



Article Indirect Contact Chamber with Dielectric Layers for Pulsed Electric Field Treatment of Microorganisms

Si Qin¹, Zixin Zhang¹, Jiangang Han¹, Shihai Huang^{2,*} and Jianzong Meng²

- ¹ School of Electrical Engineering, Guangxi University, Nanning 530004, China
- ² College of Life Science and Technology, Guangxi University, Nanning 530004, China
- * Correspondence: shihai_huang@sina.com; Tel.: +86-188-7898-1227

Abstract: Investigation of pulsed electric field (PEF) treatment of yeast at 20 kV/cm using chambers with $BaTiO_3$ dielectric layers was conducted in this study. The sterile rate as well as concentrations of metallic ions and hydroxyl radicals were measured to assess the PEF performance. The results indicated that generation of metallic ions could be reduced by 90%. However, a much higher field strength would be required for satisfactory sterilization due to the Maxwell-Wagner field relaxation, and reactions between the dielectric barriers and liquid could also occur. It was also proven that the continuous presence of a sufficient electric field is the main factor that inactivates the microorganism.

Keywords: pulsed electric field; indirect contact; dielectric barrier; ceramic

1. Introduction

High voltage impulses, which create an intensive pulsed electric field, can generate electrical potential across microbiological membranes [1]. Due to this trans-membrane potential and the induced electromechanical stress, pores can be formed on the membrane. This electroporation process can become irreversible once a critical trans-membrane potential, ~1 V, is reached [2]. As a result, functions of the membrane are disrupted and the microbial cells are inactivated. The inactivation of microorganisms is a non-thermal process, which makes pulsed electric field (PEF) treatment a promising technology in liquid food sterilization. Various studies have been made in recent decades to investigate the effects of PEF treatment on microorganisms and their products [3–7].

Normally, metallic materials, stainless steel for example, are selected as the electrodes of the treatment chamber in traditional PEF systems. In such systems, the liquid food inside the PEF chamber contacts the electrodes directly, which can bring some undesired drawbacks. Electrochemical reactions can occur between the liquid and the metallic material when high voltage is applied, which produces metallic ions or even toxic chemical species [8]. In addition, these electrochemical reactions between the metallic electrodes and the liquid normally generates gaseous byproducts, which form bubbles in the liquid sample [9]. The appearance of gaseous bubbles increases the probability of undesired dielectric breakdown in the PEF treatment chamber. Furthermore, high conduction current flowing through the liquid being treated generates significant Joule heating during the PEF process [10,11]. In some cases, an additional cooling system may be required to keep the liquid temperature within a satisfactory range [11,12].

In order to eliminate these drawbacks, researchers have attempted to use different materials and PEF chamber layouts to avoid direct contact between the metallic electrodes and the liquid. Lubicki et al. used a glass tube to separate the liquid sample and the electrodes [13]. However, the space between the glass tube and the electrodes needed to be filled with conductive fluid of similar permittivity as the liquid sample in order to achieve sterilization in this design. Qin et al. coated copper electrodes with thin TiO_2 film to a thickness of 2 μ m, and used these to construct a PEF treatment chamber [14].



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Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Successful inactivation of *Saccharomyces cerevisiae* was reported, and the conduction current was reduced by 23% compared with an un-coated PEF chamber with identical topology. Tanino et al. also covered the electrodes with carbon cloths and successful inactivation of *E. coli* and *L. homohiochii* was reported [15]. However, strong conduction current, up to ~55 A, could still be observed during the PEF treatment using this design. A conceptual PEF chamber design was proposed and discussed in [16,17], in which the metallic electrodes and the liquid would be separated by a layer of dielectric material. Figure 1 demonstrates the basic layout of this design. Ceramic was selected as the dielectric layer in [17] and inactivation of microorganisms was observed. However, the electric field strength required for successful sterilization was greater than 130 kV/cm, which was much higher than the field strength used in normal PEF treatment. The reason for using such high field strength was not mentioned in [17] and PEF inactivation of microorganisms using such a design with relatively low electric field strength has not been reported.



Figure 1. Concept of a PEF chamber with dielectric layers.

The design proposed by [16,17] demonstrated great potential, as such a design could theoretically eliminate both conduction current and electrochemical reactions completely. Since using high electric fields could bring extra difficulties in practical applications, it is necessary to investigate the performance of PEF inactivation of microorganisms using such design at normal field strength levels (<30 kV/cm). In the present study, two PEF chambers with different geometries were designed and constructed using BaTiO₃ ceramic layers. The chambers were used to PEF-process a yeast *Saccharomyces cerevisiae* suspension using 20 kV/cm electric field strength. The sterile rate of the yeast suspension was measured in order to assess the PEF performance of the ceramic chambers. In addition, the concentrations of metallic ions and hydroxyl radicals (\cdot OH) were also measured in order to investigate the effect of limiting electrochemical reactions in the designed chambers.

2. Experimental Procedures

2.1. PEF Chambers with BaTiO₃ Layers

As shown in Figure 1, the dielectric layers and the metallic electrodes are connected ideally. In this case, the static electric field strength inside the treatment region can be calculated using the following equation:

$$E_0 = U_0 \left(z_l + 2 \frac{\varepsilon_l}{\varepsilon_d} z_d \right)^{-1}$$
(1)

where E_0 is the static electric field strength inside the treatment region; U_0 is the applied voltage across the chamber; z_1 and z_d are the width of the liquid sample and the dielectric layers, respectively; ε_1 and ε_d are the relative dielectric constant of the liquid sample and the dielectric layers, respectively.

According to Equation (1), the thickness and permittivity of the dielectric layer would affect the static field strength inside the treatment region. Dielectric material with higher permittivity or thinner thickness should be selected in order to achieve satisfactory field strength in the treatment region. Therefore, $BaTiO_3$ ceramic plates, which have a dielectric constant above 2000 and relatively good machinability, were selected as the material for the dielectric layer. As a result, the effective static field strength could be approximated using the following equation:

 E_0

$$\approx U_0/z_1$$
 (2)

In practice, it was not possible to achieve ideal contact between the $BaTiO_3$ plates and the electrodes, which created a gap between the two materials. The existence of these gaps formed a very thin layer of air, which triggered the occurrence of dielectric barrier discharge (DBD) between the $BaTiO_3$ layer and the electrodes when external high voltage was applied. Once the DBD started, the actual electric field in the treatment region collapsed. In order to avoid this situation, two approaches were adopted in the present study, as demonstrated in Figure 2.



Figure 2. Schematic of the PEF chamber designs. (a) Chamber with $BaTiO_3$ plates and electrodes joined together using silver paste; (b) Chamber with extended gaps between $BaTiO_3$ plates and electrodes.

(a) The BaTiO₃ ceramic plates and the stainless-steel plates were joined together using silver paste, which is a good conducting material. The air gaps between the plates were eliminated by doing so. The width of the treatment region was set to 5 mm.

(b) The gaps between the plates were extended intentionally and would be filled with the same liquid sample. Therefore, three treatment regions were created, namely G (between the ground electrode and a $BaTiO_3$ plate), M (between the two $BaTiO_3$ plates), and H (between the HV electrode and a $BaTiO_3$ plate). The three regions shared the same static electric field strength, which can be approximated as:

$$E_0 \approx U_0 / (z_G + z_M + z_H) \tag{3}$$

where z_G , z_M and z_H are the width of the three regions, respectively, and which were all set to 3 mm in the present study. Other supporting and spacing structures of the PEF chambers were made out of Polytetrafluoroethylene (PTFE). Additional PEF chambers with identical geometry were assembled, with BaTiO₃ plates replaced by stainless-steel plates in order to provide reference results for the BaTiO₃ ceramic chambers. Figure 3 shows the actual assembled chambers.



Figure 3. Assembled PEF chambers. (**A**) Chambers with BaTiO₃ plates and electrodes joined together by silver paste. (**a1**): with BaTiO₃ plates; (**a2**): BaTiO₃ plates replaced by stainless steel plates; (**B**) Chambers with extended gaps between BaTiO₃ plates and electrodes. (**b1**): with BaTiO₃ plates; (**b2**): BaTiO₃ plates replaced by stainless steel.

2.2. PEF System Setups

An exponential decay pulse-generating system was constructed to supply HV impulses for the present study. This RC-based pulse-generating system, as shown in Figure 4, consisted of a DC-positive high voltage supply (TRC2020, Dalian Teslaman Technology Co. Ltd., Dalian, China), a 24 M Ω protective resistor, a self-breakdown spark-gap switch and a 40 nF HV charging capacitor. The DC power supply had a 50 W rated power with a maximum output voltage of 50 kV and maximum current output of 1 mA. The breakdown voltage of the switch was controlled by adjusting the distance of the inter-electrodes gap, which consequently controlled the voltage across the PEF chamber. The pulse repetition rate was regulated by adjusting the current output of the DC supply. A monitoring system, which consisted of a voltage probe (P6015A, Tektronix, Beaverton, OR, USA), a current probe (Pearson 6585), and a digital oscilloscope (TDS2024C, Tektronix, Beaverton, OR, USA), was used to monitor and record the voltage and current signals during the PEF treatment.



Figure 4. Circuit diagram of the PEF system.

2.3. Preparation of Yeast Suspension

Yeast *Saccharomyces cerevisiae* was used in the present study. A total of 0.5 g of *S. cerevisiae* was cultured in 100 mL sterilized Yeast Extract Peptone Dextrose Medium (YEPD) solution at 30 °C in a shaking incubator, which rotated at 120 rpm. After incubating for 24 h, the yeast suspension was diluted into 500 mL using sterilized deionized water which reduced the conductivity of the suspension to around 86 μ S/cm. The resulting yeast suspension was used for the PEF treatment at a population density of ~10⁶ colony forming units per milliliter (CFU/mL). Triplicate tests were conducted for each experiment.

2.4. Assessment of PEF Inactivation

The prepared yeast suspension was put into the PEF chambers and subjected to 2000 high voltage impulses at 20 kV/cm electric field strength and a pulse repetition rate of 1 pulse per second (1 pps). As discussed in Section 2.1, the approximate static electric field strength differed within treatment regions of different PEF chambers. The actual voltage applied to the PEF chambers was calculated according to Equations (2) and (3). The population density of yeast cells in the suspension was calculated before and after the treatment to evaluate the sterile rate of the PEF treatment. The spread-plate method was used to plate 0.1 mL of the sample onto YEPD agar plates. The agar plates were then incubated at 30 °C for 24 h in a stationary incubator before the number of yeast colonies were counted. The sterile rate can be calculated according to the following equation:

Sterile Rate =
$$\frac{N_C - N_T}{N_C} \times 100\%$$
 (4)

where N_C is the number of yeast colonies from the control (untreated) group and N_T is the number of yeast colonies from the PEF-treated group.

2.5. Determination of Metallic Ions and (•OH) Concentrations

In the experiments conducted using type (b) PEF chambers, measurements of metallic ions and (\cdot OH) concentrations were taken. The concentrations of iron and Ba²⁺ ions in the yeast suspension sample were measured using an ICP-MS (NexION 350X, PerkinElmer, Waltham, MA, USA) following standard operation procedures.

As a short-lived, highly reactive substance, extra trapping agents were needed in order to quantify the concentration of (\cdot OH) hydroxyl radicals [18]. In the present study, methylene blue (C₁₆H₁₈CIN₃S) was selected as the trapping agent. This solution was put into different PEF chambers before undergoing the same PEF treatment. The absorbance at 664 nm of the treated methylene blue solution was then measured using an UV-visible spectrophotometer (UVmini-1240, Shenzhen Lxyee Electronic Technology Co., Ltd., Shenzhen, China). The concentration of the (\cdot OH) in treated samples was evaluated by comparing their respective absorbances with the standard absorbance curve.

3. Results and Discussion

3.1. Voltage and Current Waveforms

As mentioned in Section 2.2, the voltage and current waveforms were monitored and recorded during PEF treatments using different chambers. As demonstrated in Figure 5, the voltage waveforms had simple, exponentially decaying waveshapes as expected from a RC-based pulse generator, despite some oscillations being observed in chambers with BaTiO₃ layers. However, the current waveforms indicate an interesting phenomenon: the current waveforms generally followed the similar tendency as the voltage waveform in the same PEF chamber. However, the peak current in chambers with BaTiO₃ layers was roughly half of that obtained in the traditional stainless-steel chambers.



Figure 5. Actual voltage and current waveforms obtained during PEF treatment using different PEF chambers.

As PEF chambers with/without the $BaTiO_3$ layers shared the same topology and geometry parameters, this result indicated that the voltage across the suspension in the $BaTiO_3$ chambers was around 50% of that in the stainless-steel chambers. Therefore, the field strengths across the same amount of suspension should also have the same ratio for the two types of chambers. As a result, an inferior sterile rate should be expected in the PEF chambers with the dielectric layers.

This unusual phenomenon revealed that something different was happening in the chamber with BaTiO₃ layers during PEF treatment, which could be the Maxwell–Wagner field relaxation process or some unknown chemical–electrical reaction. Such a phenomenon was not reported in [17], where a similar indirect-contact PEF chamber was assembled and used in practical experiments. Therefore, further investigation of the causes of this phenomenon, together with the direct measurement of electric field strength inside the PEF chamber, could be helpful for understanding the exact transient processes and PEF mechanisms involved in indirect-contact PEF chambers.

3.2. Effects on Sterile Rate

The sterile rates of the yeast suspension in different PEF chambers and treatment regions, and the statistical analyses of the results are shown in Figure 6 and Table 1, respectively. As can be seen from the figure, literally no inactivation of yeast was observed in PEF treatment using all the chambers with BaTiO₃ plates. As a comparison, an average of 63% sterile rate was achieved in the type a) chamber and all regions in the type b) chamber when the BaTiO₃ plates were replaced by stainless steel. The statistical analysis also suggested that the sterile rate of the BaTiO₃ groups was statistically no different from the untreated sample, but was significantly different (with 99% confidence) from the stainless-steel groups. The results conflicted with the experimental results reported in [17], in which ceramic material was also used as the dielectric barriers. However, the electric field strength used in [17] was more than six times the field strength used in the present study.



Figure 6. Effects of PEF treatment on the sterile rate of yeast using different types of chambers. Error bars show standard deviation (n = 3).

Table 1. Statistical analysis of the experimental results of yeast sterilization (%).

Chamber	Regions	Mean	SD	SEM	Significant Difference (From Control Group)		Significant Difference (From Stainless Steel Group)	
					At 0.05	At 0.1	At 0.01	
BaTiO ₃	Type a)	0.73	0.407	0.235	No	Yes	Yes	
	Type b) H	1.14	1.255	0.724	No	No	Yes	
	Type b) M	4.12	2.680	1.547	No	No	Yes	
	Type b) G	5.40	1.864	1.076	Yes	Yes	Yes	
	Type a)	64.60	2.485	1.435	Yes	Yes	N/A	
Stainless steel	Type b) H	56.86	5.007	2.891	Yes	Yes	N/A	
	Type b) M	62.00	2.238	1.292	Yes	Yes	N/A	
	Type b) G	62.96	6.816	3.935	Yes	Yes	N/A	

As reported in [16], the transient electric field strength in the treatment region could be affected by the Maxwell–Wagner field relaxation process in PEF chambers with dielectric barriers. According to an analytical transient model developed in [16], the electric field strength inside the treatment region could collapse quickly due to the Maxwell–Wagner surface-polarization mechanism. In addition, the peak field strength and the peak transmembrane potential could also be significantly lower than that in a traditional conducting PEF chamber, which could explain why successful inactivation was achieved in [17] while no inactivation was observed in the present study. Based on the analytical model in [16], and the practical experimental results obtained in [17] and the present study, it could be concluded that PEF inactivation of microorganisms using chambers with dielectric barriers may be possible, but significantly higher nominal electric field strength would be required due to the Maxwell–Wagner field relaxation process.

3.3. Effects on Concentrations of Metallic Ions

Figures 7 and 8 demonstrate the increase of iron and Ba²⁺ ion concentrations in the yeast solution samples respectively. The statistical analyses of these results are shown in Tables 2 and 3, respectively. It can be seen from Figure 6 that the presence of the BaTiO₃ plates limited the release of iron ions significantly. Statistically, the changes in the concentration of iron ions in all regions were significantly different (with at least 90% confidence) in the BaTiO₃ groups compared with the respective regions in the stainless-steel groups. The concentration of iron ions in the yeast suspension samples were significantly lower in all the regions in the chambers with BaTiO₃ layers compared with the traditional stainless-steel chambers. In particular, no iron ions were produced in the samples in

region M of the type b) chamber, which was formed by the two $BaTiO_3$ plates. The results confirmed that the application of dielectric layers could effectively restrict electrochemical reactions between the metallic electrodes and the liquid samples and limit the generation of resulting metallic ions.



Figure 7. Effects of PEF treatment on the concentration of iron ions in yeast suspension using different types of chambers. Error bars show standard deviation (n = 3).



Figure 8. Effects of PEF treatment on the concentration of Ba^{2+} ions in yeast suspension using different types of chambers. Error bars show standard deviation (n = 3).

Table 2. Statistical analysis of the experimental results of iron ions concentration (mg/L).

Chamber	Regions	Mean	SD	SEM	Significant Difference (From Control Group)		Significant Difference (From Stainless Steel Group)	
					At 0.05	At 0.1	At 0.05	At 0.1
BaTiO ₃	Type a)	0.029	0.005	0.003	Yes	Yes	Yes	Yes
	Type b) H	0.018	0.011	0.007	No	No	No	Yes
	Type b) M	0.015	0.024	0.014	No	No	No	Yes
	Type b) G	0.100	0.044	0.026	No	Yes	No	Yes
	Type a)	0.292	0.014	0.008	Yes	Yes	N/A	N/A
Stainless steel	Type b) H	0.322	0.148	0.085	No	Yes	N/A	N/A
	Type b) M	0.177	0.069	0.040	Yes	Yes	N/A	N/A
	Type b) G	0.519	0.220	0.127	No	Yes	N/A	N/A

Chamber	Regions	Mean	SD	SEM	Significant Difference (From Control Group)		Significant Difference (From Type a) Group)	
					At 0.05	At 0.1	At 0.01	At 0.05
BaTiO ₃	Type a)	0.005	0.003	0.002	No	No	N/A	N/A
	Type b) H	1.217	0.434	0.251	Yes	Yes	No	Yes
	Type b) M	1.101	0.248	0.143	Yes	Yes	No	Yes
	Type b) G	0.955	0.176	0.102	Yes	Yes	No	Yes

Table 3. Statistical analysis of the experimental results of Ba^{2+} ion concentration (mg/L).

It can also be noted that in region G of the type b) chambers, where the liquid samples were in contact with the ground electrodes, more iron ions were produced during the PEF process compared with other regions in both chambers with/without the dielectric layer. This phenomenon suggested that the electrochemical reactions around the ground electrodes were more intense in the present study. It is widely accepted that reactions on the anode surface are more severe than those on the cathode surface when high voltage impulses are applied [19–21]. In the present study, the pulse-generating system produced negative impulses, as can be seen from Figure 4, which made the ground electrodes effectively the anode. Therefore, the finding obtained in the present study is in line with the experimental results reported in other studies.

However, as can be seen from Figure 8, significant amount of Ba^{2+} ions were detected in the type b) chambers after the PEF treatment. On average, the concentration of Ba^{2+} ions increased by 1.1 mg/L in the 3 regions of the chambers with a $BaTiO_3$ layer. However, such changes in the Ba^{2+} ion concentration were not observed in the type a) chamber with $BaTiO_3$ layers, as can be seen from Figure 8 and Table 3. This result suggested that reactions involving the $BaTiO_3$ ceramic could have occurred during the PEF treatment, causing the release of barium ions. The exact type of reaction is unclear and further investigation would be required to identify its mechanisms. However, the possibility of such a reaction, involving dielectric material, needs to be considered in future chamber designs and material selection in order to avoid unnecessary contamination.

3.4. Effects on (•OH) Concentrations

As discussed in Section 1, the existence of hydroxyl radicals can reflect the occurrence of electrochemical reactions during the PEF process. Studies also indicate that this substance can play an important part in the inactivation of microorganisms in the liquid [22,23]. Therefore, an approach was adapted to measure the (\cdot OH) concentration as described in Section 2.4; the results and its statistical analysis are displayed in Figure 9 and Table 4, respectively. The results show that the existence of (\cdot OH) was observed in the yeast suspension in both type b) chambers. The contents of the (\cdot OH) were more notable in the chamber with BaTiO₃ barriers, but the difference between the stainless-steel groups and BaTiO₃ groups were not so significant statistically. This result provided another evidence that some PEF-triggered reactions had taken place, in which BaTiO₃ plates were involved.

It has long been suggested that the PEF inactivation of microorganisms could be the combined effect of the electric field and PEF-induced by-products, such as metallic ions and free radicals [8,24]. The experimental results from the present study indicated that while both metallic ions and hydroxyl radicals appeared in both BaTiO₃ and stainless-steel chambers after PEF treatment, inactivation of yeast was only achieved in the stainless-steel chamber. Considering that the actual transient electric field strength inside the chamber with BaTiO₃ layer could be much lower, as discussed in Section 3.1, it can be concluded that the dominant PEF inactivation mechanism was the application of intensive electric field in the present study.



Figure 9. Effects of PEF treatment on the concentration of (·OH) in yeast suspensions using different types of chambers. Error bars show standard deviation (n = 3).

Table 4. Statistical ana	lysis of the ex	perimental	results of hy	vdroxyl	l radical	concentration ((mol/	Ľ)
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Chamber	Regions	Mean	SD	SEM	Significant (From Cont	Significant Difference (From Control Group)		Significant Difference (From Stainless Steel Group)	
	-				At 0.05	At 0.1	At 0.05	At 0.1	
BaTiO ₃	Type b) H Type b) M Type b) G	0.010 0.067 0.108	0.022 0.030 0.020	0.012 0.017 0.011	Yes No Yes	Yes Yes Yes	No No No	No No Yes	
Stainless steel	Type b) H Type b) M Type b) G	0.106 0.125 0.184	0.023 0.038 0.041	0.013 0.022 0.024	Yes Yes Yes	Yes Yes Yes	N/A N/A N/A	N/A N/A N/A	

4. Conclusions

In the present study, indirect-contact PEF chambers with dielectric layers made of $BaTiO_3$ ceramic were designed and constructed. PEF inactivation of yeast *S. cerevisiae* using these chambers, and its effects on the metallic ions and hydroxyl radical concentration in the liquid samples, were investigated. The results obtained in the present study show that the inactivation of yeast in the indirect-contact chambers was unsatisfactory when a 20 kV/cm static electric field strength was applied. Based on the modeling analysis and experimental results of other studies, a much higher external static electric field would be required when this type of chamber structure is adopted due to the Maxwell–Wagner field relaxation process. The unusual current pattern observed in the PEF chamber with $BaTiO_3$ ceramic plates also suggested that this field relaxation process could have taken place. Direct measurement of the transient electric field strength inside the chamber would be helpful in confirming this phenomenon in future research.

It was confirmed in the present study that the presence of the dielectric barriers could limit the occurrence of electrochemical reactions between the liquid and the metallic electrodes during the PEF process. However, evidence suggests that other reactions involving the ceramic layers also took place during the PEF treatment in the present study, and that barium ions and hydroxyl radical were produced as a result. Further studies are required to identify the exact type of reaction observed in the present study.

The experimental results from the present study also indicates that the main factor that inactivates the microorganisms during the PEF process is the intensive external electric field. Free radicals and metallic ions, without the continuous presence of sufficient electric field strength, cannot cause the death of microbial cells in the liquid suspension. Author Contributions: Conceptualization, S.Q. and S.H.; methodology, S.Q.; validation, J.M. and J.H.; formal analysis, Z.Z.; investigation, Z.Z. and J.H.; resources, S.H.; data curation, Z.Z. and J.H.; writing—original draft preparation, S.Q.; writing—review and editing, S.H. and J.M.; visualization, J.H.; supervision, S.H.; project administration, S.Q.; funding acquisition, S.Q. All authors have read and agreed to the published version of the manuscript.

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References

- Toepfl, S.; Heinz, V.; Knorr, D. High intensity pulsed electric fields applied for food preservation. *Chem. Eng. Process* 2007, 46, 537–546. [CrossRef]
- Zimmermann, U. Electrical breakdown, electropermeabilization and electrofusion. *Rev. Physiol. Biochem. Pharmacol.* 1986, 105, 175–256.
- 3. Brito, P.S.; Canacsinh, H.; Mendes, J.P.; Redondo, L.M.; Pereira, M.T. Comparison between monopolar and bipolar microsecond range pulsed electric fields in enhancement of apple juice extraction. *IEEE Trans. Plasma Sci.* **2012**, *40*, 2348–2354. [CrossRef]
- Jaeger, H.; Meneses, N.; Knorr, D. Impact of PEF treatment inhomogeneity such as electric field distribution, flow characteristics and temperature effects on the inactivation of *E. coli* and milk alkaline phosphatase. Innov. *Food Sci. Emerg. Technol.* 2009, 10, 470–480. [CrossRef]
- 5. Guionet, A.; Fujiwara, T.; Sato, H.; Takahashi, K.; Takaki, K.; Matsui, M.; Tanino, T.; Ohshima, T. Pulsed electric fields act on tryptophan to inactivate α-amylase. *J. Electrost.* **2021**, *112*, 103597. [CrossRef]
- 6. Ohshima, T.; Tamura, T.; Sato, M. Influence of pulsed electric field on various enzyme activities. *J. Electrost.* **2007**, *65*, 156–161. [CrossRef]
- Min, S.; Evrendilek, G.A.; Zhang, H.Q. Pulsed electric fields: Processing system, microbial and enzyme inhibition, and shelf life extension of foods. *IEEE Trans. Plasma Sci.* 2007, 35, 59–73. [CrossRef]
- 8. Jayaram, S.H. Sterilization of liquid foods by pulsed electric fields. IEEE Electr. Insul. Mag. 2000, 16, 17–25. [CrossRef]
- Zhang, Q.; Barbosa-Cánovas, G.V.; Swanson, B.G. Engineering aspects of pulsed electric field pasteurization. J. Food Eng. 1995, 25, 261–281. [CrossRef]
- Narsetti, R.; Curry, R.D.; McDonald, K.F.; Clevenger, T.E.; Nichols, L.M. Microbial inactivation in water using pulsed electric fields and magnetic pulse compressor technology. *IEEE Trans. Plasma Sci.* 2006, 34, 1386–1393. [CrossRef]
- Qin, B.-L.; Zhang, Q.; Barbosa-Cánovas, G.V.; Swanson, B.G.; Pedrow, P.D. Inactivation of microorganisms by pulsed electric fields of different voltage waveforms. *IEEE Trans. Dielectr. Electr. Insul.* 1994, 1, 1047–1057.
- Aronssona, K.; Lindgrena, M.; Johanssonb, B.R.; Rönner, U. Inactivation of microorganisms using pulsed electric fields: The influence of process parameters on *Escherichia coli*, *Listeria innocua*, *Leuconostoc mesenteroides* and *Saccharomyces cerevisiae*. *Innov. Food Sci. Emerg. Technol.* 2001, 2, 41–54. [CrossRef]
- 13. Lubicki, P.; Jayaram, S. High Voltage Pulse Application for the Destruction of the Gram-negative Bacterium Yersinia enterocolitica. *Bioelectrochem. Bioenerg.* **1997**, *43*, 135–141. [CrossRef]
- Qin, S.; Timoshkin, I.; Maclean, M.; MacGregor, S.; Wilson, M.; Given, M.; Wang, T.; Anderson, J. TiO2-Coated Electrodes for Pulsed Electric Field Treatment of Microorganisms. *IEEE Trans. Plasma Sci.* 2016, 44, 2121–2128. [CrossRef]
- 15. Tanino, T.; Hirosawa, M.; Moteki, R.; Matsui, M.; Ohshima, T. Engineering of pulsed electric field treatment using carbon materials as electrode and application to pasteurization of sake. *J. Electrost.* **2020**, *104*, 103424. [CrossRef]
- 16. Timoshkin, I.V.; MacGregor, S.J.; Fouracre, R.A.; Crichton, B.H.; Anderson, J.G. Transient electrical field across cellular membranes: Pulsed electric field treatment of microbial cells. *J. Phys. D Appl. Phys.* **2006**, *39*, 569–603. [CrossRef]
- 17. Novac, B.M.; Banakhr, F.A.; Smith, I.R.; Pecastaing, L. Demonstration of a novel pulsed electric field technique generating neither conduction currents nor Joule effects. *IEEE Trans. Plasma Sci.* 2014, 42, 216–228. [CrossRef]
- 18. Peralta, E.; Roa, G.; Hemandez-Servin, J.A. Hydroxyl radicals quantification by UV spectrophotometry. *Electrochim. Acta* 2014, 129, 137–141. [CrossRef]
- 19. Roodenburg, B.; Morren, J.; Berg, H.E. Metal release in a stainless steel pulsed electric field (PEF) system: Part II. The treatment of orange juice; related to legislation and treatment chamber lifetime. *Innov. Food Sci. Emerg. Technol.* 2005, *6*, 337–345. [CrossRef]
- Saulis, G.; Riseviciene, R.; Snitka, V. Increase of the roughness of the stainless-steel anode surface due to the exposure to high-voltage electric pulses as revealed by atomic force microscopy. *Bioelectrochemistry* 2007, 70, 519–523. [CrossRef] [PubMed]
- 21. Rodaite-Riseviciene, R.; Saule, R.; Snitka, V. Release of Iron Ions From the Stainless Steel Anode Occurring During High-Voltage Pulses and Its Consequences for Cell Electroporation Technology. *IEEE Trans. Plasma Sci.* 2014, 42, 249–254. [CrossRef]

- 22. Bai, M.D.; Bai, X.; Zhang, Z. Treatment of Red Tide in Ocean Using Non-Thermal Plasma Based Advanced Oxidation Technology. *Plasma Chem. Plasma Process.* 2005, 25, 539–550. [CrossRef]
- 23. Takatsuji, Y.; Ishikawa, S.; Haruyama, T. Efficient sterilization using reactive oxygen species generated by a radical vapor reactor. *Process Biochem.* **2017**, *54*, 140–143. [CrossRef]
- 24. Hulsheger, H.; Niemann, E.G. Lethal Effect of High Voltage Pulses on *E. coli* K12. *Radiat. Eniron. Biophys.* **1980**, *18*, 281–288. [CrossRef] [PubMed]