.J. Modeling the interactions of particulates with epithelial lining fluid antioxidants. *Am. J. Physiol. Content* **1999**, *277*, L719–L726. [\[CrossRef\]](#)
19. Gao, D.; Pollitt, K.J.G.; Mulholland, J.A.; Russell, A.G.; Weber, R.J. Characterization and comparison of PM2.5 oxidative potential assessed by two acellular assays. *Atmos. Chem. Phys. Discuss.* **2020**, *20*, 5197–5210. [\[CrossRef\]](#)
20. Pipolo, S.; Percudani, R.; Cammi, R. Absolute stereochemistry and preferred conformations of urate degradation intermediates from computed and experimental circular dichroism spectra. *Org. Biomol. Chem.* **2011**, *9*, 5149–5155. [\[CrossRef\]](#) [\[PubMed\]](#)
21. Yang, A.; Jedynska, A.; Hellack, B.; Kooter, I.; Hoek, G.; Brunekreef, B.; Kuhlbusch, T.A.; Cassee, F.R.; Janssen, N.A. Measurement of the oxidative potential of PM2.5 and its constituents: The effect of extraction solvent and filter type. *Atmos. Environ.* **2014**, *83*, 35–42. [\[CrossRef\]](#)

22. Visentin, M.; Pagnoni, A.; Sarti, E.; Pietrogrande, M.C. Urban PM_{2.5} oxidative potential: Importance of chemical species and comparison of two spectrophotometric cell-free assays. *Environ. Pollut.* **2016**, *219*, 72–79. [[CrossRef](#)]
23. Bradshaw, M.P.; Barril, C.; Clark, A.; Prenzler, P.D.; Scollary, G.R. Ascorbic Acid: A Review of its Chemistry and Reactivity in Relation to a Wine Environment. *Crit. Rev. Food Sci. Nutr.* **2011**, *51*, 479–498. [[CrossRef](#)]
24. Lyu, Y.; Guo, H.; Cheng, T.; Li, X. Particle Size Distributions of Oxidative Potential of Lung-Deposited Particles: Assessing Contributions from Quinones and Water-Soluble Metals. *Environ. Sci. Technol.* **2018**, *52*, 6592–6600. [[CrossRef](#)] [[PubMed](#)]
25. Biswas, S.; Verma, V.; Schauer, J.J.; Cassee, F.R.; Cho, A.K.; Sioutas, C. Oxidative Potential of Semi-Volatile and Non Volatile Particulate Matter (PM) from Heavy-Duty Vehicles Retrofitted with Emission Control Technologies. *Environ. Sci. Technol.* **2009**, *43*, 3905–3912. [[CrossRef](#)]
26. Cho, A.K.; Sioutas, C.; Miguel, A.H.; Kumagai, Y.; Schmitz, D.A.; Singh, M.; Eiguren-Fernandez, A.; Froines, J.R. Redox activity of airborne particulate matter at different sites in the Los Angeles Basin. *Environ. Res.* **2005**, *99*, 40–47. [[CrossRef](#)] [[PubMed](#)]
27. Xiong, Q.; Yu, H.; Wang, R.; Wei, J.; Verma, V. Rethinking Dithiothreitol-Based Particulate Matter Oxidative Potential: Measuring Dithiothreitol Consumption versus Reactive Oxygen Species Generation. *Environ. Sci. Technol.* **2017**, *51*, 6507–6514. [[CrossRef](#)]
28. Zomer, B.; Collé, L.; Jedyńska, A.; Pasterkamp, G.; Kooter, I.; Bloemen, H. Chemiluminescent reductive acridinium triggering (CRAT)—Mechanism and applications. *Anal. Bioanal. Chem.* **2011**, *401*, 2945–2954. [[CrossRef](#)]
29. Venkatachari, P.; Hopke, P.K.; Grover, B.D.; Eatough, D.J. Measurement of Particle-Bound Reactive Oxygen Species in Rubidoux Aerosols. *J. Atmos. Chem.* **2005**, *50*, 49–58. [[CrossRef](#)]
30. Perrone, M.G.; Zhou, J.; Malandrino, M.; Sangiorgi, G.; Rizzi, C.; Ferrero, L.; Dommen, J.; Bolzacchini, E. PM chemical composition and oxidative potential of the soluble fraction of particles at two sites in the urban area of Milan, Northern Italy. *Atmos. Environ.* **2016**, *128*, 104–113. [[CrossRef](#)]
31. Fuller, S.; Wragg, F.; Nutter, J.; Kalberer, M. Comparison of on-line and off-line methods to quantify reactive oxygen species (ROS) in atmospheric aerosols. *Atmos. Environ.* **2014**, *92*, 97–103. [[CrossRef](#)]
32. Venkatachari, P.; Hopke, P.K.; Brune, W.H.; Ren, X.; Leshner, R.; Mao, J.; Mitchell, M. Characterization of Wintertime Reactive Oxygen Species Concentrations in Flushing, New York. *Aerosol Sci. Technol.* **2007**, *41*, 97–111. [[CrossRef](#)]
33. Pietrogrande, M.C.; Russo, M.; Zagatti, E. Review of PM Oxidative Potential Measured with Acellular Assays in Urban and Rural Sites across Italy. *Atmosphere* **2019**, *10*, 626. [[CrossRef](#)]
34. Lu, S.; Duffin, R.; Poland, C.; Daly, P.; Murphy, F.; Drost, E.; MacNee, W.; Stone, V.; Donaldson, K. Efficacy of Simple Short-Term in Vitro Assays for Predicting the Potential of Metal Oxide Nanoparticles to Cause Pulmonary Inflammation. *Environ. Health Perspect.* **2009**, *117*, 241–247. [[CrossRef](#)]
35. Senlin, L.; Zhang, W.; Zhang, R.; Liu, P.; Wang, Q.; Shang, Y.; Wu, M.; Donaldson, K.; Wang, Q. Comparison of cellular toxicity caused by ambient ultrafine particles and engineered metal oxide nanoparticles. *Part. Fibre Toxicol.* **2015**, *12*, 5. [[CrossRef](#)]
36. Shi, T.; Schins, R.P.F.; Knaapen, A.M.; Kuhlbusch, T.; Pitz, M.; Heinrich, J.; Borm, P.J.A. Hydroxyl radical generation by electron paramagnetic resonance as a new method to monitor ambient particulate matter composition. *J. Environ. Monit.* **2003**, *5*, 550–556. [[CrossRef](#)]
37. Shi, T.; Knaapen, A.; Begerow, J.; Birmili, W.; Borm, P.; Schins, R.P.F. Temporal variation of hydroxyl radical generation and 8-hydroxy-2'-deoxyguanosine formation by coarse and fine particulate matter. *Occup. Environ. Med.* **2003**, *60*, 315–321. [[CrossRef](#)]
38. Boogaard, H.; Janssen, N.A.; Fischer, P.H.; Kos, G.P.; Weijers, E.P.; Cassee, F.R.; Van Der Zee, S.C.; De Hartog, J.J.; Brunekreef, B.; Hoek, G. Contrasts in Oxidative Potential and Other Particulate Matter Characteristics Collected Near Major Streets and Background Locations. *Environ. Health Perspect.* **2012**, *120*, 185–191. [[CrossRef](#)]
39. Yang, A.; Hellack, B.; Leseman, D.; Brunekreef, B.; Kuhlbusch, T.A.; Cassee, F.; Hoek, G.; Janssen, N.A. Temporal and spatial variation of the metal-related oxidative potential of PM_{2.5} and its relation to PM_{2.5} mass and elemental composition. *Atmos. Environ.* **2015**, *102*, 62–69. [[CrossRef](#)]

40. Tong, H.; Lakey, P.S.J.; Arangio, A.M.; Socorro, J.; Shen, F.; Lucas, K.; Brune, W.H.; Pöschl, U.; Shiraiwa, M. Reactive Oxygen Species Formed by Secondary Organic Aerosols in Water and Surrogate Lung Fluid. *Environ. Sci. Technol.* **2018**, *52*, 11642–11651. [\[CrossRef\]](#)
41. Dikalov, S.; Skatchkov, M.; Bassenge, E. Quantification of Peroxynitrite, Superoxide, and Peroxyl Radicals by a New Spin Trap Hydroxylamine 1-Hydroxy-2,2,6,6-tetramethyl-4-oxo-piperidine. *Biochem. Biophys. Res. Commun.* **1997**, *230*, 54–57. [\[CrossRef\]](#) [\[PubMed\]](#)
42. Janssen, N.A.; Yang, A.; Strak, M.; Steenhof, M.; Hellack, B.; Gerlofs-Nijland, M.E.; Kuhlbusch, T.; Kelly, F.; Harrison, R.; Brunekreef, B.; et al. Oxidative potential of particulate matter collected at sites with different source characteristics. *Sci. Total. Environ.* **2014**, *472*, 572–581. [\[CrossRef\]](#)
43. Weichenthal, S.; Shekarzifard, M.; Traub, A.; Kulka, R.; Al-Rijleh, K.; Anowar, S.; Evans, G.J.; Hatzopoulou, M. Within-City Spatial Variations in Multiple Measures of PM_{2.5} Oxidative Potential in Toronto, Canada. *Environ. Sci. Technol.* **2019**, *53*, 2799–2810. [\[CrossRef\]](#) [\[PubMed\]](#)
44. Fang, T.; Verma, V.; Bates, J.T.; Abrams, J.; Klein, M.; Strickland, M.J.; Sarnat, S.E.; Chang, H.H.; Mulholland, J.A.; Tolbert, P.E.; et al. Oxidative potential of ambient water-soluble PM_{2.5} in the southeastern United States: Contrasts in sources and health associations between ascorbic acid (AA) and dithiothreitol (DTT) assays. *Atmos. Chem. Phys. Discuss.* **2016**, *16*, 3865–3879. [\[CrossRef\]](#)
45. Godri, K.J.; Harrison, R.M.; Evans, T.; Baker, T.; Dunster, C.; Mudway, I.S.; Kelly, F.J. Increased Oxidative Burden Associated with Traffic Component of Ambient Particulate Matter at Roadside and Urban Background Schools Sites in London. *PLoS ONE* **2011**, *6*, e21961. [\[CrossRef\]](#)
46. Molina, C.; Toro, R.; Manzano, C.A.; Canepari, S.; Massimi, L.; Leiva-Guzmán, M.A. Airborne Aerosols and Human Health: Leapfrogging from Mass Concentration to Oxidative Potential. *Atmosphere* **2020**, *11*, 917. [\[CrossRef\]](#)
47. Chirizzi, D.; Cesari, D.; Guascito, M.R.; Dinoi, A.; Giotta, L.; Donato, A.; Contini, D. Influence of Saharan dust outbreaks and carbon content on oxidative potential of water-soluble fractions of PM_{2.5} and PM₁₀. *Atmos. Environ.* **2017**, *163*, 1–8. [\[CrossRef\]](#)
48. Perrone, M.R.; Bertoli, I.; Romano, S.; Russo, M.; Rispoli, G.; Pietrogrande, M.C. PM_{2.5} and PM₁₀ oxidative potential at a Central Mediterranean Site: Contrasts between dithiothreitol- and ascorbic acid-measured values in relation with particle size and chemical composition. *Atmos. Environ.* **2019**, *210*, 143–155. [\[CrossRef\]](#)
49. Simonetti, G.; Conte, E.; Perrino, C.; Canepari, S. Oxidative potential of size-segregated PM in an urban and an industrial area of Italy. *Atmos. Environ.* **2018**, *187*, 292–300. [\[CrossRef\]](#)
50. Fang, T.; Zeng, L.; Gao, D.; Verma, V.; Stefaniak, A.B.; Weber, R.J. Ambient Size Distributions and Lung Deposition of Aerosol Dithiothreitol-Measured Oxidative Potential: Contrast between Soluble and Insoluble Particles. *Environ. Sci. Technol.* **2017**, *51*, 6802–6811. [\[CrossRef\]](#)
51. Gao, D.; Mulholland, J.A.; Russell, A.G.; Weber, R.J. Characterization of water-insoluble oxidative potential of PM_{2.5} using the dithiothreitol assay. *Atmos. Environ.* **2020**, *224*, 117327. [\[CrossRef\]](#)
52. Liu, W.; Xu, Y.; Liu, W.; Liu, Q.; Yu, S.; Liu, Y.; Wang, X.; Tao, S. Oxidative potential of ambient PM_{2.5} in the coastal cities of the Bohai Sea, northern China: Seasonal variation and source apportionment. *Environ. Pollut.* **2018**, *236*, 514–528. [\[CrossRef\]](#) [\[PubMed\]](#)
53. Cesari, D.; Merico, E.; Grasso, F.M.; Decesari, S.; Belosi, F.; Manarini, F.; De Nuntiis, P.; Rinaldi, M.; Volpi, F.; Gambaro, A.; et al. Source Apportionment of PM_{2.5} and of its Oxidative Potential in an Industrial Suburban Site in South Italy. *Atmosphere* **2019**, *10*, 758. [\[CrossRef\]](#)
54. Calas, A.; Uzu, G.; Kelly, F.J.; Houdier, S.; Martins, J.M.F.; Thomas, F.; Molton, F.; Charron, A.; Dunster, C.; Ollie, A.; et al. Comparison between five acellular oxidative potential measurement assays performed with detailed chemistry on PM₁₀ samples from the city of Chamonix (France). *Atmos. Chem. Phys. Discuss.* **2018**, *18*, 7863–7875. [\[CrossRef\]](#)
55. Taghvaei, S.; Sowlat, M.H.; Diapouli, E.; Manousakas, M.I.; Vasilatou, V.; Eleftheriadis, K.; Sioutas, C. Source apportionment of the oxidative potential of fine ambient particulate matter (PM_{2.5}) in Athens, Greece. *Sci. Total. Environ.* **2019**, *653*, 1407–1416. [\[CrossRef\]](#)
56. Yu, S.; Liu, W.; Xu, Y.; Yi, K.; Zhou, M.; Tao, S.; Liu, W. Characteristics and oxidative potential of atmospheric PM_{2.5} in Beijing: Source apportionment and seasonal variation. *Sci. Total. Environ.* **2019**, *650*, 277–287. [\[CrossRef\]](#)

57. Janssen, N.A.; Strak, M.; Yang, A.; Hellack, B.; Kelly, F.J.; Kuhlbusch, T.A.J.; Harrison, R.M.; Brunekreef, B.; Cassee, F.R.; Steenhof, M.; et al. Associations between three specific α -cellular measures of the oxidative potential of particulate matter and markers of acute airway and nasal inflammation in healthy volunteers. *Occup. Environ. Med.* **2014**, *72*, 49–56. [\[CrossRef\]](#)
58. Wang, Y.; Plewa, M.J.; Mukherjee, U.K.; Verma, V. Assessing the cytotoxicity of ambient particulate matter (PM) using Chinese hamster ovary (CHO) cells and its relationship with the PM chemical composition and oxidative potential. *Atmos. Environ.* **2018**, *179*, 132–141. [\[CrossRef\]](#)
59. Bates, J.T.; Weber, R.J.; Abrams, J.; Verma, V.; Fang, T.; Klein, M.; Strickland, M.J.; Sarnat, S.E.; Chang, H.H.; Mulholland, J.A.; et al. Reactive Oxygen Species Generation Linked to Sources of Atmospheric Particulate Matter and Cardiorespiratory Effects. *Environ. Sci. Technol.* **2015**, *49*, 13605–13612. [\[CrossRef\]](#)
60. Yang, A.; Janssen, N.A.H.; Brunekreef, B.; Cassee, F.R.; Hoek, G.; Gehring, U. Children's respiratory health and oxidative potential of PM_{2.5}: The PIAMA birth cohort study. *Occup. Environ. Med.* **2016**, *73*, 154–160. [\[CrossRef\]](#)
61. Maikawa, C.L.; Weichenthal, S.; Wheeler, A.J.; Dobbin, N.A.; Smargiassi, A.; Evans, G.; Liu, L.; Goldberg, M.S.; Pollitt, K.J.G. Particulate Oxidative Burden as a Predictor of Exhaled Nitric Oxide in Children with Asthma. *Environ. Health Perspect.* **2016**, *124*, 1616–1622. [\[CrossRef\]](#)
62. Fang, T.; Guo, H.; Verma, V.; Peltier, R.E.; Weber, R.J. PM_{2.5} water-soluble elements in the southeastern United States: Automated analytical method development, spatiotemporal distributions, source apportionment, and implications for health studies. *Atmos. Chem. Phys. Discuss.* **2015**, *15*, 17189–17227. [\[CrossRef\]](#)
63. Wang, Y.; Wang, M.; Li, S.; Sun, H.; Mu, Z.; Zhang, L.; Li, Y.; Chen, Q. Study on the oxidation potential of the water-soluble components of ambient PM_{2.5} over Xi'an, China: Pollution levels, source apportionment and transport pathways. *Environ. Int.* **2020**, *136*, 105515. [\[CrossRef\]](#)
64. Patel, A.; Rastogi, N. Oxidative potential of ambient fine aerosol over a semi-urban site in the Indo-Gangetic Plain. *Atmos. Environ.* **2018**, *175*, 127–134. [\[CrossRef\]](#)
65. Forman, H.J.; Finch, C.E. A critical review of assays for hazardous components of air pollution. *Free. Radic. Biol. Med.* **2018**, *117*, 202–217. [\[CrossRef\]](#)
66. Valko, M.; Jomova, K.; Rhodes, C.J.; Kuča, K.; Musílek, K. Redox- and non-redox-metal-induced formation of free radicals and their role in human disease. *Arch. Toxicol.* **2016**, *90*, 1–37. [\[CrossRef\]](#)
67. Yu, H.; Wei, J.; Cheng, Y.; Subedi, K.; Verma, V. Synergistic and Antagonistic Interactions among the Particulate Matter Components in Generating Reactive Oxygen Species Based on the Dithiothreitol Assay. *Environ. Sci. Technol.* **2018**, *52*, 2261–2270. [\[CrossRef\]](#)
68. Pietrogrande, M.; Perrone, M.R.; Manarini, F.; Romano, S.; Udisti, R.; Becagli, S. PM₁₀ oxidative potential at a Central Mediterranean Site: Association with chemical composition and meteorological parameters. *Atmos. Environ.* **2018**, *188*, 97–111. [\[CrossRef\]](#)
69. Ma, Y.; Cheng, Y.; Qiu, X.; Cao, G.; Fang, Y.; Wang, J.; Zhu, T.; Yu, J.; Xinghua, Q. Sources and oxidative potential of water-soluble humic-like substances (HULISWS) in fine particulate matter (PM_{2.5}) in Beijing. *Atmos. Chem. Phys. Discuss.* **2018**, *18*, 5607–5617. [\[CrossRef\]](#)
70. Pietrogrande, M.; Dalpiaz, C.; Dell'Anna, R.; Lazzeri, P.; Manarini, F.; Visentin, M.; Tonidandel, G. Chemical composition and oxidative potential of atmospheric coarse particles at an industrial and urban background site in the alpine region of northern Italy. *Atmos. Environ.* **2018**, *191*, 340–350. [\[CrossRef\]](#)
71. Shirmohammadi, F.; Wang, D.; Hasheminassab, S.; Verma, V.; Schauer, J.J.; Shafer, M.M.; Sioutas, C. Oxidative potential of on-road fine particulate matter (PM_{2.5}) measured on major freeways of Los Angeles, CA, and a 10-year comparison with earlier roadside studies. *Atmos. Environ.* **2017**, *148*, 102–114. [\[CrossRef\]](#)
72. Wei, J.; Yu, H.; Wang, Y.; Verma, V. Complexation of Iron and Copper in Ambient Particulate Matter and Its Effect on the Oxidative Potential Measured in a Surrogate Lung Fluid. *Environ. Sci. Technol.* **2018**, *53*, 1661–1671. [\[CrossRef\]](#)
73. Charrier, J.G.; Anastasio, C. On dithiothreitol (DTT) as a measure of oxidative potential for ambient particles: Evidence for the importance of soluble transition metals. *Atmos. Chem. Phys. Discuss.* **2012**, *12*, 9321–9333. [\[CrossRef\]](#)
74. Senlin, L.; Win, M.S.; Zeng, J.; Yao, C.; Zhao, M.; Xiu, G.; Lin, Y.; Xie, T.; Dai, Y.; Rao, L.; et al. A characterization of HULIS-C and the oxidative potential of HULIS and HULIS-Fe(II) mixture in PM_{2.5} during hazy and non-hazy days in Shanghai. *Atmos. Environ.* **2019**, *219*, 117058. [\[CrossRef\]](#)

75. Tapparo, A.; Di Marco, V.; Badocco, D.; D'Aronco, S.; Soldà, L.; Pastore, P.; Mahon, B.M.; Kalberer, M.; Giorio, C. Formation of metal-organic ligand complexes affects solubility of metals in airborne particles at an urban site in the Po valley. *Chemosphere* **2019**, *241*, 125025. [[CrossRef](#)] [[PubMed](#)]
76. Xie, T.; Senlin, L.; Zeng, J.; Rao, L.; Wang, X.; Win, M.S.; Zhang, D.; Lu, H.; Liu, X.; Wang, Q. Soluble Fe release from iron-bearing clay mineral particles in acid environment and their oxidative potential. *Sci. Total. Environ.* **2020**, *726*, 138650. [[CrossRef](#)]
77. Verma, V.; Fang, T.; Guo, H.; King, L.; Bates, J.T.; Peltier, R.E.; Edgerton, E.S.; Russell, A.G.; Weber, R.J. Reactive oxygen species associated with water-soluble PM_{2.5} in the southeastern United States: Spatiotemporal trends and source apportionment. *Atmos. Chem. Phys. Discuss.* **2014**, *14*, 12915–12930. [[CrossRef](#)]
78. Samet, J.M.; Chen, H.; Pennington, E.R.; Bromberg, P.A. Non-redox cycling mechanisms of oxidative stress induced by PM metals. *Free. Radic. Biol. Med.* **2019**, *151*, 26–37. [[CrossRef](#)] [[PubMed](#)]
79. Guo, H.; Nenes, A.; Weber, R.J. The underappreciated role of nonvolatile cations in aerosol ammonium-sulfate molar ratios. *Atmos. Chem. Phys. Discuss.* **2018**, *18*, 17307–17323. [[CrossRef](#)]
80. Wang, Q.; Kobayashi, K.; Senlin, L.; Nakajima, D.; Wang, W.; Zhang, W.; Sekiguchi, K.; Terasaki, M. Studies on size distribution and health risk of 37 species of polycyclic aromatic hydrocarbons associated with fine particulate matter collected in the atmosphere of a suburban area of Shanghai city, China. *Environ. Pollut.* **2016**, *214*, 149–160. [[CrossRef](#)]
81. Kumagai, Y.; Koide, S.; Taguchi, K.; Endo, A.; Nakai, Y.; Yoshikawa, T.; Shimojo, N. Oxidation of Proximal Protein Sulfhydryls by Phenanthraquinone, a Component of Diesel Exhaust Particles. *Chem. Res. Toxicol.* **2002**, *15*, 483–489. [[CrossRef](#)]
82. Shang, Y.; Fan, L.; Feng, J.; Lv, S.; Wu, M.; Li, B.; Zang, Y.-S. Genotoxic and inflammatory effects of organic extracts from traffic-related particulate matter in human lung epithelial A549 cells: The role of quinones. *Toxicol. In Vitro* **2013**, *27*, 922–931. [[CrossRef](#)] [[PubMed](#)]
83. Walgraeve, C.; Demeestere, K.; Dewulf, J.; Zimmermann, R.; Van Langenhove, H. Oxygenated polycyclic aromatic hydrocarbons in atmospheric particulate matter: Molecular characterization and occurrence. *Atmos. Environ.* **2010**, *44*, 1831–1846. [[CrossRef](#)]
84. Li, L.J.; Ho, S.S.H.; Feng, B.; Xu, H.; Wang, T.; Wu, R.; Huang, W.; Qu, L.; Wang, Q.; Cao, J. Characterization of particulate-bound polycyclic aromatic compounds (PACs) and their oxidations in heavy polluted atmosphere: A case study in urban Beijing, China during haze events. *Sci. Total. Environ.* **2019**, *660*, 1392–1402. [[CrossRef](#)]
85. Vreeland, H.; Weber, R.J.; Bergin, M.; Greenwald, R.; Golan, R.; Russell, A.G.; Verma, V.; Sarnat, J.A. Oxidative potential of PM 2.5 during Atlanta rush hour: Measurements of in-vehicle dithiothreitol (DTT) activity. *Atmos. Environ.* **2017**, *165*, 169–178. [[CrossRef](#)]
86. Weinstein, J.R.; Asteria-Peñaloza, R.; Diaz-Artiga, A.; Davila, G.; Hammond, S.K.; Ryde, I.T.; Meyer, J.N.; Benowitz, N.; Thompson, L.M. Exposure to polycyclic aromatic hydrocarbons and volatile organic compounds among recently pregnant rural Guatemalan women cooking and heating with solid fuels. *Int. J. Hyg. Environ. Health* **2017**, *220*, 726–735. [[CrossRef](#)]
87. Jiang, H.; Jang, M. Dynamic Oxidative Potential of Atmospheric Organic Aerosol under Ambient Sunlight. *Environ. Sci. Technol.* **2018**, *52*, 7496–7504. [[CrossRef](#)]
88. Wang, S.; Ye, J.; Soong, R.; Wu, B.; Yu, L.; Simpson, M.J.; Chan, A.W.H. Relationship between chemical composition and oxidative potential of secondary organic aerosol from polycyclic aromatic hydrocarbons. *Atmos. Chem. Phys. Discuss.* **2018**, *18*, 3987–4003. [[CrossRef](#)]
89. Win, M.S.; Tian, Z.; Zhao, H.; Xiao, K.; Peng, J.; Shang, Y.; Wu, M.; Xiu, G.; Lu, S.; Yonemochi, S.; et al. Atmospheric HULIS and its ability to mediate the reactive oxygen species (ROS): A review. *J. Environ. Sci.* **2018**, *71*, 13–31. [[CrossRef](#)] [[PubMed](#)]
90. Charrier, J.G.; Anastasio, C. Rates of Hydroxyl Radical Production from Transition Metals and Quinones in a Surrogate Lung Fluid. *Environ. Sci. Technol.* **2015**, *49*, 9317–9325. [[CrossRef](#)]
91. Kuang, X.M.; Gonzalez, D.H.; Scott, J.A.; Vu, K.; Hasson, A.S.; Charbouillot, T.; Hawkins, L.; Paulson, S.E. Cloud Water Chemistry Associated with Urban Aerosols: Rapid Hydroxyl Radical Formation, Soluble Metals, Fe(II), Fe(III), and Quinones. *ACS Earth Space Chem.* **2019**, *4*, 67–76. [[CrossRef](#)]

92. Wang, Q.; Wang, W. Size characteristics and health risks of inorganic species in PM_{1.1} and PM_{2.0} of Shanghai, China, in spring, 2017. *Environ. Sci. Pollut. Res.* **2020**, *27*, 14690–14701. [[CrossRef](#)] [[PubMed](#)]
93. Daher, N.; Ning, Z.; Cho, A.K.; Shafer, M.; Schauer, J.J.; Sioutas, C. Comparison of the Chemical and Oxidative Characteristics of Particulate Matter (PM) Collected by Different Methods: Filters, Impactors, and BioSamplers. *Aerosol Sci. Technol.* **2011**, *45*, 1294–1304. [[CrossRef](#)]

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